



# Seasonal dynamics of phytoplankton community and functional groups in a tropical river

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Received: 7 July 2021 / Accepted: 30 September 2021 / Published online: 8 October 2021  
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**Abstract** Functional classification of phytoplankton could be a valuable tool in water quality monitoring in the eutrophic riverine ecosystems. This study is novel from the Bangladeshi perspective. In this study, phytoplankton cell density and diversity were studied with particular reference to the functional groups (FGs) approach during pre-monsoon, monsoon, and

post-monsoon at four sampling stations in Karatoya River, Bangladesh. A total of 54 phytoplankton species were recorded under four classes, viz. Chlorophyceae (21 species) Cyanophyceae (16 species), Bacillariophyceae (15 species), and Euglenophyceae (2 species). A significantly higher total cell density of phytoplankton was detected during the pre-monsoon season ( $24.20 \times 10^3$  cells/l), while the lowest in monsoon ( $9.43 \times 10^3$  cells/l). The Shannon–Wiener diversity index varied significantly ( $F=16.109$ ,  $P=000$ ), with the highest value recorded during the post-monsoon season. Analysis of similarity (ANOSIM) identified significant variations among the three seasons ( $P<0.0001$ ,  $R=0.9518$ ). The similarity percentage (SIMPER) analysis pinpointed *Ulothrix* spp. (*Melosira granulate* and *Cymbella* spp.) as the most contributory species are causing such a noticeable difference. Fifty-four phytoplankton species recorded during the study period were classified into 20 functional groups, whereas D/J/M/MP/X1 was considered the most abundant FG in the Karatoya River. FGs of the Karatoya River were influenced mainly by the nutrients ( $PO_4$ -P and  $NO_3$ -N) enrichments. As a novel investigation on FGs of phytoplankton in Bangladesh, this study recommends additional surveys in other rivers and floodplains to improve our understanding of phytoplankton diversity and functional groups.

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**Keywords** Diversity index · Functional classification · Nutrient enrichment · Periodic flooding · Phytoplankton

## Introduction

Phytoplankton are remarkable aquatic organisms that are the primary producers in the aquatic food web. They constitute the rudimentary biological components of the aquatic ecosystem from where the energy is transferred to higher ranks via the food chain (Sharma et al., 2016). The diversity and abundance of phytoplankton in freshwater ecosystems are directly linked with the physicochemical water chemistry features of the waterbody (Ahmed & Wanganeo, 2015). Any changes in the physicochemical factors, particularly nutrients, elicit a quick response of the phytoplankton community (Atique & An, 2020; Haque et al., 2020). Therefore, water quality monitoring and management emerge as essential components of water's physicochemical and biological features (Atique et al., 2020; Hara et al., 2020; Kim et al., 2021). The use of phytoplankton as bioindicator organisms can serve the purpose cost-effectively. Therefore, the presence and absence of phytoplankton in a water body can provide vital information and prevalent connections within an aquatic ecosystem (Thakur et al., 2013).

Using functional groups (FGs) is a relatively modern approach in exploring the role of phytoplankton communities in a waterbody. These FGs are comparatively more effective and efficient in the ecological depiction of a water body than the phylogenetic groups, where long lists of species or dominant taxonomical groups are used for several days (Kruk et al., 2002). These groups often share similar tolerance and sensitivity levels. According to their description, FGs are comprised of species with similar morphology, physiology, and ecology and frequently coexist (Díaz & Cabido 2001; Mouillot et al., 2013; Salmaso et al., 2015; Qiu et al., 2016). They have identical ecological requirements but are not necessarily from the same phylogenetic group.

Investigations into the algal communities from a functional perspective can provide additional valuable information on the community-level response to environmental changes. Therefore, the role of FGs regarding their community characteristics can be determined and managed more precisely if the species are grouped into classes showing similar phenomena. FGs provide an easier way to examine and differentiate various seasonal variations in the

phytoplankton community and help understand the effect of environmental fluctuations (Kruk et al., 2002; Naselli-Flores et al., 2003; Weithoff et al., 2001). Occasionally, examination of FGs could ratify the latest most quantitative method of describing the phytoplankton community (Kruk et al., 2002). Therefore, our understanding of FGs can improve the species' selection dynamics in the aquatic environment (Bovo-Scomparin & Train, 2008; Sarmento et al., 2009). At present, 38 functional groups are described so far using alphanumeric codes (Padišák et al., 2009).

Although several studies have been conducted in different Bangladesh rivers, little is known about seasonality and ecology based on phytoplankton functional groups (Haque et al., 2019; Islam & Huda, 2016; Khondker & Abed, 2013). However, these groups have been discovered in lakes from several regions of the world (Celik & Ongun, 2008; Maraşlıoğlu & Gönülol, 2014; Soylu & Gönülol, 2010), rivers (Okogwu & Ugwumba, 2012; Devercelli & O'Farrell, 2013; Bortolini et al., 2014; Zanco et al., 2017; Huang et al., 2018), and artificial reservoirs (Becker et al., 2010; Borges et al., 2008).

Karatoya River is an essential river in Bangladesh that can be a suitable study area for studying the functional groups of phytoplankton. This historic waterbody has changed its course in the past and meanders into Bangladesh from Indian territory. This river is near to eutrophication due to reduced water flow and higher nutrient enrichment (Akhi et al., 2020). The continuous siltation process increasingly intensifies the threat of eutrophication. Furthermore, this river is contaminated by various metallic and nonmetallic chemicals originating from different industrial units, including textile, dyeing, pharmaceuticals, and leather (Zakir et al., 2012). Thus, determining the eutrophication level in this river using the phytoplankton FGs approach could help answer some critical questions.

Therefore, this study was designed to investigate the phytoplankton community of Karatoya River using the taxonomical and FGs approaches. The main objective of this study was to find out links between the present water quality condition and the persistent phytoplankton population. We also applied the FGs approach to investigate the phytoplankton species and finally provided a relationship between FGs and water quality parameters to describe the ecological status of

the Karatoya River, which is also a representative of the tropical rivers in Bangladesh.

**Materials and methods**

**Study area and duration**

This study sampling was performed for 12 months from July 2016 to June 2017 at four study stations, viz. Nurina, Sontola, Garodoho, and Ghatina, in the Karatoya River (KR) at Sirajgong District, Bangladesh (Fig. 1). The 12 months’ samples were subdivided into three distinct seasons based on rainfall intensity. They were termed pre-monsoon (February to May), monsoon (June to September), and post-monsoon (October to January) seasons. We recorded all the major water quality and phytoplankton parameters during this study period.

**Measurement of water quality parameters**

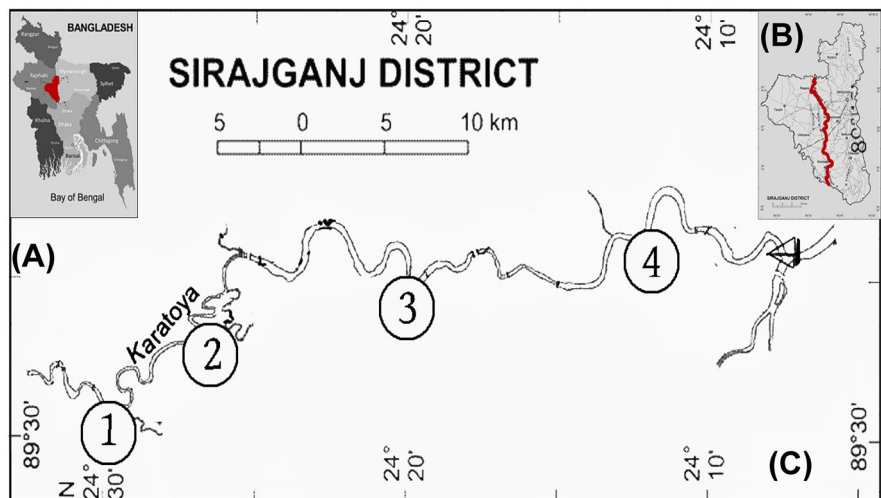
A black-colored plastic bottle (capacity 2 l) was used for water sample collection. Approximately 500 ml of water sample from each sampling station was collected, and the measurements of water quality parameters were performed on the bank of the river between 9:00 AM and 12:00 PM. Water temperature was measured by a Celsius thermometer, while pH was determined with an electronic pH meter (Jenway 3020, UK). Transparency was measured with a blank and white color-coded Secchi disk. A HACH Kit Box

(Model DR-2010, USA) was used to estimate the dissolved oxygen (DO), free carbon dioxide (CO<sub>2</sub>), alkalinity, and hardness. Phosphate-phosphorus (PO<sub>4</sub>-P) and nitrate-nitrogen (NO<sub>3</sub>-N) were measured using HACH Kit (DR-2020, USA) with high range chemicals (Phos. Ver. 3 Phosphate Reagent Powder Pillows for 25-ml sample for PO<sub>4</sub>-P analysis and Nitra Ver. 5 Nitrate Reagent Powder Pillows for 25-ml sample for NO<sub>3</sub>-N). Total dissolved solids (TDS) and electrical conductivity (EC) were measured by an Adwa AD31 waterproof EC/TDS tester.

**Determination of phytoplankton species and functional groups (FGs)**

Phytoplankton samples were collected from each study site during pre-monsoon, monsoon, and post-monsoon seasons. One hundred liters of water sample was filtered through a plankton net of 25-µm mesh size using a container of 10 l. Then, filtered samples were collected into a sample bottle and preserved immediately in 10% alcohol. The bottle was labeled and transferred to the laboratory for microscopic examination and identification. The concentrated sample vials were shaken to mix phytoplankton uniformly before microscopic examination. Each time, 1 ml of sample was drawn with the help of a dropper on a Sedge-wick Rafter Counting cell (S-R cell). The coverslips are placed, avoiding any air bubble formation (Zakir et al., 2012). Finally, the S-R counting cell was placed under the light microscope for identification and counting of phytoplankton. According

**Fig. 1** Study area map showing the sampling stations along the Karatoya River gradient (C), geographical position of the study area (B), and its placement in Bangladesh (A). The shown legends include (1) Ghatina, (2) Sontola, (3) Garodoho, and (4) Nurina in the Karatoya River, Sirajgong, Bangladesh



to Cooke (1960), Needham and Needham (1962), and Mackenthun et al. (1964), the qualitative valuation of the phytoplankton was done. The quantitative cell density of phytoplankton was expressed as cells per liter of the water sample using the following formula (Stirling & Wilsey, 2001).

$$N = \frac{A \times 1000 \times C}{V \times F \times L}$$

where  $N$  is the number of phytoplankton cells per liter of the original water,  $A$  is the total number of phytoplankton counted,  $C$  is the volume of the final concentration of the sample in ml,  $V$  is the volume of a field,  $F$  is the number of the field counted, and  $L$  is the volume of original water in liter.

All phytoplankton individuals, whether single cells, colonies, or filaments, were counted to estimate the phytoplankton cell density.

The Shannon–Wiener diversity index was calculated to understand the seasonal diversity of phytoplankton using the following formula (Shannon–Wiener 1963):

$$H = - \sum_i \frac{n_i}{N} \ln \frac{n_i}{N}$$

$H$  is the diversity index,  $n$  is the relative abundance ( $s/N$ ),  $S$  is the number of individuals for each species, and  $N$  is the total number of individuals.

The functional groups (FGs) of Phytoplankton were described according to Padišák et al. (2009).

### Statistical analyses

Seasonal variation in environmental variables was analyzed using one-way analysis of variance (ANOVA) at a 5% significance level using SPSS (Statistical Package for Social Sciences, version 20.0) software. The relationships between environmental variables and FGs were determined using Pearson's correlation in SPSS 20.0. Canonical correspondence analysis (CCA) was also performed to evaluate the relationship between the environmental variables and phytoplankton classes. Before the examination,  $\log_{10}(x+1)$  transformation standardized environmental variables and phytoplankton cell density. One-way analysis of similarities (ANOSIM) was tested to evaluate the significant variations in temporal scale and visualized through non-metric multi-dimensional scaling (NMDS) analysis. Finally, similarity percentages

(SIMPER) analyses (Clarke & Warwick, 1994) were performed to observe the percentage contribution and average dissimilarity among the three seasons. Similarity matrices were examined using the Bray–Curtis similarity index, which is used to quantify the differences in species composition. The Bray–Curtis index is always a number between 0 and 1. If 0, the samples share all the same species; if 1, they do not share any species. ANOSIM, SIMPER, and Shannon–Wiener diversity index were analyzed using Paleontological Statistics software version 3, while NMDS was performed using the vegan package in R software version 3.6.3.

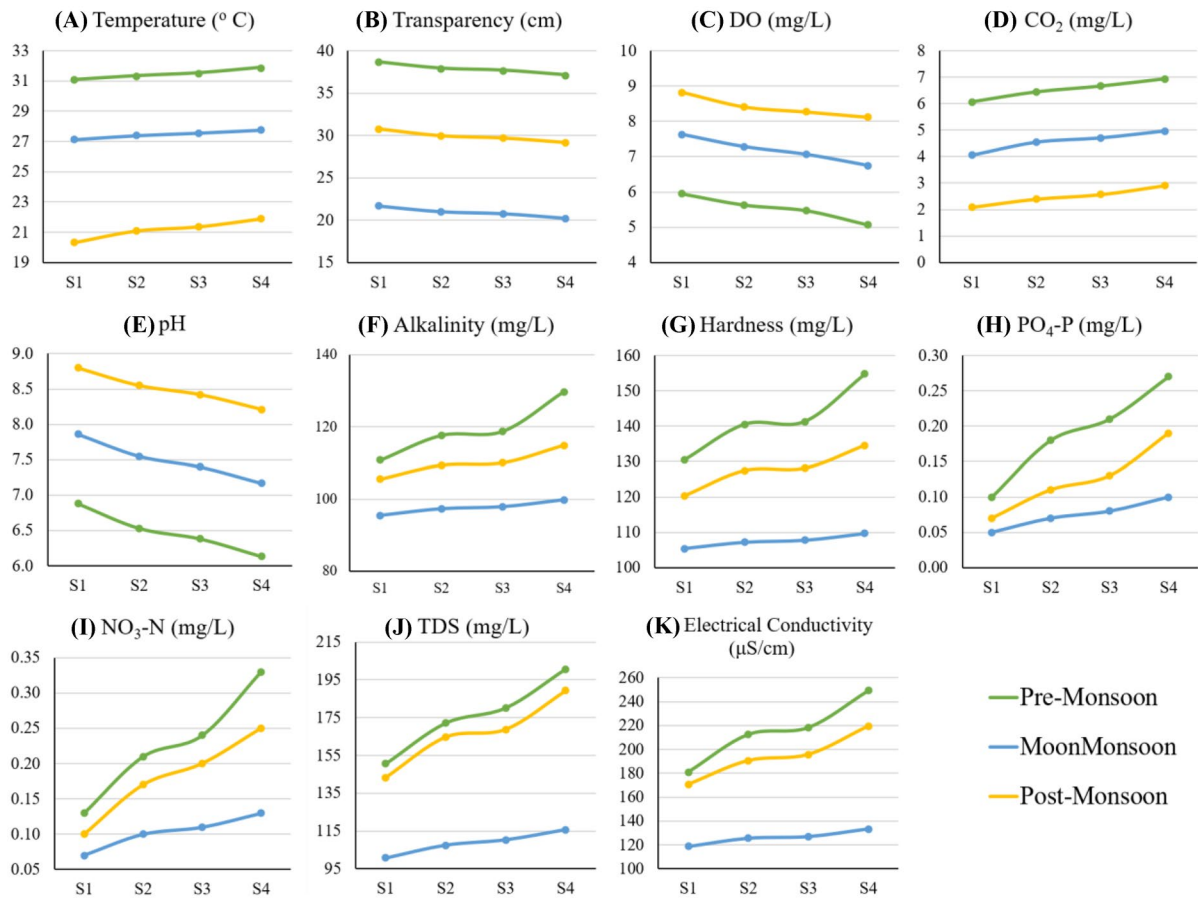
## Results

### Environmental variables

Environmental variables varied significantly based on seasonal comparisons (Fig. 2). Temperature (Fig. 2A), transparency (Fig. 2B), DO (Fig. 2C), CO<sub>2</sub> (Fig. 2D), alkalinity (Fig. 2F), hardness (Fig. 2G), TDS (Fig. 2J), and EC (Fig. 2K) showed evident seasonal variabilities at each study site (Table 1). Furthermore, significantly higher ( $P < 0.05$ ) values of most water quality parameters were observed during the pre-monsoon period. Conversely, significantly higher ( $P < 0.05$ ) nutrients, i.e., NO<sub>3</sub>-N (Fig. 2I) and PO<sub>4</sub>-P (Fig. 2L) as well as pH (Fig. 2E), were observed during the post-monsoon season. Most importantly, all the environmental variables displayed the lowest values during the monsoon season (Fig. 2).

### Phytoplankton composition and assemblage structure

Fifty-four species of phytoplankton belonging to 4 classes were identified. Chlorophyceae, Cyanophyceae, Bacillariophyceae, and Euglenophyceae constituted 21, 16, 15, and 2 species with the percentage contribution of approximately 38.89%, 29.63%, 27.78%, and 3.70%, respectively. The total cell density of phytoplankton varied significantly ( $F = 67.475$ ,  $P = 0.000$ ) between the seasons. In contrast, the highest total cell density ( $24.20 \times 10^3$  cells/l) was observed during the post-monsoon season and the lowest ( $9.43 \times 10^3$  cells/l) during monsoon (Fig. 3). Among the four phytoplankton groups, Chlorophyceae ( $5.76 \times 10^3$  cells/l) was the most abundant, while



**Fig. 2** Physicochemical water quality parameters on the site basis during the pre-monsoon, monsoon, and post-monsoon seasons

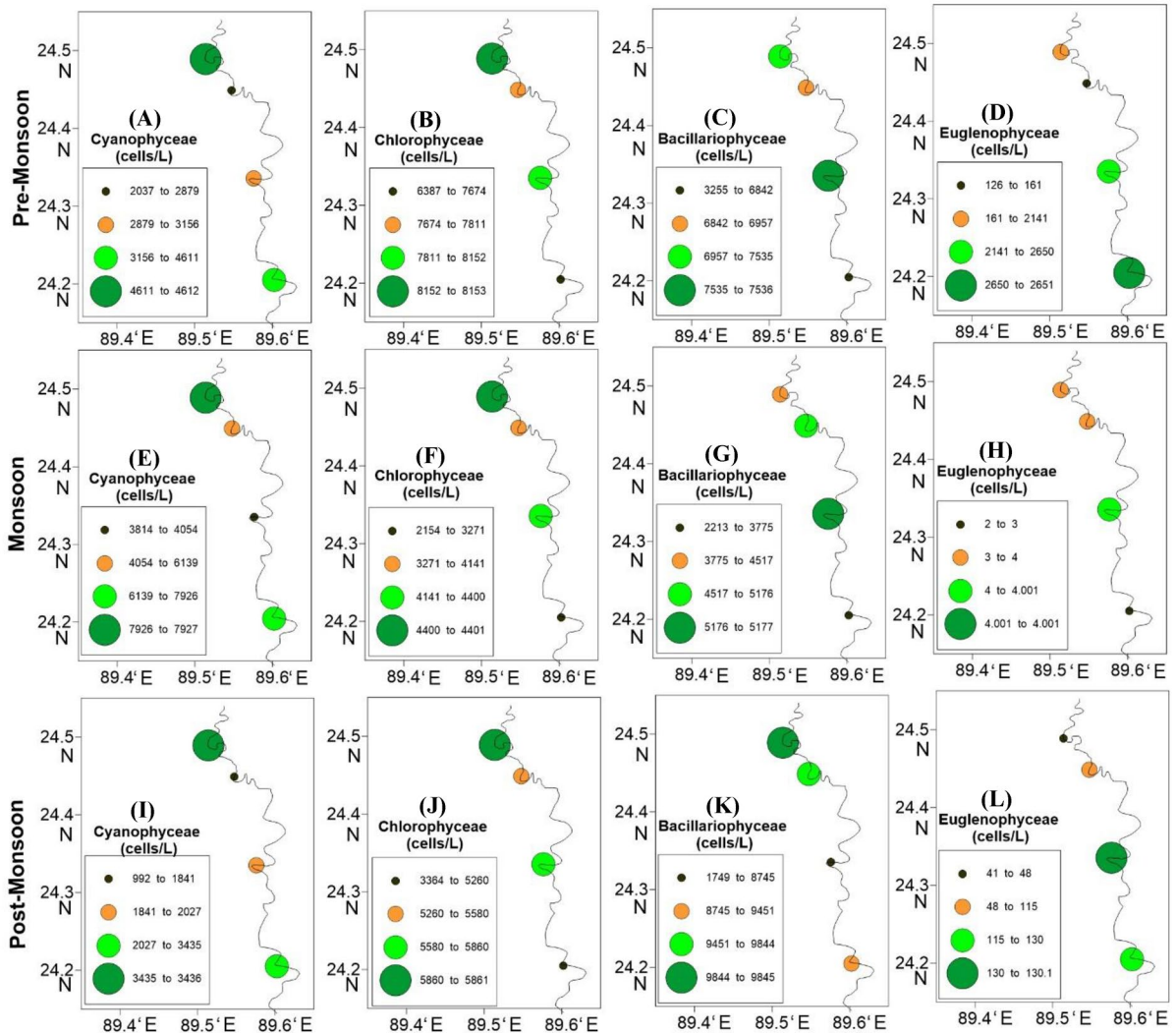
Euglenophyceae ( $0.45 \times 10^3$  cells/l) was the least abundant group. According to all seasons, sampling station 1 was dominated by Chlorophyceae and Cyanophyceae groups (Fig. 3A–B, E–F, I–J). On the other

hand, Bacillariophyceae was the highest at station three during pre-monsoon (Fig. 3C) and monsoon (Fig. 3G). Station 4 showed a higher concentration of Euglenophyceae during pre-monsoon only (Fig. 3D).

**Table 1** Monsoon-based seasonal comparisons of environmental variables in Karatoya River, Bangladesh

Parameter	Pre-monsoon	Monsoon	Post-monsoon
Temperature (°C)	31.47 ± 0.31 <sup>a</sup>	27.46 ± 0.25 <sup>b</sup>	21.16 ± 0.60 <sup>c</sup>
Transparency (cm)	37.89 ± 0.59 <sup>a</sup>	20.90 ± 0.56 <sup>c</sup>	29.89 ± 0.61 <sup>b</sup>
Dissolved oxygen (mg/l)	5.53 ± 0.34 <sup>c</sup>	7.18 ± 0.34 <sup>b</sup>	8.41 ± 0.27 <sup>a</sup>
Carbon dioxide (mg/l)	6.53 ± 0.34 <sup>a</sup>	4.57 ± 0.35 <sup>b</sup>	2.49 ± 0.31 <sup>c</sup>
pH	6.48 ± 0.28 <sup>c</sup>	7.50 ± 0.26 <sup>b</sup>	8.50 ± 0.22 <sup>a</sup>
Alkalinity (mg/l)	119.17 ± 7.07 <sup>a</sup>	97.58 ± 1.63 <sup>c</sup>	109.92 ± 3.47 <sup>b</sup>
Hardness (mg/l)	141.76 ± 9.02 <sup>a</sup>	107.49 ± 1.65 <sup>c</sup>	127.61 ± 5.29 <sup>b</sup>
PO <sub>4</sub> -P (mg/l)	0.13 ± 0.05 <sup>b</sup>	0.08 ± 0.02 <sup>c</sup>	0.19 ± 0.07 <sup>a</sup>
NO <sub>3</sub> -N (mg/l)	0.18 ± 0.06 <sup>b</sup>	0.10 ± 0.02 <sup>c</sup>	0.23 ± 0.08 <sup>a</sup>
Total dissolved solids (mg/l)	175.88 ± 18.71 <sup>a</sup>	108.50 ± 5.61 <sup>c</sup>	166.52 ± 17.07 <sup>b</sup>
Electrical conductivity (µS/cm)	215.35 ± 25.39 <sup>a</sup>	126.14 ± 5.45 <sup>c</sup>	194.12 ± 18.25 <sup>b</sup>

Values in the same raw having different superscript letters (“a” indicates significantly higher, “b” indicates an intermediate situation, and “c” describes the significantly lower values) differ significantly ( $P < 0.05$ )



**Fig. 3** Phytoplankton cell densities observed in different classes in the Karatoya River, Bangladesh

Phytoplankton, especially Euglenophyceae, significantly declined in monsoon (Fig. 3H).

The most abundant phytoplankton species recorded with a relative abundance > 10% was *Volvox* spp. (35.07%), followed by *Melosira granulata* (18.70%), *Euglena* spp. (18.54%), *Oedogonium* spp. (12.27%), *Ulothrix* spp. (11.56%), and *Synedra ulna* (10.10%) as shown in Table 2.

The Shannon–Wiener diversity index significantly varied ( $F=16.109$ ,  $P=000$ ) among the seasons. The highest diversity index value (3.90) was recorded during the post-monsoon season, while the lowest (3.77) was during the monsoon period (Fig. 4).

The phytoplankton assemblage significantly differed during the three seasons (ANOSIM,  $P=0.0001$ ,  $R=0.9518$ ). A two-dimensional NMDS based on Bray–Curtis’s similarity index divided the phytoplankton abundance during the pre-monsoon period from the monsoon and post-monsoon seasons (Fig. 5). SIMPER analysis exhibited an overall average dissimilarity of 23.10% among the seasons. The five species most significantly responsible for this difference in seasons are *Melosira granulata* (3.86%), *Ulothrix* spp. (3.65%), *Oedogonium* spp. (3.55%), *Melosira varians* (3.45%), and *Amphora ovalis* (3.39%). However, the average dissimilarity percentage between

**Table 2** List of phytoplankton species categorized based on their respective classes identified during this study along with their ascribed species code, relative seasonal abundance

Species	Species code	Relative abundance (%)			
		Pre-monsoon	Monsoon	Post-monsoon	Total
<b>Chlorophyceae</b>					
<i>Actinastrum</i> spp.	Ac	1.27	1.02	1.47	3.76
<i>Actinastrum hantzschii</i>	Ah	1.22	1.01	1.42	3.65
<i>Ankistrodesmus falcatus</i>	Af	0.83	0	1.08	2.65
<i>Chlorella</i> spp.	Chl	0.65	0.57	0.96	2.18
<i>Closterium eboracense</i>	Ce	0.77	1.18	0.99	2.94
<i>Closterium parvulum</i>	Cp	0.72	1.16	1.06	2.94
<i>Coelastrum cambricum</i>	Cc	0	0.83	1.05	2.38
<i>Cosmarium</i> spp.	Co	0.53	1.41	1.06	3.00
<i>Euastrum binale</i>	Eb	1.97	1.48	0.31	3.76
<i>Microspora</i> spp.	Mic	0.01	0.24	0.73	0.98
<i>Oedogonium</i> spp.	Oe	2.58	5.89	3.80	12.27
<i>Pediastrum biradiatum</i>	Pb	1.22	0	1.66	5.66
<i>Rhizoclonium</i> spp.	Rhi	1.14	0	1.62	5.46
<i>Scenedesmus abundans</i>	Sa	0.42	0	0.98	2.35
<i>Scenedesmus arcuatus</i>	Sca	0.49	0	1.00	2.72
<i>Scenedesmus dimorphus</i>	Sd	0.26	0.92	0.73	1.91
<i>Spirogyra</i> spp.	Spi	0.01	0	0.87	1.33
<i>Tetraedron minimum</i>	Tm	0.83	0	0.91	2.14
<i>Ulothrix</i> spp.	Ulo	2.37	5.72	3.47	11.56
<i>Volvox</i> spp.	Vol	14.54	11.35	9.18	35.07
<i>Zygnema stellium</i>	Zs	1.24	0	1.29	3.71
<b>Bacillariophyceae</b>					
<i>Achnanthes minutissima</i>	Am	1.03	1.19	0.91	3.13
<i>Amphora ovalis</i>	Ao	3.62	0	2.02	5.66
<i>Asterionella</i> spp.	Ast	0	1.26	0.94	2.21
<i>Cymbella</i> spp.	Cym	1.18	1.33	0.89	3.40
<i>Epithema</i> spp.	Epi	1.35	0	0.86	3.25
<i>Fragilaria</i> spp.	Fra	0.01	2.53	2.58	5.12
<i>Gomphonema</i> spp.	Gom	0	1.64	1.08	3.35
<i>Melosira granulata</i>	Mg	6.54	6.26	5.90	18.70
<i>Melosira varians</i>	Mv	1.05	1.17	1.47	3.69
<i>Navicula radiososa</i>	Nr	0.93	0.65	1.01	2.59
<i>Navicula</i> spp.	Nav	1.60	0	1.01	4.08
<i>Nitzschia palea</i>	Np	0.79	1.44	0.86	3.09
<i>Synedra rumpens</i>	Nr	0.99	0.99	0.89	2.87
<i>Synedra ulna</i>	Su	3.75	0.01	6.34	10.10
<i>Tabellaria</i> spp.	Ta	0.84	0	0.82	2.46
<b>Cyanophyceae</b>					
<i>Anabaena spiroides</i>	As	0.48	1.07	1.37	2.92
<i>Anabaena</i> spp.	An	0	0	1.48	1.51
<i>Aphanocapsaakoordesii</i>	Ak	0.01	0.57	0.95	1.53
<i>Aulosira</i> spp.	Au	0	2.61	1.51	4.13
<i>Calothrix</i> spp.	Ca	2.99	0	2.03	5.04

**Table 2** (continued)

Species	Species code	Relative abundance (%)			
		Pre-monsoon	Monsoon	Post-monsoon	Total
<i>Chroococcus</i> spp.	Ch	3.77	2.50	1.49	7.76
<i>Lynghya limnetica</i>	Ll	3.71	0	3.36	7.09
<i>Merismopedia</i> spp.	Me	2.09	2.27	1.58	5.94
<i>Microcystis aeruginosa</i>	Ma	0.01	3.99	4.38	8.38
<i>Microcystis pseudofilamentosa</i>	Mp	0.48	0	3.53	4.03
<i>Nostoc</i> spp.	No	4.65	0	1.67	9.10
<i>Oscillatoria acuminata</i>	Oa	4.78	0	1.73	8.28
<i>Oscillatoria perornata</i>	Op	4.41	2.67	1.70	8.78
<i>Phormidium</i> spp.	Ph	3.76	1.89	1.64	7.29
<i>Rivularia</i> spp.	Ri	3.73	2.35	1.62	7.70
<i>Spirulina major</i>	Sm	2.38	0	1.54	3.94
<b>Euglenophyceae</b>					
<i>Euglena</i> spp.	Eu	8.38	4.08	6.08	18.54
<i>Phacus</i> spp.	Pc	0	0	1.72	1.94

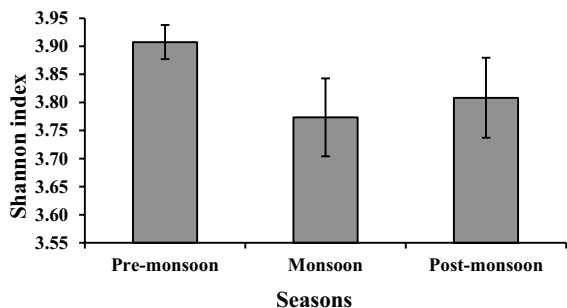
pre-monsoon to monsoon, pre-monsoon to post-monsoon, and monsoon to post-monsoon was 22.87%, 21.66%, and 24.77%, respectively. The most significant contributory species potentially responsible for this difference were *Ulothrix* spp., *Melosira granulata*, and *Cymbella* spp. The combined seasonal contributions were 5.21% (pre-monsoon to monsoon), 5.83% (pre-monsoon to post-monsoon), and 4.71% (monsoon to post-monsoon).

After considering seasonal segmentation and stations, the CCAs were performed to understand the complex relations of phytoplankton groups with environmental parameters (Fig. 6). The distance point of phytoplankton classes from environmental parameter

directional tendencies indicated the perceptual ties between them. For instance, Cyanophyceae showed a closer association with DO, pH, and transparency during pre-monsoon (Fig. 6A). Conversely, Bacillariophyceae showed more relative links to alkalinity, phosphate, and conductivity during the post-monsoon (Fig. 6C). Considering average CCA, Cyanophyceae and Bacillariophyceae are affected closely by the associated environmental parameters at the sampling area (Fig. 6).

#### Phytoplankton functional group

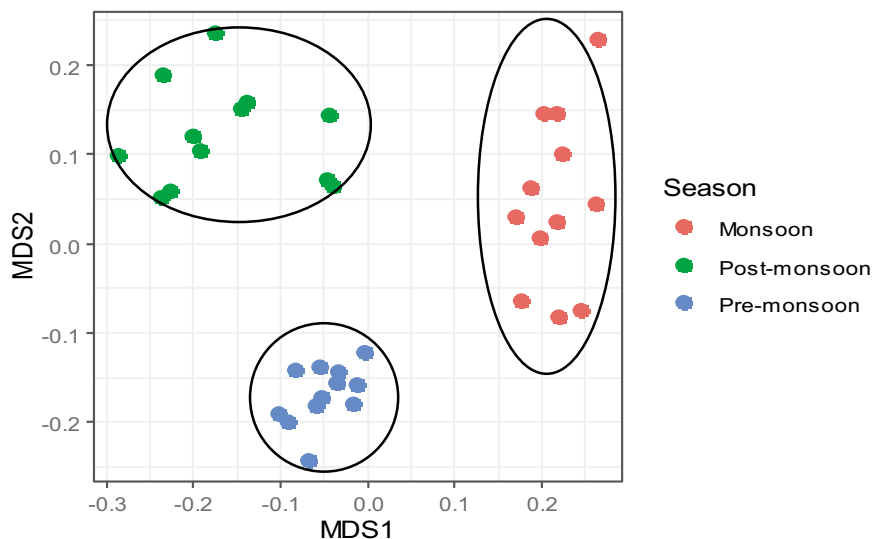
The 54 phytoplankton species recorded during this investigation were classified into 20 functional groups (FGs), as presented in Table 3. Due to the absence of previous records as parts of the FGs, *Calothrix* spp., *Nostoc* spp., *Rivularia* spp., *Euastrum binale*, *Microspora* spp., *Oedogonium* spp., *Spirogyra* spp., *Zygnema stellium*, and *Epithema* spp. were excluded from functional groups classification. The seasonal dynamics observed by the relative abundance of the functional groups displayed a relative abundance of >5% during the pre-monsoon season, while the representative FGs were D/J/M/MP/X1. However, during monsoon and post-monsoon periods, the dominant FGs were characterized as D/J/M/MP/S1/T<sub>B</sub>/X1 and D/J/M/MP/T<sub>B</sub>/X1/W1, respectively. Therefore, D/J/M/MP/X1 could be interpreted as the



**Fig. 4** Seasonal comparisons of the Shannon–Wiener diversity index calculated for the phytoplankton species in Karatoya River, Bangladesh



**Fig. 5** Seasonal classification of phytoplankton assemblages with the help of NMDS plot in Karatoya river, Bangladesh



most abundant functional group in all the seasons (Table 3).

The correlation of water quality parameters and FGs showed a significantly strong relationship among and with the selected water quality parameters (Table 4). However, most dominant FGs (D/J/M/MP/X1) were negatively influenced by water temperature. At the same time, they showed a positive correlation with the ambient nutrients (PO<sub>4</sub>-P and NO<sub>3</sub>-N), and such conditions prevailed as favorable during the post-monsoon season (DO and pH).

## Discussion

### Seasonal environmental variations

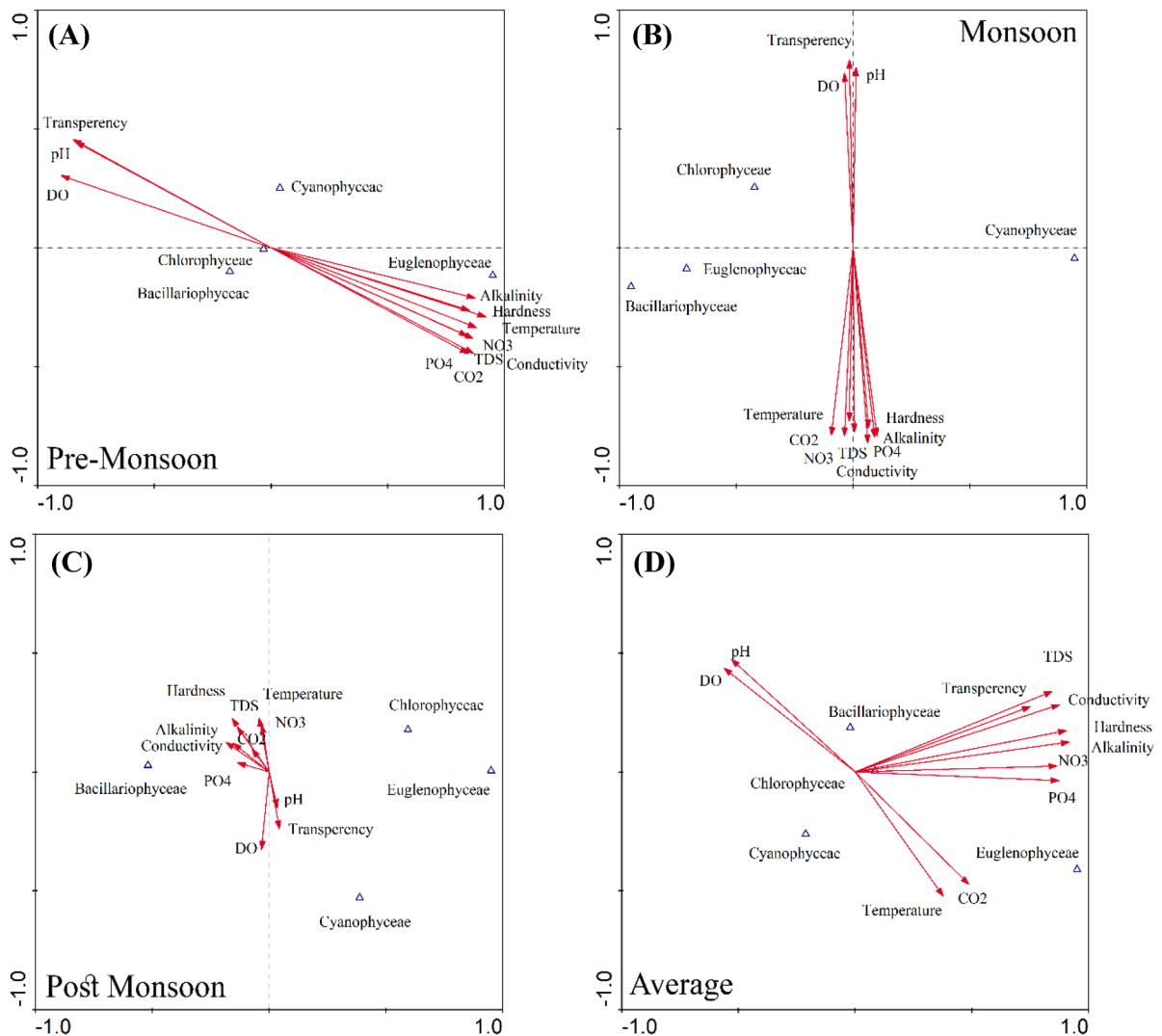
The freshwater physicochemical status of an aquatic ecosystem significantly impacts the occurrence and distribution of the aquatic biota (Sharif et al., 2017). In this study, the water temperature fluctuated between 23.54 and 30.25 °C, with the lowest during the post-monsoon period that reached the maximum value during the monsoon season. Similar observations were reported by Bera et al. (2014), who also observed a seasonal water temperature variation during a comparable study period in an identical pattern. In addition, the intensive monsoonal rainfall triggered a higher amount of washing slits, sediments, debris, and organic and inorganic suspended particles into the river channel reducing the overall water

transparency. The post-monsoon increase in DO reported in this study corroborated with the findings of Sharif et al. (2017).

Similarly, we observed the highest value of free carbon dioxide (CO<sub>2</sub>) in the pre-monsoon season. Such increased free CO<sub>2</sub> could be attributed to the decomposition of the prevalent organic matter during this season at a higher rate. The seasonal impact was also apparent in the varying pH, alkalinity, hardness, PO<sub>4</sub>-P, NO<sub>3</sub>-N, TDS, and EC, as their values were recorded lower during the monsoon period. Various researchers have reported similar findings (Ali et al., 2020; Briola et al., 2010; Sayeed et al., 2015; Varol & Şen, 2018; Venkateshwarlu et al., 2011).

### Seasonal patterns of phytoplankton assemblages

The total phytoplankton abundance in the Karotoya River demonstrated remarkable temporal variations, with a significantly higher ( $F=67.475, P=000$ ) total cell density ( $24.20 \times 10^3$  cells/l) observed during the post-monsoon season, while the lowest ( $9.43 \times 10^3$  cells/l) during monsoon. The potential underlying reasons for the maximum total cell density recorded during the post-monsoon season could be their ability to flourish under weaker light availability, lower water temperature, and higher nutrient load, as Naik et al. (2009) have reported. Vajravelu et al. (2018) observed the post-monsoon nutrient enrichment was potentially accountable for the higher phytoplankton abundance in the Parangipettai coastal waters of the southeast



**Fig. 6** Seasonal and average CCA biplots constructed for the environmental variables and phytoplankton classes during the study period

Indian Coast. However, a lower abundance of phytoplankton has also been reported during the monsoon season characterized by heavy rainfall, decreased salinity, water temperature, pH, and higher turbidity which could be the leading causative factors for this phenomenon (Babu et al., 2013; Thillai et al., 2010).

Among the four phytoplankton groups, Chlorophyceae ( $5.76 \times 10^3$  cells/l) was the most abundant, while Euglenophyceae ( $0.45 \times 10^3$  cells/l) was the least abundant group. This river is characterized by reduced water flow and increasing siltation (Akhi et al., 2020). Therefore, stagnant or near to zero flow

of the riverine water during the post-monsoon period triggered nutrient retention in the water column, increasing the nutrient load. This could be one of the principal reasons for a higher enrichment of the Chlorophyceae group during this study. Similarly, a higher density of Chlorophyceae was corroborated during low water flow by Flura et al. (2016) in Padma River, Bangladesh. The Padma River has displayed identical river flow characteristics to our studied river. The Chlorophyceae dominance has also been reported by Maraşlıoğlu and Gönülol (2014) in a nutrient-rich eutrophic Yedikır Dam Lake in Turkey.

**Table 3** Percent relative and seasonal abundance of phytoplankton FGs observed in the Karotoa River, Bangladesh

Functional groups	Species	Relative abundance (%)		
		Pre-monsoon	Monsoon	Post-monsoon
A	<i>Rhizosolium</i> spp.	0.96	1.80	1.02
C	<i>Asterionella</i> spp.	0.01	3.25	1.80
D	<i>Fragilaria</i> spp., <i>Nitzschia palea</i> , <i>Synedra ulna</i> , <i>Synedra rumpens</i>	24.60	3.68	13.19
G	<i>Volvox</i> spp.	1.02	0.99	0.98
H1	<i>Anabaena spiroides</i> , <i>Anabaena</i> spp.	3.02	2.53	3.46
J	<i>Closterium eboracense</i> , <i>Closterium parvulum</i> , <i>Coelastrum cambricum</i> , <i>Scenedesmus abundans</i> , <i>Scenedesmus arcuatus</i> , <i>Scenedesmus dimorphus</i> , <i>Pediastrum biradiatum</i> , <i>Actinastrum</i> spp., <i>Actinastrum hantzschii</i> , <i>Tetradron minimum</i>	8.89	0.84	11.36
K	<i>Aphanocapsa koordesi</i>	1.00	0.93	1.28
L <sub>M</sub>	<i>Achnanthes minutissima</i>	0.01	0.02	1.77
Lo	<i>Merismopedia</i> spp., <i>Chroococcus</i> spp.	3.20	1.52	2.52
M	<i>Microcystis aeruginosa</i> , <i>Microcystis pseudofilamentosa</i>	6.00	4.45	8.68
MP	<i>Navicula radiosa</i> , <i>Navicula</i> spp., <i>Ulothrix</i> spp., <i>Lyngbya limnetica</i> , <i>Amphora ovalis</i> , <i>Cymbella</i> spp.	8.56	6.76	24.15
N	<i>Tabellaria</i> spp.	2.88	0.03	1.84
Na	<i>Cosmarium</i> spp.	0.01	1.56	1.12
P	<i>Aulacoseira</i> spp.	0.79	0.71	1.15
S1	<i>Oscillatoria acuminata</i> , <i>Oscillatoria perornata</i>	2.86	6.81	3.93
S2	<i>Spirulina major</i>	0.32	1.15	0.87
T <sub>C</sub>	<i>Phormidium</i> spp.	0.51	1.19	1.17
T <sub>B</sub>	<i>Gomphonema</i> spp., <i>Melosira granulata</i> , <i>Melosira varians</i>	2.57	1.68	11.60
X1	<i>Ankistrodesmus falcatus</i> , <i>Chlorella</i> spp.	7.65	1.41	21.84
W1	<i>Euglena</i> spp., <i>Phacus curvicauda</i>	0.83	0.04	6.27

Phytoplankton assemblages and diversity

Phytoplankton diversity indices are widely used in connection with water quality (Ahmed & Wanganeo, 2015). The Shannon–Wiener diversity index varied significantly ( $F=16.109$ ,  $P=000$ ) among the seasons, and the highest value (3.90) was recorded during the post-monsoon season, while the lowest (3.77) record was observed during monsoon. Therefore, the observed highest value during the post-monsoon period could be linked to the diverse species composition observed during this study. Moreover, the advantageous environmental circumstances with an elevated nutrient level in water could have explained another reason for the higher diversity indices during the post-monsoon season, as Dupuis and Hann (2009) reported.

On the contrary, a lower value of the diversity shown by the index during monsoon was potentially

impacted by the higher and fluctuating water level augmented by the unfavorable environmental conditions. Roozen et al. (2003) also deemed flooding a disturbance factor that may cause water column instability, fluctuations in water level, and reduced water retention time (WRT). Thus, a strong flood sweeping away all the nutrients during the monsoon period and causing destabilized ecological settings could be one of the compelling factors for accelerated species loss and reduced number of plankton communities with a few individuals left (Wojciechowska et al., 2007).

The significantly differing phytoplankton community assemblage during the three seasons was due to the fluctuation in the available limiting nutrients in the riverine water (Danger et al., 2008). Furthermore, the phytoplankton community could have undergone a dilution effect during the monsoon season, potentially causing a lower phytoplankton cell density. As reported by Rahman and Huda (2012) and Ahmed

**Table 4** Correlation analysis on the phytoplankton FGs and environmental variables observed in the Karatoya River, Bangladesh

	Temp	Trans	DO	CO <sub>2</sub>	pH	Alk	Hard	PO <sub>4</sub> -P	NO <sub>3</sub> -N	TDS	EC
A	-0.56**	-0.19	0.52**	-0.53**	0.50**	0.42	-0.04	0.03	0.09	0.12	0.50**
C	-0.75**	-0.46**	0.13	-0.79**	0.23*	-0.47	-0.47**	-0.51**	-0.44**	0.50**	-0.36*
D	-0.19	0.59**	0.46**	-0.13	0.82**	-0.03	0.08	0.42**	0.80**	-0.31	0.07
G	-0.48**	0.18	0.48**	-0.50**	0.54**	-0.004	0.023	-0.248	-0.154	0.098	0.07
H1	-0.61**	0.20	0.55**	-0.58**	0.60**	0.04	0.06	-0.17	-0.12	0.18	0.15
J	-0.83**	0.10	0.78**	-0.82**	0.82**	-0.08	-0.03	0.78**	0.59**	0.19	0.11
K	-0.73**	0.18	0.65**	-0.60**	0.69**	0.06	0.08	-0.14	-0.05	0.25	0.20
LM	-0.79**	0.04	0.71**	-0.75**	0.74**	0.03	0.04	-0.12	-0.03	0.23	0.17
Lo	-0.83**	-0.13	0.83**	-0.83**	0.85**	-0.24	-0.23	-0.39*	-0.31	-0.04	-0.09
M	-0.77**	-0.25	0.81**	-0.80**	0.82**	-0.39*	-0.38*	0.52**	0.46**	-0.22	-0.27
MP	-0.25	0.67**	0.59**	-0.17	0.60**	0.30	0.28	0.53**	0.49**	0.17	0.12
N	-0.29	0.63**	0.17	-0.22	0.23	0.45**	0.48**	0.21	0.24	0.55**	0.54**
Na	-0.87**	-0.40*	0.88**	-0.89**	0.88**	-0.40*	-0.39*	-0.46**	-0.35*	-0.19	-0.26
P	-0.76**	0.17	0.69**	-0.73**	0.74**	0.03	0.06	-0.17	-0.09	0.24	0.18
S1	-0.44**	-0.25	0.35*	-0.38*	0.32	0.09	0.05	0.25	0.29	0.23	0.17
S2	-0.81**	-0.23	0.84**	-0.83**	0.85**	-0.34*	-0.33	-0.47**	-0.40*	-0.16	-0.21
TC	-0.83**	-0.11	0.82**	-0.83**	0.85**	-0.19	-0.18	-0.34*	-0.25	0.02	-0.04
TB	-0.86**	-0.35*	0.85**	-0.86**	0.84**	-0.29	-0.29	-0.32	-0.22	-0.06	-0.14
X1	-0.56**	0.54**	0.39*	-0.48**	0.46**	0.45**	0.48**	0.22	0.31	0.62**	0.58**
W1	-0.54**	0.03	0.42*	-0.48**	0.43**	0.21	0.22	0.27	0.37*	0.44**	0.36*

\*Correlation is significant at the 0.05 level (2-tailed)

\*\*Correlation is significant at the 0.01 level (2-tailed)

Temp. temperature, Trans. transparency, DO dissolved oxygen, CO<sub>2</sub> carbon dioxide, Alk. alkalinity, Hard. hardness, PO<sub>4</sub>-P phosphate-phosphorus, NO<sub>3</sub>-N nitrate-nitrogen, TDS total dissolved solids, EC electrical conductivity .

and Alfasane (2004) in Padma River, higher water flow largely impacted the phytoplankton density. They have reported a lower phytoplankton cell density during the monsoon season. It is further supported by greater distances of phytoplankton from other environmental variables as illustrated in the CCA plot during monsoon (Fig. 6B). Typical river phytoplankton *Melosira granulata*, as reported by Reynolds (1988), was found as the most critical species responsible for higher dissimilarity among the seasons during this study. The same species has been reported as abundant in another study conducted by Ahmed and Alfasane (2004) in a Bangladeshi river. Other critical species identified through SIMPER analysis include *Ulothrix* spp., *Oedogonium* spp., *Amphora ovalis*, and *Cymbella* spp. cannot be discussed in further detail owing to the lack of species-specific studies conducted previously in Bangladesh. However, most of these species showed a relative abundance of > 10% during this investigation. Therefore, these species are

critical in taking further in-depth monitoring of phytoplankton diversity and abundance.

Seasonal trends in the functional groups and water quality

The classification of the phytoplankton community into FGs and morphology-based functional groups (MBFGs) represents an essential tool for understanding the behavior and species dynamics in relation to environmental conditions (Salmaso & Padisák, 2007). The most dominant FGs (D/J/M/MP/X1) mainly abound at the low flow or stagnant water with higher nutrient enrichment of the water body. However, the abundance of these FGs was lower during the monsoon season and was primarily affected by turbulent floodwater currents. Thorp (2009) reported that stochastic processes are more intense in rivers, while the phytoplankton are influenced primarily by flow and flood pulse. Therefore, it is challenging

to determine a compelling relationship between the environmental conditions and the phytoplankton distribution (Rodrigues et al., 2018).

Furthermore, the wash-out effect in a river during monsoon creates difficulty establishing the phytoplankton species in a river (Fraisse et al., 2013; Stanković et al., 2012). During the study period, the FGs (D/J/M/MP/X1) were sensitive to elevated temperature and benefited from increased nutrient loads in the water column during the post-monsoon season. The CCA plot of post-monsoon also supported this conclusion. Following heavy floods, the Karotoya River was enriched with nutrients during the post-monsoon season. However, during this season, water flow was significantly reduced. The river turned into a stagnant, nutrient-rich, and eutrophic waterbody, favoring an increased enrichment of FGs like D, L<sub>0</sub>, LM, M, MP, W1, and X1. Several researchers also reported similar observations (Padisák et al., 2009; Wu et al., 2011; Xiao et al., 2011).

The correlation analysis revealed consistency between FGs and water quality parameters in corroboration with the findings reported by Calijuri et al. (2002), Donald et al. (2013), and Fernández et al. (2014). These researchers have reported a negative influence of water temperature on the FGs D and a positive relationship with the nutrients (NO<sub>3</sub>-N). Furthermore, higher PO<sub>4</sub>-P was also found advantageous to the growth of FGs M, as was endorsed by the findings of Okogwu and Ugwumba (2012). Therefore, it was evident from the assessment of FGs in the Karotoya River that the periodic flooding triggering the seasonal hydrological alterations was the most critical factor influencing the phytoplankton community. Furthermore, the nutrient regime played a crucial role in shaping the phytoplankton community, similar to the other eutrophic water bodies across the globe (Fariñas et al., 2015).

## Conclusion

This study was performed in a river primarily impacted by various factors, including intensive rainfall, rampant stagnations, higher nutrient enrichment, and increasing siltation. All these factors present an ecosystem conducive to specific phytoplankton communities and their functional groups. In conclusion, our findings illustrated hydrodynamics as the most

responsible factor in determining the phytoplankton community formation in the Karotoya River. The phytoplankton diversity was the highest (3.90) during the post-monsoon season, while the lowest (3.77) during monsoon. Results also showed a significant difference in the phytoplankton community among the seasons brought by a typical river plankton *Melosira granulata*. Again, assessment of FGs in the Karotoya River and its correlation with environmental variables exposed that the periodic flooding was the most critical factor influencing the phytoplankton community.

**Acknowledgements** The authors would like to thank the Government of the People's Republic of Bangladesh for providing available funding through the National Science and Technology (NST) fellowship.

**Data availability** The datasets used and/or analyzed during the current study are available from the corresponding author on justifiable request.

## Declarations

**Ethics approval** Not applicable.

**Consent to participate** Not applicable.

**Consent for publication** Not applicable.

**Competing interests** The authors declare no competing interests.

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