REVIEW ARTICLE

What explains the invading success of the aquatic mud snail Potamopyrgus antipodarum (Hydrobiidae, Mollusca)?

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Abstract The spread of non-native species is one of the most harmful and least reversible disturbances in ecosystems. Species have to overcome several filters to become a pest (transport, establishment, spread and impact). Few studies have checked the traits that confer ability to overcome these steps in the same species. The aim of the present study is to review the available information on the life-history and ecological traits of the mud snail, Potamopyrgus antipodarum Gray (Hydrobiidae, Mollusca), native from New Zealand, in order to explain its invasive success at different aquatic ecosystems around the world. A wide tolerance range to physico-chemical factors has been found to be a key trait for successful transport. A high competitive ability at early stages of succession can explains its establishment success in human-altered ecosystems. A high reproduction rate, high capacity for active and passive dispersal, and the escape from native

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predators and parasites explains its spread success. The high reproduction and the ability to monopolize invertebrate secondary production explain its high impact in the invaded ecosystems. However, further research is needed to understand how other factors, such as population density or the degree of human perturbation can modify the invasive success of this aquatic snail.

Keywords Potamopyrgus antipodarum · Snail · Tolerance · Life-history traits · Colonization · Spread

Introduction

The spread of non-native species can be one of the most harmful and least reversible disturbances in ecosystems (Strayer, [1999;](#page-9-0) Ricciardi & MacIsaac, [2000;](#page-8-0) Rahel, [2002](#page-8-0)). Biological invasions may alter the properties of the invaded habitat, decline biodiversity and induce biotic homogenization (Enserink, [1999;](#page-7-0) Kolar & Lodge, [2001](#page-8-0); Cambray, [2003](#page-7-0); Mills et al., [2003\)](#page-8-0). Nowadays, human activities, such us agriculture, aquaculture, recreation and international trade are increasing the range of some species (Leppa¨koski & Olenin, [2000](#page-8-0); Ricciardi & MacIsaac, [2000;](#page-8-0) Kolar & Lodge, [2001;](#page-8-0) Darrigran, [2002](#page-7-0); Grigorovich et al., [2003\)](#page-7-0). Moreover, global change may increase the chance of success of exotic species by declining the fitness of local species to the new environment (Dukes & Mooney, [1999](#page-7-0)).

Before a species becomes a pest in an ecosystem, it must successfully overcome several filters (Kolar & Lodge, [2001](#page-8-0); Sakai et al., [2001\)](#page-9-0). First, the species must travel from its native range to a new ecosystem (transport). Second, it must survive, grow and reproduce under the new environmental conditions (establishment). Third, it must acquire a high rate of population growth, invading new regions (spread). Finally, the alien species must alter the structure and functioning of the invaded ecosystem (impact) (Parker et al., [1999\)](#page-8-0). Different traits may confer success to overcome each step, so all of them must coincide in the same species to assure its invasive success (Williamson & Fitter, [1996;](#page-9-0) Sakai et al., [2001\)](#page-9-0). Most studies on invasion processes have searched for functional traits explaining the spreading success (Table 1). However, fewer studies have searched for traits explaining success in the previous invasive phases (Kolar & Lodge, [2001\)](#page-8-0), partly because exotic species do not catch the attention of researches before being widely spread. Therefore, it remains unknown whether the traits explaining spread success of a given species also contribute to explain its success in transport, establishment and impact, or different sets of traits are required to pass each filter. In order to answer this question, detailed information of the whole invasive process of a species is needed.

The New Zealand mud snail, Potamopyrgus antipodarum Gray (=P. jenkinsi Smith) (Hydrobiidae, Mollusca), meets the above condition. This invertebrate, native from New Zealand and adjacent islands (Gangloff, [1998](#page-7-0); Ponder, [1988](#page-8-0)), has successfully spread through fast rivers, slow-flowing and brackish water ecosystems of four continents (Heywood & Edwards, [1962;](#page-7-0) Quinn et al., [1998](#page-8-0); Leppäkoski & Olenin, [2000](#page-8-0); Shimada & Urabe, [2003;](#page-9-0) Cada, [2004](#page-7-0); Kerans et al., [2005](#page-8-0); Strzelec, [2005;](#page-9-0) ANS, [2007\)](#page-7-0). Its first occurrence in Europe was dated in England in 1859 (Ponder, [1988\)](#page-8-0). In Australia, the species was first reported in Tasmania in 1892, and then in Victoria in 1895 (Ponder, [1988](#page-8-0)). In North America, this species was first cited in the Middle Snake River (Idaho, north-west of USA) in 1987, probably escaped from a fish farm (Bowler, [1991\)](#page-7-0). In 1991, mud snails were found in Lake Ontario (North-East USA) and in 1997 in Columbia River (Oregon, north-west of USA), where they probably arrived via ballast water from commercial ships (Zaranko et al., [1997;](#page-9-0) Gangloff, [1998](#page-7-0)). Recently, this species has been cited in Japan (Shimada & Urabe, [2003](#page-9-0)). In spite of this rapid and well-documented spread, little is known about the potential effect of mud snail on the native communities. Moreover, most of the available information has been gathered at a local scale, and no attempt has been done to put these pieces together to gain insight on the whole process of invasion.

The aim of this work was to investigate whether the attributes which confer mud snail success in transport, spread, establishment and impact are the same or not by reviewing all published information on this mollusc.

Table 1 Functional traits reported in bibliography to explain the spreading success of alien species

Trait	References	
Rapid growth rate	Baruch & Bilbao (1999), Pattison et al. (1998)	
Early sexual maturity	Richardson et al. (1990)	
High fecundity	Richardson et al. (1990), Møller et al. (1994), Zaranko et al. (1997), Richards (2002)	
Low size of propagules	Rejmánek & Richardson (1996)	
Vegetative and parthenogenetic reproduction	Huenneke & Vitousek (1990), Lively (1992), Lake & Leishman (2004), Reichard & Hamilton (1997)	
Low susceptibility to natural enemies	Gérard et al. (2003), Maron et al. (2004), Vilà et al. (2005), Vinson & Baker (2008)	
Ability to tolerate wide range of abiotic conditions	Alonso & Camargo $(2003, 2004)$, Baruch & Bilbao (1999) , Morton (1996), Gérard et al. (2003), Strayer (1999)	
High evolutionary potential to adapt to new environments	Hänfling & Kollmann (2002) , Lee (2002)	

Bold letters highlight those functional traits present in P. antipodarum and the studies reporting them

Life-history traits and colonized habitats

P. antipodarum is a prosobranch snail (Hydrobiidae, Mollusca), which reaches a maximum size of 6– 7 mm in invaded regions, but can be up to 12 mm in New Zealand (Winterbourn, [1970](#page-9-0)). This snail has a solid operculum and its shell is long (Duft et al., [2003a](#page-7-0), [b\)](#page-7-0). Although, in its natural range, both sexual and asexual reproduction coexists, non-native populations are parthenogenetic, consisting almost exclusively of females (Lively, [1987;](#page-8-0) Jokela et al., [1997;](#page-8-0) Gangloff, [1998](#page-7-0); Jensen et al., [2001](#page-7-0); Duft et al., [2003a](#page-7-0), [b\)](#page-7-0). This invertebrate is ovoviviparous, and females brood their offsprings to the ''crawl-away'' developmental stage in a brood pouch (Jokela et al., [1997\)](#page-8-0). It reaches sexual maturity at 3–3.5 mm of shell length (Møller et al., [1994](#page-8-0); Richards, [2002](#page-8-0)). The number of generations per year ranges from 1 to 6, and one adult individual can produce an average of 230 juveniles per year (Møller et al., [1994](#page-8-0); Richards, [2002\)](#page-8-0). Its diet includes periphyton, macrophytes and detritus (Dorgelo & Leonards, [2001](#page-7-0); Jensen et al., [2001;](#page-7-0) Alonso & Camargo, [2003](#page-7-0); Duft et al., [2003a,](#page-7-0) [b;](#page-7-0) Alonso, [2005](#page-7-0)). This snail can dwell on different substrata, such as aquatic macrophytes, clay, fine sand and mud (Heywood & Edwards, [1962](#page-7-0); Marshall & Winterbourn, [1979](#page-8-0); Weatherhead & James, [2001](#page-9-0)). In addition, it buries itself in the sediment to stand dry or cold periods (Duft et al., [2003a\)](#page-7-0).

Within its native range mud snail lives in freshwater ecosystems, except temporary ponds, and also inhabits brackish waters (Winterbourn, [1973](#page-9-0)). However, in the invaded regions, it can be found in a higher variety of habitats (Table [2](#page-3-0)). Although in most cases P. antipodarum lives in freshwater habitats, it has also been found in brackish and even salty water. Regarding water speed, mud snail has colonized streams, lakes, reservoirs, estuaries and even open seas (Table [2](#page-3-0)).

Transport

There are several transport mechanisms that have been reported in literature for P. antipodarum (Table [3](#page-4-0)). The most frequently cited long-distance transport is through ballast water of commercial ships (Zaranko et al., [1997;](#page-9-0) Gangloff, [1998](#page-7-0); Leppäkoski & Olenin, 2000 ; Leppäkoski et al., 2002 ; Richards, 2002), which may explain transoceanic transport (e.g. from Australia to Europe). Other reported long- or short-distance transport means relate with commercial movements of aquaculture products, or aquatic ornamental plants; mud snails may also travel within freshwater tanks and water pipes, or within the mud attached to bills or legs of birds, or even inside the gut of birds or fishes (Haynes et al., [1985](#page-7-0); Ponder, [1988;](#page-8-0) Aarnio & Bonsdorff, [1997](#page-6-0); Zaranko et al., [1997](#page-9-0); Gangloff, [1998;](#page-7-0) Leppäkoski & Olenin, [2000](#page-8-0); Richards, [2002](#page-8-0)). Finally, other transport mechanisms are recreational vessels (e.g. kayaks and rafts) and sport fishing tools (e.g. waders and boots), where mud snails may adhere (Hosea & Finlayson, [2005;](#page-7-0) ANS, [2007](#page-7-0)).

Although the arrival to any of these transport means may be a matter of chance, the survival during the journey requires wide tolerance to physicochemical conditions. For example, a successful transport via ballast water requires high tolerance to salinity (Leppäkoski & Olenin, [2000;](#page-8-0) Richards, [2002](#page-8-0); Gérard et al., [2003](#page-7-0)). In fact, this snail has been reported to survive after short-term exposures to salinities as high as 32%, and it can feed, grow and reproduce at 15‰ salinity (Jacobsen & Forbes, [1997;](#page-7-0) Costil et al., [2001](#page-7-0); Gérard et al., [2003\)](#page-7-0).

Long distance travels also require wide tolerance to temperature change. Some authors have found that this species tolerates temperature from 0 to 28° C (Winterbourn, [1969](#page-9-0); Hylleberg & Siegismund, [1987](#page-7-0)). In an experimental study, Vareille-Morel [\(1985](#page-9-0)a, [b\)](#page-9-0) found a range of temperature tolerance between 9 and 27-C among individuals of different populations. In other experimental study, Dybdahl & Kane ([2005\)](#page-7-0) tested the growth of P. antipodarum to a range of temperatures from 12 to 24° C, finding the highest growth rate at 18°C. But to our knowledge, no study has assessed the reproductive success after exposure to extreme temperatures.

Successful transport on mud, fishing tools or recreational vessels requires a high tolerance to desiccation. Several authors have reported that this snail can survive after short desiccation periods (Bowler, [1991](#page-7-0); Zaranko et al., [1997;](#page-9-0) Cada, [2004;](#page-7-0) Lysne & Koetsier, [2006\)](#page-8-0), although desiccation tolerance declines at increasing temperature, and with decreasing snail size (Richards et al., [2004\)](#page-9-0). Fewer studies have tested the reproductive success of *P. antipodarum* after desiccation and for different environmental conditions (Vareille-Morel, [1985a,](#page-9-0) [b;](#page-9-0) Bowler, [1991;](#page-7-0) Quinn et al., [1994](#page-8-0); Zaranko

Table 2 Habitats colonized by the mud snail Potamopyrgus antipodarum. Habitat type, water salinity and population densities are shown for each site whenever available

et al., [1997\)](#page-9-0). These studies suggest that transport means implying desiccation are only effective for short distance movements, therefore contributing to snail spread once the species has reached a new area. The same can be argued for travels in other animal's gut, as mud snail can only stand such conditions for a few hours (Haynes et al., [1985;](#page-7-0) Bowler, [1991;](#page-7-0) Aarnio & Bonsdorff, [1997](#page-6-0); Zaranko et al., [1997](#page-9-0); Cada, [2004](#page-7-0); Lysne & Koetsier, [2006](#page-8-0)). In summary, wide tolerance range to physico-chemical variables is a key trait assuring travel success of mud snail, both at long and short distances.

Establishment

A wide tolerance range to multiple environmental factors also increases the chances of an exotic species survival once it has arrived in a new environment.

Invasion step	Mechanism	Reference
Transport	Ship ballast water	Zaranko et al. (1997), Gangloff (1998), Leppäkoski & Olenin, (2000), Richards (2002), Leppäkoski et al. (2002)
	Holding on aquatic ornamental plants	Ribi (1986)
	Movements of aquaculture products	Bowler (1991)
	Freshwater tanks	Ponder (1988)
	Water pipes	Ponder (1988)
	Bird and fish guts; Bills or legs of birds	Haynes et al. (1985), Aarnio & Bonsdorff (1997), Zaranko et al. (1997)
	Others: holding on recreational vessels, sport fishing tools, etc.	Hosea & Finlayson (2005); ANS (2007)
	Establishment Tolerance to wide range of environmental conditions	
	High tolerance to salinity	Leppäkoski & Olenin (2000), Richards (2002), Gérard et al. (2003)
	High tolerance to extreme temperatures	Winterbourn (1969) , Hylleberg & Siegismund (1987)
	High tolerance to human perturbations	Alonso & Camargo (2003) , Alonso (2005)
	High competition ability as compared with invertebrate native fauna (dependent on snail density)	Gangloff (1998), Schreiber et al. (2002, 2003), Hall et al. (2006)
Spread	Passive methods	
	Dispersal by birds and fish	Lassen (1975), Haynes et al. (1985)
	Drift	Richards et al. (2001)
	Holding in floating aquatic macrophytes	Ribi (1986)
	Active methods	
	Positive rheotactic	Haynes et al. (1985)
Impact	High consumption rate of primary production	Hall et al. (2003)
	High secondary production	Hall et al. (2006)
	Reduction in the colonization ability of other invertebrates	Ponder (1988), Kerans et al. (2005)
	Asymmetrical competition with native snail	Riley et al. (2008)

Table 3 Summary of transport, establishment, spread and impact mechanisms reported in scientific bibliography for *Potamopyrgus* antipodarum

However, the species still have to overcome the biological resistance opposed by the local community (competition, predation, diseases, etc.) to assure a long-term successful establishment. Theoretically, the exotic species can take advantage of two different, but not exclusive, strategies to do so: to possess a high potential to overcome biological resistance (by means of high competitive potential, escape from natural enemies, etc.), and/or to be a successful colonizer of empty spaces, where disturbances have reduced or eliminated local populations.

In non-native regions, mud snail has been mainly found in human-disturbed environments (Zaranko et al., [1997](#page-9-0); Mouthon & Charvet, [1999;](#page-8-0) Gérard et al., [2003;](#page-7-0) Schreiber et al., [2003](#page-9-0); Cada, [2004](#page-7-0); Richards et al., [2004](#page-9-0); Alonso, [2005\)](#page-7-0), as occurs with other exotic species (Rejmánek & Richardson, [1996](#page-8-0); Almasi, [2000;](#page-7-0) Lake & Leishman, [2004](#page-8-0)). Humaninduced disturbances increases the chance of success for recently arrived species, either by increasing resource availability (i.e. eutrophication), or by releasing resources capitalized by local populations (Thompson et al., [2001](#page-9-0); Schreiber et al., [2003](#page-9-0)). In habitats altered by human activities, P. antipodarum performs as a successful early colonizer (Quinn et al., [1998\)](#page-8-0) dominating the incipient community (Schreiber et al., [2003](#page-9-0); Strzelec, [2005](#page-9-0); Strzelec et al., [2005](#page-9-0); Lewin & Smolinski, [2006\)](#page-8-0), probably due to the low biotic resistance exerted by the remaining simplified native communities. The escape from parasites can additionally contribute to explain mud snail successful establishment, as it seems to leave their trematode

parasites behind when invading new regions (Gérard et al., [2003\)](#page-7-0). Experimental studies have shown that P. antipodarum growth was reduced by the presence of trematodes (Krist & Lively, [1998](#page-8-0)). Finally, mud snails are also resistant to many native predators, because of its hard shell and solid operculum (Zaranko et al., [1997](#page-9-0); Vinson & Baker, [2008](#page-9-0)). All these traits can help mud snail for a successful establishment in a new area.

Spread

Mud snail may disperse both by passive and active methods (Table [3](#page-4-0)). Passive methods have been described as the principal way of spread in European waters for P. antipodarum (Hubendick, [1950;](#page-7-0) Lassen, [1975\)](#page-8-0). Among them, several authors reported birds and fish as dispersal agents (Haynes et al., [1985](#page-7-0); Ribi, [1986;](#page-8-0) Aarnio & Bonsdorff, [1997;](#page-6-0) Zaranko et al., [1997](#page-9-0)), while others reported passive drift or dispersal by holding in floating aquatic macrophytes (Ribi, [1986](#page-8-0); Richards et al., [2001](#page-9-0)). Mud snail was found to be one of the most abundant macroinvertebrate in drift net samples in Banbury Springs (Idaho, USA) (Richards et al., [2001](#page-9-0)). These authors also showed that P. antipodarum used floating vegetation mats to colonize a lake. However, these mechanisms are only effective to colonize lakes or currents downstream from the initial population. In Australia, Loo et al. ([2007a](#page-8-0)) found that fish stocking and anglers were two passive spread mechanisms to P. antipodarum.

Regarding active dispersal mechanisms, some authors have found that positive rheotactic response can facilitate spread in invaded streams and rivers (Adam, [1942](#page-7-0); Haynes et al., [1985](#page-7-0)), and that high water speed produces a more consistent upstream movement (Haynes et al., [1985\)](#page-7-0). Adam ([1942\)](#page-7-0) found in Belgium that mud snails can spread 60 m in three months by active upstream movements. At this spread rate a single mud snail might can move upstream up to 240 m in just one year. Furthermore, as each snail can produce more than 230 juveniles per year, the number of snails in the reach can dramatically increase, as it moves upstream in a reach (Møller et al., [1994](#page-8-0); Richards, [2002\)](#page-8-0). By contrast, Richards et al. [\(2001](#page-9-0)) reported that P. antipodarum is prone to detach from substrate in high-speed waters, suggesting that fast waters can limit colonization. They also found that aquatic macrophytes are a good refuge for juveniles of P. antipodarum, which are more sensitive to velocity than adults. According to these authors, low-speed waters with high densities of macrophytes are more susceptible to mud snail spread than high-speed waters. These contradictory results indicate the necessity of further research on the active dispersal method of mud snail.

All these ecological traits help mud snail to a rapid spread around different aquatic ecosystems. Recent predictive models developed for P. antipodarum have shown that the future spread of mud snail through Australia and North America could be very fast unless prevention measures are rapidly implemented (Loo et al., [2007b\)](#page-8-0).

Impact

The principal impact of mud snail in invaded ecosystem can be attributed to its high reproductive capacity, which leads to an explosive population growth, a fast spread and a high consumption rate of the available primary production of the ecosystem. P. antipodarum is a successful early colonizer (Quinn et al., [1998\)](#page-8-0) dominating the incipient community (Schreiber et al., [2003;](#page-9-0) Strzelec, [2005](#page-9-0); Strzelec et al., [2005;](#page-9-0) Lewin & Smolinski, [2006\)](#page-8-0). In a highly productive stream of Wyoming (USA), Hall et al. [\(2003](#page-7-0)) found that mud snail consumed 75% of the primary production and excreted about 65% of the total NH_4^+ demanded by microbes and plants, therefore dominating both C and N cycles. These authors compared the effects of mud snail to those of zebra mussel (Dreissena polymorpha), an invasive bivalve which can consume nearly all the primary production of the community (Strayer, [1999](#page-9-0); Hall et al., [2003](#page-7-0)). The same authors found in another study that the secondary productivity of P. antipodarum was one of the highest ever reported for a stream invertebrate (194 g AFDM m^{-2} yr⁻¹), being 7-40 times higher than that of any macroinvertebrate in Greater Yellowstone area (Wyoming, USA) (Hall et al., [2006\)](#page-7-0). In a tributary of Snake River (Idaho, USA), mud snail densities were reported to be higher than those of native snails in three different habitats (run, edge and vegetation) (Richards et al., [2001](#page-9-0)). In ditches and canals of the Basin of the Mont St-Michel Bay

(France), mud snail dominates the gastropod communities in fresh- and salt-water ecosystems (Gérard et al., [2003](#page-7-0)). Lewin & Smolinski [\(2006](#page-8-0)) reported that mud snail made up 83% of the mollusc community in a reservoir near an industrial area in Poland. Experimental studies also showed that mud snail reduced colonization by other invertebrates in the early stages of succession (Kerans et al., [2005](#page-8-0); Ponder, [1988\)](#page-8-0).

Several authors have compared the densities of mud snail between degraded and intact stream. Alonso [\(2005](#page-7-0)) found high densities of P. antipodarum in human-polluted reaches of the Henares River, whereas no snails were found in well-preserved areas. Similarly, Schreiber et al. ([2003\)](#page-9-0) found higher density of P. antipodarum in areas with multiple human land uses than in low-impact sites of Victoria streams (Australia). Moreover, at low population density, P. antipodarum has been even found to facilitate some native invertebrates, as its faeces, which contain processed cellulases and chitinases plus mucoproteins and mucopolysaccharides, constitute a suitable food for native grazers and depositfeeders (Gangloff, [1998](#page-7-0); Schreiber et al., [2002](#page-9-0)).

According to the five-level framework put forward by Parker et al. ([1999\)](#page-8-0) to assess the impact of an invader, mud snail impacts on the invaded ecosystems are related mainly with two levels: (1) the population effects, given that P. antipodarum possess a population density in the invaded ecosystem higher than most native invertebrates and (2) effects on ecosystem processes, as the mud snail can consume most of the primary production of the stream, and therefore it can dominate the secondary production of the invertebrate community.

This revision shows that the invasion success of mud snail can be largely dependent on the conservation state of the invaded habitat, and that P. antipodarum is a very successful colonizer of empty spaces, typically occurring at early stages of succession, but less successful at overcoming the biological resistance of an intact native community.

Concluding remarks

The revised bibliography has shown that a wide tolerance to physico-chemical conditions contributes to explain the success of mud snail in the two former steps to become an invasive species (transport and

establishment). However, a successful establishment also relies on a high capacity to overcome biotic resistance, either by successfully colonizing early stages of succession in human-altered habitats, or by leaving behind parasite and predator control. Its high reproductive rate, together with its ability to disperse by active and passive mechanisms, explains mud snail potential for an efficient spread. Finally, mud snail ability to alter the structure and function of invaded ecosystems (impact) is again due to the high reproductive rate, which leads to extremely high population density and to the consumption of most of the primary productivity of the ecosystem. Therefore, the coincidence of wide tolerance to abiotic factors and high reproductive capacity on the same species may have allowed it overcome most of the filters to become a pest. Human-disturbed ecosystems are more susceptible to mud snail invasion than intact ones, although the latter may be also affected by mud snail.

Future research on mud snail invasion should address several open questions: (1) To assess the reproductive viability of mud snail after exposure to gradients of different conditions (humidity, temperature, etc.) to understand its transport-spread potential, (2) to study the potential impacts of mud snail on native faunas at different densities, especially in perturbed ecosystems, where it apparently shows higher success and (3) to identify the ecosystems that are susceptible to invasion in order to prevent spread of P. antipodarum into these regions.

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References

Aarnio, K. & E. Bonsdorff, 1997. Passing the gut of juvenile flounder, Platichthys fesus (l.)–diferential survival of zoobenthic prey species. Marine Biology 129: 11–14.

- Adam, W., 1942. Sur la répartition et la biologie de Hydrobia jenkinsi Smith en Belgique. Bulletin du Museé royal d'Histoire naturell de Belgique 18: 1–18.
- Almasi, K. N., 2000. A Non-native Perennial Invades a Native Forest. Biological Invasions 2: 219–230.
- Alonso, A., 2005. Valoración de la degradación ambiental y efectos ecotoxicológicos sobre la comunidad de macroinvertebrados bentónicos en la cabecera del río Henares. Dissertation, Universiy of Alcalá, Spain.
- Alonso, A. & J. A. Camargo, 2003. Short-term toxicity of ammonia, nitrite and nitrate to the aquatic snail Potamopyrgus antipodarum (Hidrobiidae, Mollusca). Bulletin of Environmental Contamination and Toxicology 70: 1006–1012.
- Alonso, A. & J. A. Camargo, 2004. Sub-lethal responses of the aquatic snail Potamopyrgus antipodarum (Hydrobiidae, Mollusca) to unionized ammonia: a tolerant invading species. Fresenius Environmental Bulletin 13: 607–615.
- ANS, 2007. National management and control plan for the New Zealand mudsnail (Potamopyrgus antipodarum). United States Federal Aquatic Nuisance species Task Force. [www.anstaskforce.gov/Documents/NZMS_Mgmt](http://www.anstaskforce.gov/Documents/NZMS_MgmtControl_Final.pdf) [Control_Final.pdf.](http://www.anstaskforce.gov/Documents/NZMS_MgmtControl_Final.pdf)
- Baruch, Z. & B. Bilbao, 1999. Effects of fire and defoliation on the life history of native and invader C-4 grasses in a Neotropical savanna. Oecologia 119: 510–520.
- Bowler, P. A., 1991. The rapid spread of the freshwater Hydrobiid snail Potamopyrgus antipodarum (Gray) in the Middle Snake River, Southern Idaho. Proceeding of the Desert Fishes Council 21: 173–182.
- Brzezinski, T. & A. Kolodziejczyk, 2001. Distribution of Potamopyrgus antipodarum (Gray, 1843) in waters of the Wigry National Park and the effect of selected habitat factors on its occurrence. Folia Malacologica 9: 125–135.
- Cada, C. A., 2004. Interactions between the invasive New Zealand mud snail, Potamopyrgus antipodarum, baetid mayflies, and fish predators. Dissertation, University of Montana State
- Cambray, J. A., 2003. Impact on indigenous species biodiversity caused by globalisation of alien recreational freshwater fisheries. Hydrobiologia 500: 217–230.
- Costil, K., G. B. J. Dussart & J. Daguzan, 2001. Biodiversity of aquatic gastropods in the Mont St-Michel basin (France) in relation to salinity and drying of habitats. Biodiversity and Conservation 10: 1–18.
- Darrigran, G., 2002. Potential impact of filter-feeding invaders on temperate inland freshwater environments. Biological Invasions 4: 145–156.
- Dorgelo, J., 1987. Density fluctuations in populations (1982– 1986) and biological observations of Potamopyrgus jenkinsi in two trophically differing lakes. Hydrobiol Bull Amsterdam 21: 95–110.
- Dorgelo, J. & P. E. G. Leonards, 2001. Relationship between C/N ratio of food types and growth rate in the snail Potamopyrgus jenkinsi (E. A. Smith). Journal of the North American Benthological Society 20: 60–67.
- Duft, M., U. Schulte-Oehlmann, M. Tillman, B. Markert & J. Oehlmann, 2003a. Toxicity of triphenyltin and tributyltin to the freshwater mudsnail Potamopyrgus antipodarum in a new sediment biotest. Environmental Toxicology and Chemistry 22: 145–152.
- Duft, M., U. Schulte-Oehlmann, L. Weltje, M. Tillmann & J. Oehlmann, 2003b. Stimulated embryo production as a parameter of estrogenic exposure via sediments in the freshwater mudsnail Potamopyrgus antipodarum. Aquatic Toxicology 64: 437–449.
- Dukes, J. S. & H. A. Mooney, 1999. Does global change increase the success of biological invaders? Trends in Ecology and Evolution 14: 135–139.
- Dybdahl, M. F. & S. L. Kane, 2005. Adaptation vs. phenotypic plasticity in the success of a clonal invader. Ecology 86: 1592–1601.
- Enserink, M., 1999. Biological invaders sweep in. Science 285: 1834–1836.
- Gangloff, M. M., 1998. The New Zealand mud snail in Western North America. Aquatic Nuisance Species 2: 25–30.
- Gérard, C., A. Blanc & K. Costil, 2003. Potamopyrgus antipodarum (Mollusca:Hydrobiidae) in continental aquatic gastropod communities: impact of salinity and trematode parasitism. Hydrobiologia 493: 167–172.
- Grant, A. & A. D. Briggs, 1998. Toxicity of ivermectin to estuarine and marine invertebrates. Marine Pollution Bulletin 36: 540–541.
- Grigorovich, I. A., A. V. Korniushin, D. K. Gray, I. C. Duggan, R. I. Colautti & H. J. MacIsaac, 2003. Lake Superior: an invasion coldspot? Hydrobiologia 499: 191–210.
- Hall, Jr., R. O., M. F. Dybdahl & M. C. VanderLoop, 2006. Extremely high secondary production of introduced snails in rivers. Ecological Applications 16: 1121–1131.
- Hall, Jr., R. O., J. L. Tank & M. F. Dybdahl, 2003. Exotic snails dominate nitrogen and carbon cycling in a highly productive stream. Frontiers in Ecology and Environment 1: 407–411.
- Hänfling, B. & J. Kollmann, 2002. An evolutionary perspective of biological invasions. TREE 17: 545–546.
- Haynes, A., B. J. R. Taylor & M. E. Varley, 1985. The influence of the mobility of Potamopyrgus jenkinsi (Smith, E.A.) (Prosobranchia: Hydrobiidae) on its spread. Archives für Hydrobiologie 100: 479-491.
- Heywood, J. & R. W. Edwards, 1962. Some aspects of the ecology of Potamopyrgus jenkinsi Smith. Journal of Animal Ecology 31: 239–250.
- Hosea, R. C., & B. Finlayson, 2005. Controlling the spread of New Zealand mud snails on wading gear. State of California, The Resources Agency. Administrative Report 2005-02
- Hubendick, B., 1950. The effectiveness of passive dispersal in Hydrobia jenkinsi. Zoologiska Bidrag fran Uppsala 28: 493–504.
- Huenneke, L. F. & P. M. Vitousek, 1990. Seedling and clonal recruitment of the invasive tree Psidium cattleianum: Implications for management of native Hawaiian forests. Biological Conservation 53: 199–211.
- Hylleberg, J. & H. R. Siegismund, 1987. Niche overlap in mud snail (Hydrobiidae): frezing tolerance. Marine Biology 94: 403–407.
- Jacobsen, R. & V. E. Forbes, 1997. Clonal variation in lifehistory traits and feeding rates in the gastropod, Potamopyrgus antipodarum: performance across a salinity gradient. Functional Ecology 11: 260–267.
- Jensen, A., V. E. Forbes & D. Parker Jr., 2001. Variation in cadmium uptake, feeding rate, and life-history effects in

the gastropod Potamopyrgus antipodarum: linking toxicant effects on individuals to the population level. Environmental Toxicology and Chemistry 20: 2503–2513.

- Jokela, J., C. M. Lively, M. E. Dybdahl & J. A. Fox, 1997. Evidence for a cost of sex in the freshwater snail Potamopyrgus antipodarum. Ecology 78: 452–460.
- Katayama, M. & S. Ryoji, 2004. The first record of a freshwater snail Potamopyrgus antipodarum (Mollusca, Gastropoda) in Gunna Prefecture. Field Biology 13: 46– 48. in Japan, with English abstract.
- Kerans, B. L., M. E. Dybdahl, M. M. Gangloff & J. E. Jannot, 2005. Potamopyrgus antipodarum: distribution, density, and effects on native macroinvertebrate assemblages in the Greater Yellowstone Ecosystem. Journal of the North American Benthological Society 24: 123–138.
- Kolar, C. S. & D. M. Lodge, 2001. Progress in invasion biology: predicting invaders. Trends in Ecology and Evolution 16: 199–204.
- Krist, A. C. & C. M. Lively, 1998. Experimental exposure of juvenile snails (Potamopyrgus antipodarum) to infection by trematode larvae (Microphallus sp.): infectivity, fecundity compensation and growth. Oikos 116: 575–582.
- Lake, J. C. & M. R. Leishman, 2004. Invasion success of exotic plants in natural ecosystems: the role of disturbance, plant attributes and freedom from herbivores. Biological Conservation 117: 215–226.
- Lassen, H. H., 1975. Diversity of freshwater snails in view of equilibrium theory of island biogeography. Oecologia 19: 1–8.
- Lee, C. E., 2002. Evolutionary genetics of invasive species. TREE 17: 386–391.
- Leppäkoski, E., 1984. Introduced species in the Baltic Sea and its coastal ecosystems. Ophelia 3: 123–135.
- Leppäkoski, E., S. Gollasch, P. Gruszka, H. Ojaveer, S. Olenin & V. Panov, 2002. The Baltic—a sea of invaders. Canadian Journal of Fisheries and Aquatic Sciences 59: 1175–1188.
- Leppäkoski, E. & S. Olenin, 2000. Non-native species and rates of spread: lessons from the brackish Baltic Sea. Biological Invasions 2: 151–163.
- Lewin, I. & A. Smolinski, 2006. Rare and vulnerable species in the mollusc communities in the mining subsidence reservoirs of an industrial area (The Katowicka Upland, Upper Silesia, Southern Poland). Limnologica 36: 181–191.
- Lively, C. M., 1987. Evidence from a New Zealand snail for the maintenance of sex by parasitism. Nature 328: 519–521.
- Lively, C. M., 1992. Parthenogenesis in a freshwater snail reproductive assurance versus parasitic release. Evolution 46: 907–913.
- Loo, S. E., R. P. Keller & B. Leung, 2007a. Freshwater invasions: using historical data to analyse spread. Diversity and Distributions 13(1): 23–32.
- Loo, S. E., R. M. Nally & P. S. Lake, 2007b. Forecasting New Zealand mudsnail invasion range: Model comparisons using native and invaded ranges. Ecological Applications 17: 181–189.
- Lysne, S. & P. Koetsier, 2006. Experimental studies on habitat preference and tolerances of three species of snails from the Snake River of southern Idaho, USA. American Malacological Bulletin 21: 77–85.
- Maron, J. L., M. Vilà & J. Arnason, 2004. Loss of enemy resistance among introduced populations of St. John's wort (Hypericum perforatum). Ecology 85: 3243–3252.
- Marshall, J. W. & M. J. Winterbourn, 1979. An ecological study of a small New Zealand stream with particular reference to the oligochaeta. Hydrobiologia 65: 199–208.
- Mills, E. L., J. M. Casselman, R. Dermott, J. D. Fitzsimons, G. Gal, K. T. Holeck, J. A. Hoyle, O. E. Johannsson, B. F. Lantry, J. C. Makarewicz, E. S. Millard, I. F. Munawar, M. Munawar, R. O'Gorman, R. W. Owens, L. G. Rudstam, T. Schaner & T. J. Stewart, 2003. Lake Ontario: food web dynamics in a changing ecosystem (1970– 2000). Canadian Journal of Fisheries and Aquatic Sciences 60: 471–490.
- Møller, V., V. E. Forbes & M. H. Depledge, 1994. Influence of acclimation and exposure temperature on the acute toxicity of cadmium to the freshwater snail Potamopyrgus antipodarum (Hydrobiidae). Environmental Toxicology and Chemistry 13: 1519–1524.
- Morton, B., 1996. The aquatic nuisance species: a global perspective and review. In D'itri, F. (ed.), Zebra Mussels and Other Aquatic Species. Ann Arbor Press, Ann Arbor, Michigan.
- Mouthon, J. & S. Charvet, 1999. Compared sensitivity of species, genera and families of Molluscs to biodegradable pollution. Annales de Limnologie 35: 31–39.
- Parker, I. M., D. Simberloff, W. M. Lonsdale, K. Goodell, M. Wonham, P. M. Kareiva, M. H. Williamson, B. Von Holle, P. B. Moyle, J. E. Byers & L. Goldwasser, 1999. Impact: toward a framework for understanding the ecological effects of invaders. Biological Invasions 1: 3–19.
- Pattison, R. R., G. Goldstein & A. Ares, 1998. Growth, biomass allocation and photosynthesis of invasive and native Hawaiian rainforest species. Oecologia 117: 449–459.
- Ponder, W. F., 1988. Potamopyrgus antipodarum, a molluscan colonizer of Europe and Australia. Journal of Molluscan Studies 54: 271–286.
- Quinn, G. P., P. S. Lake & S. G. Schreiber, 1998. A comparative study of colonization by benthos in a lake and its outflowing stream. Freshwater Biology 39: 623–635.
- Quinn, J. M., G. L. Steele, C. W. Hickey & M. L. Vickers, 1994. Upper thermal tolerances of twelve New Zealand stream invertebrate species. New Zealand Journal of Marine and Freshwater 28: 391–397.
- Rahel, F. J., 2002. Homogenization of freshwater faunas. Annual Review of Ecology and Systematics 33: 291–315.
- Reichard, S. J. & C. W. Hamilton, 1997. Predicting invasions of woody plants introduced into North America. Conservation Biology 11: 193–2003.
- Rejma´nek, M. & D. M. Richardson, 1996. What attributes make some plant species more invasive? Ecology 77: 1655–1661.
- Ribi, G., 1986. Within-lake dispersal of the prosobranch snails, Viviparus ater and Potamopyrgus jenkinsi. Oecologia 69: 60–63.
- Ricciardi, A. & H. J. MacIsaac, 2000. Recent mass invasion of the North American Great Lakes by Ponto-Caspian species. Trends in Ecology and Evolution 15: 62–65.
- Richards, D. C., 2002. The New Zealand mudsnail invades the Western United States. Aquatic Nuisance Species 4: 42–44.
- Richards, D. C., L. D. Cazier & G. T. Lester, 2001. Spatial distribution of three snail species, including the invader Potamopyrgus antipodarum, in a freshwater spring. Western North American Naturalist 61: 375–380.
- Richards, D. C., P. O'Connell & D. C. Shinn, 2004. Simple control method to limit the spread of the New Zealand mudsnail Potamopyrgus antipodarum. North American Journal of Fisheries Management 24: 114–117.
- Richardson, D. M., L. Cowling & D. Lemaitre, 1990. Assessing the risk of invasive success in Pinus and Banksia in South-African mountain Fynbos. Journal of Vegetation Science 1: 629–642.
- Riley, L. A., M. F. Dybdahl & R. O. Hall, 2008. Invasive species impact: asymmetric interactions between invasive and endemic freshwater snails. Journal of the North American Benthological Society 27: 509–520.
- Sakai, A. K., F. W. Allendorf, J. S. Holt, D. M. Lodge, J. Molofsky, K. A. With, S. Baughman, R. J. Cabin, J. E. Cohen, N. C. Ellstrand, D. E. McCauley, P. O'Neil, I. M. Parker, J. N. Thompson & S. G. Weller, 2001. The population biology of invasive species. Annual Review of Ecology and Systematics 32: 305–332.
- Schreiber, E. S. G., P. S. Lake & G. P. Quinn, 2002. Facilitation of native stream fauna by an invading species? Experimental investigations of the interaction of the snail, Potamopyrgus antipodarum (Hydrobiidae) with native benthic fauna. Biological Invasions 4: 317–325.
- Schreiber, E. S. G., G. P. Quinn & P. S. Lake, 1998. Life history and population dynamics of the exoctic snail Potamopyrgus antipodarum (Prosobranchia: Hydrobiidae) in Lake Purrumbete, Victoria, Australia. Marine and Freshwater Research 49: 73–78.
- Schreiber, E. S. G., G. P. Quinn & P. S. Lake, 2003. Distribution of an alien aquatic snail in relation to flow variability, human activities and water quality. Freshwater Biology 48: 951–961.
- Shimada, K. & M. Urabe, 2003. Comparative ecology of the alien freshwater snail Potamopyrgus antipodarum and the indigenous snail Semisulcospira spp. Venus 62: 39–53. in Japan English abstract.
- Strayer, D. L., 1999. Effects of alien species on freshwater mollusks in North America. Journal of the North American Benthological Society 18: 74–98.
- Strzelec, M., 2005. Impact of the introduced Potamopyrgus antipodarum (Gastropoda) on the snail fauna in postindustrial ponds in Poland. Biologia 60: 159–163.
- Strzelec, M., A. Spyra, M. Krodkiewska & W. Serafinski, 2005. The long-term transformations of Gastropod

communities in dam-reservoirs of Upper Silesia (Southern Poland). Malacologica Bohemoslovaca 4: 41–47.

- Thompson, K., J. G. Hodgson, J. P. Grime & M. J. W. Burke, 2001. Plant traits and temporal scale: Evidence from a 5-year invasion experiment using native species. Journal of Ecology 89: 1054–1060.
- Van den Berg, M., H. Coops, R. Noordhuis, J. van Schie & J. Simons, 1997. Macroinvertebrate communities in relation to submerged vegetation in two Chara-dominated lakes. Hydrobiologia 342(343): 143–150.
- Vareille-Morel, C., 1985a. Resistance of the prosobranch mollusc, Potamopyrgus jenkinsi (E. A. Smith 1889 to increasing temperatures: an experimental study. Annales de Limnologie 21: 19–24.
- Vareille-Morel, C., 1985b. Resistance of the prosobranch mollusc, Potamopyrgus jenkinsi (E. A. Smith 1889 to decreasing temperatures: an experimental study. Annales de Limnologie 21: 221–226.
- Vila`, M., J. L. Maron & L. Marco, 2005. Evidence for the enemy release hypothesis in Hypericum perforatum. Oecologia 142: 474–479.
- Vinson, M. R. & M. A. Baker, 2008. Poor growth of Rainbow Trout fed New Zealand Mud Snails Potamopyrgus antipodarum. North American Journal of Fisheries Management 28: 701–709.
- Weatherhead, M. A. & M. R. James, 2001. Distribution of macroinvertebrates in relation to physical and biological variables in the littoral zone of nine New Zealand lakes. Hydrobiologia 462: 115–129.
- Williamson, M. & A. Fitter, 1996. The varying success of invaders. Ecology 77: 1661–1666.
- Winterbourn, M. J., 1969. Water temperature as a factor limiting the distribution of Potamopyrgus antipodarum (Gastropoda-Prosobranchia) in the New Zealand thermal region. New Zealand Journal of Marine and Freshwater 3: 453–458.
- Winterbourn, M. J., 1970. The New Zealand species of Potamopyrgus (Gastropoda: Hydrobiidae). Malacologia 10: 283–321.
- Winterbourn, M. J., 1973. A guide to the freshwater mollusca of New Zealand. Tuatara 20: 141–159.
- Zaranko, D. T., D. G. Farara & F. G. Thompson, 1997. Another exotic mollusc in the Laurentian Great Lakes: the New Zealand native Potamopyrgus antipodarum (Gray 1843) (Gastropoda, Hydrobiidae). Canadian Journal of Fisheries and Aquatic Sciences 54: 809–814.