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Manipulating the expression of a cell wall invertase gene increases grain yield in maize

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Abstract Cell wall invertases play an important role in plant growth and development, especially in the grain filling of crop plants. However, their potential in high yield crop breeding has not been investigated. In this study, the main hybrid maize cultivar Zheng Dan 958 (ZD958) was used as a basic variety to assess whether ZmGIF1, a cell wall invertase from maize, can be used to breed new cultivars with higher grain yields. ZmGIF1 expression, cell wall invertase activity, and sugar content in different parental inbred cultivars were compared with those in Zheng 58 and Chang 7-2, the parental inbred cultivars of ZD958. Parental cultivars which showed higher ZmGIF1 expression and invertase activity were selected and intercrossed to improve the expression of *ZmGIF1*. Compared with the basic cultivar ZD958, higher *ZmGIF1* expression and cell wall invertase activity were observed in most hybrid F1 lines, leading to increased grain yield in them. All these results suggest that the expression of *ZmGIF1* can be manipulated using different parental cultivars to increase the grain yield of their hybrid F1 progenies.

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Introduction

In higher plants, the transport of sucrose, the major form of photosynthates, between source and sink through the phloem is largely dependent on its initial cleavage in the sink tissues and organs by invertase or sucrose synthsase (Sonnewald et al. 1997). The expression of invertase has been investigated in different plants (Albaceteet al. 2014; Canam et al. 2008; Chandra et al. 2015; Chourey et al. 2006; Kim et al. 2000; Taliercio et al. 1999; Wang et al. 2008a, b; Yao et al. 2014). The biological functions of different invertases in sink strength have been studied in several plant species such as potato (Zrenner et al. 1995), rice (Wang et al. 2008a, b) and maize (Chourey and Nelson 1976). Transgenic potato plants ectopically expressing a yeast invertase produced larger tubers than did the wild type plants (Sonnewald et al. 1997). The loss of function rice mutant of OsGIF1 exhibited decreased grain filling rate and an up to 24% reduction in seed weight. Whereas OsGIF1 overexpressing plants showed increased grain filling rate, grain size and grain weight (Wang et al. 2008a, b). In maize, ZmGIF1, a cell wall invertase (also known as Mn1 or INCW2) encoded by Mn1 (the ortholog of OsGIF1) and abundantly expressed in endosperm, works as the major determinant for sink strength in developing seeds (Chourey et al. 2012; Lowe and Nelson 1946). Similarly, the loss of function maize mutant of mnl showed an up to 70% reduction in seed weight (Cheng et al. 1996).

Genetic transformation has been used as a powerful technique to increase the grain yield in different crop plants, such as rice (*Oryza sativa*) (Jeong et al. 2013; Smidansky et al. 2003; Wu et al. 2008), wheat (*Triticum aestivum*) (Munns et al. 2012; Smidansky et al. 2002), potato (*Solanum tuberosum*)(Stark et al. 1992), cotton (Zhang et al. 2011), and aspen (Eriksson et al. 2000). An up to



25% grain weight increase in maize was also achieved by the ectopic expression of AGPase, a glgC-16 ADP glucose pyrophosphorylase from *E. coli* (Wang et al. 2007), or the overexpression of Sh2 (shrunken-2), the large subunit of AGPase, and Bt2, the endosperm small subunit of AGPase, from maize (Li et al. 2011). Recently, an up to 64% grain yield increase was obtained by overexpressing a modified Sh2 enzyme (Hannah et al. 2012).

Previously, we constitutively expressed the cell wall invertase encoding genes from *Arabidopsis*, rice and maize in the maize inbred line Ye478 (Li et al. 2013). Introduction of the cell wall invertase genes from different sources all significantly increased cell wall invertase activity in transgenic plants, and brought about an up to 145.3% increase in the grain yield of transgenic plants. In this study, we demonstrate for the first time that *ZmGIF1* can be engineered using different parental cultivars to breed high yield non-transgenic maize plants.

Materials and methods

Plant materials

Maize (*Zea mays* L.) cultivars used in this study were Zheng 58 and Chang 7-2 provided by Longping Hitech (Hefei, China), Ping An 24 provided by Yantai Qingyuan Hitech (Yantai, China), and different parental inbred cultivars bred at College of Agriculture, Ludong Unversity (Yantai, China). Plants were grown in green house and field as described previously (Li et al. 2013). For *ZmGIF1* expression analyses, total RNA was extracted from the leaves of different cultivars as described previously (Li et al. 2013).

Quantitative real-time PCR analyses

For ZmGIF1 expression analyses, total RNA was extracted from the leaves of different cultivars as described previously (Li et al. 2013). Real-time PCR was performed using ZmGIF1 forward (5'-GAACGGCAAGATATCCCT GA-3') and reverse (5'-CATGACCG GCTTCTTCATCT -3) primers and SYBR Green Real-time PCR Master Mix (TOYOBO, Osaka, Japan), as described previously (Li et al. 2013). The expression level of ZmActin1 in different cultivars was also determined as a quantitative control with forward (5'-ATCACCATTGGGTCAGAAAGG-3') and reverse (5'-GTGCTGAGAG AAGCCAAAATAG AG-3') primers. The expression of ZmGIF1 in the fifth leaves of different parental cultivars and hybrid F1 lines at six-leaf-stage was examined.

Cell wall invertase activity and sugar content assays

Seeds of different cultivars were sown in both greenhouse and field. At six-leaf-stage, the fifth fully expanded leaves of different hybrid F1 lines were collected for invertase activity assays as described previously (Li et al. 2013). The developing grains at 20 DAP (days after pollination) of different parental cultivars grown on Nanbin Farm (Sanya, Hainan Province, China) was also collected for invertase activity and sugar content analyses as described previously (Hampp et al. 1994; Li et al. 2013).

Generation of hybrid F1 lines

To produce the hybrid F1 lines, seeds of parental cultivars which showed higher *ZmGIF1* expression and cell wall invertase activity were sown on Wuyi Farm (Urmuqi, Xinjiang Province, China) and intercrossed by hand pollination with each other in 2014.

Grain yield assays

For grain yield analyses, Zheng Dan 958 (ZD958), Ping An 24 (PA24), and different hybrid F1 lines were grown under natural condition in Dehui (Jilin Province, China) in 2015 and 2016. Seeds were sown in a population density of approximately $5500/\mu$. PA24 is a local main cultivar to be used as a control variety.

Results and discussion

Selection of parental inbred cultivars

To assess whether ZmGIF1 can be used for high grain yield cultivar breeding in maize, we first performed quantitative real-time PCR and cell wall invertase activity analyses. Based on our previous studies that ZmGIF1 was predominantly expressed in sink tissues and organs (Li et al. 2013), the transcriptional level of ZmGIF1 in the fifth leaves of different parental inbred cultivars at six-leaf-stage were compared with that of Zheng 58 and Chang 7-2, the parental inbred cultivars of ZD958, which is being widely grown in China. A total number of 458 inbred cultivars were screened, and those which showed higher ZmGIF1 expression and/or invertase activity were selected for further study. Initially, most cultivars, especially female parental cultivars, which produced more biomass during their vegetative growth, also showed higher ZmGIF1 expression and invertase activity. As shown in Fig. 1, the control parental cultivars Zheng 58 and Chang 7-2 exhibited less shoot growth than did the other selected ones (Fig. 1a). Consistently, ZmGIF1 expression in their leaves and invertase activity in their developing



Fig. 1 Phenotype, ZmGIF1 expression, invertase activity, and sugar content analyses. Ten representative parental inbred lines, including Chang 7-2 and Zheng 58, used in this study were shown. **a** Phenotypes of 16-day-old seedlings (scale *bar* = 10 cm) and kernels (scale *bar* = 1 cm) of the indicated parental inbred lines. **b** Relative expression of *ZmGIF1* in the fifth leaves of parental lines in (**a**) at six-leaf-

stage. Real-time PCR was performed using ZmGIF1- or ZmActin1-specific primers. The house-keeping gene ZmActin1 was used as an internal control. The expression level of ZmGIF1 in line T-828 was assigned a value of 1.0. (**c**, **d**) Cell wall invertase activity and sugar content analyses in the developing seeds of parental lines in (**a**) at 20 DAP (days after pollination)

seeds were also lower (Fig. 1b, c). We also examined sugar contents in the developing seeds of different parental cultivars at 20 DAP, and no significant correlation between shoot growth and glycometabolism was observed (Fig. 1d). Possible explanation is that the difference of *ZmGIF1* expression and/or invertase activity between these cultivars is not significant enough. This is consistent with our previous study that higher glucose, fructose and sucrose contents were only observed in transgenic maize lines which showed very high expression of *AtGIF1*, *ZmGIF1* or *OsGIF1* (Li et al. 2013).

Manipulation of ZmGIF1 expression in hybrid F1 lines

To further explore whether *ZmGIF1* expression and invertase activity from different parental cultivars can be combined in their hybrid F1 generation, a total number of 12 parental cultivars which showed higher *ZmGIF1* expression and/ or invertase activity than did Zheng 58 or Chang 7-2 were selected, grown on Wuyi farm (Urmuqi, X·injiang Province, China), and intercrossed with each other (including Zheng 58 and Chang 7-2) in July, 2014. A total number of 182

hybrid F1 lines (combinations) were generated. As expected, the expression level of *ZmGIF1*, as well as invertase activity, was successfully combined in most hybrid F1 lines. For example, the transcriptional level of *ZmGIF1* in the leaves of hybrid lines Zheng $58 \times T316$, Zheng $58 \times T379$, Zheng $58 \times T6893$ and Zheng $58 \times T133$ was higher than that in the leaves of Zheng $58 \times Chang 7-2$ (ZD958). Consistently, the invertase activity was also significantly higher in these hybrid lines (Fig. 2a, b). Therefore, expression of *ZmGIF1* in



Fig. 2 ZmGIF1 expression and cell wall invertase activity analyses in different hybrid F1 lines. Nine representative lines including Zheng Dan 958 (ZD958), the hybrid of Zheng 58 and Chang 7-2, were shown. **a** Relative expression of ZmGIF1 in fifth leaves of six-leafstage seedlings. Real-time PCR was performed using ZmGIF1- or ZmActin1-specific primers. The house-keeping gene ZmActin1 was used as an internal control. The expression level of ZmGIF1 in hybrid line Zheng 58 and T-876 was assigned a value of 1.0. **b** Cell wall invertase activities in the fifth leaves of six-leaf-stage seedlings in (**a**)

these parental cultivars was successfully combined in their hybrid F1 generation.

Engineering of high grain yield maize plants

Previously, genetic expression of cell wall invertases from Arabidopsis, rice and maize all improved the growth rate and grain yield of transgenic maize plants. Transgenic plants grew more rapidly and produced larger cobs with increased seed size and weight than did the wild type plants (Li et al. 2013). To determine whether increased expression of ZmGIF1 and invertase activity would also increase grain yield in the generated hybrid lines, hybrid F1 plants were grown in Dehui (Jilin Province, China) for field trials in 2015 and 2016. Again, hybrid F1 lines with higher ZmGIF1 expression and invertase activity produced larger grain ears (Fig. 3), resulting in increased grain yield per plant (Table 1). Since all the hybrid lines were grown under natural condition (no irrigation), the grain yield increase in 2015 was lower (with an up to 13.9% increase) than that in 2016 (with an up to 28.6% increase) due to the local drought weather in 2015 (Table 2). Similar to transgenic plants expressing AtGIF1, ZmGIF1 or OsGIF1, the higher grain yields in them were resulted from both increased seed number and weight per ear (Table 1). This is consistent with the observation in transgenic maize plants (Li et al. 2013).

Taken together, we reported here a new strategy for the breeding of non-transgenic high grain yield maize plants



Fig. 3 Corn cobs of selected hybrid F1 lines from the field trial of Dehui (Jilin Province, China) in 2015. Zheng Dan 958 (ZD958) and a local main cultivar Ping An 24 (PA24) were included as controls

Table 1 Mean comparisons for cob and grain yield of Zheng Dan 958 (ZD958) and different hybrid F1 lines

Year Line	2015				2016			
	Ear length (cm)	Grain row per ear	Grain number per ear	Grain weight per ear (Kg)	Ear length (cm)	Grain row per ear	Grain number per ear	Grain weight per ear (Kg)
1	$17.34 \pm 0.20*$	18.2±0.3**	37.5±0.4**	$0.254 \pm 0.005 **$	18.24±0.21**	16.2 ± 0.3	39.9 ± 0.5	0.305±0.017**
3	$18.11 \pm 0.22^{**}$	$17.0 \pm 0.4*$	$40.3 \pm 0.5 **$	$0.265 \pm 0.005^{**}$	$18.04 \pm 0.18^{**}$	15.3 ± 0.4	$42.9 \pm 0.6^{**}$	$0.277 \pm 0.004^{**}$
17	$18.29 \pm 0.20^{**}$	$17.8 \pm 0.4^{**}$	$38.0 \pm 0.5^{**}$	$0.261 \pm 0.004^{**}$	$19.15 \pm 0.17^{**}$	$17.4 \pm 0.3^{**}$	$43.3 \pm 0.5^{**}$	$0.289 \pm 0.005^{**}$
28	$20.40 \pm 0.22^{**}$	15.7 ± 0.3	$41.3 \pm 0.7^{**}$	$0.256 \pm 0.005^{**}$	$19.47 \pm 0.18^{**}$	15.1 ± 0.3	$42.1 \pm 0.5^{**}$	$0.259 \pm 0.002^{**}$
40	$18.02 \pm 0.24^{**}$	$17.0 \pm 0.3*$	$40.2 \pm 0.5^{**}$	$0.261 \pm 0.005^{**}$	$18.3 \pm 0.18^{**}$	16.6 ± 0.4	$41.8 \pm 0.5^{**}$	$0.298 \pm 0.004^{**}$
42	$20.69 \pm 0.27^{**}$	16.6 ± 0.4	$43.0 \pm 0.5^{**}$	$0.276 \pm 0.006^{**}$	$18.95 \pm 0.21^{**}$	16.1 ± 0.3	$40.6 \pm 0.6*$	$0.264 \pm 0.004^{**}$
45	$18.75 \pm 0.22^{**}$	$17.2 \pm 0.4^{**}$	$42.4 \pm 0.7 **$	$0.254 \pm 0.004^{**}$	$19.71 \pm 0.16^{**}$	$17.0 \pm 0.2^{**}$	$46.0 \pm 0.4^{**}$	$0.284 \pm 0.003^{**}$
48	$19.24 \pm 0.22^{**}$	$18.1 \pm 0.4^{**}$	$37.3 \pm 0.6^{**}$	$0.255 \pm 0.004^{**}$	$20.78 \pm 0.21^{**}$	$18.1 \pm 0.3^{**}$	$43.7 \pm 0.5^{**}$	$0.292 \pm 0.005^{**}$
55	$19.05 \pm 0.23^{**}$	$18.1 \pm 0.3^{**}$	$42.3 \pm 0.6^{**}$	$0.274 \pm 0.005^{**}$	$19.66 \pm 0.15^{**}$	$17.1 \pm 0.2^{**}$	$43.8 \pm 0.3^{**}$	$0.290 \pm 0.004^{**}$
60	$20.66 \pm 0.25^{**}$	$18.1 \pm 0.4^{**}$	$41.9 \pm 0.6^{**}$	$0.281 \pm 0.006^{**}$	$20.04 \pm 0.20^{**}$	$17.1 \pm 0.4^{**}$	$41.0 \pm 0.5^{**}$	$0.266 \pm 0.003^{**}$
93	$18.53 \pm 0.23^{**}$	15.7 ± 0.3	$39.5 \pm 0.7 **$	$0.240 \pm 0.005 *$	$19.70 \pm 0.19^{**}$	15.7 ± 0.4	$45.3 \pm 0.5^{**}$	$0.270 \pm 0.004^{**}$
153	$21.83 \pm 0.39^{**}$	16.7 ± 0.4	$47.6 \pm 1.0^{**}$	$0.246 \pm 0.007^{**}$	$25.81 \pm 0.88^{**}$	$17.5 \pm 0.3^{**}$	$53.1 \pm 0.6^{**}$	$0.308 \pm 0.006^{**}$
ZD 958	16.69 ± 0.19	15.9 ± 0.3	34.7 ± 0.5	0.223 ± 0.004	16.75 ± 0.15	15.7 ± 0.3	39.0 ± 0.4	0.237 ± 0.003

Data are from plants in randomized complete block design with three replications grown under natural condition in Dehui (Jilin Province, China) in 2015 and 2016. Seeds of ZD958 and hybrid F1 lines were sown in a population density of approximately 5500/ μ . Values shown are means \pm SEM. Values of hybrid F1 lines significantly different from those of ZD958 at *0.01 < P < 0.05 and **P < 0.01 were indicated using the *t* test (n=20)

Table 2Mean comparisons forcob and grain yield of ZhengDan 958 (ZD958), Ping An 24(PA24), and different hybridF1 lines

Line	Ear length (cm)	Grain row per ear	Grain number per ear	Grain weight per ear (Kg)
1	$17.34 \pm 0.20*$	18.2±0.3**	$37.5 \pm 0.4 **$	$0.254 \pm 0.005 **$
3	$18.11 \pm 0.22^{**}$	$17.0 \pm 0.4*$	$40.3 \pm 0.5^{**}$	$0.265 \pm 0.005^{**}$
15	$21.00 \pm 0.17 ^{**}$	16.3 ± 0.3	$42.1 \pm 0.6^{**}$	$0.285 \pm 0.004 **$
17	$18.29 \pm 0.20 **$	$17.8 \pm 0.4^{**}$	$38.0 \pm 0.5^{**}$	$0.261 \pm 0.004 **$
22	$18.72 \pm 0.20 **$	16.6 ± 0.3	35.2 ± 0.5	0.231 ± 0.002
23	$19.74 \pm 0.31^{**}$	$17.4 \pm 0.3^{**}$	$38.1 \pm 0.8^{**}$	$0.265 \pm 0.006^{**}$
24	$19.89 \pm 0.24 **$	$16.9 \pm 0.3^*$	$40.8 \pm 0.6^{**}$	$0.252 \pm 0.004 **$
28	$20.40 \pm 0.22^{**}$	15.7 ± 0.3	$41.3 \pm 0.7^{**}$	$0.256 \pm 0.005 **$
40	$18.02 \pm 0.24 **$	$17.0 \pm 0.3^*$	$40.2 \pm 0.5^{**}$	$0.261 \pm 0.005^{**}$
42	$20.69 \pm 0.27^{**}$	16.6 ± 0.4	$43.0 \pm 0.5^{**}$	$0.276 \pm 0.006^{**}$
45	$18.75 \pm 0.22^{**}$	$17.2 \pm 0.4^{**}$	$42.4 \pm 0.7^{**}$	$0.254 \pm 0.004 **$
48	$19.24 \pm 0.22^{**}$	$18.1 \pm 0.4^{**}$	$37.3 \pm 0.6^{**}$	$0.255 \pm 0.004 **$
49	$19.82 \pm 0.21 **$	$17.5 \pm 0.2^{**}$	$40.2 \pm 0.6^{**}$	$0.245 \pm 0.004 **$
53	$18.69 \pm 0.24 **$	$17.6 \pm 0.3 **$	$39.0 \pm 0.6^{**}$	$0.254 \pm 0.006 **$
55	$19.05 \pm 0.23^{**}$	$18.1 \pm 0.3^{**}$	$42.3 \pm 0.6^{**}$	$0.274 \pm 0.005 **$
60	$20.66 \pm 0.25^{**}$	$18.1 \pm 0.4^{**}$	$41.9 \pm 0.6^{**}$	$0.281 \pm 0.006^{**}$
93	$18.53 \pm 0.23 **$	15.7 ± 0.3	$39.5 \pm 0.7 **$	$0.240 \pm 0.005*$
109	$19.02 \pm 0.25^{**}$	16.4 ± 0.4	35.9 ± 0.4	$0.242 \pm 0.0004^{**}$
118	19.13±0.39**	15.9 ± 0.3	34.3 ± 0.9	0.211 ± 0.006
137	$18.92 \pm 0.16^{**}$	16.0 ± 0.2	$37.8 \pm 0.4^{**}$	0.231 ± 0.003
153	$21.83 \pm 0.39^{**}$	16.7 ± 0.4	$47.6 \pm 1.0^{**}$	$0.246 \pm 0.007 **$
ZD958	16.69 ± 0.19	15.9 ± 0.3	34.7 ± 0.5	0.223 ± 0.004
PN24	$18.94 \pm 0.22^{**}$	16.0	$38.4 \pm 0.6^{**}$	0.233 ± 0.004

Data are from plants in randomized complete block design with three replications grown under natural condition in Dehui (Jilin Province, China) in 2015. Seeds of ZD958, PA24 and different hybrid F1 lines were sown in a population density of approximately $5500/\mu$. Values shown are means \pm SEM. Values of PA24 and hybrid F1 lines significantly different from that of ZD958 at *0.01 < P < 0.05 and **P < 0.01 were indicated using the *t* test(n=20). PA24 is a local main cultivar to be used as an internal control

(US Patent No.9139840 authorized on November 4, 2015). The cell wall invertase gene ZmGIF1 can be used as a selection marker, and its expression can be combined between different parental cultivars to increase the invertase activity, and as a result, increase the grain yield of their hybrid progenies.

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Compliance with ethical standards

Conflict of interest We declare that we do not have any commercial or associative interest that represents a conflict of interest in connection with the work submitted.

References

- Albacete A, Cantero-Navarro E, Balibrea ME et al (2014) Hormonal and metabolic regulation of tomato fruit sink activity and yield under salinity. J Exp Bot 65:6081–6095
- Canam T, Unda F, Mansfield SD (2008) Heterologous expression and functional characterization of two hybrid poplar cell-wall invertases. Planta 228:1011–1019
- Chandra A, Verma PK, Islam MN et al (2015) Expression analysis of genes associated with sucrose accumulation in sugarcane (Saccharum spp. hybrids) varieties differing in content and time of peak sucrose storage. Plant Biol (Stuttg) 17:608–617
- Cheng WH, Taliercio EW, Chourey PS (1996) The *miniature1* seed locus of maize encodes a cell wall invertase required for normal development of endosperm and maternal cells in the pedicel. Plant Cell 8:971–983
- Chourey PS, Nelson O (1976) The enzymatic deficiency conditionged by the *shrunken-1* mutation in maize. Biochem Genet 14:1041–1055

- Chourey PS, Jain M, Li QB et al (2006) Genetic control of cell wall invertases in developing endosperm of maize. Planta 223:159-167
- Chourey PS, Li QB, Cevallos-Cevallosc J (2012) Pleiotropy and its dissection through a metabolic gene *Miniature1* (*Mn1*) that encodes a cell wall invertase in developing seeds of maize. Plant Sci 184:45–53
- Eriksson ME, Israelsson M, Olsson O et al (2000) Increased gibberellin biosynthesis in transgenic trees promotes growth, biomass production and xylem fiber length. Nat Biotechnol 18:784–788
- Hampp R, Egger B, Effenberger S et al (1994) Carbon allocation in developing spruce needles-enzymes and intermediates of sucrose metabolism. Physiol Plant 90:299–306
- Hannah LC, Futch B, Bing J et al (2012) A *shrunken-2* transgene increases maize yield by acting in maternal tissues to increase the frequency of seed development. Plant Cell 24:2352–2363
- Jeong JS, Kim YS, Redillas MC et al (2013) OsNAC5 overexpression enlarges root diameter in rice plants leading to enhanced drought tolerance and increased grain yield in the field. Plant Biotechnol J 11(1):101–114
- Kim JY, Mahé A, Guy S et al (2000) Characterization of two members of the maize gene family, Incw3 and Incw4, encoding cellwall invertases. Gene 245:89–102
- Li N, Zhang S, Zhao Y et al (2011) Over-expression of AGPase genes enhances seed weight and starch content in transgenic maize. Planta 233:241–250
- Li B, Liu H, Zhang Y et al (2013) Constitutive expression of cellwall invertase genes increase grain yield and starch content in maize. Plant Biotechnol J 11:1080–1091
- Lowe J, Nelson JR (1946) Miniature seed: a study in the development of a defective caryopsis in maize. Genetics 31:525–533
- Munns R, James RA, Xu B (2012) Wheat grain yield on saline soils is improved by an ancestral Na⁺ transporter gene. Nat Biotechnol 30:360–364
- Smidansky ED, Clancy M, Meyer FD et al (2002) Enhanced ADP glucose pyrophosphorylase activity in wheat endosperm increases seed yield. Proc Natl Acad Sci USA 99:1724–1729
- Smidansky ED, Martin JM, Hannah LC et al (2003) Seed yield and plant biomass increases in rice are conferred by deregulation of endosperm ADP-glucose pyrophosphorylase. Planta 216:656–664
- Sonnewald U, Hajirezaei MR, Kossmann J et al (1997) Increased potato tuber size resulting from apoplastic expression of a yeast invertase. Nat Biotechnol 15:794–797
- Stark DM, Timmerman KP, Barry GF et al (1992) Regulation of the amount of starch in plant tissues by ADP glucose pyrophosphorylase. Science 258:287–292
- Taliercio EW, Kim JY, Mahé A et al (1999) Isolation, characterization and expression analyses of two cell wall invertase genes in maize. J Plant Physiol 155:197–204
- Wang ZY, Chen ZP, Wang JH et al (2007) Increasing maize seed weight by enhancing the cytoplasmic ADP-glucose pyrophosphorylase activity in transgenic plants. Plant Cell Tissue Organ Cult 88:83–92
- Wang ET, Wang JJ, Zhu XD et al (2008a) Control of rice grain-filling and yield by a gene with a potential signature of domestication. Nat Genet 40:1370–1374
- Wang YQ, Wei XL, Xu HL et al (2008b) Cell-wall invertases from rice are differentially expressed in Caryopsis during the grain filling stage. J Integr Plant Biol 50:466–474
- Wu CY, Trieu A, Radhakrishnan P et al (2008) Brassinosteroids regulate grain filling in rice. Plant Cell 20:2130–2145
- Yao Y, Geng MT, Wu XH et al (2014) Genome-wide identification, 3D modeling, expression and enzymatic activity analysis of cell wall invertase gene family from cassava (*Manihot esculenta* Crantz). Int J Mol Sci 15:7313–7331

Zrenner R, Salanoubat M, Willmitaer L et al (1995) Evidence of the crucial role of sucrose synthase for sink strength using transgenic potato plants (*Solanum tuberosum* L.). Plant J 7:97–107