

To share or not to share: clonal integration in a submerged macrophyte in response to light stress

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Abstract The ability of clonal plant species to share resources has been studied in many experiments. The submerged macrophyte *Potamogeton perfoliatus* produces interconnected ramets within short time intervals and hence may or may not share resources with ramets growing in less favourable microhabitats. From a genet point of view, sharing with ramets growing under less favourable conditions might not be an optimal strategy when photosynthates could be used to establish other ramets growing under more favourable conditions. To analyse the plasticity in clonal integration of *P. perfoliatus*, we set up a factorial aquaria experiment with unshaded or shaded recipient ramets (offspring), which were connected to or separated from donor ramets (parents). Increased biomass production of offspring in parent–offspring systems compared with severed offspring in both light and shade showed that ramets share resources through clonal integration. The relative translocation to the first- and second-offspring generation was influenced by habitat quality: If first-offspring ramets grew in a shaded microhabitat, second-offspring ramets clearly profited. This may be at least partially because of the fact that resources are shifted from first-offspring to

second-offspring ramets, indicating controlled senescence of the first-offspring. This complex sharing behaviour might be relevant when plants produce ramets within a dense patch of macrophytes, where support of a shaded ramet might not pay off.

Keywords Clonal architecture · Habitat heterogeneity · Biomass allocation · *Potamogeton* · Plant senescence

Introduction

Clonal plants are characterised by the reiteration of potentially independent modules, called ramets, which consist of shoots, rhizomes or stolons, and roots. Clonal integration involves resource sharing through rhizomes or stolons and plays an important role in the regulation of shoot growth. The transport of water, nutrients, and photosynthates has been shown to increase the capacity of plants to tolerate resource heterogeneity, to colonise different microhabitats, and to recover from herbivory (Ong & Marshall, 1979; Schmid et al., 1988; Alpert, 1999). The degree of resource sharing of a clonal plant species is under both genetical and environmental control (Alpert, 1999; van Kleunen et al., 2000). The primary motor behind clonal integration might be either the resource export from parents acting as a source (push model) or the demand of offspring acting as a sink (pull model) (Pitelka & Ashmun, 1985; Marshall & Price, 1997).

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Among aquatic plants, clonal integration is known from emergent macrophytes (Hester et al., 1994; Amsberry et al., 2000), floating stoloniferous species (Methy et al., 1990; Li & Wang, 2011), submerged macrophytes (Xiao et al., 2007), and marine seagrasses (Tomasko & Dawes, 1989; Marbà et al., 2002).

Although clonal integration might enable ramets to develop at less suitable microhabitats (e.g., Tomasko & Dawes, 1989; Methy et al., 1990), modelling analyses suggest that support of such ramets might not always be beneficial for the genet, especially when there are other ramets in more suitable microhabitats (Gardner & Mangel, 1999). However, until recently most studies analysing clonal integration almost exclusively focussed on resource sharing, but did not examine conditions when resource sharing with a specific ramet might not be profitable (but see Hellström et al., 2006).

Here, we analyse whether resource sharing occurs with a ramet growing at a less suitable microhabitat of the submerged rhizomatous freshwater species *Potamogeton perfoliatus* L. This species can form large patches that can facilitate other macrophyte species when threatened by herbivory or eutrophication (Le Bagousse-Pinguet et al., 2012a, b). In situ surveys in Lake Constance revealed that each *P. perfoliatus* plant sprouts from a turion in spring and produces horizontal rhizomes of up to 1.5-m length, bearing up to 15 ramets during a short vegetation period (Wolfer & Straile, 2004a, b). Because of the rapid establishment of subsequent ramets, several newly established ramets can occur, and it is not clear whether a ramet in a less suitable habitat will be supported.

We tested for the effects of shading of one offspring (i.e., growing in a less suitable microhabitat) on the growth of two offspring generations in a 2×2 factorial aquarium experiment. The first offspring was either left connected to or severed from the parental ramet. Hence, we analysed the relative performance of two ramets in a 3-ramet system (connected) and in a 2-ramet system (severed). At the start of the experiment, the second of the focal ramets was not yet established. Consequently, in the 2-ramet system where no support from the parent is possible, the success of the clonal fragment depends on the photosynthesis of the shaded ramet. In contrast, in the 3-ramet system, the shaded ramet is not crucial for the success of the clonal fragment as the parent

ramet can share resources with the unshaded offspring. Hence, we tested the following two hypotheses:

- (1) Total biomass of offspring ramets in the 3-ramet (parent–offspring) system is enhanced as compared with biomass of offspring ramets in the 2-ramet system (severed offspring system), i.e., there is clonal integration between parent and offspring.
- (2) The shaded first–offspring ramet benefits less from clonal integration than the unshaded second–offspring ramet in the 3-ramet system.

Materials and methods

Origin and pre-cultivation of plant material

Potamogeton perfoliatus shoots were collected at the Lower Basin of Lake Constance, a large meso-oligotrophic lake in central Europe (9°18'E, 47°39'N). All shoots originated from the same patch with a diameter of approximately 15 m and had an intermediate developmental age. Because they were cut off above the sediment, they had no rhizomes and no roots when planted. The shoots were planted across three aquaria (length: 80 cm, width: 40 cm, height: 50 cm), filled with 10 cm of natural sediment from Lake Constance, and supplied with 125 l of filtered lake water. The water was exchanged twice a week; light was provided by pairs of white and plant-grow tubes (light intensity: $40 \mu\text{E m}^{-2} \text{s}^{-1}$) at 14 h a day. The experiment was started after the ramets were well established and had formed short second-offspring shoots.

Experimental design

The experiment started when all planted shoots (parent plants, P) had established one offspring ramet (O_1), i.e., the first focal ramet. About 50% of the focal ramets were left connected to their parent, 50% were severed from their parent. In addition, habitat heterogeneity with regard to light conditions was introduced in 50% of the connected and severed observation units by shading O_1 with cylinder-shaped nets (Agroflor, height: 40 cm, diameter: 6 cm, shading effect: 63%) (Fig. 1). Eight replicates of each treatment were distributed randomly across the three

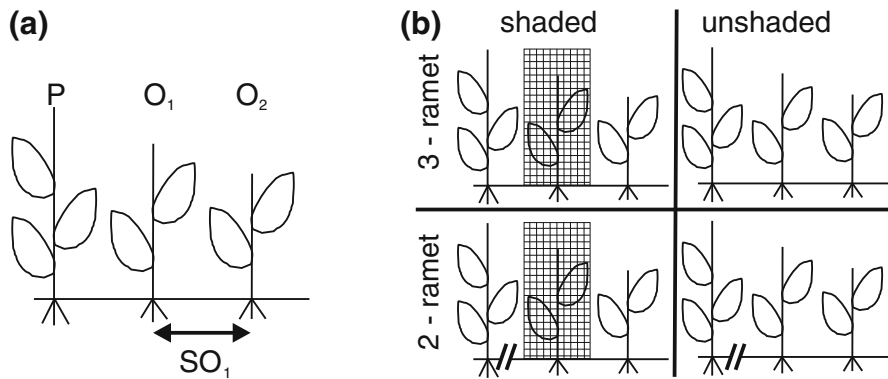


Fig. 1 Sketch of **a** an experimental unit **b** of the experimental design. Abbreviations as used in the text: *P* parent ramet; *O*₁ first-offspring ramet, and *O*₂ second-offspring ramet as the two focal ramets; *SO*₁ spacer length between focal ramets. Rhizomes

between *P* and *O*₁ were either left connected or severed midway between the shoots. *O*₁ ramets, but not *O*₂ ramets, were either kept unshaded or shaded

aquaria. At the beginning of the experiment, neither shoot lengths of *P* ramets (31 ± 8 cm) nor shoot lengths of *O*₁ ramets (12 ± 5 cm) differed between treatments. The experiment was ended after 3 weeks when plants had produced the second focal ramet (*O*₂). The plants were carefully removed from the sediment with their rhizomes and roots and thoroughly washed. After the measurement of shoot lengths and lengths of the rhizomes between *O*₁ and *O*₂—called spacer length (*SO*₁) below—shoot, rhizome, and root fractions were dried at 105°C, cooled down, and weighed on an analytic scale.

Resource sharing was inferred from increased shoot length and biomass in connected compared with severed focal ramets (Tomasko & Dawes, 1989; Amsberry et al., 2000), but results are only presented for biomass, because analyses based on shoot measurements yielded consistent results.

Statistical analysis

Before analyses, shoot biomasses were log- and proportional data arcsine square root transformed to ensure normality and homogeneity for variances. Effects of shading and severing on total offspring biomass and the biomass ratio of *O*₁ versus *O*₂ were analysed with a linear mixed model with shading and severing and their interaction as fixed factors. Treatment effects on biomasses of *O*₁ and *O*₂ were separately analysed with a mixed model with four treatment levels and subsequent Tukey's post hoc

comparison (Hothorn et al., 2008). In addition, we performed a linear mixed model with shading, severing, and ramet order as fixed factors to explicitly test for interactions of ramet order with severing and shading.

Root, shoot, and rhizome allocation of *O*₁ were calculated by dividing the respective biomasses by total biomass of *O*₁ (shoot biomass + root biomass + rhizome biomass). Effects of severing and shading on biomass allocation and on spacer length (*SO*₁) were analyzed with mixed models with shading and severing and their interaction as fixed factors. To investigate whether observed variability in *SO*₁ was primarily because of allometry, i.e., differences in shoot biomass of *O*₁, we additionally ran models using *O*₁ shoot biomass as a covariate in models.

All statistical models were set up as mixed models in the nlme package of R (Pinheiro et al., 2011). All models considered besides fixed factors (see above) the random factor “aquarium” to account for slight differences in growth conditions between aquaria. Likelihood ratio tests were used to assess the significance of the random factor and indicated significance levels between 0.01 and 0.6 in the different models. However, to present conservative significance levels for the fixed effects, the random factor was included in all models. We do not report individual likelihood ratio test with the exception of Table 1. Likelihood ratio tests were also used in the model with shoot biomass as covariate to analyse which fixed factors significantly contribute to the model.

Table 1 Effects of shading, severing, and ramet order on shoot biomass

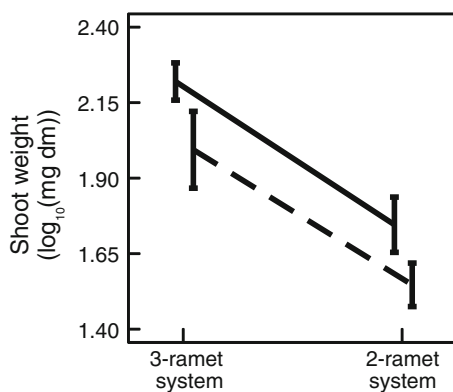
	DF	F value	P value
Severing	1,48	59.5	0.0001
Shading	1,48	13.6	0.0006
Ramet order	1,48	35.5	0.0001
Severing × shading	1,48	0.9	0.3449
Severing × ramet order	1,48	15.9	0.0002
Shading × ramet order	1,48	0.8	0.3748
Severing × shading × ramet order	1,48	6.4	0.0147

Linear mixed model with shoot biomass (log transformed) as dependent variable and severing, shading, ramet order (first or second offspring) and their interactions as fixed factors. To account for possible aquaria effects on shoot biomass, “aquarium” was included as a random factor in the model. A likelihood ratio test (LR = 2.999) suggests the inclusion of this random factor with $P = 0.083$

Results

Clonal integration

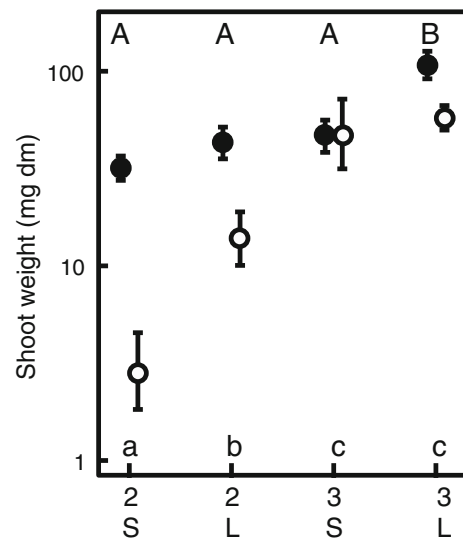
After 3 weeks of growth, severing had reduced total shoot biomass of focal ramets ($O_1 + O_2$) strongly (63% reduction, mixed model; $F_{1,23} = 30.8$, $P < 0.0001$, Fig. 2), whereas shading of the first offspring (O_1) had a smaller effect (30% reduction, $F_{1,23} = 5.8$, $P < 0.05$). Shading did not influence the response of focal ramet shoot biomass to being severed (severing × shading interaction: $F_{1,23} = 0.03$, $P = 0.86$). Biomass of parent ramets was affected neither by severing nor by shading of O_1 ($F_{1,20} = 2.3$, $P = 0.15$, and $F_{1,20} = 0.09$,

**Fig. 2** Total shoot biomass of both focal ramets (mean ± SE) of *P. perfoliatus* in the 2-ramet versus 3-ramet system. Solid lines indicate “light,” hatched lines indicate “shade” treatments

$P = 0.77$) nor by their interaction ($F_{1,20} = 0.02$, $P = 0.88$).

Focal ramets were differently affected by severing and by shading (Fig. 3). Biomass of O_1 was highest in the 3-ramet light treatment as compared with all other treatments (Fig. 3). When shaded, there was no significant biomass difference between the 2- and 3-ramet systems, meaning that O_1 performance was not significantly increased by the presence of a parent. In contrast, O_2 did profit from the 3-ramet system under both light and shade conditions (of O_1). Furthermore, its biomass increase because of clonal integration was higher when O_1 was shaded than when O_1 was unshaded. Shading of O_1 did not influence the biomass of O_2 in the 3-ramet system. As a consequence, biomass of O_2 was similar to the biomass of O_1 in the 3-ramet shade treatment, but was less than O_1 in all other treatments (Fig. 3).

As a consequence of different responses of O_1 and O_2 to severing and shading, the growth of O_2 relatively to O_1 (O_2/O_1) was significantly influenced by the interaction between severing and shading ($F_{1,23} = 9.6$,

**Fig. 3** Shoot biomass of the two focal ramets O_1 (filled circles) and O_2 (open circles) in the different treatments. Numbers 2 and 3 refer to the 2-, 3-ramet systems, respectively, S shade, L (light) refers to the two light conditions. There are significant treatment differences for O_1 biomass (linear mixed model: $F_{3,23} = 9.99$, $P < 0.0002$) and for O_2 biomass ($F_{3,23} = 18.0$, $P < 0.0001$). Capital letters at the top of the graph indicate treatment differences in O_2 biomass, lower case letters at the bottom of the graph show treatment differences in O_1 biomass. Different letters indicate significant differences between treatments (Tukey’s post hoc test, $P < 0.05$)

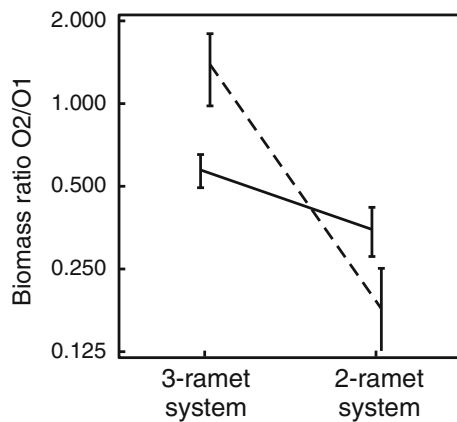


Fig. 4 Biomass ratio of second offspring and first offspring (O_2/O_1) (mean \pm SE) of *P. perfoliatus* grown in aquaria. Solid lines indicate “light,” hatched lines indicate “shade” treatments

$P = 0.005$, Fig. 4): Shading of O_1 increased the O_2/O_1 biomass ratio when O_1 was connected to a parent ramet, but decreased it when the connection was severed. This difference in response of the two focal ramets to shading in the 2- versus 3-ramet system is also supported by a significant three-way interaction: severing \times shading \times ramet order in a mixed model distinguishing the two focal ramets with the factor “ramet order” (Table 1).

Biomass allocation and spacer length

Root allocation of O_1 was affected by the interaction between the shading and severing (Table 2): Shading increased root allocation of connected O_1 but reduced it in severed O_1 (Fig. 5a). No significant treatment effects (Table 2) were observed for shoot allocation (Fig. 5b) and rhizome allocation (Fig. 5c).

Spacer length (SO_1) was close to significantly related to the interaction between severing and shading interaction ($F_{1,25} = 3.69$, $P = 0.07$), significantly to

severing ($F_{1,25} = 10.9$, $P < 0.003$), but not to shading ($F_{1,25} = 0.4$, $P = 0.55$). With shoot biomass as a covariate, SO_1 was significantly related to the interaction of shoot biomass and severing ($F_{1,25} = 19.1$, $P = 0.0002$), shoot biomass ($F_{1,25} = 11.6$, $P = 0.0022$), and severing ($F_{1,5} = 8.2$, $P = 0.0084$). While SO_1 was negatively related to shoot biomass in 3-ramet systems, SO_1 was positively related to shoot biomass in 2-ramet systems (Fig. 6). This suggests that the 3-ramet system O_1 responded to growth conditions by increasing spacer length, whereas in the 2-ramet system, O_1 responded to growth conditions by decreasing spacer length. Inclusion of shading and its interactions with shoot biomass or severing did not significantly improve the model with severing and shoot biomass as fixed factors (Likelihood ratio tests, all $P > 0.05$). This suggests that possible effects of shading on SO_1 may be because of effects of shading on O_1 shoot biomass.

Discussion

Clonal integration

The substantially higher biomass of offspring in the 3-ramet systems as compared with the 2-ramet systems shows that parent ramets of *P. perfoliatus* were capable of sharing resources acropetally through clonal integration. However, clonal integration was not uniform, but rather sensitive to the growth conditions of the first focal ramet: O_1 ramets substantially benefited from integration only when in the light but were not supported when shaded. In contrast, many previous studies have found physiological integration especially where recipient ramets experienced stress through resource limitation in heterogeneous environments (Alpert & Stuefer, 1997; Hutchings & Wijesinghe, 1997; Alpert, 1999). For example, connected shaded

Table 2 Effects of shading and severing on root, rhizome, and shoot allocation

	Severing	Shading	Severing \times shading
Root allocation	$F_{1,26} = 4.62$, $P = 0.04$	$F_{1,26} = 3.48$, $P = 0.07$	$F_{1,26} = 7.6$, $P = 0.01$
Rhizome allocation	$F_{1,26} = 0.12$, $P = 0.73$	$F_{1,26} = 0.92$, $P = 0.34$	$F_{1,26} = 0.02$, $P = 0.88$
Shoot allocation	$F_{1,26} = 0.19$, $P = 0.67$	$F_{1,26} = 0.21$, $P = 0.65$	$F_{1,26} = 2.47$, $P = 0.13$

Linear mixed model results with allocations (arcsine square root transformed) as dependent variable and severing, shading, their interaction as fixed factors. To account for possible aquaria effects on shoot growth, “aquarium” was included as a random factor in the models

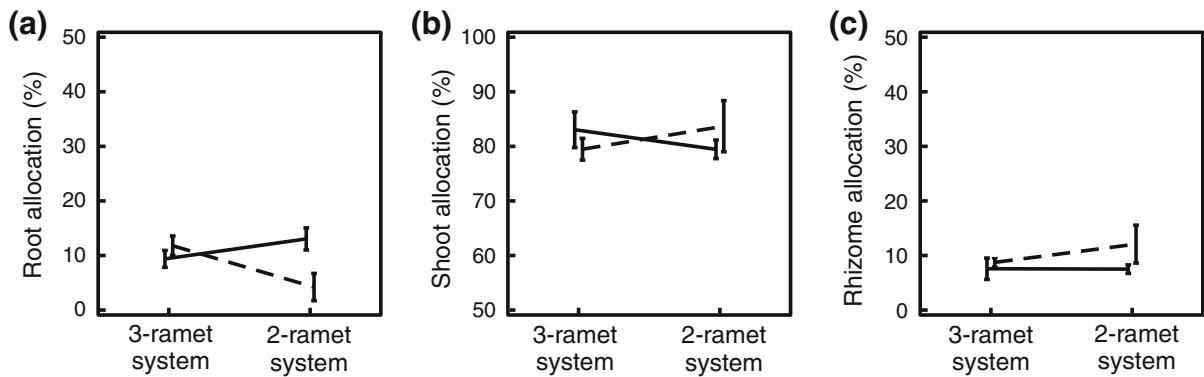


Fig. 5 a Root, b shoot, and c rhizome allocation \pm SE of 2- and 3-ramet system of *P. perfoliatus* first offspring (O_1). Solid lines indicate “light,” hatched lines indicate “shade” treatments

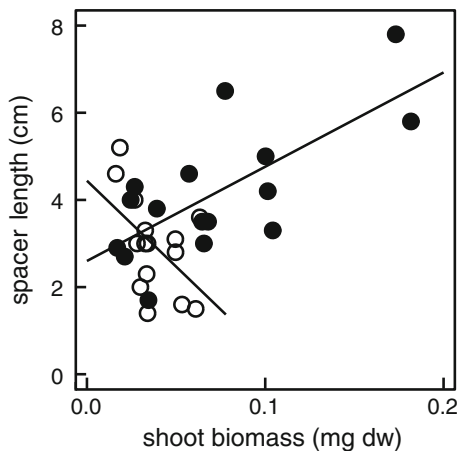


Fig. 6 Spacer length between focal ramets (SO_1) in relation to first offspring (O_1) shoot biomass. Filled symbols indicate 3-ramet system treatments, open symbols indicate 2-ramet system treatments

shoots of the seagrass *Thalassia testudinum* achieved growth rates equal to non-shaded controls (Tomasko & Dawes, 1989). Likewise, severely shaded ramets of *Lolium perenne* continued to grow and produced new leaves as a result of physiological integration (Ong & Marshall, 1979), and shaded shoots of *Eichhornia crassipes* suffered less when connected to parent shoots growing in the light (Methy et al., 1990). However, in our study, the proximal offspring (O_1) benefited from the 3-ramet system, i.e., from the connection to its parental ramet only under full light, but was not supported when shaded. In the latter situation, the shaded ramet O_1 was passed by and resources were directed to the distal offspring O_2 . Indeed, O_2 ramets of *P. perfoliatus* benefited from

resource sharing relatively more strongly when O_1 ramets were shaded. This shows that clonal integration in *P. perfoliatus* can also occur between non-adjacent shoots (see also Terrados et al., 1997) and that resources are shared with distal offspring depending on the microhabitat, i.e., growth conditions, of the proximate offspring. Likewise, resource sharing has often been observed with ramets suffering from herbivory (Marshall & Sagar, 1965; Schmid et al., 1988). However, lack of support for damaged ramets of a perennial herb has been demonstrated and attributed to competition between sibling ramets (Hellström et al., 2006).

Competition within branches of a plant has also been demonstrated in pine trees (Honkanen & Haukioja, 1994) and pea plants (Novoplansky et al., 1989), and has been termed the branch-competition hypothesis (Sachs & Novoplansky, 1997). This hypothesis predicts that a plant module that is inferior because of, e.g., herbivory damage or microhabitat unsuitability should be left out of support when more viable sinks are available. Our results are in line with this prediction. However, shaded O_1 in the 3-ramet system showed even strong signs of chlorosis that was not the result of shading per se, as shaded O_1 in the 2-ramet system appeared vigorous and retained green leaves. This observation suggests that the branch-competition hypothesis is unlikely to give a complete explanation of our results: At the end of the experiment, shoot biomass of O_1 and O_2 was similar (Fig. 3). Assuming a faster growth rate of unshaded O_2 relative to shaded O_1 suggests that during most of the experimental time, biomass of O_1 was larger than biomass of O_2 . It is,

hence, difficult to believe that despite these biomass differences, competitive superiority of O_2 was large enough to result into chlorosis of O_1 . Rather, the large differences in performance of O_1 suggest controlled senescence of shaded O_1 in the 3-ramet treatment possibly associated with remobilization of resources (Ong & Marshall, 1979; Stapel & Hemminga, 1997) towards O_2 . Hence, integration seems to have qualitatively altered the response of shaded O_1 by inducing a novel response, i.e., chlorosis (de Kroon et al., 2005). Interestingly, controlled senescence of *P. perfoliatus* has also been suggested as a response to intense herbivory (Miler & Straile, 2010): High herbivory pressure of a lepidopteran larvae resulted in re-translocation of nutrients from shoots to overwintering organs and consequently to senescence of shoots. This suggests that controlled senescence is a behaviour highly important for the response of the species to biotic and abiotic factors, and should be considered in future studies aiming to model the temporal and spatial dynamics of this species (e.g., Wolfer et al., 2006).

Biomass allocation and spacer length

Shading increased root allocation of connected offspring but reduced root allocation in severed offspring. Our analyses suggest that the differences in biomass allocation are not because of allometric growth rules, which predict that smaller plants show a higher biomass allocation to belowground structures (Müller et al., 2000). In contrast to this prediction, O_1 in 2-ramet shade systems had the lowest root allocation despite their small biomass. This might result from a shortage of carbohydrates and the need to invest in shoot biomass (Alcoverro et al., 1997). The lower root allocation in the severed shaded offspring compared with higher root allocation in connected shaded offspring is also in line with the foraging hypothesis proposing that single plants specialise in the most limiting resource (here light), and integrated plants specialise in the most abundant resource (here probably nutrients) (Stuefer et al., 1996). In a clonal ramet system with “division of labour” (Hutchings & Wijesinghe, 1997), ramets may continue to take up nutrients by roots even when they are non-photosynthetic (Jonsdottir & Callaghan, 1990) and in our experiment, shaded O_1 may still contribute to plant growth by supplying nutrients to unshaded O_2 .

As in some terrestrial plants (Wijesinghe & Handel, 1994; van Kleunen et al., 2000), severing significantly reduced spacer lengths of *P. perfoliatus* offspring. This might be attributed to the stronger effect of severing on offspring shoot biomass, and to the overall positive relationship between shoot biomass and spacer length. In situ, spacer length of *P. perfoliatus* strongly increases with distance from the primary ramet, possibly as a consequence of an increasing biomass and production of an interconnected clonal fragment with the number of produced ramets (Wolfer & Straile, 2004b). As a result of shading, O_1 in the 3-ramet system responded to the unfavourable growth conditions by decreasing spacer length. However, O_1 in the 2-ramet system responded to shade conditions by increasing spacer length, even though shoot biomass was slightly decreased. This is in line with the predictions of the foraging hypothesis: Shaded shoots are expected to produce longer rhizomes to “escape” from the unfavourable habitat (Hartnett & Bazzaz, 1983; Sutherland & Stillman, 1988). Possibly, foraging is only expressed in the growth patterns when growth is strongly impaired by shading, and integration is not possible (but see de Kroon & Hutchings, 1995 for a critical discussion of the plant foraging hypothesis).

To conclude, our experiment has shown that (1) *P. perfoliatus* parent ramets support their clonal offspring ramets through translocation of resources, i.e., there is clonal integration within a genet and (2) the relative translocation of resources to different offspring generations depends on the habitat quality of the individual ramets: Ramets in unfavourable microhabitats, e.g., under light stress, are not integrated when support of other ramets provides higher benefits for the genet. This behaviour might be highly relevant when plants produce new ramets within dense patches of macrophytes (Wolfer & Straile, 2004a, b). In such a case, it might not benefit the plant to support a severely shaded ramet, but rather to invest in rhizome growth and new ramets at the outer perimeter of the patch, where microhabitats are more suitable.

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References

- Alcoverro, T., J. Romero, C. M. Duarte & N. I. Lopez, 1997. Spatial and temporal variations in nutrient limitation of seagrass *Posidonia oceanica* growth in the NW Mediterranean. *Marine Ecology-Progress Series* 146: 155–161.
- Alpert, P., 1999. Clonal integration in *Fragaria chiloensis* differs between populations: ramets from grassland are shelfish. *Oecologia* 120: 69–76.
- Alpert, P. & J. F. Stuefer, 1997. Division of labour in clonal plants. In van Groenendael, J. & H. deKroon (eds), *The Ecology and Evolution of Clonal Plants*. Backhuys Publishers, Leiden: 137–154.
- Amsberry, L., M. A. Baker, P. J. Ewanchuk & M. D. Bertness, 2000. Clonal integration and the expansion of *Phragmites australis*. *Ecological Applications* 10: 1110–1118.
- de Kroon, H. & M. J. Hutchings, 1995. Morphological plasticity in clonal plants: the foraging concept reconsidered. *Journal of Ecology* 83: 143–152.
- de Kroon, H., H. Huber, J. F. Stuefer & J. M. van Groenendael, 2005. A modular concept of phenotypic plasticity in plants. *New Phytologist* 166: 73–82.
- Gardner, S. N. & M. Mangel, 1999. Modeling investments in seeds, clonal offspring, and translocation in a clonal plant. *Ecology* 80: 1202–1220.
- Hartnett, D. C. & F. A. Bazzaz, 1983. Physiological integration among intraclonal ramets in *Solidago canadensis*. *Ecology* 64: 779–788.
- Hellström, K., M. M. Kytoviita, J. Tuomi & P. Rautio, 2006. Plasticity of clonal integration in the perennial herb *Linaria vulgaris* after damage. *Functional Ecology* 20: 413–420.
- Hester, M. W., K. L. McKee, D. M. Burdick, M. S. Koch, K. M. Flynn, S. Patterson & I. A. Mendelsohn, 1994. Clonal integration in *Spartina patens* across a nitrogen and salinity gradient. *Canadian Journal of Botany* 72: 767–770.
- Honkanen, T. & E. Haukioja, 1994. Why does a branch suffer more after branch-wide than after tree-wide defoliation. *Oikos* 71: 441–450.
- Hothorn, T., F. Bretz & P. Westfall, 2008. Simultaneous inference in General Parametric Models. *Biometrical Journal* 50: 346–363.
- Hutchings, M. J. & D. K. Wijesinghe, 1997. Patchy habitat, division of labour and growth dividends in clonal plants. *Trends in Ecology & Evolution* 12: 390–394.
- Jonsdottir, I. S. & T. V. Callaghan, 1990. Intraclonal translocation of ammonium and nitrate nitrogen in *Carex bigelowii* Torr Ex Schwein using N-15 and nitrate reductase assays. *New Phytologist* 114: 419–428.
- Le Bagousse-Pinguet, Y., E. M. Gross and D. Straile, 2012a. Release from competition and protection determine the outcome of plant interactions along a grazing gradient. *Oikos* 121: 95–101.
- Le Bagousse-Pinguet, Y., P. Liancourt, N. Gross and D. Straile, 2012b. Indirect facilitation promotes macrophyte dominance in freshwater ecosystems threatened by eutrophication. *Journal of Ecology*. doi:10.1111/j.1365-2745.2011.01931.x.
- Li, W. G. & J. B. Wang, 2011. Influence of light and nitrate assimilation on the growth strategy in clonal weed *Eichhornia crassipes*. *Aquatic Ecology* 45: 1–9.
- Marbà, N., M. A. Hemminga, M. A. Mateo, C. M. Duarte, Y. E. M. Mass, J. Terrados & E. Gacia, 2002. Carbon and nitrogen translocation between seagrass ramets. *Marine Ecology Progress Series* 226: 287–300.
- Marshall, C. & E. A. C. Price, 1997. Sectoriality and its implications for physiological integration. In deKroon, H. & J. van Groenendael (eds), *The Ecology and Evolution of Clonal Plants*. Backhuys Publishers, Leiden: 79–107.
- Marshall, C. & G. R. Sagar, 1965. Influence of defoliation on distribution of assimilates in *Lolium multiflorum* Lam. *Annals of Botany* 29: 365–370.
- Methy, M., P. Alpert & J. Roy, 1990. Effects of light quality and quantity on growth of the clonal plant *Eichhornia crassipes*. *Oecologia* 84: 265–271.
- Miler, O. & D. Straile, 2010. How to cope with a superior enemy? Plant defence strategies in response to annual herbivore outbreaks. *Journal of Ecology* 98: 900–907.
- Müller, I., B. Schmid & J. Weiner, 2000. The effect of nutrient availability on biomass allocation patterns in 27 species of herbaceous plants. *Perspectives in Plant Ecology, Evolution and Systematics* 3: 115–127.
- Novoplansky, A., D. Cohen & T. Sachs, 1989. Ecological implications of correlative inhibition between plant shoots. *Physiologia Plantarum* 77: 136–140.
- Ong, C. K. & C. Marshall, 1979. Growth and survival of severely-shaded tillers in *Lolium perenne* L. *Annals of Botany* 43: 147–155.
- Pinheiro, J., D. Bates, S. DebRoy, D. Sarkar and the R Development Core Team, 2011. nlme: linear and nonlinear mixed effects models. R package version 3.1-100.
- Pitelka, L. & J. Ashmun, 1985. Physiology and integration of ramets in clonal plants. In Jackson, J. B. C., L. W. Buss & R. E. Cook (eds), *Population Biology and Evolution of Clonal Organisms*. Yale University Press, New Haven, Connecticut: 399–436.
- Sachs, T. & A. Novoplansky, 1997. What does a clonal organization suggest concerning clonal plants? In van Groenendael, J. & H. deKroon (eds), *The Ecology and Evolution of Clonal Plants*. Backhuys Publishers, Leiden: 55–78.
- Schmid, B., G. M. Puttick, K. H. Burgess & F. A. Bazzaz, 1988. Clonal integration and effects of simulated herbivory in old-field perennials. *Oecologia* 75: 465–471.
- Stapel, J. & M. A. Hemminga, 1997. Nutrient resorption from seagrass leaves. *Marine Biology* 128: 197–206.
- Stuefer, J. F., H. de Kroon & H. J. During, 1996. Exploitation of environmental heterogeneity by spatial division of labour in a clonal plant. *Functional Ecology* 10: 328–334.
- Sutherland, W. J. & R. A. Stillman, 1988. The foraging tactics of plants. *Oikos* 52: 239–244.
- Terrados, J., C. M. Duarte & W. J. Kenworthy, 1997. Is the apical growth of *Cymodocea nodosa* dependent on clonal integration? *Marine Ecology Progress Series* 158: 103–110.
- Tomasko, D. A. & C. J. Dawes, 1989. Evidence for physiological integration between shaded and unshaded short shoots of *Thalassia testudinum*. *Marine Ecology Progress Series* 54: 299–305.
- van Kleunen, M., M. Fischer & B. Schmid, 2000. Clonal integration in *Ranunculus reptans*: by-product or adaptation? *Journal of Evolutionary Biology* 13: 237–248.

- Wijesinghe, D. K. & S. N. Handel, 1994. Advantages of clonal growth in heterogenous habitats: an experiment with *Potentilla simplex*. *Journal of Ecology* 82: 495–502.
- Wolfer, S. R. & D. Straile, 2004a. Density control of clonal growth of *Potamogeton perfoliatus*. *Limnologica* 34: 98–104.
- Wolfer, S. R. & D. Straile, 2004b. Spatio-temporal dynamics and plasticity of clonal architecture in *Potamogeton perfoliatus*. *Aquatic Botany* 78: 307–318.
- Wolfer, S. R., E. H. van Nes & D. Straile, 2006. Modelling the clonal growth of the rhizomatous macrophyte *Potamogeton perfoliatus*. *Ecological Modelling* 192: 67–82.
- Xiao, K. Y., D. Yu, X. W. Xu & W. Xiong, 2007. Benefits of clonal integration between interconnected ramets of *Valisneria spiralis* in heterogeneous light environments. *Aquatic Botany* 86: 76–82.