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Net ecosystem CO₂ exchange and carbon cycling in tropical lowland flooded rice ecosystem

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Abstract The seasonal fluxes of CO_2 and its characteristics with relation to environmental variables were investigated under tropical lowland flooded rice paddies employing the open path eddy covariance technique. The seasonal net ecosystem carbon budget was quantified by empirical modelling approach. The integrated net ecosystem exchange (NEE), gross primary production (GPP) and ecosystem respiration (RE) in the flooded rice field was -448, 811 and 363 g C m⁻² in wet season. Diurnal variations of mean NEE values during the season varied from +3.99 to $-18.50 \text{ }\mu\text{mol CO}_2 \text{ }m^{-2} \text{ s}^{-1}$. The daily average NEE over the cropping season varied from +2.73 to $-7.74 \text{ g C m}^{-2} \text{ day}^{-1}$. The net ecosystem CO₂ exchange reached its maximum in heading to flowering stage of rice with an average value of -5.67 g C m⁻² day⁻¹. On daily basis the flooded rice field acted as a net sink for CO₂ during most of the times in growing season except few days at maturity when it became a net CO_2 source. The rate of CO_2 uptake by rice as observed from negative NEE values increased proportionally with air temperature up to 34 °C. The carbon distribution in different component of soil-plant system namely, soil organic carbon, dissolved organic carbon, methane emission,

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rhizodeposition, carbon in algal biomass, crop harvest and residues were quantified and carbon balance sheet was prepared for the wet season in tropical rice. Carbon balance sheet for tropical rice revealed 7.12 Mg C ha⁻¹ was cycled in the system in wet season.

 $\label{eq:covariance} \begin{array}{l} \mbox{Keywords} & \mbox{Open path eddy covariance} \\ \mbox{Net ecosystem CO}_2 \mbox{ exchange} \\ \mbox{Flooded rice field} \\ \mbox{Rice-ecosystem carbon cycling} \end{array}$

Introduction

Quantification of greenhouse gas exchanges between the ecosystem and the atmosphere is one of the key issues to assess the global budget of greenhouse gases. Rice is the major food crop in Asia and about 80 % of it is grown under flooded conditions (Towprayoon et al. 2005). Rice is grown in different environments ranging from tropical to temperate regions with varying climatic, edaphic and biological conditions and under different agricultural management practices which naturally affect the rates of CH₄ and CO₂ emissions. According to the Food and Agricultural Organization of the United Nations (FAO), the area of rice paddies in Asia is about 87 % of the world's total rice cultivated area (Pakoktom et al. 2009). Rice paddies in tropical low land flooded soil play a crucial

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role in the global budget of greenhouse gases such as CO_2 and CH_4 (IPCC 2007). Many of the factors controlling gas exchanges between rice paddies and the atmosphere are different from dry land agriculture and other ecosystems because rice is flooded during most of its cultivation period. Therefore, field studies to measure net CO_2 fluxes from flooded soil and to improve the understanding of the factors controlling the fluxes are important.

The eddy covariance (EC) technique is widely employed as the standard micrometeorological method to monitor fluxes of CO₂, water vapour and heat, which are bases to determine CO₂ and heat balances of land surfaces (Aubinet et al. 2000). Longterm observations of CO₂ flux have been carried out in various ecosystems in the world. These observation sites are mostly located in forest ecosystems (Saigusa et al. 2002; Carrara et al. 2003). On the other hand, some studies in non-forest ecosystems, viz. grasslands, wetlands and agricultural fields have also been observed because of their contribution to regional and global CO₂ budgets (Saito et al. 2005; Tsai et al. 2006). In Asia, EC flux measurements were conducted in Japan (Miyata et al. 2000, 2005; Saito et al. 2005), Thailand (Pakoktom et al. 2009), China (XiuE et al. 2007; Hossen et al. 2010), South Korea (Moon et al. 2003), Bangladesh (Hossen et al. 2007; Hossen et al. 2011), Philippines (Alberto et al. 2009) and Taiwan (Tseng et al. 2010) to monitor seasonal, annual and inter-annual variations in CO₂ fluxes in irrigated and aerobic rice fields. Most of the above studies were limited to the temporal variation of CO₂ exchange in relation to meteorological parameters and crop harvest. But understanding the processes and components of net ecosystem carbon budget (NECB) in lowland flooded rice paddy is essential to know whether the system is behaving as net C sink (net carbon accumulation in the system) or C source (net carbon depletion or loss from the system). Agricultural activities involving the addition of C inputs in the form of organic manures and other associated phenomena (rhizodeposition, aquatic biomass, CO₂-C fixation in the form of NEP, decayed roots and stubbles left in the field) and C output through crop harvest and gaseous C emission (CH_4 and CO_2). It is very important to quantify the balance between these two processes in order to get a clear insight of C cycling under lowland flooded rice paddy ecosystem for maintaining ecosystem health and sustainability.

To address these issues, the present study was undertaken to understand the season-long net ecosystem exchange (NEE), gross primary production (GPP) and ecosystem respiration (RE) and its characteristics in relation to environmental variables and to quantify the carbon budget of the ecosystem by considering the possible inflow and outflow of carbon in tropical lowland flooded rice paddies.

Materials and methods

Site description

The study was conducted at the experimental farm of the Central Rice Research Institute (CRRI), Cuttack, India ($85^{\circ}56'25''$ E, 20°27'6'' N, and 24 m above mean sea level). Mean annual highest and lowest temperatures were 39.2 and 22.5 °C, respectively and mean annual temperature is 27.7 °C. Average annual rainfall is 1,500 mm, of which 75–80 % is received during June to September. The difference between mean summer and mean winter soil temperature is more than 5 °C, thus qualifying as a hypothermic temperature class. The soil is an Aeric Endoaquept with sandy clay loam texture (25.9 % clay, 21.6 % slit, 52.5 % sand), bulk density 1.41 Mg m⁻³, pH (1:2.5 soil: solution ratio) 6.2, electrical conductivity 0.40 dSm⁻¹, total carbon (TC) 1.12 % and total nitrogen (TN) 0.08 %.

Crop establishment

The study was conducted during wet season (July to October) in rice-fallow-rice cropping sequence. Flooded rice field (2.25 ha (150 m \times 150 m)) was uniformly planted with low land rice variety. Thirty days old seedlings were transplanted on 8th July, 2010 with a spacing of $15 \text{ cm} \times 15 \text{ cm}$. The field was remained flooded with 12 ± 3 cm water depth throughout the growing season up to 2 weeks before harvest. Three split doses of N (40–20–20 kg ha^{-1}) fertilizer were applied at basal, maximum tillering (MT) and panicle initiation (PI) stages. Compost was applied to the experimental plot at a rate of 5 t ha^{-1} during field preparation. After harvest the field was kept fallow for 2 months allowing the rations to grow on the field till mid January just prior to the next dry season's field preparation. The site was flat and had enough fetch for micrometeorological flux

measurements which is required for foot print of the measured fluxes.

Eddy covariance flux measurement, quality control, gap-filling of NEE data and flux partitioning

The eddy covariance system was installed in the middle of a 2.25 ha lowland flooded rice field. Flux densities of CO₂ over the rice canopy were measured by the open path eddy covariance technique. A sonic anemometer (CSAT3, Campbell Scientific. Corp., Canada) measured three-dimensional wind speed and sonic temperature. To measure the fluctuations in CO_2 and water vapour densities, an open path infrared gas analyzer (LI-7500, LICOR Inc., USA) was used. Both the sensors, CSAT3 and LI-7500 were installed at 3.0 m height on a tripod aluminium mast. The LI-7500 was set back from CSAT3 to minimize flow distortions and the head was tilted about 12° from vertical to minimize the amount of precipitation that accumulated on the window. The prevalent wind was blowing from south west direction to the flux measurement sensors. Net ecosystem CO₂ exchange (NEE) was calculated as the sum of eddy CO_2 flux (F_c) and CO_2 storage change (Fs) within air space below fluxmeasuring height. The F_s was neglected for the NEE calculation because canopy height was relatively low at less than 1.3 m. The net gain or loss of carbon from an ecosystem is defined as net ecosystem production (NEP) and results from the gain of carbon from autotrophic organisms (gross primary production, GPP) minus its loss from autotrophic (Ra) and heterotrophic (Rh) respiration:

$$NEP = GPP - Ra - Rh \tag{1}$$

Eddy covariance measurements and meteorological data were recorded for the entire 24-h throughout the season. The mean vertical flux density of CO_2 was obtained as the 30 min covariance between vertical fluctuations (ω') and the CO_2 mixing ratio (C') (Baldocchi 2003):

$$\mathbf{F}_{\mathbf{z}} = \overline{\rho \mathbf{a}} \ast \omega' \mathbf{C}' \tag{2}$$

where ρa refers to air density, the overbars denote time averaging and the primes represent fluctuations of average value. A positive covariance between ω' and C' indicates net CO₂ transfer into the atmosphere and a negative value indicates net CO₂ absorption by the vegetation. The 30 min average of NEE was used for calculation (Massman and Lee 2002).

The footprint analysis was done to confirm and ensuring that the measured fluxes were representative of the plots of interest. The model of Schuepp et al. (1990) was used which measured the cumulative normalized contribution to the surface flux from the upwind location. The eddy flux in the flooded field came from an average of 141 m during daytime and extended up to an average of 145 m during night.

The NEE were sampled at 10 Hz using a data logger programme performed all the processing online and in real time. It applied cross products (second moments) required for offline coordinate rotation following Kaimal and Finnigan (1994) and Tanner and Thurtell (1969) as well as Webb et al. (1980) term for CO₂ flux. Quality checks of eddy covariance flux data were done to eliminate the badly affected data by rainfall, instrument malfunction and inappropriate meteorological conditions. The spike detection and storage correction (Papale et al. 2006), U* filtering (Reichstein et al. 2005; Papale et al. 2006) were performed on the datasets. The friction velocity (U*) threshold for night time data was 0.1 m s^{-1} . Gap-filling of missing and or discarded data were done by "look-up" table approach (Falge et al. 2001) in order to estimate seasonal NEE.

For flux partitioning (i.e., NEE into GPP and RE) of eddy data, rectangular hyperbola (Ruimy et al. 1995) method was used which depended on α (apparent ecosystem quantum yield), β (NEE at light saturation maximum CO₂ uptake rate), γ (estimate of ecosystem respiration, RE), and Q (PAR, photosynthetically active radiation) with the help of following equation:

 $NEE_{night+day} = -[(\alpha.\beta.Q)/(\alpha.Q+\beta)] + \gamma$ (3)

The GPP was calculated by using the equation

$$NEE = GPP - RE \tag{4}$$

Meteorological, soil and plant parameters

Air temperature and relative humidity were measured at 3 m height with temperature-humidity sensors (HMP45C, Campbell Scientific Corp., Canada) on half-hourly basis. The net radiation was measured with the help of net radiometer (NR LITE, KIPP and ZONEN, the Netherlands) installed at 2 m height above the soil surface. Soil temperature, soil heat flux and volumetric water content at 15 cm soil depth were measured on a half-hourly basis using soil and water temperature probe (107 B), heat flux probe (HFT3 transducer) and water content reflectometer (CS 616-L) (Campbell Scientific Corp., Canada), respectively. Vapour pressure deficit (VPD) was estimated from the difference between saturated vapour pressure and actual vapour pressure which were monitored (LI-7500) on half-hourly basis. Leaf area index (LAI) was periodically measured by digital LAI meter (M/S Delta T devices, UK) at different crop growth stages.

Estimation of parameters for net ecosystem carbon budgeting

The net ecosystem carbon budget (NECB) was estimated by partitioning organic carbon into several components in flooded rice paddy ecologies to quantify carbon accumulation and loss from the ecosystem using following equation (Smith et al. 2010):

$$NECB = NEP - D - F - H - VOC - CH_4 - E + I$$
(5)

where, NEP was net ecosystem production, D was the carbon loss as dissolved organic carbon, F was the loss of carbon by fire, H was the carbon loss by harvest, VOC was the carbon loss by volatile organic compound, methane (CH_4) was the carbon loss as microbially produced methane, E was the carbon loss by erosion and eluviation, I was the addition of carbon from organic manure and other sources (compost, rice roots and stubbles of previous season, rhizodeposition and algal biomass of present season).

Carbon inputs

The NEP was estimated through eddy covariance system (described in previous section). Total carbon content of the added compost on dry weight basis was measured by using the organic elemental analyzer (Thermo Scientific, Flash 2000). Carbon input from rhizodeposition was estimated by using conversion factor of 15 % of total above ground biomass at harvest (Mandal et al. 2008). The rice roots and stubble left over in field of previous season, and plant biomass (root, shoot and grain) of present season were measured from ten replications (1 m² area each), randomly selected from fetch area of eddy covariance system. The default value of algal C load

i.e. 0.70 Mg C ha⁻¹ (Mandal et al. 2008) in tropical rice was used for carbon balance calculation.

Carbon output

To estimate dissolved organic carbon (DOC), the soilwater solution were collected at 3–6 days intervals coinciding the date of CH_4 estimation throughout the cropping season in ten replications. The samples were passed through 0.45 µm filter and air dried. The total C of the samples was measured by organic elemental analyzer (Thermo Scientific, Flash 2000) (Zsolnay 2003).

Methane fluxes were measured by manual closed chambers at close periodical intervals of 3-6 days from study site. Samplings for CH₄ flux measurements (3 replications) were done from flooded rice field in the morning (09:00-09:30 h) and afternoon (15:00-15:30 h), and the average of the morning and afternoon fluxes was used as the flux for the day. The samplings were done at 6, 11, 16, 20, 25, 31, 37, 41, 46, 52, 57, 61, 67, 73, 79, 85, 91, 97, 103, 108, 112, and 116 days after transplanting (DATs) of the rice crop encompassing the all growth phase of the crop. For measuring CH₄ emissions, six rice hills were covered with a locally fabricated Perspex chamber (53 cm \times length \times width \times height $37 \text{ cm} \times 51 \text{ cm}$, from seedling to tillering, and 53 cm \times 37 cm \times 71 cm, length \times width \times height from maximum tillering to maturity stages). A battery-operated air circulation pump with an air displacement of 1.5 Lmin^{-1} (M/s Aerovironment Inc., Monrovia, CA, USA) and connected to polyethylene tubing was used to mix the air inside the chamber and draw the air samples into Tedlar gas-sampling bags (M/s Aerovironment Inc.) at fixed intervals of 0, 15 and 30 min. Methane concentrations of gas samples from the sampling bags were analyzed by gas chromatography (Chemito, CERES 800 plus, M/s Thermo Scientific) equipped with a flame ionization detector (FID) and Porapak Q column (6 feet long, 1/8 inch outer diameter, 80/100 mesh size, stainless steel column). The temperature of the injector, column and detector was maintained at 150, 50 and 230 °C, respectively. The carrier gas (nitrogen) flow was maintained at 15 mL min⁻¹. The gas chromatograph was calibrated before and after each set of measurements by using 1.2 and 1.8 μ L CH₄ L⁻¹ in N2 (M/s Chemtron Science Laboratories, India) as the primary standard. Fluxes of CH₄ were calculated by successive linear interpolation of the average emissions on the sampling days, assuming that the emissions followed a linear trend during the periods when no sampling was done (Datta et al. 2009; Bhattacharyya et al. 2012a, b). Cumulative CH₄ emissions for the entire cropping period were computed by plotting the flux values against the days of sampling and were expressed as Mg CH₄–C ha⁻¹.

Total harvest C removal was estimated from shoot and grain yield after harvest (mentioned in 'carbon input' section). Eluviation, erosion and leaching component were not estimated it was derived from net ecosystem C budget equation (Eq. 4).

Total organic carbon of soil

Total organic carbon in soil (TOC) before and after the season were measured by taking composite soil samples (ten replications in the study area) by a sample probe (8 cm diameter) from a depth of 0–15 cm. Immediately after sampling, excess water was allowed to drain off, visible root fragments and stones were removed manually and transferred to the laboratory for analyses. The fresh soil was air-dried until constant weight reached, sieved through a 2 mm mesh, mixed and stored in sealed plastic jars for analyses. The TOC of the soil samples were measured by dry combustion method using organic elemental analyzer (Thermo Scientific, Flash 2000).

Results and discussion

Diurnal and seasonal variation of NEE, GPP and RE

Over the season, diurnal variations of mean NEE varied from +3.99 to -18.50μ mol CO₂ m⁻² s⁻¹, where, positive sign indicated net CO₂ emission into the atmosphere and negative sign denote net CO₂ assimilation or uptake by the crop. The rice paddy ecosystem was behaving as a CO₂ source during night hours and a CO₂ sink during the day. Almost over the entire season, on daily basis, rice crop behaved as net CO₂ sink except few days during the maturity period. Throughout the cropping season on the basis of average diurnal NEE, maximum CO₂ assimilation or uptake by rice crop was found at 11.00 h and maximum emission was observed at 4.00 h. The daily

average NEE over the cropping season varied from +2.63 to -7.47 μ mol CO₂ m⁻² s⁻¹, i.e., +2.73 to -7.74 g C m⁻² day⁻¹. The NEE was lower in stage I (vegetative stage, average $-2.36 \text{ g C m}^{-2} \text{ day}^{-1}$), then gradually decreased in stage II (tillering to panicle initiation stage, average -3.98 g C m^{-2} day^{-1}) and III (reproductive stage, average -5.46g C $m^{-2} day^{-1}$) and reached its maximum in the stage IV (heading to flowering stage, average -5.67g C m⁻² day⁻¹), and then started to increase in stage V (ripening stage, average $-4.00 \text{ g C m}^{-2} \text{ day}^{-1}$) and VI (harvest stage, average $-3.15 \text{ g C m}^{-2} \text{ day}^{-1}$) (Fig. 1a-f). The eddy covariance based CO₂ flux monitoring was done for 111 consecutive days (6-116 days after transplanting) and the integrated season-long NEE value was-448 g C m⁻². The GPP and RE varied from -5.74 to -8.99 g C m⁻² day⁻¹ and 2.89 to 3.86 g C m⁻² day⁻¹, respectively at different growth stages. The highest GPP (-8.99 g C m⁻² day⁻¹) and RE (3.86 g C m⁻² day⁻¹) were recorded at the reproductive stage and maximum tillering to panicle initiation stage, respectively (Fig. 1a–f).

It was evident from the study that the amplitude of the daily variation in NEP increased as leaf area index (LAI) at different growth stages increased (vegetative stage LAI 1.2, NEP $+2.36 \text{ g C m}^{-2}$ day⁻¹; tillering to panicle initiation stage LAI 3.5, NEP +3.98 g C m⁻² day^{-1;} reproductive stage LAI 4.8, NEP +5.46 g C m⁻² day^{-1;} heading to flowering stage LAI 5.5, NEP $+5.67 \text{ g C m}^{-2} \text{ day}^{-1}$; ripening stage LAI 4.6, NEP +4.00 g C m⁻² day⁻¹ and harvest stage LAI 2.9, NEP +3.15 g C m⁻² day^{-1}) and reached its peak around the heading to flowering stage and, then on, decreased gradually till maturity due to leaf senescence or reduction in leaf greenness during maturity. This is supported by the study of Pakoktom et al. (2009) and Patel et al. (2011). Net exchanges of CO_2 between rice paddies and the atmosphere are controlled by several biological and physical processes. During the day time, NEE is the difference between the CO_2 uptake from plant photosynthesis and the CO₂ emitted through respiration of plant and soil. Respiration at night led to an efflux of CO₂ to the atmosphere in absence of photosynthesis. Same type of results were found by Alberto et al. (2009) in Philippines, Tseng et al. (2010) in Taiwan and Miyata et al. (2000) in Japan in rice ecologies. CO₂ uptake declined through Fig. 1 NEE at different growth stages of rice viz.
(a) vegetative stage,
(b) tillering to panicle initiation, (c) reproductive stage, (d) heading to flowering stage, (e) ripening stage and (f) harvesting stage



the afternoon hours due to reduction in leaf level gas exchange (Larcher 1995; Goulden et al. 2004). Afternoon depression might be due to stomatal response to vapour pressure deficit, low leaf water potential or retarding photosynthesis due to relatively increased temperature (Jones 1992; Larcher 1995).

Effect of net radiation, PAR on GPP

During the study period diurnal mean net radiation varied from -36.89 (19.00 h) to 448.98 W m⁻² (11.00 h) (mean value 106.67 W m⁻²) (Fig. 2a). After sun rise, due to incoming solar radiation it gradually started to increase and reached its peak at 11.00 h and then started to decline gradually due to inclined radiation. Significantly (p < 0.01) positive correlation (r = 0.69**, 0.61**) was found between net radiation and PAR with GPP (Table 1a). As a whole, the NEE becomes more negative during the day because of increasing CO₂ uptake through

photosynthesis (increase in GPP) as net radiation increases (Alberto et al. 2009; Saito et al. 2005).

Effect of vapour pressure deficit on GPP

Vapour pressure deficit significantly influenced the GPP during the study. The daily average VPD in the study area was 0.722 kPa (varied between 0.234 to 1.136 kPa) (Fig. 2c). The lowland rice field was flooded with 12–15 cm of standing water that increased the vapour pressure in the canopy-soil system of rice (Fig. 2b), which in turn, influenced VPD and consequently the GPP. Increasing VPD caused partial stomatal closure which resulted in reduction in photosynthesis and thereby reducing GPP (Saitoh and Ishihara 1987; Mahrt and Vickers 2002; Pakoktom et al. 2009). On the other hand, VPD increases air temperature that leads to increase in respiration and thereby reduces the NEE and GPP (Alberto et al. 2009).



Fig. 2 Diurnal changes in mean net radiation (a), vapour pressure deficit (b), vapour pressure (c), soil temperature at 15 cm soil depth (d), soil heat flux at 15 cm soil depth (e) and air temperature (f) with respect to local time observed during the wet season, 2010

Effect of soil moisture on GPP

Volumetric soil moisture content was not significantly correlated with GPP as there was no moisture stress most of the time of crop growing season (Table 1a). Soil CO_2 efflux might have been enhanced due to

microbial decomposition of soil organic matter in submerged soils. The magnitude and duration of CO_2 efflux depends on the amount and duration of precipitation (Huxman et al. 2004; Sponseller 2007), during moisture stress condition, which in our study was only limited to last 15 days before harvest of the crop. It

VPD AT SM NR PAR GPP (a) VPD 1.00 AT 0.81** 1.00 0.04^{ns} -0.09^{**} SM 1.00 0.54** NR 0.56** -0.04*1.00 -0.04^{ns} PAR 0.38** 0.39** 0.86** 1.00 0.01 ns GPP 0.31** 0.36** 0.69** 0.61** 1.00 AT ST SM RE SHF (b) AT 1.00 ST 0.56** 1.00 -0.24 **-0.42**SM 1.00 RE 0.12** 0.05** -0.05**1.000.25** 0.02 ns SHF 0.46** 0.45** 1.00

Table 1 Correlation coefficients between GPP (a), RE (b) and related environmental variables (n = 5,328)

Here, *RE* ecosystem respiration (µmol CO₂ m⁻² s⁻¹), *GPP* gross primary productivity (µmol CO₂ m⁻² s⁻¹), *SM* soil moisture content (%), *AT* air temperature (°C), *SHF* soil heat flux (W m⁻²), *VPD* vapour pressure deficit (kPa), *PAR* photo active radiation (µmol m⁻² s⁻¹), *NR* net radiation (W m⁻²). ** 1 % level of significance (p < 0.01); * 5 % level of significance (p < 0.05); ^{ns} non significant

actually depend on antecedent soil moisture conditions and reflects trade-offs between autotrophic (net CO_2 uptake) and heterotrophic (net CO_2 release) contributions (Potts et al. 2006). In this study, as the rice fields remained submerged most of the times during the crop season, the magnitude of the CO_2 efflux strongly depended on the ratio of activated autotrophic to heterotrophic soil organisms and on the actual wetting depth.

Effect of soil temperature on RE

The diurnal mean soil temperature at 15 cm soil depth varied between 28.9 to 30.6 °C (Fig. 2d) depending on the intensity of the incoming solar radiation, soil characteristics and moisture conditions. Standing water of 12–15 cm was maintained in the field most of the times during the cropping season except few days during maturity. Due to standing water in the lowland (flooded) rice fields, the soil temperature did not vary significantly throughout the cropping season up to 14 days before harvest as standing soil water acted as an insulator (Mowjood et al. 1997). Significant relationship $(r = 0.05^{**})$ was existed between soil temperature and RE (Table 1b).

Effect of soil heat flux on RE

Diurnal variation of average soil heat flux at 15 cm soil depth showed a negative trend from 21.00 h to 11.30 h and positive trend from 12.00 h to 20.30 h and the average value ranged from -8.53 W m⁻² (7.30 h) to +12.04 W m⁻² (15.30 h) (Fig. 2e). In tropical flooded rice ecology the heat flux became important as high temperatures prevailed with high relative humidity (71–98 %, in the study site) at most of the growth stages during *kharif.* However, in this study nonsignificant correlation was found between soil heat flux and RE (Table 1b).

Effect of air temperature on NEE and RE

During the study period diurnal mean air temperature was in the range of 28.4–32.1 °C (Fig. 2f). Whereas, the daily mean values of air temperature varied from 24.6–32.2 °C (data not presented). Maximum CO₂ uptake was observed between 22 and 34 °C and its uptake gradually decreased beyond 34 °C (Fig. 3). The increase of the air temperature above 34 °C increased NEE (less negative value) by decreasing photosynthetic rate in flooded rice ecology (Fig. 3). Decline in photosynthesis due to onset of stress triggering into circadian rhythm or stomatal response to higher temperature showed significant positive correlation (r = 0.12**) with RE during the course of study (Table 1b).



Fig. 3 NEE (μ mol $m^{-2}~s^{-1})$ as a function of air temperature (°C)

Photosynthesis is particularly sensitive to heat stress. The sensitivity of photosynthesis to heat stress is associated with the inactivation of the enzyme Rubisco involved in the Calvin cycle. The enzyme Rubisco catalyzes the first major step of carbon fixation and is temperature sensitive (Crafts-Brandner and Salvucci 2000). Warm temperature stimulates photorespiration and ordinary respiration in plants. Thus, most plants drop their photosynthetic activity at higher temperature. Decline in CO_2 uptake may also be due to stomatal closure in response to evaporative demand or a change in photosynthetic biochemistry in response to increased temperature or a circadian rhythm.

The regression model between selected environmental variables on GPP and RE

The environmental factors primarily affecting GPP are PAR, VPD, temperature and soil moisture, whereas, RE is influenced by air or soil temperature, soil moisture and soil heat flux. The observed results showed that the GPP and RE varied in diurnal and seasonal time scales due to leaf gas exchange and pattern of light interception by the canopy (Ruimy et al. 1995; Jarvis and Leverenz 1983). The analysis of the observed results of GPP, RE and environmental factors over flooded rice paddy ecosystem in the growing season revealed that CO₂ absorption in the daytime (morning) was higher than that of in the afternoon hours. Plant photosynthesis during the afternoon hours was inhibited by the higher temperatures (resulting in low enzyme activity) and vapour pressure deficit (causing stomatal closure) and thus carbon absorption was restrained during afternoon. Mid-day photosynthesis inhibition is a self-protection mechanism of plants adapting to drought stress (Matos et al. 1998).

The regression models of GPP and RE with the environmental variables like PAR, VPD, air temperature (AT), soil moisture (SM) and soil heat flux (SHF) were:

$$GPP = 0.77(AT) + 0.01(PAR) - 0.90(VPD) - 15.31$$
(6)

$$RE = 0.13(AT) + 0.56(SM) - 0.09(SHF) - 1.46$$
(7)

The model implies that GPP was positively correlated with AT and PAR, whereas, it was negatively correlated with VPD. However, RE was positively correlated with AT and SM but negatively correlated with SHF.

Partitioning of carbon cycling and budgeting in rice-ecosystem

The net ecosystem production estimated during the season was in the order of $+4.48 \text{ Mg C ha}^{-1}$. The NEP is equal but opposite in sign to NEE (NEE = -NEP; Schmid et al. 2000; Barker and Griffis 2005; Black et al. 2007). As plants exchanged most of their carbon as CO₂, so eddy flux-derived NEP was an ideal variable for C budgeting as well as C cycling in flooded rice paddy ecosystem (Baldocchi 2003). Therefore, NEP (4.48 Mg C ha^{-1}) after completion of wet season was carbon input in the system. Addition of carbon inputs from stubble and root biomass (residue left in the field of the previous season), rhizodeposition, algal biomass, compost addition and NEP were 0.17, 0.87, 0.68, 0.70, 0.22, 4.48 Mg C ha⁻¹, respectively totalling of about 7.12 Mg C ha⁻¹ (Fig. 4). But, the stored carbon was released and lost from the system due to several farm practices and crop biochemical and physiological processes. Carbon removal after harvest through rice grain, straw, stubble and root was 5.76 Mg C ha⁻¹ (Fig. 4). The cumulative CH_4 -C and DOC losses of the season were in the order of 0.08, 0.93 Mg C ha⁻¹ (Fig. 5). Therefore, C output (as grain and biomass yield plus CH₄-C and DOC) from the system during the study period was $6.77 \text{ Mg C ha}^{-1}$. As crop residue, stubble burning was not done, the gaseous carbon loss due to fire was not included during balancing the input and output components in NECB model (Eq. 5). Volatile organic carbon (VOC) losses from lowland flooded rice ecosystem were assumed to be negligible (Smith et al. 2010). As, total organic carbon content in the soil (upper 15 cm surface) was not changed significantly before and after the experiment (data not presented), the balancing between the carbon input and output $[7.12 - (5.76 + 0.93 + 0.08) = 0.35 \text{ Mg C ha}^{-1}]$ of the system were done by considering the losses from erosion and eluviations (leached from upper layer of profile to lower layer) (Smith et al. 2010).



Fig. 4 Schematic diagram of the carbon budget of lowland (flooded) rice paddy ecosystem



Conclusion

The season-long study revealed that the low land rice fields have the capacity to sequester carbon from the atmosphere. NEE exhibited a clear diurnal pattern throughout the season. It showed day time uptake and night time release of carbon dioxide by the rice canopy. It is evident that the flooded rice paddy ecosystem behaved as net CO_2 sinks. Carbon distribution in different soil-plant system revealed that 63, 9.5 and 10 % of total carbon entered into the system through net ecosystem production, rhizodeposition and algal biomass, respectively. Whereas, 81 and 14 % of total C removal from the system through crop harvest and combined process of dissolved organic carbon leaching and methane emission, respectively. Whereas, most of the studies in Bangladesh, Philippines, Japan, India and other Asian countries concentrated on temporal and spatial variation of GPP, NEE, RE in relation to meteorological variable, the present study gave emphasis on system based C balancing incorporating NEP as a major component. However, a long term study incorporating different cultivars and exhaustive soil and gaseous C component analysis could give clearer picture of net ecosystem carbon budget. Acknowledgments The work has been partially supported by the grant of ICAR-NAIP, Component-4 (2031), project entitled "Soil organic carbon dynamics vis-à-vis anticipatory climatic changes and crop adaptation strategies", ICAR-NICRA and CRRI. The suggestion given by Dr. D. C. Uprety, Dr. S. N. Singh, Dr. V. R. Rao and Dr. T. K. Adhya is gratefully acknowledged. Part of the result is the PhD work of Mr. Suvadip Neogi. Technical support was provided by the technical staffs of the Division of Crop Production, CRRI.

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