ORIGINAL PAPER

Effects of elevated CO₂ and water stress on physiological responses of *Perilla frutescens* var. *japonica* HARA

Seong Han Lee · Su Young Woo · Sun Mi Je

Received: 13 June 2014/Accepted: 27 November 2014/Published online: 11 December 2014 © Springer Science+Business Media Dordrecht 2014

Abstract The aim of the present study was to investigate the interactive effects of expected environmental constraints, specifically elevated CO₂ and drought conditions, on the physiological responses of Perilla frutescens var. japonica Arum. Perilla frutescens var. japonica 'Arum' was exposed to 700 μ mol mol⁻¹ of CO₂ under both wellwatered and water-stressed conditions. Photosynthetic rate was higher under elevated CO₂ conditions. Stomatal resistance increased while transpiration rates declined, which suggests that water-use efficiency rose under elevated CO₂ conditions. Under water-stressed conditions, elevated CO₂ concentrations induced much higher stomatal resistance than ambient CO₂ levels. This result implies that elevated CO₂ concentrations might increase plant sensitivity to water stress, thereby providing plants with increased protection against drought. Furthermore, elevated CO₂ concentrations alleviated drought-induced photosynthetic decline in the early stages of drought, although this effect was not sustainable. Water stress also elevated ascorbate peroxidase (APX) activity but had little effect on glutathione reductase activity. Water stress-induced APX activity was much lower under elevated CO₂ conditions compared to ambient CO₂ levels. Therefore, elevated CO₂ concentrations might increase plant resistance to water stress. Moreover, there is a possibility that increased ethylene evolution under elevated CO₂ conditions could

S. H. Lee (🖂)

ASEAN-Korea Forest Cooperation Secretariat, Seoul 150-874, Korea e-mail: earlymay1004@yahoo.com

S. Y. Woo · S. M. Je Department of Environmental Horticulture, University of Seoul, Seoul 130-743, Korea improve the capacity of plants to scavenge reactive oxygen species by enhancing APX activity.

Keywords Elevated $CO_2 \cdot Water stress \cdot Ethylene \cdot Antioxidative enzymes \cdot Stomatal resistance$

Introduction

Many concerns have arisen concerning the potential impact of climatic change induced by increasing greenhouse gas concentrations and rising annual temperatures on the ecological performance of both natural and artificial eco-systems (such as agriculture). Atmospheric CO₂ concentration has an indirect effect on plants through induction of climatic change as well as a direct effect on plant growth and physiology (Bettinger et al. 2013; Luong et al. 2013). It is commonly expected that the greenhouse effect will result in the modification of precipitation regimes. In fact, changing patterns of precipitation such as increased duration of drought periods in mid-summer as well as concentrated rainfall in autumn have been observed, especially in middle latitude regions (Ruiz-Sánchez et al. 2007). These shifts in precipitation patterns might increase the seasonal and local frequencies of drought stress in plants.

Water availability from nearby surroundings is one of the most important factors in terms of plant productivity and survival, especially in natural eco-systems. Innes (1992) reported that the most important factor affecting the growth of European beech is water availability from soil. Dry weather in 1976 and between 1983 and 1984 is regarded to be the major reason for the decline of the European beech (Ling et al. 1993). As present climatic conditions will assumedly be maintained in the future, it is important to investigate the interactive effects between high atmospheric CO_2

concentrations and drought stress on plant eco-systems, including agriculture. There have been many reports demonstrating the positive effects of elevated CO₂ concentrations on plant productivity mediated by improved energy efficiency caused by increased C assimilation rates (Boese et al. 1997; Reddy et al. 2010; Thomas et al. 1993) as well as reduced photorespiration (Havir and McHale 1989; Robredo et al. 2010; Roden and Ball 1996). It has also been reported that stomatal conductivity decreases under elevated CO₂ conditions (Ainsworth and Rogers 2007; Zhu et al. 1998), suggesting that water-use efficiency (WUE) of plants can be increased (Bunce 2004; Malmström and Field 1997; Oliveira et al. 2013). Therefore, among the factors influencing drought stress in plants, elevation of atmospheric CO₂ is considered to be one of the most important factors. Although elevation of atmospheric CO₂ concentrations may potentially increase drought stress in forest eco-systems by increasing total leaf area, it is generally assumed that photosynthetic limitations caused by drought are alleviated under high atmospheric CO₂ conditions, and leaf water potential under drought conditions can be maintained by increased water use efficiency under high atmospheric CO₂ levels (Ge et al. 2011; Oliveira et al. 2013; Roger et al. 1984).

On the other hand, an elevated atmospheric CO_2/O_2 ratio can reduce the rate of photorespiration, thereby decreasing generation of internal reactive oxygen species (ROS). This suggests that the capacity of plants to detoxify ROS might be improved under elevated atmospheric CO₂ conditions (Polle et al. 1993, 1997; Schwanz and Polle 2001). Both CO₂ and air pollutants such as SO₂ and NO₂ are byproducts of fossil fuel combustion, and ROS are regarded as a major substance inducing plant damage under air pollution or drought stress. On that point, the effect of CO_2 on plant oxidative stress is no less important than its effect on plant productivity under changing climatic conditions. Thus, the aim of the present study was to investigate the interactive effects of elevated CO₂ concentration and drought on the physiological responses of plant using Perilla frutescens var. japonica 'Arum' which has been known as an indicator species of environmental fluctuations having an important economic value in many of East Asian countries (Hur et al. 1995; Park et al. 2001; Kim et al. 2013).

Perilla frutescens var. japonica 'Arum' was grown from seeds in plastic pots (top diameter 15 cm) containing a

mixture of 1:1(v/v) of peat (Klasman, Potground-H, Ger-

many): perlite for 60 days in a greenhouse maintained at

 27 ± 0.5 °C during the day and 20 ± 0.5 °C at night. The

Materials and methods

Plant culture

Macroelement (meL ⁻¹)								
N	Р	К	Ca	Mg	S			
11.7	2.0	6.7	3.5	2.0	2.0			
Microel	ement (ppm)							
Fe	Mn	В	Zn	Cu	Мо			
2.0	0.3	0.5	0.3	0.1	0.05			

pots were flushed once a day and fertilized twice a week with a nutrient solution developed for leafy vegetables by the University of Seoul (UOS) (Table 1).

CO₂ treatment and water regime

Plants were placed in a controlled environment chamber maintained at 27 ± 0.5 °C during the day, 20 ± 0.5 °C at night, 50 ± 5 % relative humidity, and $250 \pm 30 \ \mu\text{mol} \ \text{m}^{-2} \ \text{s}^{-1}$ light intensity. After acclimation for a week, plants were treated with either ambient or elevated CO₂ concentrations under both well-watered and drought-stressed conditions. CO₂ concentrations of 350 \pm 20 µmol mol⁻¹ (ambient) and $680 \pm 30 \text{ }\mu\text{mol mol}^{-1}$ (elevated) were used. The elevated CO₂ concentration was maintained for 8 h per day for 1 week and was monitored by a computer-controlled NDIR (nondispersive infrared) CO₂ analyzer (Mapo Technomax Ltd. Korea) and injection system (Genie, USA). Two watering regimes, drought-stressed and well-watered, were imposed under both CO₂ concentrations by withholding water from the beginning of CO₂ treatment for the drought-stressed regime and watering once a day for the well-watered regime. Plant water status was monitored by measuring the relative water content (RWC) of leaves.

Gas exchange measurement

All photosynthetic parameters of plants were measured without removing the plants from growth chambers to ensure that conditions during measurement did not differ from experimental conditions. Measurements were taken between 11:00 and 14:00 h. Leaf photosynthetic rate, CO_2 absorption rate, and stomatal resistance were measured using a Li-6200 (Li-Cor, USA), and transpiration rate was measured using a porometer (Li-1600, Li-Cor, USA) on fully expanded leaves.

Ethylene evolution measurement

The amount of ethylene evolution was determined according to the methods by Mathooko et al. (1998). Samesized leaf discs were collected using a cork borer and enclosed in airtight vials containing wet filter paper to prevent leaf desiccation. Following incubation at 25 °C for 2 h in the dark, 1 mL of headspace was withdrawn. Ethylene concentration in the gas sample was measured using a gas chromatograph (model GC-14B, Shimadzu, Japan) equipped with a flame-ionization detector and an activated alumina column (110 × 0.32 cm). A standard curve was used for quantification of ethylene, and total ethylene evolution was normalized to the fresh weight of leaves.

Antioxidative enzyme assays

The activities of two antioxidative enzymes, ascorbate peroxidase (APX) and glutathione reductase (GR), were measured according to the methods by Asada (1992) and Kondo and Saji (1992) respectively. The overall procedure was carried out at 0-4 °C. Randomly sampled mature leaf tissues were ground in a chilled mortar using specific buffers and pH values for each enzyme. To measure APX activity, leaf tissue samples (0.2 g) were ground with 2 mL of ice-cold extraction buffer (1 M AsA, 100 mM K-P buffer pH 7.4). The homogenate was centrifuged at 4 °C for 3 min at 12,000 rpm. The supernatant (60 μ L) was added to a reaction mixture containing 2.34 mL of H₂O, 300 µL of 1 MK-P buffer (pH 6.5), 180 µL of 10 mM AsA, and 120 µL of 5 mM H₂O₂. Enzyme activity was calculated based on the decrease in A₂₉₀ of the mixture over a 30 s interval, as detected by a UV spectrophotometer (UV2100, Shimadzu, Japan). To measure GR activity, leaf tissue samples (0.15 g) were ground with 2 mL of cold extraction buffer containing 50 mM K-P buffer (pH 7.0), 5 % PVP, 5 mM AsA, 5 mM DTT, 5 mM EDTA, and 0.1 M NaCl. The homogenate was centrifuged at 4 °C for 5 min at 15,000 rpm, after which the supernatant (150 µL) was added to a reaction mixture containing 2.43 mL of H₂O, 300 µL of 1 MK-P buffer (pH 7.8), 60 µL of 10 mM GSSG, and 60 µL of 10 mM NADPH. Enzyme activity was calculated based on the decrease in A₃₄₀ of the mixture over a 90 s interval. The protein content was quantified following the methods by Bradford (1976).

Statistical analysis

Statistical analysis was carried out on each measurement with SAS (9.13). Data was submitted to Duncan's multiple range test and the significance of differences between treatments was considered at the $P \le 0.05$ level.

Results and discussion

Effect of high CO₂ on photosynthetic parameters

Single treatments of elevated CO_2 under well-watered conditions increased the photosynthetic rate of *P*.



Fig. 1 Effect of elevated CO₂ on photosynthetic rate in *P. frutescens* var. *japonica* 'Arum' under both well-watered and drought-stressed conditions. *Each symbol* represents the mean of 10 observations \pm SE (*CW* well-watered control at ambient CO₂ level, *CD* drought-stressed at ambient CO₂ level, *TW* well-watered at 700 µmol mol⁻¹ CO₂ level, *TD* drought-stressed at 700 µmol mol⁻¹ CO₂ level)

frutescens var. japonica 'Arum' by 39 and 30 % on the 2nd and 5th days of treatment, respectively (Fig. 1). As elevation of atmospheric CO₂ generally shifts the activity of RubisCO in favor of carboxylation in most C₃ species, net photosynthesis initially increases in response to increased CO₂ levels (Bowes 1991; Robredo et al. 2010; Stitt 1991). Along with an increase in photosynthetic rate, the stomatal response could be affected by elevated CO₂ concentrations (Ainsworth and Rogers 2007; Berryman et al. 1994; Kellomäki and Wang 1998; Robredo et al. 2010). Stomatal resistance of P. frutescens var. japonica 'Arum' increased by 29 and 19 % on the 2nd and 5th days of treatment, respectively (Fig. 2). WUE of a plant is the ratio of carbon gain to water loss. Thus, increased stomatal resistance under elevated CO₂ conditions might increase WUE (Gunderson et al. 1993; Idso and Idso 1994; Malmström



Fig. 2 Effect of elevated CO₂ on stomatal resistance in *P. frutescens* var. *japonica* 'Arum' under both well-watered and drought-stressed conditions. ^zMean separation (n = 10) within treatments by Duncan's multiple range test at $P \le 0.05$ (*CW* well-watered control at ambient CO₂ level, *CD* drought-stressed at ambient CO₂ level, *TW* well-watered at 700 µmol mol⁻¹ CO₂ level, *TD* drought-stressed at 700 µmol mol⁻¹ CO₂ level)



Fig. 3 Effect of elevated CO₂ on transpiration rate in *P. frutescens* var. *japonica* 'Arum' under both well-watered and drought-stressed conditions. *Each symbol* represents the mean of 10 observations \pm SE (*CW* well-watered control at ambient CO₂ level, *CD* drought-stressed at ambient CO₂ level, *TW* well-watered at 700 µmol mol⁻¹ CO₂ level, *TD* drought-stressed at 700 µmol mol⁻¹ CO₂ level)

and Field 1997) as the result of a reduced transpiration rate. Transpiration rates in well-watered plants under elevated CO_2 conditions decreased significantly during treatment (Fig. 3). This result shows that the WUE of *P. frutescens* var. *japonica* 'Arum' might be improved by increased net photosynthesis and a decreased transpiration rate, suggesting resistance to drought might be increased. Robredo et al. (2007) and Rogers et al. (1984) also reported reduction of the transpiration rate due to high atmospheric CO_2 levels even under drought conditions as well as delayed onset of irreversible water stress.

Interactive effect of high CO₂ and drought on photosynthetic parameters

Leaf RWC was reduced by only 3 % during the 7 days of drought period (data not shown), which suggests that *P. frutescens* var. *japonica* 'Arum' is relatively resistant to drought. However, there were significant differences in the physiological responses, including photosynthetic parameters. Photosynthetic rate decreased by 15 and 31 % on the 2nd and 5th days of drought treatment, respectively, under ambient CO₂ levels (CD) (Fig. 1). The photosynthetic rate on the 2nd day of treatment in plants exposed to both

drought and elevated CO₂ levels (TD) was higher than that in drought-stressed plants under ambient CO₂ conditions (Fig. 1). As drought stress progressed, the photosynthetic rate decreased by 23 % by day 5 even under elevated CO₂ conditions. This rate was similar to that in plants subjected to drought treatment under ambient CO₂ levels. That is, elevated CO₂ concentrations alleviated drought-induced reduction of photosynthesis in the early stages of drought, although this effect was not sustainable. Drought treatment under ambient CO₂ conditions increased stomatal resistance by about 36 % by the 5th day of treatment (Fig. 2). It is known that both stomatal closing induced by the ABAmediated signal transduction pathway (Wilkinson et al. 1998) as well as decreased ATP-synthase activity (Tezara et al. 1999, 2008) are involved in reduction of photosynthesis under drought conditions. In plants subjected to combined elevated CO₂ and drought treatment, stomatal resistance was 64 % higher by day 2 and twofold higher by day 5 compared to plants subjected to ambient CO₂ and drought treatment (Fig. 2). This result suggests that elevated atmospheric CO₂ concentrations increase the sensitivity of stomata to drought stress, which helps plants respond to drought stress more effectively. Rogers et al. (1984) also reported that elevation of stomatal resistance with decreasing leaf water potential in soybeans is exacerbated under elevated CO₂ conditions. Similar results have been observed in tropical tree species (Berryman et al. 1994; Heath 1998). The CO_2 absorption rate was higher in plants exposed to both drought and elevated CO₂ than in drought-stressed plants under ambient CO₂ conditions, although stomatal resistance was much higher in plants subjected to combined drought and elevated CO₂ treatment (Table 2). Therefore, the high photosynthetic rates under drought and elevated CO₂ conditions in the early stages of drought can be attributed to increased carbon fixation capacity resulting from higher WUE and CO₂ absorption rates in response to elevated CO₂ concentrations. Malmström and Field (1997) reported that increased WUE resulting from reduced transpiration rates in response to elevated CO₂ concentrations results in higher whole-plant carbon gain. However, after 5 days of drought treatment, the CO_2 absorption rate under high CO_2 conditions had

Table 2 Effect of elevated CO₂ concentrations on CO₂ absorption rate in *P. frutescens* var. *japonica* 'Arum' under well-watered and drought-stressed conditions

CO_2 level (µmol mol ⁻¹)	CO_2 absorption rate (µg cm ⁻² min ⁻¹)				
	3rd day		7th day		
	Well-watered	Drought	Well-watered	Drought	
Ambient	$1.05\pm0.14^{\rm z}$	0.87 ± 0.15	0.84 ± 0.09	0.52 ± 0.12	
700	1.38 ± 0.23	1.10 ± 0.23	1.06 ± 0.13	0.49 ± 0.12	

^z Means \pm SE of result from 10 observations are shown for each treatment

decreased to the same rate as that seen under ambient CO_2 levels (Table 2) along with reduction of the photosynthetic rate. Tezara et al. (1999, 2008) reported that drought stress decreases leaf ATP and RuBP contents as a result of reduced ATP-synthase activity. This photosynthetic limitation is not compensated for by elevation of the atmospheric CO_2 concentration. This suggests that expected improvement of plant productivity via increased atmospheric CO_2 concentrations might be limited by shifts in precipitation patterns induced by the greenhouse effect.

Effect of high CO₂ on evolution of secondary toxic substances

ROS are some of the most important secondary toxic substances inducing oxidative damage to plants (such as lipid peroxidation) under environmental stress. Metabolic changes induced by increased atmospheric CO₂ concentrations might affect the activities of antioxidative enzymes and thus plant resistance to environmental stresses (Polle et al. 1997; Schwanz and Polle 2001). Therefore, the effects of high atmospheric CO₂ concentrations on activities of GR and APX, which play a central role in protecting chloroplasts and other cellular components from oxidative damage, were examined. The results show that GR activity was not affected significantly by CO₂ concentration under well-watered conditions during treatment (Fig. 4). APX activity increased in response to high CO₂ concentrations under well-watered conditions, although the difference was not very significant (Fig. 4). After they observed that activities of superoxide dismutase (SOD) and catalase were decreased in the response to high atmospheric CO₂ concentrations, Polle et al. (1997) suggested that the ROS detoxification capacity of plants might be improved under high atmospheric CO₂ concentrations by providing increased amounts of substrate for detoxification and repair processes. However, peroxidase is known to be related to various metabolic processes in plants such as the biosynthesis of hormones and lignification, which suggests its activity might increase in response to metabolic changes related to increasing atmospheric CO₂ concentrations (Polle et al. 1993). Ethylene can also affect the activities of various antioxidative enzymes such as APX (Ievinsh et al. 1995; Lafuente et al. 2004; Mehlhorn 1990), and ethylene biosynthesis can increase under high atmospheric CO₂ concentrations (Mathooko et al. 1998). It was therefore, suspected that APX activity might be increased under high atmospheric CO₂ concentrations. Ethylene evolution in well-watered plants also increased in response to elevated CO₂ concentrations, especially in the early stage of treatment (Table 3). CO_2 can stimulate ACC oxidase activity so that an increase in intercellular CO₂ concentration under high atmospheric CO₂ enhances ethylene biosynthesis in



Fig. 4 Effects of elevated CO₂ on activities of glutathione reductase (GR) and ascorbate peroxidase (APX) in *P. frutescens* var. *japonica* 'Arum' under both well-watered and drought-stressed conditions. ^zMean separation (n = 5) within treatments by Duncan's multiple range test at $P \le 0.05$ (*CW* well-watered control at ambient CO₂ level, *CD* drought-stressed at ambient CO₂ level, *TW* well-watered at 700 µmol mol⁻¹ CO₂ level, *TD* drought-stressed at 700 µmol mol⁻¹ CO₂ level)

Table 3 Effect of elevated CO₂ on ethylene evolution in *P. frutescens* var. *japonica* 'Arum' under well-watered and drought-stressed conditions

CO ₂ level	Ethylene evolution (nL $g^{-1} h^{-1}$)					
(µmol mol ⁻¹)	3rd day		7th day			
	Well-watered	Drought	Well-watered	Drought		
Ambient 700	3.07 b ^z 5.10 a	4.51 ab 4.93 ab	2.35 b 3.82 a	3.87 a 3.79 a		

^z Different letters indicate significant differences in mean separation among treatments at $P \le 0.05$ by Duncan's multiple range test

plants (Mathooko et al. 1998). Mehlhorn (1990) reported that plants pre-exposed to ethylene showed increased resistance to ozone and that this response was related to the ethylene-induced stimulation of APX activity. Therefore, it is suspected that an increase in APX activity under high CO_2 concentration in the present study does not result from an increase in ROS but rather from metabolic changes induced by elevated CO_2 concentration or increased amounts of ethylene evolution. In the latter case, increased APX activity could allow plants to become more resistant to oxidative damage induced by environmental stresses such as air pollution. On the other hand, ethylene is known to be a major secondary toxic substance to induce lipid peroxidation under stress conditions. However, it has also been reported that ACC treatment, which increases ethylene evolution, induces no significant changes in ion leakage as an index of membrane damage by lipid peroxidation. This suggests that the actions of ethylene may vary depending on the cause of induction. Thus, the effect of increased ethylene evolution due to elevated atmospheric CO_2 levels on plant responses to environmental stress remains unclear.

Interactive effect of high CO₂ and drought on evolution of secondary toxic substances

Drought stress induces damage to the electron transport system in thylakoid membranes, resulting in increased superoxide production (Kitao et al. 2003; Price and Hendry 1991). The rates of ROS production induced by drought stress under both ambient and elevated CO₂ levels were measured in this experiment based on changes in GR and APX activities. The activity of GR in plants exposed to drought under ambient CO₂ levels showed no significant increase during treatment, whereas APX activity increased by 50 % by day 7 of drought treatment (Fig. 4). The antioxidant defense system of plants consists of a variety of antioxidant enzymes but their activities are not all increased under stress conditions that lead to the formation of ROS in plants. For example, GR activity in response to salt or drought stress might not increase until the stress level becomes severe (Hernandez et al. 1999; Zhang and Kirkham 1996). In our study, drought stress, elevated CO₂ levels, as well as their combined treatment all had no significant effect on GR activity, whereas APX activity in plants exposed to combined drought and elevated CO₂ treatment was much lower compared to plants under drought at ambient CO₂ conditions (Fig. 4). Therefore, elevated CO₂ conditions might allow plants to become more resistant to drought stress with respect to oxidative damage. The activities of antioxidative enzymes in response to drought stress generally increase with decreases in leaf water potential (Hernandez et al. 1999; Zhang and Kirkham 1996). Price and Hendry (1991) reported that reduction of chlorophyll content is strongly correlated to reduction of RWC. Production of ROS in response to drought stress is the major cause of chlorophyll destruction. In the present study, reduction of RWC and chlorophyll content as the result of drought treatment was not significant (data not shown), suggesting P. frutescens var. japonica 'Arum' could respond to drought stress properly by increasing stomatal resistance and ROS scavenging metabolisms. Drought stress also increases ethylene evolution, thereby inducing leaf senescence and retarding the growth rate. Drought treatment increased ethylene evolution under both ambient and elevated CO₂ conditions (Table 3). Under elevated CO_2 conditions, ethylene evolution was higher in well-watered plants than in droughtstressed plants. The effect of elevated CO₂ levels on ethylene evolution might be limited under drought conditions as the result of elevated stomatal resistance. Leaf abscission induced by drought stress imposed by withholding water generally appears after re-watering since the transpiration stream is restricted under drought conditions, which prevents the transfer of ACC, the precursor of ethylene, to the leaf (Nilsen and Orcutt 1996). Therefore, ethylene evolution could immediately increase after plants are released from drought stress. This may be possible under elevated CO₂ conditions, considering that CO₂ may activate ACC oxidase. These phenomena might affect plant responses to environmental stresses stress in relation to the action of ethylene mentioned above. In summary, elevation of atmospheric CO₂ levels increased the photosynthetic rate as well as decreased the transpiration rate, resulting in elevated plant WUE. Under elevated CO₂ conditions, stomata of P. frutescens var. japonica 'Arum' showed much higher sensitivity to drought. This result suggests that plants are afforded increased protection against drought stress under elevated CO₂ conditions. The extent of these responses will be important to determine plant productivity and survival in relation to climatic changes induced by increasing atmospheric CO₂ concentrations. Furthermore, elevated CO₂ concentrations reduced drought-induced oxidative damage and seemed to have an indirect effect on APX activity by enhancing ethylene evolution. Therefore, this effect of elevated CO2 levels on antioxidative defense systems should be considered as an important factor in the present environment in which plant stressors are gradually increasing.

Acknowledgments This research was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (Grant No. NRF-2012R1A1A2005498).

References

- Ainsworth EA, Rogers A (2007) The response of photosynthesis and stomatal conductance to rising [CO₂]: mechanisms and environmental interactions. Plant Cell Environ 30:258–270
- Asada K (1992) Ascorbate peroxidase—a hydrogen peroxide-scavenging enzyme in plants. Physiol Plant 85:235–241
- Berryman CA, Eamus D, Duff GA (1994) Stomatal responses to a range of variable in two tropical tree species grown with CO₂ enrichment. J Exp Bot 45:539–546

- Bettinger P, Siry J, Merry K (2013) Forest management planning technology issues posed by climate change. Forest Sci Tech 9:9–19
- Boese SR, Wolfe DW, Melkonian JJ (1997) Elevated CO₂ mitigates chilling-induced water stress and photosynthetic reduction during chilling. Plant, Cell Environ 20:625–632
- Bowes G (1991) Growth at elevated CO₂: photosynthetic responses mediated through rubisco. Plant, Cell Environ 14:795–806
- Bradford MM (1976) A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem 72:248–254
- Bunce JA (2004) Carbon dioxide effects on stomatal responses to the environment and water use by crops under field conditions. Oecologia 140:1–10
- Ge ZM, Zhou X, Kellomäki S, Wang KY, Peltola H, Martikainen PJ (2011) Responses of leaf photosynthesis, pigments and chlorophyll fluorescence within canopy position in a boreal grass (*Phalaris arundinacea* L.) to elevated temperature and CO₂ under varying water regimes. Photosynthetica 49:172–184
- Gunderson CA, Norby RJ, Wullschleger SD (1993) Foliar gas exchange responses of two deciduous hardwoods during 3 years of growth in elevated CO₂: no loss of photosynthetic enhancement. Plant, Cell Environ 16:797–807
- Havir EA, Mchale NA (1989) Regulation of catalase activity in leaves of *Nicotiana sylvestris* by high CO₂. Plant Physiol 89:952–957
- Heath H (1998) Stomata of trees growing in CO₂-enriched air show reduced sensitivity to vapour pressure deficit and drought. Plant, Cell Environ 21:1077–1088
- Hernandez J, Campillo A, Jimenez A, Alarcon JJ, Sevilla F (1999) Response of antioxidant system and leaf water relations to NaCl stress in pea plants. New Phytol 141:241–251
- Hur JS, Hur YK, Lee CI (1995) Evaluation of SO₂ or O₃ exposure durations requiring foliar damage development by using bioindicating plants. Korean J Plant Pathol 11:107–115
- Idso KE, Idso SB (1994) Plant responses to atmosperic CO₂ enrichment in the face of enironmental constraints: a review of the past 10 years' research. Agric For Meteorol 69:153–203
- Ievinsh G, Valcina A, Ozola D (1995) Induction of ascorbate peroxidase activity in stressed pine (*Pinus Sylvestris* L.) needles—a putative role for ethylene. Plant Sci 112:167–173
- Innes JL (1992) Observations on the condition of beech (Fagus sylvatica L.) in Britain in 1990. Forestry 65:35–60
- Kellomäki S, Wang KY (1998) Sap flow in Scots pines growing under conditions of year-round carbon dioxide enrichment and temperature elevation. Plant, Cell Environ 20:995–1006
- Kim JA, Sa KJ, Choi SH, Lee JK (2013) Morphological variation of cultivated types of Perilla crop and their weedy types in East and Southeast Asia. Korean J Crop Sci 58:408–415
- Kitao M, Lei TT, Koike T, Tobita H, Maruyama Y (2003) Higher electron transport rate observed at low intercellular CO₂ concentration in long-term drought-acclimated leaves of Japanese mountain birch (*Betula ermanii*). Physiol Plant 118:406–413
- Kondo N, Saji H (1992) Tolerance of plants to air pollutants. J Japan Soc Air Pollut 27:273–288
- Lafuente MT, Sala JM, Zacarias L (2004) Active oxygen detoxifying enzymes and phenylalanine ammonia-lyase in the ethyleneinduced chilling tolerance in citrus fruit. J Agric Food Chem 52:3606–3611
- Ling KA, Power SA, Ashmore MR (1993) A survey of the health of Fagus sylvatica in southern Britain. J Appl Ecol 30:295–306
- Luong TH, Jang KS, Lim HW, Choi WJ, Lee KH (2013) Correlation of tree ring growths of four major species with climate changes in South Korea. Forest Sci Tech 9:180–186
- Malmström CM, Field CB (1997) Virus-induced difference in response of oat plants to elevated carbon dioxide. Plant, Cell Environ 20:178–188

- Mathooko FM, Inaba A, Nakamura R (1998) Characterization of carbon dioxide stress induced ethylene biosynthesis in cucumber (*Cucumis sativus* L.) fruit. Plant Cell Physiol 39:285–293
- Mehlhorn H (1990) Ethylene-promoted ascorbate peroxidase activity protects plants against hydrogen peroxide, ozone and paraquat. Plant, Cell Environ 13:971–976
- Nilsen ET, Orcutt DM (1996) Water limitation. In: Nilsen ET, Orcutt DM (eds) The physiology of plants under stress. Wiley, New York, pp 322–361
- Oliveira VF, Silva EA, Zaidan LBP, Carvalho MAM (2013) Effects of elevated CO₂ concentration and water deficit on fructan metabolism in *Viguiera discolor* Baker. Plant Biol 15:471–482
- Park DS, Lee KI, Park KY (2001) Quantitative analysis of dietary fibers from *Perilla frutescens* seeds and antimutagenic effect of its extracts. J Korean Soc Food Sci Nutr 30:900–905
- Polle A, Pfirrmann T, Chakrabarti S, Rennenberg H (1993) The effects of enhanced ozone and enhanced carbon dioxide concentrations in biomass, pigments and antioxidative enzymes in spruce needles (*Picea abies* L.). Plant, Cell Environ 16:311–316
- Polle A, Eiblmeier M, Sheppard L, Murray M (1997) Responses of antioxidative enzymes to elevated CO₂ in leaves of beech (*Fagus* syvatica L.) seedlings grown under a range of nutrient regimes. Plant, Cell Environ 20:1317–1321
- Price AH, Hendry GAF (1991) Iron-catalysed oxygen radical formation and its possible contribution to drought damage in nine native grasses and three cereals. Plant, Cell Environ 14:477–484
- Reddy AR, Rasineni GK, Raghavendra AS (2010) The impact of global elevated CO₂ concentration on photosynthesis and plant productivity. Curr Sci 99:46–57
- Robredo A, Pérez-López U, de la Maza HS, González-Moro B, Lacuesta M, Mena-Petite A, Muñoz-Rueda A (2007) Elevated CO₂ alleviates the impact of drought on barley improving water status by lowering stomatal conductance and delaying its effect on photosynthesis. Environ Exp Bot 59:252–263
- Robredo A, Pérez-López U, Lacuesta M, Mena-Petite A, Muñoz-Rueda A (2010) Influence of water stress on photosynthetic characteristics in barley plants under ambient and elevated CO₂ concentrations. Biol Plant 54:285–292
- Roden JS, Ball MC (1996) The effect of elevated [CO₂] on growth and photosynthesis of two eucalyptus species exposed to high temperatures and water deficits. Plant Physiol 111:909–919
- Rogers HH, Siont N, Cure JD, Smith JM, Bingham GE (1984) Influence of elevated carbon dioxide on water relations of soybeans. Plant Physiol 74:233–238
- Ruiz-Sánchez MC, Domingo R, Pérez-Pastor A (2007) Daily variations in water relations of apricot trees under different irrigation regimes. Biol Plant 51:735–740
- Schwanz P, Polle A (2001) Differential stress responses of antioxidative systems to drought in pendunculate oak (*Quercus robur*) and maritime pine (*Pinus pinaster*) grown under high CO₂ concentrations. J Exp Bot 52:133–143
- Stitt M (1991) Rising CO₂ levels and their potential significance for carbon flow in photosynthetic cells. Plant, Cell Environ 14:741–762
- Tezara W, Mitchell VJ, Driscoll SD, Lawlor DW (1999) Water stress inhibits plant photosynthesis by decreasing coupling factor and ATP. Nature 401:914–917
- Tezara W, Driscoll S, Lawlor DW (2008) Partitioning of photosynthetic electron flow between CO_2 assimilation and O_2 reduction in sunflower plants under water deficit. Photosynthetica 46:127–134
- Thomas RB, Reid CD, Ybema R, Strain BR (1993) Growth and maintenance components of leaf respiration of cotton grown in elevated carbon dioxide partial pressure. Plant, Cell Environ 16:539–546

- Wilkinson S, Corlett JE, Oger L, Davies WJ (1998) Effect of xylem pH on transpiration from wild type and flacca tomato leaves. Plant Physiol 117:703–709
- Zhang J, Kirkham MB (1996) Enzymatic responses of the ascorbateglutathione cycle to drought in sorghum and sunflower plants. Plant Sci 113:139–147
- Zhu J, Talbott LD, Jin X, Zeiger E (1998) The stomatal response to CO₂, is linked to change in guard cell zeaxanthin. Plant, Cell Environ 21:813–820