# Simultaneous spectrophotometric determination of trace amount of polycyclic aromatic hydrocarbons in water samples after magnetic solid-phase extraction by using projection pursuit regression

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Abstract Magnetic solid-phase extraction based on coated nano-magnets Fe<sub>3</sub>O<sub>4</sub> was applied for the preconcentration of four polycyclic aromatic hydrocarbons (PAHs; anthracene, phenanthrene, fluorine, and pyrene) in environmental water samples prior to simultaneous spectrophotometric determination using multivariate calibration method. Magnetic nanoparticles, carrying target metals, were easily separated from the aqueous solution by applying an external magnetic field so, no filtration or centrifugation was necessary. After elution of the adsorbed PAHs, the concentration of PAHs was determined spectrophotometrically with the aid of a new and efficient multivariate spectral analysis base on principal component analysis-projection pursuit regression, without separation of analytes. The obtained results revealed that using projection pursuit regression as a flexible modeling approach improves the predictive quality of the developed models compared with partial least squares and least squares support vector machine methods. The method was used to determine four PAHs in environmental water samples.

**Keywords** Polycyclic aromatic hydrocarbons · Magnetic solid-phase extraction · Projection pursuit regression

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# Introduction

Polycyclic aromatic hydrocarbons (PAHs) represent an important class of hazardous organic chemicals consisting of two or more fused benzene rings in linear, angular, or cluster arrangements (Bjorseth 1983). They are ubiquitous pollutants present in all compartments of the environment (atmosphere, soil and water) and produced in a variety of industrial processes and are usually associated with incomplete combustion of fossil fuels. Some of PAHs exhibit carcinogenic and mutagenic properties and the wide distribution of them in the environment pose serious health risks to all living organisms (Neff 1979; Cavalieri and Rogan 1998). Moreover, PAHs have attracted increasingly attention as biochemical, immunologic, genetic, and reproductive effects have been widely observed in fish and other animals from marine and freshwater environments upon exposure to PAHs (Pagnout et al. 2006; Geffard et al. 2003). Therefore, reliable analytical methods for the continuous monitoring of these compounds are of great interest. Owing to the complexity of environmental sample matrices and the relative low concentration of PAHs in these samples, separation and preconcentration procedure were necessary in the analysis of PAHs in the environmental samples to improve the sensitivity of method (Pena et al. 2009; Shariati-Feizabadi et al. 2003; Lim et al. 2007; Li et al. 2008b; Oliferova et al. 2005; Saleh et al. 2009).

Solid-phase extraction (SPE) is a routine extraction method for trace levels of contaminants in environmental samples. Recently, use of nanoparticles (NPs) for sample extraction in SPE is gaining research interest (Lin et al. 2007; Zhao et al. 2008; Huang and Hu 2008a, b; Suleiman et al. 2009; Zargar et al. 2009). Compared with traditional SPE sorbents, NPs offer a significantly higher surface area-to-volume ratio and a short diffusion route, resulting in high extraction capacity, rapid extraction dynamics, and high extraction efficiencies (Hu et al. 2007; Yao et al. 2008; Huang and Hu 2008b). Among different kinds of NPs, magnetic NPs, mainly including Fe<sub>3</sub>O<sub>4</sub> NPs, appears as an interesting advanced composite material. It has received increasing attention in the past decades due to its unique physical and chemical properties and high potential applications in various fields such as cell separation, magnetically assisted drug delivery, enzyme immobilization, and protein separation (Nandigala et al. 1997; Li et al. 2008a, b, c; Li et al. 2007; Whiteaker et al. 2007; Horak et al. 2001). The magnetic NPs with adsorbed sample can be easily collected by using an external magnetic field placed outside of the extraction container without additional centrifugation or filtration of the sample, which makes sampling and collection easier and faster. Moreover, the magnetic NPs may be reused or recycled. These NPs can be coated with surfactants and as a result prevent their aggregation in liquids and improve their chemical stability. The repulsive interactions between particles can be created by coating a surfactant layer on particle surfaces (Li, et al. 2008a, b, c; Sun et al. 2009).

Recently, method development for simultaneous determination of solutes without prior separation has been markedly increased. This was attributed to the availability of powerful instrumentation and robust numerical analytical methods. Multivariate calibration methods such as multiple linear regression (Haaland and Thomas 1988), principal component regression (Al-Degs et al. 2008), and partial least squares regression (Ghasemi and Niazi 2005) are widely used for simultaneous determination with multivariate data, such as those provided by ultraviolet (UV)/visible (vis) absorption (Ghasemi and Zolfonoun 2010), infrared (Kansiz et al. 2000), or fluorescence spectrometry (Madrakian et al. 2009). However, these methods are not suitable when nonlinearity is observed in the system. Artificial neural networks and support vector machines are nonlinear modeling techniques that have attracted increasing interest in recent years. These methods make it possible to eliminate or reduce the effects of the analyte–analyte interaction, the synergistic effect, the multistep process and any other unknown nonlinearity, and often produce superior models compared with models developed by the linear approaches (Hasani et al. 2007).

Projection pursuit regression (PPR) is a nonlinear multivariate regression procedure, which was proposed by Friedman and Stuetzle (1981). It seeks the "interesting" projections of data from high- to lower-dimensional space and tries to find the intrinsic structural information hidden in the high-dimensional data (Huber 1985). With the obtained interesting projections direction, it can be used for further study of visual pattern recognition and regression (Du et al. 2008a; Ren et al. 2007). At present, it has been applied successfully to tackle some chemical problems (Liu et al. 2007; Du et al. 2008b).

In this paper, a SPE method based on sodium dodecyl sulfate (SDS)-coated magnetic nanoparticles is developed for extraction and preconcentration of PAHs from environmental water samples, prior to their simultaneous spectrophotometric determination using principal component analysis-projection pursuit regression (PC-PPR) algorithm. For comparison, partial least squares (PLS), and least squares support vector machine (LS-SVM) methods are also employed for determination of PAHs.

# Theory

### Projection pursuit regression

The PPR method is based on an iterative two-stage process of projection (reduction of parameter space) and smoothing (establishing nonlinear relation). The reduction of the parameter space is essential for the application of smoothing; smoothing in high-dimensional spaces quickly becomes impossible because of data sparsity (Huber 1985). In principle, PPR consists of linear combinations of nonlinear transformations of linear combinations of explanatory variables. The basic model takes the form:

$$y = \overline{y} + \sum_{i=1}^{M} f_i(\alpha_i^T X) + \varepsilon$$

where,  $\overline{y}$  is the average output variable, M is the number of incorporated smooths, f is the smooth function (ridge function),  $\alpha$  is the parameter of projection direction, and  $\varepsilon$  is the residual error after fitting *M* smooths. The vector  $\alpha_i^T X$  is the projection of *X* onto the unit vector  $\alpha_i$ , where the directions  $\alpha_i$  are chosen to optimize model fit. The functions  $f_i$  are unspecified by the model and estimated using a flexible smoothing method. This allows the PPR to be very general as it fits non-linear functions  $f_i$  of any class of linear combinations in *X*. Then it can produce a nonlinear regression model by the summation of a number of ridge functions. The PPR algorithm estimates  $\alpha_i$  by maximum of an index and  $f_i$  by a low dimensional nonparametric regression estimate based on the projected data, iteratively. The procedure is repeated forward until the residual sum of squares (RSS) is less than a predetermined value.

$$RSS = \sum_{i=1}^{n} \left[ y_i - \sum_{i=1}^{M} f_i(\alpha_i^T x_i) \right]^2$$

# Experimental

#### Reagents

All reagents used were of analytical grade and were used as supplied. SDS, methanol, ammonia solution,  $FeCl_3 \cdot 6H_2O$ , and  $FeCl_2 \cdot 4H_2O$ , were purchased from Merck (Germany). Standard stock solutions (1,000 µg mL<sup>-1</sup>) of anthracene, phenanthrene, fluorine, and pyrene were prepared by dissolving the pure solid (Merck) in methanol and stored in dark at 4 ° C. These Standard stock solutions were diluted with methanol and working solutions were prepared daily by adequate dilution with pure water.

# Preparation of magnetic Fe<sub>3</sub>O<sub>4</sub> NPs

The Fe<sub>3</sub>O<sub>4</sub> NPs were prepared by chemical coprecipitation method (Li et al. 2008a, b, c). Briefly, FeCl<sub>3</sub>·6H<sub>2</sub>O (11.68 g) and FeCl<sub>2</sub>·4H<sub>2</sub>O (4.30 g) were dissolved in 200 mL deionized water under nitrogen gas with vigorous stirring at 85 °C. Then, 20 mL of 30 % aqueous ammonia was added to the solution. The color of bulk solution changed from orange to black immediately. After the reaction, the obtained Fe<sub>3</sub>O<sub>4</sub> NPs precipitate was separated from the reaction medium by magnetic field, and washed with 200 mL deionized water four times.



Anthracene

Phenanthrene

Fluorene

300

-Pyrene

**Fig. 1** Effect of SDS concentration on the extraction efficiency of PAHs. Conditions—sample volume, 100 mL;  $Fe_3O_4$  NPs amount, 0.1 g; concentration of analytes, 20 ng mL<sup>-1</sup>; and volume of eluent, 1 mL methanol

200

SDS (mg L<sup>-1</sup>)

Instrumentation and software

100

100

80

60

40

20

0

0

Recovery (%)

A Perkin Elmer (Lambda 25) spectrophotometer with 10 mm quartz cells was used for UV–vis spectra acquisition. The data were processed on a Toshiba computer with Pentium IV as central processing unit (4 Gb RAM) using MATLAB software, version 7.7. The PPR optimization and model results were obtained using the statistical pattern recognition toolbox version 2.11. LS-SVM regression was performed using the LS-SVM lab toolbox version 1.5 (MATLAB toolbox for least-squares support vector machines). PLS and other calculations were carried out using PLS-Toolbox, version 5.8 (Eigenvectors Company).

### Magnetic SPE procedure

A 100-mL sample or standard solution containing SDS (100  $\mu$ g mL<sup>-1</sup>) was transferred in a glassware beaker. Then 50 mg Fe<sub>3</sub>O<sub>4</sub> NPs was added into the sample solution. The mixture was stirred for 15 min.



Fig. 2 Effect of the Fe<sub>3</sub>O<sub>4</sub> NPs amount on the extraction efficiency of PAHs. Conditions—sample volume, 100 mL; SDS concentration, 100 mg  $L^{-1}$ ; concentration of analytes, 20 ng mL<sup>-1</sup>; and volume of eluent, 1 mL methanol

400



Fig. 3 Absorption spectra of four PAHs after magnetic SPE

Subsequently, the  $Fe_3O_4$  NPs were isolated by placing a strong magnet and the supernatant was poured away. Finally, the preconcentrated target analyte absorbed on SDS-coated  $Fe_3O_4$  NPs was eluted with 1.0 mL of methanol and transferred to a quartz cell. The absorption spectra were recorded between 325 and 225 nm.

### Analysis of the real samples

Tap water, well water, river water and mineral water samples were filtered through 0.45  $\mu$ m membrane filter and maintained in glass containers, then stored at a temperature of 4 °C. Suitable aliquots were taken and subjected to preconcentration and determination by the procedure described above.

# **Results and discussion**

### Optimization of extraction conditions

# Influence of SDS concentration on the adsorption of PAHs

The effect of the surfactant amount on the adsolubilization of target compounds was studied in batch mode. As can been seen from Fig. 1, in the absence

 Table 1
 Analytical parameters of the proposed method

of surfactant, the proportion of target compounds that were adsorbed was relatively low. The adsorption amount of PAHs increased remarkably with the increasing amount of SDS. Maximum adsorption was obtained at a concentration of 100 mg L<sup>-1</sup> of SDS. When SDS concentration was above 100 mg L<sup>-1</sup>, the adsorption of the analyte decreased gradually, which may be attributed that the SDS molecules began to form micelles in the bulk aqueous solution and the micelles caused the PAHs to redistribute into the solution again. Given these findings, 100 mg L<sup>-1</sup> was selected as the final addition amount of SDS in the next studies.

### Effect of the sorbent amount

In comparison with traditional sorbents (microsized sorbents), NPs offer a significantly higher surface area-to-volume ratio. Therefore, satisfactory results can be achieved with fewer amounts of NPs. In order to study the effect of the sorbent, 2 to 100 mg of Fe<sub>3</sub>O<sub>4</sub> NPs was added to 100 mL of the sample solution (Fig. 2). The obtained results showed that by increasing the sorbent amounts from 2 up to 50 mg due to increasing accessible sites, extraction recovery increased and after that remained constant. A 50 mg of the Fe<sub>3</sub>O<sub>4</sub> NPs was selected for subsequent experiments.

### Effect of eluent type

According to the reported results (Li et al. 2008a, b, c; Sun et al. 2009), adsorbed analytes on the surface of mineral oxides in the presence of a surfactant can be quantitatively eluted using organic solvents. Therefore, to find the best eluent, different organic solvents were tested. Obtained results showed that among the tested organic solvents, methanol was found to be the superior solvent in comparison with other solvents for

| Analyte      | $\lambda_{\max}$ (nm) | Linear range $(ng mL^{-1})$ | Calibration equation              | $R^2$  | $\begin{array}{c} \text{LOD} \\ (\text{ng mL}^{-1}) \end{array}$ | RSD% (n=6) |
|--------------|-----------------------|-----------------------------|-----------------------------------|--------|--|------------|
| Anthracene   | 251                   | 0.8–20                      | <i>y</i> =0.0894 <i>x</i> +0.0416 | 0.9977 | 0.21   | 4.1        |
| Pyrene       | 240                   | 2-40                        | <i>y</i> =0.0363 <i>x</i> +0.0015 | 0.9984 | 0.45   | 3.9        |
| Phenanthrene | 250                   | 3–40                        | y=0.0296x+0.0234                  | 0.9982 | 0.65   | 4.2        |
| Fluorene     | 260                   | 5-120                       | <i>y</i> =0.0113 <i>x</i> +0.0015 | 0.9970 | 1.66   | 4.5        |
|              |                       |                             |                                   |        |  |            |

 Table 2
 Concentration data of the different mixtures used in the calibration set for the determination of PAHs

| Solution | Concentration (ng mL <sup>-1</sup> ) |        |              |          |  |  |  |  |  |
|----------|--------------------------------------|--------|--------------|----------|--|--|--|--|--|
| number   | Anthracene                           | Pyrene | Phenanthrene | Fluorene |  |  |  |  |  |
| 1        | 20                                   | 0      | 0            | 0        |  |  |  |  |  |
| 2        | 0                                    | 40     | 0            | 0        |  |  |  |  |  |
| 3        | 0                                    | 0      | 40           | 0        |  |  |  |  |  |
| 4        | 0                                    | 0      | 0            | 120      |  |  |  |  |  |
| 5        | 10                                   | 20     | 0            | 0        |  |  |  |  |  |
| 6        | 10                                   | 0      | 20           | 0        |  |  |  |  |  |
| 7        | 10                                   | 0      | 0            | 60       |  |  |  |  |  |
| 8        | 0                                    | 20     | 20           | 0        |  |  |  |  |  |
| 9        | 0                                    | 20     | 0            | 60       |  |  |  |  |  |
| 10       | 0                                    | 0      | 20           | 60       |  |  |  |  |  |
| 11       | 5                                    | 30     | 0            | 0        |  |  |  |  |  |
| 12       | 5                                    | 0      | 30           | 0        |  |  |  |  |  |
| 13       | 5                                    | 0      | 0            | 90       |  |  |  |  |  |
| 14       | 0                                    | 10     | 30           | 0        |  |  |  |  |  |
| 15       | 0                                    | 10     | 0            | 90       |  |  |  |  |  |
| 16       | 0                                    | 0      | 10           | 90       |  |  |  |  |  |
| 17       | 15                                   | 10     | 0            | 0        |  |  |  |  |  |
| 18       | 15                                   | 0      | 10           | 0        |  |  |  |  |  |
| 19       | 15                                   | 0      | 0            | 30       |  |  |  |  |  |
| 20       | 0                                    | 30     | 10           | 0        |  |  |  |  |  |
| 21       | 0                                    | 30     | 0            | 30       |  |  |  |  |  |
| 22       | 0                                    | 0      | 30           | 30       |  |  |  |  |  |
| 23       | 10                                   | 10     | 10           | 0        |  |  |  |  |  |
| 24       | 10                                   | 10     | 0            | 30       |  |  |  |  |  |
| 25       | 10                                   | 0      | 10           | 30       |  |  |  |  |  |
| 26       | 0                                    | 20     | 10           | 30       |  |  |  |  |  |
| 27       | 5                                    | 20     | 10           | 0        |  |  |  |  |  |
| 28       | 5                                    | 20     | 0            | 30       |  |  |  |  |  |
| 29       | 5                                    | 0      | 20           | 30       |  |  |  |  |  |
| 30       | 5                                    | 10     | 20           | 0        |  |  |  |  |  |
| 31       | 0                                    | 10     | 20           | 30       |  |  |  |  |  |
| 32       | 5                                    | 10     | 0            | 60       |  |  |  |  |  |
| 33       | 5                                    | 0      | 10           | 60       |  |  |  |  |  |
| 34       | 0                                    | 10     | 10           | 60       |  |  |  |  |  |
| 35       | 5                                    | 10     | 10           | 30       |  |  |  |  |  |

desorption of analytes from surface of the SDScoated-Fe<sub>3</sub>O<sub>4</sub> NPs. To achieve the highest recovery of the adsorbed PAHs, the effect of the volume of the eluent was also tested. The minimum volume of methanol required for quantitative elution of the retained analytes was 1.0 mL.

### Effect of solution volume

The effect of solution volume was examined by preconcentrating different volumes (50–800 mL) of aqueous solutions spiked with a constant mass of 1.0  $\mu$ g of each analytes. The obtained results showed that when aqueous solution volume was up to 500 mL, recoveries above 95 % were obtained for the four analytes. Thus, 500 mL was considered to be the maximal enrichment volume for water samples.

### Absorption spectra

Figure 3 shows the absorption spectra for individual analytes after magnetic SPE. As can be seen in Fig. 3, an important degree of spectral overlap occurs between the PAHs which may limit the application of conventional calibration methods for analysis of the current PAHs. So PC-PPR, PLS, and LS-SVM were used to obtain quantitative information from the spectra. Spectra of mixtures of PAHs solutions between 325 and 225 nm wavelengths by 1-nm intervals were recorded, and then the data were digitized and stored for late treatment.

# Univariate calibration

Under the optimum conditions calibration curves were constructed for four PAHs by enriching 100 mL aqueous solutions and measuring the absorbances at  $\lambda_{max}$  of each analyte. With the overall SPE procedure and spectrophotometric analysis method mentioned above, quantitative parameters such as linear range, correlation coefficient, detection limit



Fig. 4 Score plots for absorption data matrices of the calibration and prediction sets

and relative standard deviation were evaluated and the results are shown in Table 1. It can be seen that the present method has high sensitivity and good precision. The limits of detection were calculated by using signal-to-noise ratio of 3.

# Multivariate calibration

The first step in the simultaneous determination of the four analytes by multivariate calibration methods involves constructing the calibration and prediction sets. An experimental design was used to maximize statistically the information content in the spectra. A training set of 35 samples was taken (Table 2). To ensure that the predictions are in the subspace of training set, the score plot of first principal component versus second was sketched and all the samples are spanned with the training set scores (Fig. 4).

**Table 4** Statistical parameters obtained by applying the PC-PPR, PLS, and LS-SVM methods to the synthetic mixtures

| Method | Analyte      | NPC <sup>a</sup> | RMSEP | REP(%) |
|--------|--------------|------------------|-------|--------|
| PC-PPR | Anthracene   | 5                | 0.85  | 12.13  |
| PLS    |              | 4                | 1.16  | 15.05  |
| LS-SVM |              | _                | 1.84  | 23.88  |
| PC-PPR | Pyrene       | 2                | 1.33  | 9.34   |
| PLS    |              | 2                | 1.92  | 13.45  |
| LS-SVM |              | _                | 2.18  | 15.23  |
| PC-PPR | Phenanthrene | 5                | 3.13  | 16.87  |
| PLS    |              | 5                | 5.34  | 28.77  |
| LS-SVM |              | -                | 3.30  | 17.78  |
| PC-PPR | Fluorene     | 6                | 3.39  | 7.77   |
| PLS    |              | 4                | 3.98  | 9.13   |
| LS-SVM |              | _                | 2.95  | 6.78   |

<sup>a</sup> Number of principal component

Table 3 Added and found results of synthetic mixtures of PAHs (in nanograms per milliliter)

| Method | Anthracene |       |                 | Pyrene |       | Phenanthrene    |       | Fluorene |                 |       |       |                 |
|--------|------------|-------|-----------------|--------|-------|-----------------|-------|----------|-----------------|-------|-------|-----------------|
|        | Added      | Found | Recovery<br>(%) | Added  | Found | Recovery<br>(%) | Added | Found    | Recovery<br>(%) | Added | Found | Recovery<br>(%) |
| PC-PPR | 7.0        | 6.62  | 94.5            | 10.0   | 9.84  | 98.4            | 25.0  | 23.10    | 92.4            | 45.0  | 41.99 | 93.3            |
| PLS    |            | 6.12  | 87.4            |        | 7.78  | 77.8            |       | 18.37    | 73.5            |       | 40.92 | 90.9            |
| LS-SVM |            | 6.37  | 91.0            |        | 10.70 | 107.0           |       | 21.83    | 87.3            |       | 40.79 | 90.6            |
| PC-PPR |            | 4.99  | 99.8            |        | 14.07 | 93.8            |       | 21.30    | 85.2            |       | 62.29 | 95.8            |
| PLS    | 5.0        | 3.05  | 61.0            | 15.0   | 11.31 | 75.4            | 25.0  | 22.27    | 89.1            | 65.0  | 62.52 | 96.2            |
| LS-SVM |            | 5.67  | 113.4           |        | 14.98 | 99.9            |       | 22.92    | 91.7            |       | 63.65 | 97.9            |
| PC-PPR |            | 13.75 | 114.6           |        | 23.85 | 95.4            |       | 8.61     | 86.1            |       | 23.10 | 92.4            |
| PLS    | 12.0       | 12.42 | 103.5           | 25.0   | 22.62 | 90.5            | 10.0  | 15.46    | 154.6           | 25.0  | 22.73 | 90.9            |
| LS-SVM |            | 9.26  | 77.2            |        | 21.67 | 86.7            |       | 9.71     | 97.1            |       | 27.29 | 109.2           |
| PC-PPR |            | _     | _               |        | 22.47 | 112.4           |       | 24.48    | 81.6            |       | 48.27 | 87.8            |
| PLS    | 0          | _     | _               | 20.0   | 20.06 | 100.3           | 30.0  | 20.13    | 67.1            | 55.0  | 47.51 | 86.4            |
| LS-SVM |            | _     | _               |        | 23.84 | 119.2           |       | 22.45    | 74.8            |       | 53.06 | 96.5            |
| PC-PPR |            | 8.76  | 87.6            |        | _     | -               |       | 16.56    | 82.8            |       | 27.07 | 90.2            |
| PLS    | 10.0       | 9.01  | 90.1            | 0      | _     | -               | 20.0  | 16.83    | 84.2            | 30.0  | 26.44 | 88.13           |
| LS-SVM |            | 8.66  | 86.6            |        | _     | -               |       | 18.59    | 93.0            |       | 24.47 | 81.6            |
| PC-PPR |            | 4.61  | 92.1            |        | 21.99 | 110.0           |       | _        | -               |       | 82.45 | 97.0            |
| PLS    | 5.0        | 4.41  | 88.2            | 20.0   | 19.34 | 96.7            | 0     | _        | -               | 85.0  | 81.25 | 95.6            |
| LS-SVM |            | 4.54  | 90.8            |        | 18.80 | 94.0            |       | _        | -               |       | 83.61 | 98.4            |
| PC-PPR |            | 13.90 | 92.7            |        | 9.62  | 96.2            |       | 17.30    | 86.5            |       | _     | _               |
| PLS    | 15.0       | 13.17 | 87.8            | 10.0   | 8.91  | 89.1            | 20.0  | 16.65    | 83.3            | 0     | _     | _               |
| LS-SVM |            | 11.34 | 75.6            |        | 12.32 | 123.2           |       | 21.69    | 108.5           |       | -     | -               |

| Sample        | Anthracene                                |                 | Pyrene                       |                 | Phenanthrene                 | e               | Fluorene                                  |                 |  |
|---------------|---|-----------------|------------------------------|-----------------|------------------------------|-----------------|---|-----------------|--|
|               | Found (ng mL <sup><math>-1</math></sup> ) | Recovery<br>(%) | Found (ng mL <sup>-1</sup> ) | Recovery<br>(%) | Found (ng mL <sup>-1</sup> ) | Recovery<br>(%) | Found (ng mL <sup><math>-1</math></sup> ) | Recovery<br>(%) |  |
| Tap water     | 4.56 (0.26) <sup>a</sup>                  | 91.2            | 11.48 (0.43)                 | 114.8           | 9.39 (0.43)                  | 93.9            | 18.30 (0.44)                              | 91.5            |  |
| Mineral water | 5.03 (0.09)                               | 100.6           | 9.46 (0.25)                  | 94.6            | 8.86 (0.48)                  | 88.6            | 17.53 (0.63)                              | 87.7            |  |
| River water   | 4.83 (0.12)                               | 96.6            | 9.23 (0.45)                  | 92.3            | 9.58 (0.40)                  | 95.8            | 19.36 (0.55)                              | 96.8            |  |

**Table 5** Determination of PAHs in spiked water samples (N=3; actual concentrations of anthracene, pyrene, phenanthrene, and fluorene is 5, 10, 10, and 20 ng mL<sup>-1</sup>, respectively)

<sup>a</sup> Standard deviation

In this work, principle component analysis (PCA) was applied as a data pretreatment of the calibration data matrix of the analyte mixtures, and the PC scores were then used as the input of the PPR. When the PCA data reduction procedure is applied prior to the construction of the PPR model, its dimensional effect is to increase the numerical stability of the model construction process and reduce the amount of co-linearity between variables. In addition, PCA allows one to eliminate the irrelevant information such as noise or redundancies present in the data matrix. In this study, spectrophotometric data were employed in PPR to predict the concentrations of the corresponding analytes. The spectrophotometric data, before building the PPR models, were subjected to the principal component analysis and decomposed to PC scores. The PC-PPR models including one to ten PC scores were trained. The lowest RMSECV values were obtained with five, two, five, and six input factors for anthracene, pyrene, phenanthrene, and fluorene, respectively. The chemometrics calibrations were computed with the PC-PPR, PLS, and LS-SVM algorithms, using the correlation for the training set concentration and its current data.

For the evaluation of the predictive ability of multivariate calibration models, the root mean square error of prediction (RMSEP) and relative error of prediction (REP) can be used: (Table 4)

$$\text{RMSEP} = \left[\frac{1}{n}\sum_{i=1}^{n} (y_i - \widehat{y}_i)^2\right]^{0.5}$$

$$\operatorname{REP\%} = \frac{100}{\overline{y}} \left[ \frac{1}{n} \sum_{i=1}^{n} \left( y_i - \widehat{y}_i \right)^2 \right]^{0.5}$$

where  $\hat{y}$  is the predicted concentration in the sample *i*, *y* the observed value of the concentration in the sample *i*, and *n* is the number of samples in the prediction set.

Determination of PAHs in synthetic mixtures

The predictive ability and validation of the calibration models was assessed using seven synthetic mixtures of four PAHs. The results obtained from simultaneous determination of anthracene, pyrene, phenanthrene, and fluorene by PC-PPR and PLS methods are listed in Table 3. Table 4 shows RMSEP and REP for synthetic series of PAHs mixtures. It can be seen from this table that for anthracene, phenanthrene, and pyrene the

| Anthracene found (ng mL <sup>-1</sup> ) |  | Pyrene found (ng 1  | $mL^{-1}$ )   | Phenanthrene found   | enanthrene found (ng m $L^{-1}$ ) Fluorene found (n   |  |   |  |  |  |
|---|--|---|---|--|---|--|---|--|--|--|
| Proposed method                         | HPLC   | Proposed method   | HPLC  | Proposed method  | HPLC  | Proposed method  | HPLC  |  |  |  |
| 1.24 (0.27) <sup>a</sup>                | 1.36   | 2.68 (0.33)   | 2.83  | BDL  | BDL   | BDL  | BDL   |  |  |  |
| 4.78 (0.12)                             | 5.23   | 2.36 (0.44)   | 2.03  | 9.58 (0.40)  | 10.26   | BDL  | BDL   |  |  |  |
|   | Anthracene found (<br>Proposed method<br>1.24 (0.27) <sup>a</sup><br>4.78 (0.12) | Anthracene found (ng mL <sup>-1</sup> )           Proposed method         HPLC           1.24 (0.27) <sup>a</sup> 1.36           4.78 (0.12)         5.23 | Anthracene found (ng mL <sup>-1</sup> )Pyrene found (ng mProposed methodHPLCProposed method1.24 (0.27) <sup>a</sup> 1.362.68 (0.33)4.78 (0.12)5.232.36 (0.44) | Anthracene found (ng mL <sup>-1</sup> )Pyrene found (ng mL <sup>-1</sup> )Proposed methodHPLCProposed method1.24 (0.27) <sup>a</sup> 1.362.68 (0.33)2.834.78 (0.12)5.232.36 (0.44)2.03 | Anthracene found (ng mL <sup>-1</sup> )Pyrene found (ng mL <sup>-1</sup> )Phenanthrene foundProposed methodHPLCProposed methodHPLCProposed method1.24 (0.27) <sup>a</sup> 1.362.68 (0.33)2.83BDL4.78 (0.12)5.232.36 (0.44)2.039.58 (0.40) | Anthracene found (ng mL <sup>-1</sup> )Pyrene found (ng mL <sup>-1</sup> )Phenanthrene found (ng mL <sup>-1</sup> )Proposed methodHPLCProposed methodHPLC1.24 (0.27) <sup>a</sup> 1.362.68 (0.33)2.83BDL4.78 (0.12)5.232.36 (0.44)2.039.58 (0.40)10.26 | Anthracene found (ng mL^-1)Pyrene found (ng mL^-1)Phenanthrene found (ng mL^-1)Fluorene found (ng mL^-1)Proposed methodHPLCProposed methodHPLCProposed methodHPLCProposed method1.24 (0.27) <sup>a</sup> 1.362.68 (0.33)2.83BDLBDLBDLBDL4.78 (0.12)5.232.36 (0.44)2.039.58 (0.40)10.26BDL |  |  |  |

Table 6 Determination of PAHs in the well water samples (N=3)

*BDL* below the detection limit

<sup>a</sup> Standard deviation

statistical parameters of PC-PPR models are superior to that of LS-SVM. In the case of fluorene, LS-SVM results are slightly better than those of PC-PPR. This may be explained by the fact that the PPR method performs a flexible regression in a low-dimensional variable space, contrarily to the SVM regression which uses a fixed transfer function (radial basis function). The PC-PPR models were consistently better than PLS, as shown by the resulting figures of merit for these models (Table 4). Since the PPR analysis takes into account both linearity and nonlinearity in the model construction, therefore it yielded more predictive models than PLS analysis.

# Analysis of real water samples

The proposed methodology was applied for the determination of anthracene, phenanthrene, fluorine, and pyrene in different water samples including, tap water, mineral water and river water. The obtained results are given in Table 5. The relative recoveries for the spiked samples were in the range of 87–114 %. Also this method was applied to the determination of PAHs in two well water samples. In addition, the prediction results using PC-PPR were compared with HPLC method. Comparisons of the prediction capability of the proposed method and the HPLC method for determination of PAHs are presented in Table 6. The results demonstrated that the proposed method was suitable for the determination of PAHs in real water samples.

# Conclusions

A novel SPE method based on SDS-coated  $Fe_3O_4$ NPs was developed for the preconcentration of four PAHs in environmental water samples without the need for chromatographic separation. The use of NPs endued the SPE method with high extraction capacity and preconcentration factors. The magnetic separation greatly improved the separation rate while avoided the time-consuming column passing or filtration operation. PC-PPR calibration method was found effective for simultaneous determination of PAHs in water Samples. Multivariate calibration methods offer a simple and a reasonable substitute for expensive liquid chromatography for PAHs quantification in water samples. The results show the strong potential of PC-PPR, as a multivariate calibration method, to be applied to spectral data for multicomponent determinations.

# References

- Al-Degs, Y. S., El-Sheikh, A. H., Al-Ghouti, M. A., Hemmateenejad, B., & Walker, G. M. (2008). Solid-phase extraction and simultaneous determination of trace amounts of sulphonated and azo sulphonated dyes using microemulsion-modified-zeolite and multivariate calibration. *Talanta*, 75, 904–915.
- Bjorseth, A. (Ed.). (1983). *Handbook of polycyclic aromatic hydrocarbons*. New York: Marcel Dekker.
- Cavalieri, E., & Rogan, E. (1998). Mechanisms of tumor initiation by polycyclic aromatic hydrocarbons in mammals. In A. H. Neilson (Ed.), *The handbook of environmental chemistry, part J. PAHs and related compounds* (p. 87). Berlin: Springer.
- Du, H., Wang, J., Hu, Z., & Yao, X. (2008a). Quantitative structure-retention relationship study of the constituents of saffron aroma in SPME-GC–MS based on the projection pursuit regression method. *Talanta*, 77, 360–365.
- Du, H., Wang, J., Watzl, J., Zhang, X., & Hu, Z. (2008b). Prediction of inhibition of matrix metalloproteinase inhibitors based on the combination of projection pursuit regression and grid search method. *Chemometrics and Intelligent Laboratory Systems*, 93, 160–166.
- Friedman, J. H., & Stuetzle, W. (1981). Projection pursuit regression. Journal of the American Statistical Association, 76, 817–823.
- Geffard, O., Geffard, A., His, E., & Budzinski, H. (2003). Assessment of the bioavailability and toxicity of sediment-associated polycyclic aromatic hydrocarbons and heavy metals applied to *Crassostrea gigas* embryos and larvae. *Marine Pollution Bulletin*, 46, 481–490.
- Ghasemi, J., & Niazi, A. (2005). Spectrophotometric simultaneous determination of nitroaniline isomers by orthogonal signal correction–partial least squares. *Talanta*, 65, 1168– 1173.
- Ghasemi, J., & Zolfonoun, E. (2010). Simultaneous spectrophotometric determination of trace amounts of uranium, thorium, and zirconium using the partial least squares method after their preconcentration by α-benzoin oxime modified Amberlite XAD-2000 resin. *Talanta*, 80, 1191–1197.
- Haaland, D., & Thomas, E. (1988). Partial least-squares methods for spectral analyses. Relation to other quantitative calibration methods and the extraction of qualitative information. *Analytical Chemistry*, 60, 1193–1200.
- Hasani, M., Yaghoubi, L., & Abdollahi, H. (2007). Kinetic spectrophotometric method for simultaneous determination of glycine and lysine by artificial neural networks. *Analytical Biochemistry*, 365, 74–81.
- Horak, D., Rittich, B., Safar, J. A., Spanova, A., Lenfeld, J., & Benes, M. J. (2001). Properties of RNase A immobilized on magnetic poly(2-hydroxyethyl methacrylate) microspheres. *Biotechnology Progress*, 17, 447–452.

- Hu, J., Lo, I. M. C., & Chen, G. (2007). Comparative study of various magnetic nanoparticles for Cr(VI) removal. Separation and Purification Technology, 56, 249–256.
- Huang, C., & Hu, B. (2008a). Speciation of inorganic tellurium from seawater by ICP-MS following magnetic SPE separation and preconcentration. *Journal of Separation Science*, 31, 760–767.
- Huang, C., & Hu, B. (2008b). Silica-coated magnetic nanoparticles modified with  $\gamma$ -mercaptopropyltrimethoxysilane for fast and selective solid phase extraction of trace amounts of Cd, Cu, Hg, and Pb in environmental, and biological samples prior to their, determination by inductively coupled plasma mass spectrometry. *Spectrochimica Acta B*, 63, 437–444.
- Huber, P. J. (1985). Projection Pursuit (with discussion). *The Annals of Statistics, 13,* 435–475.
- Kansiz, M., Billan-Jacobe, H., & Mcnaughtoni, D. (2000). Quantitative determination of the biodegradable polymer poly(b-hydroxybutyrate) in a recombinant *Escherichia coli* strain by use of mid-infrared spectroscopy, and multivariative statistics. *Applied and Environmental Microbiology*, 66, 3415–3420.
- Li, Y., Liu, Y. C., Tang, J., Lin, H. Q., Yao, N. S., et al. (2007). Fe3O4-Al2O3 magnetic core-shell microspheres for rapid and highly specific capture of phosphopeptides with mass spectrometry analysis. *Journal of Chromatography. A*, *1172*, 57–71.
- Li, Y., Qi, D. W., Deng, C. H., Yang, P. Y., & Zhang, X. M. (2008a). Novel Fe<sub>3</sub>O<sub>4</sub>-TiO<sub>2</sub> core-shell microspheres for selective enrichment of phosphopeptides in phosphoproteome analysis. *Journal of Proteome Research*, 7, 1767– 1777.
- Li, C. F., Wong, J. W. C., Huiea, W., & Choia, M. M. F. (2008b). On-line flow injection-cloud point preconcentration of polycyclic aromatic hydrocarbons coupled with highperformance liquid chromatography. *Journal of Chromatography. A*, 1214, 11–16.
- Li, J., Zhao, X., Shi, Y., Cai, Y., Mou, S., & Jiang, G. (2008c). Mixed hemimicelles solid-phase extraction based on cetyltrimethylammonium bromide-coated nano-magnets Fe<sub>3</sub>O<sub>4</sub> for the determination of chlorophenols in environmental water samples coupled with liquid chromatography/spectrophotometry detection. *Journal of Chromatography. A*, *1180*, 24–31.
- Lim, L. W., Okouchi, Y., & Takeuchi, T. (2007). On-line preconcentration of trace carcinogenic polycyclic aromatic hydrocarbons (PAHs) in microcolumn liquid chromatography via large volume injection. *Talanta*, 72, 1600–1608.
- Lin, P. C., Tseng, M. C., Su, A. K., Chen, Y. J., & Lin, C. C. (2007). Functionalized magnetic nanoparticles for smallmolecule isolation, identification, and quantification. *Analytical Chemistry*, 79, 3401–3408.
- Liu, H., Yao, X., Liu, M., Hu, Z., & Fan, B. (2007). Prediction of gas-phase reduced ion mobility constants (K0) based on the multiple linear regression and projection pursuit regression. *Talanta*, 71, 258–263.
- Madrakian, T., Afkhami, A., & Mohammadnejad, M. (2009). Simultaneous spectrofluorimetric determination of levodopa and propranolol in urine using feed-forward neural networks assisted by principal component analysis. *Talanta*, 78, 1051–1055.

- Nandigala, P., Chen, T. H., & Yang, C. (1997). Immunomagnetic isolation of islets from the rat pancreas. *Biotechnology Progress*, 13, 844–851.
- Neff, J. M. (1979). *Polycyclic aromatic hydrocarbons in the aquatic environment*. London: Applied Science Publishers.
- Oliferova, L., Statkus, M., Tsysin, G., Shpigun, O., & Zolotov, Y. (2005). On-line solid-phase extraction and HPLC determination of polycyclic aromatic hydrocarbons in water using fluorocarbon polymer sorbents. *Analytica Chimica Acta*, 538, 35–40.
- Pagnout, C., Rast, C., Veber, A. M., Poupin, P., & Férard, J. F. (2006). Ecotoxicological assessment of PAHs and their dead-end metabolites after degradation by mycobacterium sp. strain SNP11. Ecotoxicology and Environmental Safety, 65, 151–158.
- Pena, M. T., Casais, M. C., Mejuto, M. C., & Cela, R. (2009). Development of an ionic liquid based dispersive liquid– liquid microextraction method for the analysis of polycyclic aromatic hydrocarbons in water samples. *Journal of Chromatography. A*, 1216, 6356–6364.
- Ren, Y., Liu, H., Yao, X., & Liu, M. (2007). Prediction of ozone tropospheric degradation rate constants by projection pursuit regression. *Analytica Chimica Acta*, 589, 150–158.
- Saleh, A., Yamini, Y., Faraji, M., Rezaee, M., & Ghambarian, M. (2009). Ultrasound-assisted emulsification microextraction method based on applying low density organic solvents followed by gas chromatography analysis for the determination of polycyclic aromatic hydrocarbons in water samples. *Journal of Chromatography. A*, 1216, 6673–6679.
- Shariati-Feizabadi, S., Yamini, Y., & Bahramifar, N. (2003). Headspace solvent microextraction and gas chromatographic determination of some polycyclic aromatic hydrocarbons in water samples. *Analytica Chimica Acta*, 489, 21–31.
- Suleiman, J. S., Hu, B., Peng, H., & Huang, C. (2009). Separation/ preconcentration of trace amounts of Cr, Cu and Pb in environmental samples by magnetic solid-phase extraction with Bismuthiol-II-immobilized magnetic nanoparticles and their determination by ICP-OES. *Talanta*, 77, 1579–1583.
- Sun, L., Zhang, C., Chen, L., Liu, J., Jin, H., Xu, H., et al. (2009). Preparation of alumina-coated magnetite nanoparticle for extraction of trimethoprim from environmental water samples based on mixed hemimicelles solid-phase extraction. *Analytica Chimica Acta*, 638, 162–168.
- Whiteaker, J. T., Zhao, L., Zhang, L. C., Feng, H. Y., Piening, B. D., Anderson, L., et al. (2007). Antibody-based enrichment of peptides on magnetic beads for mass-spectrometrybased quantification of serum biomarkers. *Analytical Biochemistry*, 362, 44–54.
- Yao, N., Chen, H., Lin, H., Deng, C., & Zhang, X. (2008). Enrichment of peptides in serum by C8-functionalized magnetic nanoparticles for direct matrix-assisted laser desorption/ionization time-of-flight mass spectrometry analysis. *Journal of Chromatography A*, 1185, 93–101.
- Zargar, B., Parham, H., & Hatamie, A. (2009). Modified iron oxide nanoparticles as solid phase extractor for spectrophotometric determination and separation of basic fuchsin. *Talanta*, 77, 1328–1331.
- Zhao, X., Shi, Y., Wang, T., Cai, Y., & Jiang, G. (2008). Preparation of silica-magnetite nanoparticle mixed hemimicelle sorbents for extraction of several typical phenolic compounds from environmental water samples. *Journal of Chromatography. A, 1188*, 140–147.