

Pattern of genetic inheritance of morphological and agronomic traits of sorghum associated with resistance to sorghum shoot fly, *Atherigona soccata*

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Received: 14 July 2017/Accepted: 4 January 2018/Published online: 27 January 2018 © Springer Science+Business Media B.V., part of Springer Nature 2018

Abstract Sorghum shoot fly, *Atherigona soccata* is an important pest of sorghum during the seedling stage, which influences both fodder and grain yield. To understand the nature of inheritance of shoot fly resistance in sorghum, we performed generation mean analysis using two crosses IS $18551 \times \text{Swarna}$ and M $35\text{-}1 \times \text{ICSV}$ 700 during the 2013-2014 cropping seasons. The F_1 , F_2 , BC₁ and BC₂ progenies, along with the parental lines were evaluated for agronomic and morphological traits associated with resistance/susceptibility to sorghum shoot fly, *A. soccata*. The cross IS $18551 \times \text{Swarna}$ exhibited significant differences between the parents for shoot fly

Electronic supplementary material The online version of this article (https://doi.org/10.1007/s10681-018-2111-9) contains supplementary material, which is available to authorized users.

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deadhearts (%) in the postrainy season. The progenies of this cross exhibited lower shoot fly damage, suggesting that at least one of the parents should have genes for resistance to develop shoot fly-resistant hybrids. Leaf glossiness, leafsheath pigmentation and plant vigor score during the seedling stage exhibited non-allelic gene interactions with dominant gene action, whereas 100 seed weight showed both additive and dominant gene interactions. Presence of awns showed recessive nature of the awned gene. Generation mean analysis suggested that both additive and dominance gene effects were important for most of the traits evaluated in this study, but dominance had a more pronounced effect.

Keywords Sorghum shoot fly resistance · *Atherigona soccata* · Genetic inheritance · Generation mean analysis

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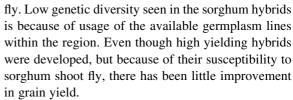
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Introduction

Sorghum [Sorghum bicolor (L.) Moench] is the fifth most important drought tolerant cereal crop after maize, rice, wheat, and barley (Doggett 2008). It is adapted to the tropical and subtropical climates of the semi-arid regions, and is the staple food for millions of people living in these regions. According to Food and Agriculture organization (FAO 2014), the grain sorghum area in India is about 5.82 m ha, with a production of 5.39 million tonnes of grain sorghum, with a productivity of 926.1 kg/ha. It is a multipurpose crop that can be utilised as food, feed, fodder, and presently, it is emerging as a bio-fuel crop (House 1985; Doggett 1988; Rooney and Waniska 2000). It is vulnerable to several biotic and abiotic constraints, resulting in decreased grain yields, and consequently leading to decline in the sorghum area under cultivation. Hence, it is important to increase the grain yields within the existing area to feed the growing population in the semi-arid regions of Asia and Africa.

During the process of breeding for high yielding sorghums, no attention was paid for insect pest resistance and as a result, most of the high yielding sorghum cultivars are susceptible to insect pests. Therefore, it is important to focus attention on the constraints that result in grain yield loss than on genetic improvement for grain yield per se. About 150 insect pests attack sorghum from seedling to physiological maturity stage. Of these, sorghum shoot fly, Atherigona soccata (Rondani) is one of the serious insect pests that attack sorghum at the seedling stage. Shoot fly infestation results in severe economic loss to the farmers (Sharma 1993; Riyazaddin et al. 2015). Sorghum shoot fly, A. soccata infests sorghum at the early seedling stage i.e., from 7 to 30 days after seedling emergence (DAE). Host plant resistance (HPR) is one of the effective methods for controlling shoot fly, A. soccata. A number of genotypes with resistance to shoot fly have been identified, but the levels of resistance are low to moderate (Pradhan and Jotwani 1978; Taneja and Leuschner 1985; Sharma et al. 2003).

The genotypes exhibiting resistance to sorghum shoot fly, *A. soccata* generally have poor grain type, low productivity and physiologically inefficient plant type, which were undesirable. Whereas, the hybrids developed for high grain yield have lower genetic diversity, and are highly susceptible to sorghum shoot



The choice of selection and breeding strategies for genetic improvement of sorghum or any other crop largely depend on the type, and relative importance of genetic components, and presence of non-allelic interactions. In view of the potential economic and environmental constraints associated with insecticide use, breeding of crop varieties with resistance to shoot fly is a promising method to control the insect pests (Sharma 1993). Hence, transferring the resistance from agronomically undesirable genotypes (resistant genotypes) into the high-yielding hybrids is essential (Rana et al. 1981) for sustainable sorghum production.

Understanding the genetic inheritance of shoot fly resistance and the agronomic, and morphological traits associated with resistance/susceptibility to shoot fly damage will be helpful in breeding sorghums with high grain yields that are acceptable to the farmers. Genetic improvement depends primarily on the effectiveness of selection among the progenies that differ in genetic value. Generation means provides information on genetic inheritance of the quantitative traits. Most of the researchers working on shoot fly resistance have focused mainly on inheritance of shoot fly resistant traits, with little information on agronomic and morphological traits. An understanding of genetic inheritance of resistance to shoot fly, A. soccata, and as well as the agronomic, morphological traits will be useful to breed sorghums with shoot fly resistance and desirable agronomic traits. Hence, the present study was aimed at understanding the type of gene interactions governing inheritance of shoot fly resistance, and the agronomic and morphological traits associated with shoot fly resistance.

Materials and methods

Experiments were conducted at the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru 502 324, Medak, Telangana, India, which is situated at 17°53′N latitude, 78°27′E longitude and at an altitude of 545 m.



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Experimental material

The experimental material consisted of six generations of two crosses of M 35-1 \times ICSV 700, and IS 18551 \times Swarna. One of the crosses involved a shoot flyresistant (IS 18551, P₁) and the susceptible (Swarna, P₂) genotypes. Back cross progenies were obtained by crossing the F₁ progeny with either of the parents [BC₁ $(F_1 \text{ crossed with } P_1), \text{ and } BC_2 (F_1 \text{ crossed with } P_2)].$ The F₁'s were selfed to obtain the F₂ progenies. Hence, segregating and non-segregating material was generated (viz. P₁, P₂, F₁, F₂, BC₁ and BC₂). All the six generations were evaluated in replicated trials using randomized complete block design (RCBD), during the rainy and postrainy seasons. The test material was sown with parents in two rows, F₁'s in a single row, F₂'s in 10 rows, and back cross progenies in four rows, with a row length of 2.0 m, row to row distance of 75 cm, and a distance of 10 cm in between the plants. A basal dose of ammonium phosphate was applied to the field @ 100 kg/ha. Normal agronomic practices were followed in raising the crop. Earthing up and top dressing with urea at 100 kg/ha was done 30 days after seedling emergence. During the postrainy season, the test plots were irrigated at 30 day intervals. Interlard fish meal technique as described by Soto (1974) and Sharma (1993) was followed for multiplication of shoot fly population and even exposure of the test genotypes to shoot fly infestation. One set of the replicated test material was grown under protected conditions by periodical spraying with cypermethrin, and applying the carbofuran 3G granules in the leaf whorls to protect the seedlings from shoot fly damage for recording data on morphological, agronomic, panicle traits, and grain yield.

Observations

Data on shoot fly damage was recorded by observing the number of shoot fly deadhearts in a test plot at 21 DAE, and expressed as percentages. The data on the agronomic, morphological and panicle traits were recorded based on the sorghum descriptors (IBPGR and ICRISAT 1993), from seedling to the harvesting stage with slight modifications (Supplementary Table 1). Days to 50% flowering was recorded when half of the panicle, and 50% of the plants in the experimental plot attained the anthesis stage, while plant height of three randomly selected plants within a

plot was recorded at maturity. Data on 100 seed weight and grain yield were recorded from the protected test plots after harvesting and threshing the panicles.

Leaf glossiness was visually scored on a 1–5 scale at 10–12 DAE (fifth leaf stage), when the expression of this trait is most apparent, in the morning hours, when there was maximum reflection of light from the leaf surface (Sharma and Nwanze 1997), leafsheath pigmentation was visually scored on a 1–3 rating scale at 7 DAE (Dhillon et al. 2006), and seedling vigor at 10 DAE on a 1–3 scale (Sharma and Nwanze 1997). Data were also recorded on waxy bloom, plant color, inflorescence exsertion, panicle compactness, panicle shape, glume color, glume coverage, awns, grain color, and grain lustre (Supplementary Table 1).

Statistical analysis

The data were subjected to the analysis of variance (ANOVA) using GenStat, 14th edition (GenStat 2010). F-test was used to test the significance of differences between the test genotypes, while least significance differences (LSD) was used to compare the genotypic means at $P \leq 0.05$. Data obtained for various morphological, agronomic and panicle traits were subjected to generation mean analysis followed by Mather (1949) and Hayman and Mather (1955) for scaling test, and Hayman (1958) approach to find the significant inter-allelic interactions, using Windostat (Indostat 2004) software.

Results

Mean performance of crosses across seasons

Analysis of variance for various agronomic, morphological, and panicle traits for the rainy and postrainy seasons are presented in Tables 1 and 2. The F-values due to generations were significant at $P \leq 0.01$ for days to 50% flowering, 100 seed weight, grain yield, glume color and glume cover across seasons for both the crosses; while plant height, waxy bloom, grain lustre and awns showed significant F-values for the cross IS $18551 \times Swarna$ across seasons. The cross M $35-1 \times ICSV$ 700 showed non-significant variation for these traits, which exhibit moderate levels of resistance to shoot fly. Plant color exhibited significant F-value at $P \leq 0.01$ for the cross M $35-1 \times ICSV$ 700



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Table 1 Mean performance of two crosses with respect to various agronomic and morphological traits of sorghum across seasons (ICRISAT, Patancheru, 2013–2014)	crosses with 1	respect to	various ag	gronomic and m	orphological traits of so	rghum across seasons	s (ICRISA)	l, Patanche	ru, 2013–20	14)
Pedigree	Generation	Shoot fly deadhearts (%)	y rts (%)	Leaf glossy score	Leaf sheath pigmentation	Plant vigor score	Days to 50% flowering	\$0% \$	Plant height (cm)	ıt (cm)
		2013R	2013PR	2013PR	2013PR	2013PR	2013R	2013PR	2013R	2013PR
Cross M 35-1 \times ICSV 700										
M 35-1	\mathbf{P}_1	70.54	7.89	1.67	1.00	1.67	74.33	72.80	306.70	182.00
$M 35-1 \times ICSV 700$	F_1	78.15	13.46	1.00	1.00	2.00	67.47	68.93	312.70	212.70
M $35-1 \times ICSV 700$	F_2	71.34	8.23	2.06	1.57	1.57	69.82	71.15	301.90	207.60
${M 35-1 \times ICSV 700} \times {M 35-1}$	BC_1	71.33	86.6	2.02	1.78	1.54	70.22	70.56	301.20	200.50
$\{M\ 35-1\times ICSV\ 700\}\times ICSV 700$	BC_2	45.20	16.90	1.81	1.46	1.51	71.70	71.39	314.80	206.50
ICSV 700	\mathbf{P}_2	64.68	20.99	1.33	1.00	1.00	78.33	75.47	295.30	213.30
Mean		28.99	12.90	1.65	1.30	1.55	71.98	71.70	305.43	203.80
SE±		7.39	3.51	0.31	0.10	0.16	0.97	0.78	7.71	8.79
Vr (10, 5)		2.40	2.22	1.81	13.11**	4.36*	15.86**	8.20**	0.92	1.75
LSD (P 0.05)		SN	SN	NS	0.30	0.49	3.05	2.45	NS	NS
Cross IS 18551 \times Swarna										
IS 18551	P_1	51.87	3.60	1.00	1.00	1.00	79.73	16.67	325.30	210.70
IS $18551 \times Swama$	\mathbf{F}_{1}	68.15	17.68	3.00	1.00	2.00	64.80	66.13	312.70	215.30
IS $18551 \times Swama$	F_2	82.44	24.27	3.06	1.70	1.51	68.29	89.99	280.50	203.60
$\{IS\ 18551 \times Swarna\} \times IS$ 18551	BC_1	81.90	16.22	2.68	1.48	1.70	70.56	70.56	303.30	212.90
$\{IS\ 18551 \times Swarna\} \times Swarna$	BC_2	91.89	38.26	4.18	1.57	1.83	69.07	64.04	252.50	194.10
Swarna	P_2	59.61	54.31	5.00	1.67	2.00	79.59	62.33	169.30	130.70
Mean		72.64	25.70	3.15	1.40	1.67	69.69	67.70	273.96	194.50
SE±		10.71	3.44	0.09	0.14	0.10	0.85	0.67	3.90	7.96
Vr (10, 5)		2.05	27.46**	253.45**	5.34**	13.58**	39.95**	**90.09	215.83**	16.37**
LSD (P 0.05)		SN	10.83	0.27	0.44	0.32	2.68	2.11	12.30	25.09



Table 1 continued

Pedigree	Generation	100 seed weight (g)	eight (g)	Grain yield (t/ha)	(t/ha)	Waxy bloom	m,	Plant color	
		2013R	2013PR	2013R	2013PR	2013R	2013PR	2013R	2013PR
Cross M 35-1 \times ICSV 700									
M 35-1	${\bf P_l}$	2.31	3.56	2.51	92.9	2.00	2.67	1.00	1.00
$M 35-1 \times ICSV 700$	${\sf F}_{\sf l}$	3.12	3.96	5.17	12.24	1.67	2.67	1.00	1.00
M $35-1 \times ICSV 700$	F_2	2.54	3.64	2.87	8.96	1.08	2.26	1.42	1.37
$\{M 35-1 \times ICSV 700\} \times M 35-1$	BC_1	2.70	3.97	4.23	11.85	1.32	2.31	1.05	1.08
$\{M 35-1 \times ICSV 700\} \times ICSV 700$	BC_2	2.44	3.44	3.55	10.63	1.12	2.20	1.43	1.57
ICSV 700	P_2	2.14	2.43	1.03	5.68	1.33	2.00	2.00	2.00
Mean		2.54	3.50	3.23	9.32	1.42	2.40	1.32	1.30
$SE\pm$		0.13	0.05	0.46	0.61	0.22	0.21	90.0	0.03
$V_{\Gamma}(10, 5)$		**62.9	161.97**	**98.6	20.11**	2.69	1.59	37.89**	155.39**
LSD (P 0.05)		0.41	0.14	1.44	1.93	NS	NS	0.20	0.10
Cross IS 18551 × Swarna									
IS 18551	\mathbf{P}_1	1.57	2.21	1.20	4.77	1.00	2.00	1.00	1.00
IS $18551 \times \text{Swarna}$	${\tt F}_1$	2.73	3.18	4.42	10.13	2.67	3.00	1.00	1.00
IS $18551 \times \text{Swarna}$	F_2	2.39	3.12	2.55	7.93	1.99	2.68	1.04	1.00
$\{IS\ 18551 \times Swarna\} \times IS\ 18551$	\mathbf{BC}_1	2.34	2.78	3.14	9.19	1.43	2.24	1.16	1.00
$\{IS\ 18551 \times Swarna\} \times Swarna$	BC_2	2.98	3.63	4.02	8.50	2.73	2.91	1.00	1.00
Swama	P_2	3.00	3.36	4.57	3.64	3.00	3.00	1.00	1.00
Mean		2.50	3.05	3.32	7.36	2.14	2.60	1.03	1.00
$SE\pm$		0.10	90.0	0.29	0.30	0.14	90.0	0.04	0.00
V_{Γ} (10, 5)		30.29**	65.48**	19.62**	71.67**	31.52**	53.96**	2.22	1.00
LSD (P 0.05)		0.31	0.19	0.92	96.0	0.45	0.18	NS	NS

NS non-significant

 * , $^{**}F$ test significant at P 0.05 and 0.01, respectively; R, rainy season; PR, postrainy season

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Table 2 Mean performance of two crosses with respect to various panicle traits of sorghum across seasons (ICRISAT, Patancheru, 2013–2014)

Pedigree	Generation	Inflores		Panicle compa		Panicle shape	Glume	color	Glume c	overage
		2013R	2013PI	2013R	2013PR	2013PR	2013R	2013PR	2013R	2013PR
Cross M 35-1 × ICSV 700										
M 35-1	P_1	2.00	2.00	2.00	3.00	4.00	3.00	3.00	1.00	1.00
M 35-1 × ICSV 700	F_1	1.33	3.00	2.00	3.00	4.00	3.00	3.00	1.00	1.00
M 35-1 × ICSV 700	F_2	1.73	2.40	2.00	2.86	3.58	2.66	2.76	1.69	1.27
{M 35-1 × ICSV 700} × M 35-1	BC_1	2.23	2.40	2.00	2.99	3.96	2.78	2.81	1.27	1.00
{M 35-1 × ICSV 700} × ICSV 700	BC_2	2.08	2.59	2.00	2.95	3.84	2.50	2.28	2.53	1.40
ICSV 700	P_2	2.33	2.00	2.00	3.00	4.00	2.00	2.00	3.67	3.00
Mean		1.95	2.40	-	3.00	3.90	2.66	2.60	1.86	1.40
SE±		0.21	0.09	-	0.01	0.04	0.06	0.06	0.34	0.07
Vr		3.17	18.11**	_	15.68**	15.42**	39.74**	43.69**	9.57**	137.28**
LSD (P 0.05)		NS	0.28	_	0.05	0.13	0.19	0.20	1.08	0.21
Cross IS 18551 × Swarna										
IS 18551	P_1	1.67	2.00	2.00	3.00	4.00	3.00	3.00	9.00	6.33
IS 18551 × Swarna	F_1	1.00	1.00	2.00	2.00	1.00	3.00	4.00	5.00	3.67
IS 18551 × Swarna	F_2	1.31	1.70	2.00	2.46	2.42	3.86	3.80	5.03	4.96
{IS 18551 × Swarna} × IS 18551	BC_1	1.57	1.73	2.00	2.80	3.40	3.32	3.87	6.39	6.33
{IS 18551 × Swarna} × Swarna	BC_2	1.36	1.32	2.00	2.36	2.10	4.44	3.67	1.80	2.44
Swarna	P_2	1.00	1.00	2.00	2.00	1.00	6.00	4.00	1.00	1.00
Mean		1.32	1.50	-	2.40	2.30	3.94	3.70	4.70	4.10
SE±		0.15	0.10	-	0.04	0.12	0.19	0.06	0.19	1.16
Vr		3.63*	18.19**	_	97.12**	97.11**	37.71**	36.36**	238.02**	3.45*
LSD (P 0.05)		0.46	0.31	-	0.13	0.39	0.59	0.20	0.60	3.66
Pedigree		Gener	ation	Grain colo	or	Grain lus	tre	A	wns	
				2013R	2013PR	2013R	2013P	R 20)13R	2013PR
Cross M 35-1 × ICSV 700										
M 35-1		P_1		1.00	1.00	2.00	2.00)	2.00	2.00
M 35-1 × ICSV 700		F_1		1.00	1.00	2.00	2.00)	2.00	2.00
M 35-1 × ICSV 700		F_2		1.00	1.00	2.00	2.00)	2.00	2.00
{M 35-1 × ICSV 700} × I	M 35-1	BC_1		1.00	1.00	2.00	2.00)	2.00	2.00
{M 35-1 × ICSV 700} × I	CSV 700	BC_2		1.00	1.00	2.00	2.00)	2.00	2.00
ICSV 700		P_2		1.00	1.00	2.00	2.00		2.00	2.00
Mean				_	=	_	_		_	_
SE±				_	=	_	_		_	_
Vr				_	=	_	_		_	_
LSD (P 0.05)				_	_	_	_		_	_
Cross IS 18551 × Swarna										
IS 18551		P_1		1.00	1.00	1.00	1.00)	2.00	2.00
IS 18551 × Swarna		F_1		1.00	1.00	2.00	2.00		1.00	1.00
IS 18551 × Swarna		F ₂		1.00	1.00	1.84	1.92		1.13	1.11
{IS 18551 × Swarna} × IS	3 18551	BC_1		1.00	1.00	1.72	1.84		1.27	1.50
$\{IS\ 18551 \times Swarna\} \times S$		BC_2		1.00	1.00	2.00	1.99		1.00	1.00



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Table 2 continued

Pedigree	Generation	Grain co	olor	Grain lusti	e	Awns	
		2013R	2013PR	2013R	2013PR	2013R	2013PR
Swarna	P_2	1.00	1.00	2.00	2.00	1.00	1.00
Mean		_	_	1.76	1.8	1.2	1.30
SE±		_	_	0.01	0.01	0.02	0.02
Vr		_	_	789.75**	1110.02**	281.83**	295.43**
LSD (P 0.05)		-	-	0.04	0.04	0.07	0.07

NS non-significant

across seasons. Inflorescence exsertion showed significant differences between the parents of the two crosses across seasons, except for M 35-1 \times ICSV 700 during the rainy season. Leaf glossy score, leafsheath pigmentation and plant vigor score, which were recorded during postrainy season, exhibited significant differences between the parents for both the crosses, except leaf glossy score, which was non-significant for the cross M 35-1 \times ICSV 700. The traits that had shown non-significant differences between the generations were not subjected to generation mean analysis.

The per se performance of the parents and their generations are given in Tables 1 and 2. The order of the generations presented in the tables is as follows P₁, F₁, F₂, BC₁, BC₂, and P₂; where P₁ and P₂ were the female and male parents, respectively. The performances of different generations is discussed below.

Shoot fly deadhearts

The parents of the cross IS $18551 \times S$ warna exhibited significant differences for percentage shoot fly deadhearts in the postrainy season (Table 1). The susceptible parent, Swarna suffered greater shoot fly damage (54.31%) as compared to the resistant parent, IS 18551 (3.60%). The progenies F_1 and BC_1 exhibited lower shoot fly deadhearts (17.68 and 16.22%, respectively), and were nearer to the resistant parent, IS 18551; whereas F_2 and BC_2 generations exhibited higher numbers of deadhearts (24.27 and 38.26%, respectively), and were closer to the susceptible parent, Swarna.

Days to 50% flowering

Days to 50% flowering exhibited significant differences between the parents across seasons (Table 1). Both the crosses flowered at the same time with a mean flowering period of 71.98 \pm 0.97 and 71.70 \pm 0.78 for the cross M 35-1 \times ICSV 700, and 69.69 \pm 0.85 and 67.70 \pm 0.67 days for the cross IS 18551 \times Swarna, respectively, during the rainy and postrainy seasons. In both the crosses, the mean performances of the progenies were on par with the early flowering parent.

Plant height

The cross M 35-1 \times ICSV 700 exhibited nonsignificant differences between the generations for plant height because of equal height attained by the parents, and their progenies across seasons (Table 1). The cross IS 18551 × Swarna showed significant performance differences, with a mean 273.96 ± 3.90 and 194.50 ± 7.96 cm, respectively, in the rainy and postrainy seasons. The F₁, F₂, and BC₁ exhibited plant height towards IS 18551, while the BC₂ exhibited moderate plant height. In both the crosses, the parents attained different heights across the seasons, with longer plants in the rainy season.

100 seed weight

There were significant differences between the generations for 100 seed weight (Table 1). In the cross M $35\text{-}1 \times ICSV~700$, F_1 has attained highest 100 seed weight of 3.12 and 3.96 g in the rainy and postrainy seasons, respectively. The remaining generations had



^{*, **}F test significant at P 0.05 and 0.01, respectively; R, rainy season; PR, postrainy season

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higher 100 seed weight than the parent M 35-1 (2.31 and 3.56 g respectively, in the rainy and postrainy seasons). In the cross IS $18551 \times \text{Swarna}$ the female parent IS 18551 recorded lower 100 seed weight of 1.57 and 2.21 g than the male parent Swarna 3.00 and 3.36 g respectively, in the rainy and postrainy seasons. The other generations recorded 100 seed weight towards the male parent, Swarna.

Grain yield

Significant differences were observed between the parents for grain yield across seasons (Table 1). In M 35-1 \times ICSV 700, the per se performance of M 35-1 was 2.51 and 6.56 t/ha, and of ICSV 700 1.03 and 5.68 t/ha, respectively, in the rainy and postrainy season. The per se performance of the F_1 's was greater than the better parent. The other generations also recorded more grain yield, tilting towards the parent with high grain yield.

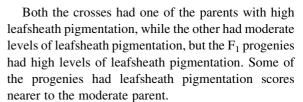
Waxy bloom and plant color

There were no significant differences in waxy bloom among the parents of the cross M 35-1 \times ICSV 700 across seasons (Table 1), but both the parents in the cross IS 18551 \times Swarna exhibited significant differences for waxy bloom across seasons. The progenies had greater amounts of waxy bloom than the female parent, IS 18551.

The parents of the cross IS 18551 \times Swarna were nontan type and their progenies also showed the nontan plant color (Table 1), but in the cross M35-1 \times ICSV 700, the parent M 35-1 was non tan, and ICSV 700 was tan type and the F_1 progenies of their cross were non-tan.

Leaf glossiness, leafsheath pigmentation and plant vigor

These traits were recorded only in the postrainy season, and there were significant differences between the parents, except in leaf glossy score of the cross M $35\text{-}1 \times ICSV$ 700, as both these parents were glossy (Table 1). In the cross IS $18551 \times Swarna$ the parent IS 18551 was highly glossy, while Swarna was nonglossy. The F_1 and F_2 progenies of these two parents was moderately glossy, with a score of 3.00 and 3.06, respectively.



One of the parents had high plant vigor (1.00), whereas the other exhibited moderate vigor (2.00). The F_1 progenies had a vigor score of 2.00.

Inflorescence exsertion

The two crosses exhibited significant differences between the parents for inflorescence exsertion across seasons, except M 35-1 \times ICSV 700 cross in the rainy season (Table 2). The F_1 progenies in the cross IS 18551 \times Swarna exhibited good panicle exsertion, while the F_1 progenies of the cross M 35-1 \times ICSV 700 did not show any particular trend.

Panicle compactness

Both the crosses did not differ significantly in panicle compactness in the rainy season, but differed significantly in panicle compactness in the postrainy season (Table 2). In the cross IS $18551 \times Swarna$, IS 18551 had a compact panicle (3.00), while Swarna had a semiloose panicle (2.00). The F_1 progenies had semiloose panicles (2.00).

Panicle shape

Panicle shape was recorded only in the postrainy season. The parents did not differ in panicle shape in the cross M 35-1 \times ICSV 700 (Table 2). The parents in the cross IS 18551 \times Swarna had the contrasting panicle shapes. Swarna had erect panicle with score of 1.00, whereas IS 18551 had elliptical panicle with a score of 4.00. Their F_1 s had erect panicle with a score of 1.00.

Glume color and glume coverage of the grain

Parents differed significantly in glume color and glume coverage of the grain (Table 2). In the cross M 35-1 \times ICSV 700, M 35-1 was red glumed and ICSV 700 was mahogany colored. Their F₁ progenies exhibited red glume color, indicating the dominance nature of red glumes across seasons.



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M 35-1 had 25% and ICSV 700 had 50% of the grain covered with glumes. F_1 progenies were with 25% of the grain covered with the glume, indicating dominant nature of glume covering of the grain in this cross. In the cross IS 18551 \times Swarna, IS 18551 grain were fully covered by the glumes, while in Swarna 25% of the grain was covered with glume, and the F_1 progenies were with 50 to 75% glume coverage.

Grain lustre and awns

There were no significant differences between the parents of the cross M 35-1 \times ICSV 700 for grain lustre and awns (Table 2). In the cross IS 18551 \times Swarna, the cross of non-lustrous and lustrous seed trait generated F_1 progenies with lustrous seed, indicating the dominance nature of the gene controlling grain lusture.

Awns were present in IS 18551, but absent in Swarna, F_1 progenies were awnless, indicating the recessive nature of the awned gene.

Gene effects and genetic parameters

The replicated data obtained from six generations of the two cross combinations for agronomic, morphological and panicle traits were subjected to generation mean analysis using scaling tests to test the fitness of additive—dominance model, and Hayman's six parameter model to find the significant inter-allelic interactions. Only the traits that showed significant F values were included for generation mean analysis and explained hereunder.

Leaf glossiness

The F value for leaf glossy score was non-significant for the cross M 35-1 \times ICSV 700. A, B, and D scales were significant for leaf glossy score in the cross IS 18551 \times Swarna, indicating the presence of non-allelic interactions for this trait (Tables 3, 4). Partitioning of the generation means showed significant mean (m) and additive (d), dominance (h), additive \times additive (i), and dominance \times dominance (l) components. The dominance and dominance \times dominance components were in opposite direction, which suggested the presence of duplicate epistasis. The dominance variance was greater than the additive variance, indicating the predominance of

dominance gene effects (Table 5). The narrow sense heritability was low and the dominance degree was negative.

Leafsheath pigmentation

The scales A, B, and C for the cross M 35-1 \times ICSV 700, and all scales for the cross IS 18551 \times Swarna were significant in the postrainy season (Tables 3, 4). The significance of the scales indicated the presence of non-allelic interactions. Generation means partitioned into six components using Hayman's method revealed the significance of mean for both the crosses, with significant additive and dominance \times dominance interactions for the cross M 35-1 \times ICSV 700; and significant dominance and additive \times additive components for IS 18551 \times Swarna. The variance due to dominance was greater than the additive variance in both the crosses, with higher degree of dominance (Table 5).

Plant vigor

The scales A and C for the cross M $35-1 \times ICSV 700$, and all the scales for IS 18551 × Swarna were significant in the postrainy season (Tables 3, 4), indicating the inadequacy of additive-dominance model and presence of non-allelic interactions. Partitioning the generation means showed significance of mean for both the crosses. The dominance component was significant for the cross M 35-1 \times ICSV 700, and the dominance, dominance × dominance, and additive x additive components were significant for the cross IS 18551 × Swarna, in which the dominance and dominance × dominance components were in opposite direction, indicating the presence of duplicative epistasis. The dominance variance was high in both the crosses, which indicated the predominance of dominance gene action (Table 5).

Days to 50% flowering

M 35-1 \times ICSV 700 exhibited significant scales of C and D for this trait, and the scales A, B, and D were significant for the cross IS 18551 \times Swarna, indicating the inadequacy of simple additive–dominance model and the presence of epistatic interactions in the rainy season (Tables 3, 4). The partitioning of generation means and estimation of the genetic components



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Table 3 Scaling tests and genetic components (Hayman's 6 parameter model) for various agronomic and morphological traits of M 35-1 × ICSV 700 in sorghum across seasons (ICRISAT, Patancheru, 2013–2014)

	Leafsheath pigmentation Plant vigor score		Days to 50% flowering	ring	100 seed weight		Grain yield	
	2013PR	2013PR	2013R	2013PR	2013R	2013PR	2013R	2013PR
Scales	Se							
A	$1.56 \pm 0.15**$	$-0.58 \pm 0.19**$	-1.37 ± 1.36	-0.61 ± 1.16	-0.04 ± 0.14	$0.42 \pm 0.16**$	0.79 ± 0.82	$4.90 \pm 0.99**$
В	$0.91 \pm 0.15**$	0.01 ± 0.12	-2.40 ± 1.52	-1.63 ± 1.28	$-0.37 \pm 0.14**$	$0.49 \pm 0.14**$	0.90 ± 0.60	$3.32 \pm 0.97**$
Ŋ	$2.27 \pm 0.18**$	$-0.39 \pm 0.19*$	$-8.31 \pm 2.10**$	-1.54 ± 1.59	-0.55 ± 0.24 *	$0.64 \pm 0.19**$	$-2.41 \pm 0.82 **$	-0.88 ± 1.04
Q	-0.11 ± 0.14	0.09 ± 0.12	$-2.27 \pm 1.17*$	0.35 ± 0.92	-0.07 ± 0.08	$-\ 0.13\pm0.11$	$-2.05 \pm 0.52 **$	$-4.55 \pm 0.80**$
Gene	Genetic components							
Ш	$1.57 \pm 0.04**$	$1.57 \pm 0.04**$	$69.82 \pm 0.40**$	$71.15 \pm 0.27**$	$2.54 \pm 0.03 **$	$3.64 \pm 0.03**$	$2.87 \pm 0.13 **$	$8.96 \pm 0.23**$
р	$0.32 \pm 0.11**$	0.04 ± 0.09	-1.483 ± 0.86	-0.83 ± 0.75	$0.25 \pm 0.06 **$	$0.53 \pm 0.09**$	0.68 ± 0.45	1.23 ± 0.66
h	0.21 ± 0.27	0.48 ± 0.24 *	-4.32 ± 2.44	$-5.90 \pm 1.93**$	$1.03 \pm 0.20 **$	$1.24 \pm 0.23 **$	$7.50 \pm 1.08 **$	$15.23 \pm 1.61**$
1	$-2.69 \pm 0.46**$	0.76 ± 0.41	-0.78 ± 4.03	2.94 ± 3.38	0.28 ± 0.34	$-1.18 \pm 0.41**$	$-5.78 \pm 1.97**$	$-17.33 \pm 2.82**$
i	0.21 ± 0.27	-0.19 ± 0.23	4.54 ± 2.35 *	-0.70 ± 1.84	0.13 ± 0.17	0.27 ± 0.22	$4.09 \pm 1.03 **$	$9.11 \pm 1.59**$
j	0.32 ± 0.11	-0.30 ± 0.11	0.52 ± 0.93	0.51 ± 0.76	0.17 ± 0.07	-0.04 ± 0.09	-0.05 ± 0.46	0.79 ± 0.68

m mean, d additive gene action, h dominant gene action, l dominance x dominance gene interaction, i additive x additive gene interaction, j additive x dominant gene interaction

*, **t test significant at P 0.05 and 0.01, respectively; R, rainy season; PR, postrainy season



Table 4 Scaling tests and genetic components (Hayman's 6 parameter model) for various agronomic and morphological traits of 1S 18551 × Swarna in sorghum across seasons (ICRISAT, Patancheru, 2013–2014)

		<i>(</i>					
	Leaf glossy score	Leafsheath pigmentation	Plant vigor score	Days to 50% flowering	gı	Plant height	
	2013PR	2013PR	2013PR	2013R	2013PR	2013R	2013PR
Scales							
A	$1.35 \pm 0.26 **$	$0.95 \pm 0.15**$	$0.39 \pm 0.12**$	$-3.41 \pm 1.14**$	-1.69 ± 1.27	$-31.33 \pm 7.30**$	-0.22 ± 8.28
В	$0.36 \pm 0.17*$	$0.48 \pm 0.19*$	$-0.34 \pm 0.13**$	$7.67 \pm 1.28**$	-0.38 ± 1.18	$23.07 \pm 10.31*$	$42.22 \pm 8.41 **$
C	0.24 ± 0.25	$2.13 \pm 0.22**$	$-0.97 \pm 0.15**$	-1.85 ± 1.42	$-4.53 \pm 1.76**$	2.17 ± 9.82	$42.33 \pm 13.40**$
О	$-0.74 \pm 0.20**$	$0.35 \pm 0.14**$	$-0.51 \pm 0.12**$	$-3.05 \pm 1.00**$	-1.23 ± 1.08	5.22 ± 7.13	0.17 ± 5.25
Genetic	Genetic components						
ш	$3.06 \pm 0.06 **$	$1.70 \pm 0.05 **$	$1.51 \pm 0.04**$	$68.29 \pm 0.29 **$	$66.68 \pm 0.37**$	$280.54 \pm 1.99**$	$203.58 \pm 1.69**$
p	$-1.51 \pm 0.16**$	-0.10 ± 0.11	-0.13 ± 0.09	1.49 ± 0.80	$6.51 \pm 0.79**$	$50.80 \pm 5.90 **$	$18.78 \pm 4.01 **$
h	$1.47 \pm 0.40**$	$-1.04 \pm 0.29**$	$1.52 \pm 0.23**$	-1.80 ± 2.04	-0.90 ± 2.21	$54.90 \pm 14.54 **$	$44.33 \pm 11.98**$
1	$-3.19 \pm 0.67**$	-0.72 ± 0.48	$-1.07 \pm 0.39**$	$-10.36 \pm 3.51**$	-0.40 ± 3.60	18.70 ± 25.56	-41.67 ± 20.90 *
i	$1.47 \pm 0.40**$	$-0.71 \pm 0.28**$	$1.02 \pm 0.23**$	$6.10 \pm 1.99**$	2.47 ± 2.16	-10.43 ± 14.25	-0.33 ± 10.50
j	0.50 ± 0.16	0.24 ± 0.12	0.37 ± 0.09	-5.54 ± 0.82	-0.66 ± 0.82	-27.20 ± 6.07	-21.22 ± 4.49
		100 seed weight			Grain yield		
		2013R	2013PR		2013R		2013PR
Scales							
Ą		$0.38 \pm 0.11**$	0.17 ± 0.11	0.11	0.67 ± 0.37	7	$3.45 \pm 0.77**$
В		0.22 ± 0.13	$0.72 \pm 0.15**$	0.15**	-0.94 ± 0.57	7	$3.21 \pm 0.75**$
C		$-0.49 \pm 0.18**$	$0.56 \pm 0.16**$	0.16**	$-4.38 \pm 0.50**$	**0	$3.04 \pm 0.94**$
D		$-0.54 \pm 0.08**$	-0.17 ± 0.10	0.10	-2.05 ± 0.34 **	* * *	-1.81 ± 0.66
Genetic	Genetic components						
В		$2.39 \pm 0.03**$	$3.12 \pm 0.03**$	0.03**	$2.55 \pm 0.09**$	**6	$7.93 \pm 0.21**$
p		$-0.64 \pm 0.06**$	$-0.85 \pm 0.08**$	**80.0	$-0.88 \pm 0.30**$	**0	0.69 ± 0.51
h		$1.53 \pm 0.17**$	$0.73 \pm 0.21 **$	0.21**	$5.64 \pm 0.71**$	**	$9.54 \pm 1.34**$
1		$-1.68 \pm 0.29**$	$-1.22 \pm 0.36**$	0.36**	$-3.83 \pm 1.30**$	**0	$-10.28 \pm 2.25**$
i		$1.09 \pm 0.15**$	0.33 ± 0.20	0.20	$4.11 \pm 0.69**$	**6	$3.62 \pm 1.32**$
j		0.08 ± 0.07	-0.27 ± 0.09	60.00	0.81 ± 0.33	3	0.12 ± 0.53

m mean, d additive gene action, h dominant gene action, l dominance × dominance gene interaction, i additive × additive gene interaction, j additive × dominant gene interaction

*, **/r test significant at P 0.05 and 0.01, respectively; R, rainy season; PR, postrainy season



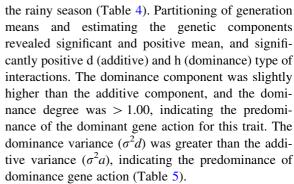
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revealed highly significant positive mean values in both the crosses. In the rainy season, M 35-1 \times ICSV 700 showed significant i type of interaction i.e., additive × additive interactions were significant and positive in the cross IS 18551 × Swarna; both i (additive \times additive) and l (dominance \times dominance) type of interactions were significant. The additive and dominance variances were also calculated and the estimates revealed that the additive variance $(\sigma^2 a)$ was greater than the dominance variance $(\sigma^2 d)$ in M 35-1 × ICSV 700, whereas IS 18551 × Swarna exhibited higher dominance variance $(\sigma^2 d)$ than the additive variance (Table 5). The narrow sense heritability for M 35-1 × ICSV 700 was moderate (0.45), indicating the presence of additive nature of gene action, whereas in the cross IS 18551 \times Swarna, narrow sense heritability was negative indicating dominance gene action.

In the postrainy season, the scales were nonsignificant for the cross M $35-1 \times ICSV 700$, indicating that additive dominance model explained the inheritance of this trait, whereas for the cross IS 18551 × Swarna, the scale C was significant, indicating the presence of non-allelic interactions, and inadequacy of additive dominance model (Tables 3, 4). Therefore, a six parameter model was adopted to test the presence of non-allelic interactions. Partitioning of the generation means and estimation of the genetic components revealed significance of mean (m) for the cross IS 18551 × Swarna, in which the additive component was significant, indicating the predominance of additive nature of gene action. The narrow sense heritability was 0.32, which was quite low, and the degree of dominance was negative (Table 5). The dominance variance $(\sigma^2 d)$ was greater than the additive variance $(\sigma^2 a)$ for both the crosses. The broadsense heritability was high (0.85) for the cross M 35-1 × ICSV 700, indicating the environmental influence on the expression of this trait.

Plant height

The F value for plant height in the cross M $35-1 \times ICSV$ 700 was non-significant, and hence, this cross was not considered for generation mean analysis across seasons. The cross IS $18551 \times Swarna$ exhibited significant A and B scales for plant height, indicating the inadequacy of simple additive–dominance model and presence of epistatic interactions in



In the postrainy season, the cross IS 18551 \times Swarna exhibited significant B and C scales for plant height (Table 4), indicating the inadequacy of additive dominance model in explaining the inheritance of this trait. Partitioning of generation means revealed positive and significant mean (m), and significant addidominance. and dominance × dominance interactions. The dominance and dominance × dominance interactions were with opposite signs, indicating duplicate epistasis. The narrow sense heritability was negative, while the broad sense heritability was moderate (0.59) with high degree of dominance (> 1.00), indicating over-dominance type of gene action (Table 5).

100 seed weight

The scales B and C for the cross M $35-1 \times ICSV 700$, and A, C and D's for the cross IS 18551 × Swarna were significant, indicating the presence of non-allelic interactions for this trait in the rainy season (Tables 3, 4). To identify the type of interactions present, the generation means were partitioned into six components, and the mean (m) for both the crosses was positive and highly significant. Both additive and dominant components were significant in both the crosses, and l and i type of interactions were significant for the cross IS 18551 × Swarna. Dominant component, and dominance × dominance interactions exhibited opposite sign for the cross IS 18551 × Swarna, indicating the presence of duplicate epistasis. The narrow sense heritability was 0.42 and 0.26, respectively, in the crosses M 35-1 \times ICSV 700 and IS 18551 × Swarna (Table 5). Variation in narrow sense heritability estimates might be because of differences in the parents involved in these crosses. The dominance degree for this trait was > 1.00 for the cross M 35-1 × ICSV 700, indicating the over-



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dominance nature of gene effects for this trait, and negative dominance degree was observed for the cross IS $18551 \times Swarna$.

In the postrainy season, 100 seed weight exhibited significant A, B, and C scales for the cross M 35-1 \times ICSV 700, and significant B and C scales for the cross IS 18551 × Swarna, indicating the presence of nonallelic interactions (Tables 3, 4). Partitioning of generation means into six components by Hayman's method revealed significant and positive means for both the crosses. The additive, dominance, dominance \times dominance components for M 35-1 \times ICSV 700, and dominance, dominance × dominance, and additive \times additive components for IS 18551 \times Swarna were significant for this trait. In both the crosses, dominance and dominance x dominance components exhibited opposite signs, indicating the presence of duplicate epistasis for this trait. The narrow sense heritability was negative in both the crosses, with high broad sense heritabilities (Table 5). The dominance degree was > 1.00 for the cross M $35-1 \times ICSV$ 700 and IS $18551 \times Swarna$, exhibiting negative dominance.

Grain yield

Scaling test for grain yield revealed that the scales C and D were significant for both the crosses in the rainy season, indicating the presence of non-allelic interactions (Tables 3, 4). Partitioning of generation means into the six components revealed positive and highly significant means (m) for both the crosses. The dominance and additive components were significant for the cross IS 18551 \times Swarna with significant l and i type of interactions. Whereas for the cross M 35-1 \times ICSV 700, the additive component (d) was nonsignificant, but dominance and l and i type of interactions were significant. The dominance component and dominance × dominance interaction was in opposite direction, indicating the presence of duplicate epistasis. The estimates of additive × additive (i) gene interactions was greater than dominance × dominance (l) interactions, suggesting predominance of additive gene action. The dominance variance was greater than the additive variance for both the crosses (Table 5). Narrow sense heritability was very low and negative in both the crosses. The dominance degree was negative for IS 18551 × Swarna, but positive and > 1.00 for M 35-1 \times ICSV 700 cross, indicating over dominance nature of gene action in this cross.

M 35-1 \times ICSV 700 cross exhibited significant A, B, and D scales, while A, B, and C scales in the postrainy season for the cross IS 18551 × Swarna indicated the presence of non-allelic interactions for this trait (Tables 3, 4). Partitioning of the generation means by Haymans's six parameter model revealed significant and positive mean (m), and dominance, dominance × dominance, and additive × additive components were significant for both the crosses. The dominance and dominance × dominance components exhibited opposite signs, indicating the presence of duplicate epistasis for grain yield in the postrainy season. The narrow sense heritability was low, but broad sense heritability was high, indicating the environmental influence for this trait in both the crosses (Table 5). The dominance variance ($\sigma^2 d$) was high for both the crosses, indicating the predominance of the dominant gene action. The narrow sense heritability was lower and negative with high degree (> 1.00) of dominance, indicating over-dominance nature of gene action.

Inflorescence exsertion

The F-value was non-significant for the cross M 35-1 × ICSV 700 in the rainy season, and hence, excluded from analysis. All the scales were significant for inflorescence exsertion in the cross IS 18551 \times Swarna, indicating the presence of non-allelic interactions, and inadequacy of additive dominance model in explaining the inheritance of this trait (Tables 5, 6, 7). The Hayman's six component analysis revealed significant mean (m) value, with a significant additive component. The dominance × dominance and additive x additive gene interactions were also significant. The estimate for additive × additive component was greater than the dominance × dominance component, indicating predominance of additive gene action. The narrow sense heritability was negative (-0.37), but broad sense heritability (0.79) was high, the dominance degree being 1.15, indicating overdominance gene action (Table 8).

In the postrainy season, the scale C for the cross M $35-1 \times ICSV$ 700, and all the scales for the cross IS $18551 \times Swarna$ were significant, indicating inadequacy of the additive dominance model and presence of non-allelic interactions for inflorescence exsertion



Table 5 Estimates of various genetic parameters for different agronomic and morphological traits of two crosses of sorghum across seasons (ICRISAT, Patancheru, 2013–2014)

		Leafsheath pigmentation	Plant vigor score	ore	Days to 5	Days to 50% flowering	ьs	100 s	100 seed weight	ıţ	Grai	Grain yield	
	2013PR	×	2013PR		2013R	2013PR	3PR	2013R		2013PR	2013R	IR.	2013PR
Cross M 35-1 \times ICSV 700	ICSV 700												
$\sigma^2 g$	0.57	7	0.33		24.24	16.	16.60	0.04	4	0.21	1.	1.94	11.53
$\sigma^2 a$	-0.03	~	- 0.06		13.03	-3.02	.02	0.06	. 9(-0.13	– 6.	6.46	- 7.75
$\sigma^2 d$	09.0		0.40		11.21	19.	19.62	-0.02	72	0.33	8	8.40	19.28
$\sigma^2 p$	0.57	7	0.39		28.71	19.	19.38	0.14	14	0.25	2.	2.77	12.27
$h_{ m b}^2$	I		0.85		0.84	0.	98.0	0.31	31	0.84	0.	0.70	0.94
$h_{\rm ns}^{-2}$	-0.04	_	-0.16		0.45	-0.16	.16	0.42		-0.50	_ 2.	2.34	-0.63
Dominance degree	e 0.80		3.56		1.71	2.	2.67	2.01)1	1.52	3.	3.31	3.52
Traits	Leaf glossy score	Leafsheath pigmentation	Plant vigor score	Days to 50% flowering	50%	Plant height	ıt.	100 seed	100 seed weight	Grain yield	ield	Waxy bloom	шоо
	2013PR	2013PR	2013PR	2013R	2013PR	2013R	2013PR	2013R	2013PR	2013R	2013PR	2013R	2013PR
Cross IS 18551 × Swarna	× Swarna												
$\sigma^2 g$	1.18	0.59	0.41	19.90	31.13	883.17	406.57	0.09	0.17	1.33	10.17	0.24	0.29
$\sigma^2 a$	- 0.20	0.14	-0.03	-6.14	10.64	-691.70	-72.46	0.04	-0.13	-3.30	-0.31	0.18	0.28
$\sigma^2 d$	1.38	0.45	0.43	26.04	20.49	1574.86	479.03	90.0	0.30	4.63	10.47	0.05	0.01
$\sigma^2 p$	1.18	0.65	0.41	21.17	33.13	959.12	687.52	0.15	0.20	1.72	10.62	0.36	0.29
$h_{ m b}^2$	1.00	0.91	1.00	0.94	0.94	0.92	0.59	0.64	0.84	0.78	96.0	0.67	1.00
$h_{ m ns}^2$	-0.17	0.21	- 0.07	-0.29	0.32	-0.72	-0.11	0.26	-0.68	-1.92	-0.03	0.52	0.98
Dominance degree	66.0 –	3.30	- 3.38	- 1.10	-0.37	1.04	1.54	- 1.55	- 0.93	- 2.53	3.72	- 0.89	- 0.38

 $\sigma^2 g$ genotypic variance, $\sigma^2 a$ additive variance, $\sigma^2 d$ dominance variance, $\sigma^2 p$ phenotypic variance, h_b^2 broadsense heritability, h_{ns}^2 narrowsense heritability; R, rainy season; PR, postrainy season



Table 6 Scaling tests and genetic components (Hayman's 6 parameter model) for panicle traits of M 35-1 × ICSV 700 in sorghum across seasons (ICRISAT, Patancheru, 2013–2014)

	Inflorescence exsertion	Panicle compactness	Panicle shape	Glume color		Glume coverage	
	2013PR	2013PR	2013PR	2013R	2013PR	2013R	2013PR
Scales							
A	-0.20 ± 0.17	-0.03 ± 0.03	-0.08 ± 0.08	$-0.43 \pm 0.11**$	$-0.37 \pm 0.09**$	$0.53 \pm 0.18**$	I
В	0.17 ± 0.15	$-0.11 \pm 0.05*$	$-0.32 \pm 0.16*$	ı	$-0.44 \pm 0.10**$	0.40 ± 0.43	$-1.20 \pm 0.19**$
C	$-0.39 \pm 0.17*$	$-0.58 \pm 0.09**$	$-1.69 \pm 0.25**$	$-0.38 \pm 0.17*$	0.04 ± 0.13	0.09 ± 0.44	$-0.90 \pm 0.17**$
О	-0.18 ± 0.14	$-0.22 \pm 0.05 **$	$-0.64 \pm 0.16**$	0.03 ± 0.13	$0.43 \pm 0.10**$	-0.42 ± 0.27	0.15 ± 0.13
Genetic	Genetic components						
ш	$2.40 \pm 0.04**$	$2.86 \pm 0.02 **$	$3.58 \pm 0.06 **$	$2.66 \pm 0.04**$	$2.76 \pm 0.03**$	$1.69 \pm 0.09**$	$1.27 \pm 0.04**$
р	-0.19 ± 0.11	0.04 ± 0.03	0.12 ± 0.09	$0.28 \pm 0.09**$	$0.53 \pm 0.07**$	$-1.27 \pm 0.19**$	$-0.40 \pm 0.09**$
h	$1.36 \pm 0.29**$	$0.44 \pm 0.10**$	$1.29 \pm 0.31 **$	0.44 ± 0.25	-0.35 ± 0.19	-0.49 ± 0.55	$-1.30 \pm 0.25**$
1	-0.33 ± 0.49	$-0.31 \pm 0.15*$	$-0.89 \pm 0.43*$	0.49 ± 0.42	$1.66 \pm 0.31 **$	$-1.78 \pm 0.90*$	$1.50 \pm 0.41 **$
i	0.36 ± 0.29	$0.44 \pm 0.10**$	$1.29 \pm 0.31 **$	-0.06 ± 0.25	$-0.85 \pm 0.19**$	0.84 ± 0.53	-0.30 ± 0.25 *
j	-0.19 ± 0.11	0.04 ± 0.03	0.12 ± 0.09	-0.22 ± 0.10	0.03 ± 0.07	0.07 ± 0.23	0.00 ± 0.09

m mean, d additive gene action, h dominant gene action, l dominance x dominance gene interaction, i additive x additive gene interaction, j additive x dominant gene interaction

*, **t test significant at P 0.05 and 0.01, respectively; R, rainy season; PR, postrainy season



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Table 7 Scaling tests and genetic components (Hayman's 6 parameter model) for various morphological and panicle traits of IS 18551 × Swarna in sorghum across seasons (ICRISAT, Patancheru, 2013–2014)

	Waxy bloom		Inflorescence exsertion	sertion	Panicle compactness	Panicle shape Glume color	Glume color		Glume coverage	
	2013R	2013PR	2013R	2013PR	2013PR	2013PR	2013R	2013PR	2013R	2013PR
Scales										
Ą	A $-0.81 \pm 0.18**$	$-0.51 \pm 0.10**$	$0.48 \pm 0.19**$	$0.47 \pm 0.16**$	$**60.0 \pm 09.0$	$1.80 \pm 0.25 **$	$0.64 \pm 0.22 **$	$0.73 \pm 0.24**$	$-1.23 \pm 0.58*$	$2.67 \pm 1.16*$
В	$-~0.2~\pm~0.17$	$-0.18 \pm 0.06**$	$0.72 \pm 0.13**$	$0.64 \pm 0.13**$	$0.71 \pm 0.10**$	$2.20 \pm 0.30 **$	$-\ 0.12\pm0.35$	$-0.67 \pm 0.14**$	$-2.40 \pm 0.30 **$	0.22 ± 0.43
Ü	$C - 1.38 \pm 0.30**$	$-0.28 \pm 0.14*$	$0.58 \pm 0.19**$	$1.78 \pm 0.21**$	$0.85 \pm 0.13**$	$2.68 \pm 0.38 **$	0.45 ± 0.35	0.22 ± 0.26	0.13 ± 0.62	$5.17 \pm 1.31**$
Ω	$-~0.19~\pm~0.11$	$0.20 \pm 0.09*$	$-0.31 \pm 0.12**$	$0.34\pm0.15*$	$-0.23 \pm 0.09**$	-0.66 ± 0.27 *	-0.04 ± 0.27	0.08 ± 0.19	$1.88 \pm 0.45 **$	$1.14 \pm 0.46**$
Geneti	Genetic components									
ш	$1.99 \pm 0.04**$	$2.68 \pm 0.04**$	$1.31 \pm 0.03**$	$1.70 \pm 0.05**$	$2.46 \pm 0.03 **$	$2.42 \pm 0.10**$	$3.86 \pm 0.09 **$	$3.80 \pm 0.07 **$	$5.03 \pm 0.16**$	$4.96 \pm 0.17**$
p	$-1.31 \pm 0.08**$	$-0.67 \pm 0.06**$	$0.21\pm0.10*$	$0.41 \pm 0.10**$	$0.44 \pm 0.07**$	$1.30 \pm 0.20 **$	$-1.12 \pm 0.21**$	0.20 ± 0.14	$4.59 \pm 0.33 **$	$3.89 \pm 0.31**$
ч	$1.04 \pm 0.26**$	0.09 ± 0.18	0.28 ± 0.24	$-1.17 \pm 0.29**$	-0.04 ± 0.19	$-~0.18\pm~0.55$	$-1.43 \pm 0.54**$	0.35 ± 0.38	$-3.76 \pm 0.90 **$	$-2.28 \pm 1.07*$
1	0.64 ± 0.45	$1.09 \pm 0.27**$	$-1.82 \pm 0.42**$	-0.44 ± 0.46	$-1.77 \pm 0.29**$	$-5.32 \pm 0.88**$	-0.59 ± 0.89	0.08 ± 0.62	$7.39 \pm 1.45 **$	-0.611 ± 1.8
i	0.37 ± 0.23	$-0.41 \pm 0.18*$	$0.62 \pm 0.23**$	$-0.67 \pm 0.29*$	$0.46 \pm 0.19**$	$1.32 \pm 0.55 *$	0.07 ± 0.54	$-\ 0.15\pm0.38$	$-3.76 \pm 0.90 **$	$-2.28 \pm 0.91*$
j	-0.31 ± 0.08	-0.17 ± 0.06	$-\ 0.12 \pm 0.11$	-0.09 ± 0.10	-0.06 ± 0.07	-0.20 ± 0.20	0.38 ± 0.21	0.70 ± 0.14	0.59 ± 0.33	1.22 ± 0.59

m mean, d additive gene action, h dominant gene action, l dominance \times dominance gene interaction, i additive \times additive \times additive \times additive \times dominant gene interaction

*, **t test significant at P 0.05 and 0.01, respectively; R, rainy season; PR, postrainy season



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(Tables 6, 7). Partitioning of the generation means using the six parameter model revealed significant and positive mean (m). The dominance (h) component was significant for the cross M 35-1 × ICSV 700, and the additive, dominance and additive × additive gene interactions were significant in the cross IS 18551 × Swarna. The additive component was greater than the dominance component, indicating the predominance of additive gene action for this trait in the cross IS 18551 × Swarna. The narrow sense heritability was low, while the dominance degree was negative in both the crosses (Table 8).

Waxy bloom

The scaling test for this trait in the cross IS $18551 \times S$ Swarna exhibited significant A and C scales in the rainy season (Table 7), indicating the presence of non-allelic interactions, and inadequacy of additive-dominance model in explaining the inheritance of this trait. Partitioning of the generation means into the six

components revealed positive and highly significant mean (m). The dominance and additive components were also significant. The dominance component was high as compared to the additive component, but the additive variance was greater than the dominance variance. The trait exhibited moderate narrow sense heritability (0.52), and dominance degree was negative (Table 8).

The F-value of this trait was non-significant for the cross M 35-1 \times ICSV 700 in the postrainy season, and hence, not included for calculating the generation means. All the scales were significant for the cross IS 18551 \times Swarna, indicating inadequacy of additive—dominance model in explaining the inheritance of this trait, and presence of non-allelic interactions (Table 7). Partitioning of the generation means into six parameter model revealed significant mean (m), while the components additive, dominance \times dominance, additive \times additive were also significant. The additive variance was high (0.28), and the narrow sense heritability (0.98) was also high (Table 8).

Table 8 Estimates of various genetic parameters for different panicle traits of two crosses of sorghum across seasons (ICRISAT, Patancheru, 2013–2014)

Traits	Inflorescer	ice exsertion	Panicle compactness	Panicle shape	Glume o	color	Glume o	coverage
	2013PR		2013PR	2013PR	2013R	2013PR	2013R	2013PR
Cross M 35-1 × ICS	SV 700							
$\sigma^2 g$	0.51		0.12	1.09	0.31	0.30	1.25	0.51
$\sigma^2 a$	0.04		0.18	1.60	0.05	0.23	0.70	0.36
$\sigma^2 d$	0.47		- 0.06	- 0.51	0.26	0.06	0.56	0.14
$\sigma^2 p$	0.51		0.12	1.09	0.31	0.30	1.49	0.51
$h_{ m b}^2$	_		_	_	1.00	1.00	0.84	1.00
$h_{ m ns}^2$	0.09		1.48	1.47	0.16	0.79	0.47	0.71
Dominance degree	-2.70		3.33	3.28	1.25	-0.81	0.62	1.80
Traits	Inflorescen	ce exsertion	Panicle compactness	Panicle shape	Glume o	color	Glume c	overage
	2013PR	2013PR	2013PR	2013PR	2013R	2013PR	2013R	2013PR
Cross IS 18551 × S	warna							
$\sigma^2 g$	0.22	0.69	0.25	2.19	1.85	1.04	5.76	2.49
$\sigma^2 a$	-0.11	0.47	0.11	0.87	0.56	0.32	3.52	5.01
$\sigma^2 d$	0.33	0.22	0.14	1.32	1.30	0.72	2.24	-2.52
$\sigma^2 p$	0.28	0.69	0.25	2.19	1.85	1.04	5.76	6.78
$h_{ m b}^2$	0.79	1.00	1.00	1.00	1.00	1.00	1.00	0.37
$h_{ m ns}^2$	- 0.37	0.68	0.42	0.40	0.30	0.31	0.61	0.74
Dominance degree	1.15	- 1.69	- 0.30	- 0.38	1.13	1.32	- 0.91	- 0.77

 $[\]sigma^2 g$ genotypic variance, $\sigma^2 a$ additive variance, $\sigma^2 d$ dominance variance, $\sigma^2 p$ phenotypic variance, h_b^2 broadsense heritability, h_{ns}^2 narrowsense heritability; R, rainy season; PR, postrainy season



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Panicle compactness

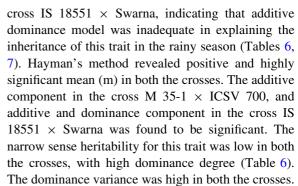
The F-value for panicle compactness was non-significant in the rainy season for both the crosses, and hence, excluded from the analysis. B, C, and D scales for M 35-1 \times ICSV 700 cross, and all the scales for the cross IS 18551 × Swarna were significant, indicating the presence of the non-allelic interactions in the inheritance of this trait in the postrainy season (Tables 6, 7). Partition of the generation means using the Haymans' six parameter model revealed significant and positive mean (m) for both the crosses. The components h, l, and i were significant for M 35-1 \times ICSV 700, indicating the presence of epistatic interactions. The dominant and dominant x dominant components were with opposite signs, indicating presence of duplicate epistasis in the postrainy season. IS $18551 \times \text{Swarna}$ showed significant mean (m), and significant d, l, and i components. High narrow sense heritabilities were observed for this trait in both the crosses, with high dominance degree (3.33) for M 35-1 × ICSV 700, but low and negative dominance degree for the cross IS $18551 \times \text{Swarna}$ (Table 8).

Panicle shape

The B, C, and D scales for the cross M 35-1 \times ICSV 700, and all the scales for IS 18551 × Swarna were significant in the postrainy season (Tables 6, 7). The significance of the scales indicated the presence of the non-allelic interactions. In order to know the type of interactions, the generation means were partitioned into six parameters, which revealed that the mean was significant for both the crosses. Dominance, dominance × dominance, and additive × additive components were significant for the cross M 35-1 \times ICSV 700, while additive, dominance × dominance, and additive × additive interactions were significant for the cross IS $18551 \times \text{Swarna}$. The dominance variance was high in the cross IS 18551 × Swarna, but high additive variance was recorded for the cross M 35-1 \times ICSV 700, with high dominance degree in the cross M 35-1 × ICSV 700, and negative dominance degree in the cross IS $18551 \times \text{Swarna}$ (Table 8).

Glume color

The A and C scales were significant for the cross M $35-1 \times ICSV 700$, while scale A was significant for the



The scales A, B, and D and A, B were significant, respectively, for the crosses M 35-1 × ICSV 700 and IS $18551 \times \text{Swarna}$ in the postrainy season (Tables 6, 7), indicating the presence of non-allelic interactions in the inheritance of this trait in the postrainy season. Partitioning of the generation means revealed the positive significant mean (m) in both the crosses. The dominance × dominance, tive × additive components were significant for the cross M 35-1 \times ICSV 700, and none of the components was significant for the cross IS $18551 \times Swarna$. The additive variance was greater for the cross M 35-1 × ICSV 700, while dominance variance was greater for the cross IS $18551 \times \text{Swarna}$ (Table 8). The cross M 35-1 × ICSV 700 exhibited higher narrow sense heritability (0.79) with negative dominance degree. The cross IS 18551 × Swarna showed lower narrow sense heritability with high dominance degree (1.32), indicating over dominance type of gene action.

Glume coverage

The scaling test for this trait in the rainy season revealed significant A scale for the cross M 35-1 × ICSV 700, and significant A and C scale for the cross IS 18551 × Swarna indicating non-allelic interactions, and inadequacy of additive-dominance model in explaining the inheritance of this trait (Tables 6, 7). Partitioning of the generation means into six components, and estimation of the genetic components revealed positive and significant means (m) for both the crosses. The additive and dominance × dominance interactions were significant for the cross M 35-1 \times ICSV 700, while in IS 18551 \times Swarna exhibited significant dominance and additive components, and significant dominance × dominance and additive × additive gene interactions. The dominance dominance × dominance interactions



opposite signs, indicating complementary gene action. The additive component was greater than the dominance component, whereas dominance \times dominance interaction was greater in magnitude than the additive \times additive gene interactions. The narrow sense heritability estimates were 0.45 and 0.61, respectively, for the crosses M 35-1 \times ICSV 700 and IS 18551 \times Swarna (Table 8). The dominance degree for the cross M 35-1 \times ICSV 700 was 0.62 indicating partial dominance, whereas the cross IS 18551 \times Swarna exhibited negative dominance degree.

In the postrainy season, glume coverage of the grain exhibited significant B, C scales for the cross M 35-1 × ICSV 700, while the cross IS 18551 × Swarna exhibited significant A, C, and D scales, indicating the presence of non-allelic interactions (Tables 6, 7). Partitioning of generation means using Hayman's six parameter model revealed significant and positive mean for both the crosses. The additive, dominance, dominance × dominance, and additive × additive interactions were significant for the cross M 35-1 \times ICSV 700, while additive, dominance, and additive × additive components were significant for IS 18551 × Swarna. The dominance and dominance x dominance components were opposite in sign, indicating the presence of duplicative epistasis for the cross M 35-1 \times ICSV 700. The additive variance was high for both the crosses, while the cross M 35-1 × ICSV 700 exhibited higher dominance degree, whereas the cross IS 18551 × Swarna exhibited negative dominance degree (Table 8).

Discussion

Sorghum shoot fly, A. soccata is an economically important pest that has a significant bearing on grain yield in sorghum. The present studies were aimed at understanding genetic parameters for different traits including the traits associated with shoot fly resistance using generation mean analysis to detect the major gene effects (additive and dominance), and their digenic (additive × additive, additive × dominance, and dominance × dominance) interactions for inheritance of quantitative traits (Kearsey and Pooni 1996). Generation mean analysis helps us in understanding the performance of the parents used in crosses, and the potential of crosses to be used either for heterosis exploitation or pedigree selection (Sharma and Sain

2003). Susceptibility/resistance of the progenies of the cross IS 18551 × Swarna, and the performance of the progenies indicated that one of the parents should have genes for shoot fly resistance (IS 18551) to develop shoot fly-resistant sorghums. The non-allelic interactions between the genes for component traits such as leafsheath pigmentation and plant vigor suggested that proper care should be taken while selecting the sorghum genotypes for the crossing program, based on these traits. Predominance of dominant gene action has been reported for the leaf glossiness, leafsheath pigmentation and plant vigor, whereas Riyazaddin et al. (2016) and Aruna et al. (2011) reported additive type of gene action for these traits. The results showed dominant nature of gene action for early flowering and early maturity. The cross between the tall and dwarf sorghum genotypes generated the progenies with tall plants, indicating the dominance nature of the tallness gene in sorghum. The morphological traits such as grain lustre and red glume color also showed dominant gene action. The cross between the awned and awnless parents produced the awnless F₁ progeny, indicating the recessive nature of the gene action for presence of awns. The earlier studies of Ravindrababu and Pathak (2000) reported that additive, dominance, and epistatic (additive × dominance) effects were important for resistance to shoot fly. The results suggested that delayed selection is the best approach for the traits governed by dominance and epistasis effects. However, the traits that were governed by additive effects should undergo thorough selection at an early stage.

Conclusion

The present studies indicated that at least one of the parents involved in the crossing program should possess genes for resistance to shoot fly to develop high-yielding shoot fly-resistant sorghum. Both the non-allelic and predominance of dominance gene action for the component traits indicated that heterosis breeding is ideal for improving shoot fly resistance in sorghum genotypes. The additive nature of gene action for most of the traits indicated the importance of heterosis breeding, followed by simple selection for developing shoot fly-resistant sorghums.

Acknowledgements We are thankful to the sorghum entomology group and the sorghum breeding group for their



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support and help in carrying out this research work. We also thank Bill and Melinda Gates Foundation for their financial support through HOPE-sorghum and millet project.

References

- Aruna C, Padmaja PG, Subbarayudu B, Seetharama N (2011) Genetics of traits associated with shoot fly resistance in post-rainy season sorghum (*Sorghum bicolor* L.) Indian. J Genet 71(1):9–16
- Dhillon MK, Sharma HC, Singh R, Naresh JS (2006) Influence of cytoplasmic male-sterility on expression of physicochemical traits associated with resistance to sorghum shoot fly, Atherigona soccata (Rondani). SABRAO J Breed Genet 38(2):105–122
- Doggett H (1988) Sorghum, 2nd edn. John Wiley and Sons Inc, New York
- Doggett H (2008) Sorghum, 2nd edn. John Wiley and Sons, New York, pp 70–117
- FAO (2014) Crops primary equivalent. www.faostat.fao.org. Accessed 25 Oct 2014
- GenStat, (2010). Introduction to GenStat for Windows Genstat, 13th edn. Lawes Agricultural Trust, Rothamsted Experimental Station
- Hayman BI (1958) The separation of epistatic from additive and dominance variation in generation means. Heredity 12:371–390
- Hayman BI, Mather K (1955) The description of genetics of interaction in continuous variation. Biometrics 51:69–82
- House LR (1985) A guide to sorghum breeding, 2nd edn. International Crops Research Institute for the Semi-Arid Tropics, Patancheru, Andhra Pradesh, India
- IBPGR and ICRISAT (1993) Descriptors for sorghum [Sorghum bicolor (L.) Moench]. International Board for Plant Genetic Resources, Rome, Italy; International Crops Research Institute for the Semi-Arid Tropics, Patancheru, Andhra Pradesh, India
- Indostat Services (2004) Windostat. Indostat Services, Hyderabad
- Kearsey MJ, Pooni HS (1996) The genetical analysis of quantitative traits. Chapman and Hall, London, p 46
- Mather K (1949) Biometrical genetics, 1st edn. Methuen, London
- Pradhan S, Jotwani MG (1978) Investigations on insect pests of sorghum and millets with special reference to host plant resistance: final Technical Report (1975–1977). Project A7

- Ent-120. Research Bulletin No. 2. New Delhi: Indian Agricultural Research Institute
- Rana BS, Jotwani MG, Rao NGP (1981) Inheritance of host plant resistance to the sorghum shoot fly. Insect Science and its Applications 2:105–109
- Ravindrababu Y, Pathak AR (2000) Combining ability analysis over environments for yield and shoot fly resistance in sorghum. J Res Maharashtra Agric Univ 25:237–239
- Riyazaddin MD, Kavi Kishor PB, Ashok Kumar A, Belum Reddy VS, Rajendra SM, Sharma HC (2015) Mechanisms and diversity of resistance to sorghum shoot fly, *Atherig-ona soccata*. Plant Breed 134:423–436. https://doi.org/10. 1111/pbr.12276
- Riyazaddin M, Are AK, Munghate RS, Bhavanasi R, Polavarapu KKB, Sharma HC (2016) Inheritance of Resistance to Sorghum Shoot Fly, *Atherigona soccata* in Sorghum, *Sorghum bicolor* (L.) Moench. Front Plant Sci 7(543):1–18. https://doi.org/10.3389/fpls.2016.00543
- Rooney LW, Waniska RD (2000) Sorghum food and industrial utilization. In: Smith CW, Frederiksen RA (eds) Sorghum: origin, history, technology, and production. John Wiley & Sons Inc, New York, pp 689–729
- Sharma HC (1993) Host-plant resistance to insects in sorghum and its role in integrated pest management. Crop Prot 12:11–34
- Sharma HC, Nwanze KF (1997) Mechanisms of resistance to insects and their usefulness in sorghum improvement. Information Bulletin no. 55, International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, Andhra Pradesh, India
- Sharma SN, Sain RS (2003) Genetic architecture of grain weight in durum wheat under normal and late sown environments. Wheat Inform Serv 96:28–32
- Sharma HC, Taneja SL, Kameswara Rao N, Prasada Rao KE (2003) Evaluation of sorghum germplasm for resistance to insect pests. Information Bulletin no. 63, International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, Andhra Pradesh, India
- Soto PE (1974) Ovipositional preference and antibiosis in relation to resistance to sorghum shoot fly. J Econ Entomol 67:165–167
- Taneja SL, Leuschner K (1985) Resistance screening and mechanisms of resistance in sorghum to shoot fly. In: Proceedings of the International Sorghum Entomology Workshop, 15–21 July, 1984, Texas A and M University, College Station, TX, USA, pp 115–129. International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, Andhra Pradesh, India

