Penetration and reproduction of root-knot nematodes on cucurbit species

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Accepted: 2 December 2013 / Published online: 8 January 2014 © KNPV 2014

Abstract Cucurbits are often cultivated in rotation with Solanaceae in double-cropping systems. Most cucurbits have been described as susceptible to root-knot nematodes (RKN) but little is known on their relative levels of susceptibility. Because RKN species differ in rates of root invasion and reproductive traits, isolates of M. arenaria, M. incognita and M. javanica were compared on five cucurbit hosts in experiments run in a climate growth chamber. They included zucchini squash cv Amalthee, cucumber cv Dasher II, melon cv Pistolero, pumpkin cv Totanera and watermelon cv Sugar Baby. All cucurbits were susceptible to the three RKN isolates although M. javanica showed higher invasion rates, faster development and higher egg production than M. arenaria on the selected cucurbits. Apparent differences among cucurbits were primarily due to root invasion rates and formation of egg masses. Both Cucumis species (cucumber and melon) were better hosts for nematode invasion and reproduction than zucchini squash, followed by watermelon. Large invasion rates followed by small reproduction traits were linked to M. incognita on zucchini squash. Reduced invasion rates and egg mass formation along with delayed early development were shown on watermelon.

Keywords Cucurbitaceae · Host suitability · Meloidogyne arenaria · Meloidogyne incognita · Meloidogyne javanica Root-knot nematodes (RKN) are important pests for many vegetable crops worldwide (Karssen 2002). The second-stage juveniles (J2) penetrate the roots and migrate through the intercellular space to the vascular cylinder to initiate and develop a permanent feeding site. Once established, J2 moult three times to become adults. Mature females lay eggs into a gelatinous matrix attached to the posterior end of the female.

Cucurbits are often cultivated in rotation with Solanaceae in double-cropping systems in several vegetable production areas. For instance, pepper is rotated with squash or cucumber (Thies et al. 2004) and tomato with melon or watermelon (Talavera et al. 2012). Most edible cucurbits are hosts of the most widespread rootknot nematodes M. arenaria, M. incognita and M. javanica, but comparative studies on their pathogenic effects on cucurbits are limited. Meloidogyne spp. differ in rates of root penetration (Arens et al. 1981; Khan and Khan 1991a; Ehwaeti et al. 1999; Dutta et al. 2011) and reproduction (Roberts and Thomason 1989) on different hosts (Carneiro et al. 2000); furthermore various plants species differ on their ability to withstand nematode damage (Ehwaeti et al. 1999). Non-host plants do not allow nematode attack, often preventing root penetration and thereby nematode development and reproduction. Resistance is used to describe the ability of a plant to suppress development or reproduction of the nematode. Susceptible plants allow normal nematode development and the expression of any associated disease (Cook and Evans 1987; Roberts 2002). Susceptible plants facilitating the building up of high nematode densities are considered good hosts (Seinhorst 1967), and this ability is generally referred as the reproduction factor (Rf) that is measured as the final

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population density (Pf) divided by the initial population density (Pi). On the contrary, poor host plant often show low Rf. Host plants to which the nematode multiplies but suffer little damage are termed tolerant (Cook and Evans 1987). Large soil infestations may result in high invasion rates that may cause great tissue injury of meristematic cells affecting initial plant growth. This situation can also be detrimental for nematode development, as nematodes will compete for available feeding sites resulting in reduced multiplication rates (Arens et al. 1981). Conversely, high nematode multiplication rates with no plant damage may be achieved with slight soil infestations (Di Vito et al. 1985). Therefore, information on the host-parasite relationship in rotational crops like members of the cucurbit family will be valuable for sustainable management of RKN in double cropping systems.

The objectives of this study were to compare penetration and reproduction of *Meloidogyne arenaria*, *M. incognita* and *M. javanica* isolates on a diversity of cucurbits including zucchini squash, cucumber, melon, pumpkin, and watermelon, and to select the most useful cultivar of cucurbits to include in double-cropping systems with Solanaceae.

Materials and methods

Nematode isolates

The nematode isolates used were *M. arenaria* (code Ma 68), *M. incognita* (code Mi PM26) and *M. javanica* (code Mj 05) from the nematode collection at IRTA, Centre of Cabrils. Nematode cultures were started from the progeny of one single female and they were maintained on susceptible tomato cv Roma in spring-summer and on celery cv D'Elne in autumn-winter in a greenhouse. Juveniles were obtained from infected tomato roots cv Roma by the Baermann tray method (Whitehead and Hemming 1965).

Inoculation process

Cucurbit seeds were soaked overnight and germinated in vermiculite trays for 3 days. Seedlings were transplanted at the cotyledon stage to 20-cm³ capacity clay pots filled with sterilized river sand. Seedlings were allowed to growth 2 weeks for watermelon and 1 week for the others species before nematode inoculation. Individual

seedlings were inoculated with 200 freshly hatched J2 (less than 72 h-old, Pi) of each isolate in approximately 0.5 ml of water.

Plants were maintained in a growth chamber at 26 ± 1 °C with a 16 h light photoperiod, watered as needed and fertilized with a slow-release fertilizer (Osmocote [®] Scotts Company, Netherlands, 15 % N+10 % P₂O₅+12 % K₂O+2 % MgO₂+micro-elements) at the beginning of the test.

Nematode root penetration

A time course experiment was conducted to compare root penetration by three RKN isolates on five cucurbit species. Each RKN isolate and cucurbit combination was replicated seven times. Tests were run separately and repeated two times. The cucurbits included zucchini squash (Cucurbita pepo L.) cv Amalthee, cucumber (Cucumis sativus L.) cv Dasher II, melon (C. melo L.) cv Pistolero, pumkin (Cucurbita maxima Duschesne) cv Totanera and watermelon [Citrullus lanatus (Thunb), Matsum & Nakai] cv Sugar Baby. For each RKN isolate, seedlings were grouped into three sets, one set per harvest date at 4, 7, and 11 days post-inoculation - dpi. At each harvesting time, plants were carefully removed from the pots and the root system washed free of soil. Roots were stained with acid fuchsin 0.05 % (Bridge and Page 1982), and examined under a stereo microscope to count the number of infection sites, and nematodes inside them. Infection sites were recognized because the root tissue was swollen and contained at least one nematode inside. Nematodes were categorized according to their post-embryonic developmental stages as J2 (vermiform), third-stage juveniles (J3, sausage-like) and fourth-stage juvenile (J4, sac-shape) (Taylor and Sasser 1978).

Nematode reproduction

A comparison of RKN reproduction on cucurbit hosts was conducted using the same nematode isolates, cucurbit cultivars except pumpkin that was not included, and experimental conditions. Seedlings were obtained and inoculated as referred before to penetration test. The experimental design was a completely randomized block that included all possible combinations corresponding to 12 treatments (four cucurbit species × three RKN species) with 12 replicates

per treatment. Five plants from each treatment were harvested 7 dpi to determine J2 penetration following the methodology previously described. The remaining seven plants/treatment were uprooted, the roots gently rinsed in water and transplanted into new pots filled with 500 cm³ of sterilized river sand to remove all J2 from root surface and prevent any further root penetration. The experiment was conducted twice.

At 45 dpi (728°-days, basal temperature of 10 °C), the root systems were washed free of soil and weighed. Egg masses (EMs) were stained with a 0.1 g 1^{-1} erioglaucine solution (Aldrich Chemical Company) for 2 h (Omwega et al. 1988) and recorded. Eggs from the entire root system were extracted by maceration in a blender containing a 0.5 % NaOCl solution for 10 min (Hussey and Barker 1973) and counted to determine Pf. Both non-hatched eggs and empty eggs (egg shells) were recorded and the hatching rate was estimated. The fecundity of the females was estimated by dividing Pf by EMs and the Rf (Rf = Pf/Pi) was calculated.

Statistical analyses

The SAS system V8 (SAS Institute Inc., Cary, NC) was used for statistical analyses. Prior to the analyses, when needed, nematode data were log transformed [log10 (x+1)] to homogenize the variances. Data from the root penetration and nematode reproduction experiments were combined because there was no significant difference between the repeated experiments and they were analyzed using analysis of variance (ANOVA). Comparisons were conducted within cucurbit host and within nematode isolate (n=14; seven replications \times two experiments). In addition, data were grouped by nematode isolate (four cucurbits \times 14 replications) and the new set of data subjected to ANOVA. Data from cucurbit species were pooled (three RKN isolates \times 14 replications) and subjected to ANOVA. When the analyses showed statistical differences (P=0.05), the means were separated according to Tukey HSD (Honestly Significant Difference) Test.

Results

Nematode root penetration

Significantly more (P < 0.05) infection sites were found on zucchini squash infected by *M. incognita* (94 ± 12 , mean \pm standard deviation) at 11 dpi than *M. arenaria* (68 \pm 7) followed by *M. javanica* (49 \pm 16). The number of infection sites on watermelon roots (25 \pm 7) at 11 dpi was significantly fewer (*P*<0.05) than on cucumber (44 \pm 6), melon (44 \pm 7) and pumpkin (59 \pm 12) regardless of the RKN isolates.

Root penetration followed a similar pattern on both Cucurbita species (zucchini squash and pumpkin) although M. incognita J2 invaded zucchini squash roots more rapidly and in numbers significantly highest (P < 0.05) than the other two isolates at all harvesting times (Fig. 1a). Final penetration rates on zucchini squash were 98 % for M. incognita, 46 % for M. arenaria and 48 % for M. javanica (Fig. 1a); on pumpkin were 99 %, 70 % and 68 %, respectively (Fig. 1b). On cucumber, M. incognita and M. javanica showed significantly higher (P < 0.05) penetration rates than M. arenaria at all harvesting times, and *M. javanica* was higher (P < 0.05) than *M. incognita* at 11 dpi (Fig. 1c). The final penetration rates were 96 %, 78 %, and 59 %, for M. javanica, M. incognita and M. arenaria, respectively (Fig. 1c). On melon, M. javanica invaded more rapidly and in numbers significantly highest (P < 0.05) than M. incognita followed by M. arenaria, and final penetration rates were 100 %, 84 % and 54 %, respectively (Fig. 1d). On watermelon, root penetration by *M. javanica* (51 %) was significantly higher (P < 0.05) than that of the other two RKN isolates (41 % to 42 %) (Fig. 1e).

At 4 dpi only vermiform J2 were found in roots of cucurbit species (data not shown). At 7 dpi, most penetrating J2 were at the J3-stage on zucchini squash, cucumber, and melon, with the exception of *M. javanica* on zucchini squash. RKN development was delayed on pumpkin and watermelon (Fig. 2a). At 11 dpi most nematodes were at J4-stage in zucchini squash, cucumber, and melon roots except for *M. arenaria* on melon (Fig. 2b). A mixture of J3 and J4 stages occurred on pumpkin, whereas on watermelon were found the three juveniles stages (J2, J3 and J4) with a dominance of the J3-stage (Fig. 2b).

Nematode reproduction

The Pi for this experiment was the number of nematodes that penetrated the roots at 7 dpi. Therefore, differences in Pi values were due to different penetration rates of the isolates on the cucurbit hosts (Table 1).

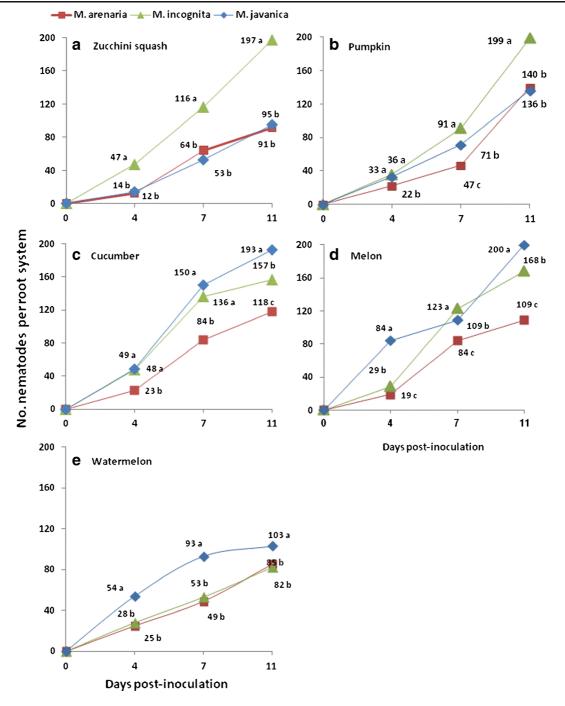


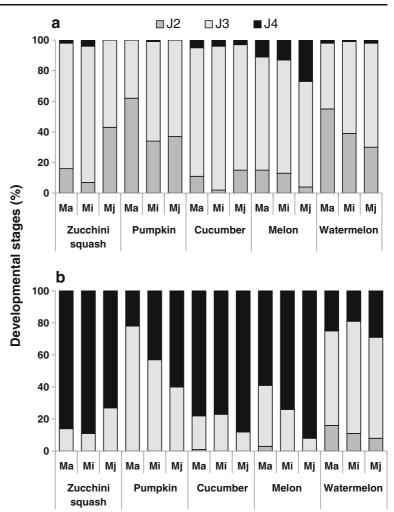
Fig. 1 Number of second-stage juveniles of *Meloidogyne arenaria*, *M. incognita* and *M. javanica* inside the roots of zucchini squash (*Cucurbita pepo*) cv Amalthee (a), pumpkin (*Cucurbita maxima*) cv Totanera (b), cucumber (*Cucumis sativus*) cv Dasher II (c), melon (*Cucumis melo*) cv Pistolero (d), and watermelon

Within cucurbit host, *M. incognita* showed higher Pi values on zucchini squash than *M. arenaria* followed by

(*Citrullus lanatus*) cv Sugar Baby (e) 4, 7, and 11 days postinoculation with 200 juveniles per plant. Values are means of 14 replicates. At each harvesting date, values sharing the same letter do not differ significantly according to the Tukey HDS Test (P=0.05)

M. javanica but there was no difference in EMs among the isolates (Table 1). However, *M. incognita* Pf was

Fig. 2 Post-embryonic development of Meloidogyne arenaria, M. incognita and M. javanica in roots of zucchini squash (Cucurbita pepo) cv Amalthee, pumpkin cv Totanera (Cucurbita maxima), cucumber (Cucumis sativus) cv Dasher II, melon (Cucumis melo) cv Pistolero, and watermelon (Citrullus lanatus) cv Sugar Baby at 7 (a) and 11 (b) days postinoculation of 200 juveniles per plant. Values are means of 14 replicates. J2 - secondstage juveniles; J3 - thirdstage juveniles; J4 - fourthstage juveniles



significantly lower (P < 0.05) than that of the other two isolates. On cucumber, *M. javanica* showed significantly higher (P < 0.05) Pi values than the other two isolates; but only *M. javanica* Pf significantly differed (P < 0.05) from *M. arenaria* Pf (Table 1). On melon, the Pi values of *M. javanica* were significantly higher (P < 0.05) than those of *M. incognita* followed by *M. arenaria*. Besides, *M. javanica* and *M. incognita* showed significantly higher (P < 0.05) Pf than *M. arenaria*. On watermelon, *M. javanica* showed significantly higher (P < 0.05) Pi values than the other two isolates, but only *M. incognita* Pf was higher (P < 0.05) than *M. arenaria* Pf (Table 1).

The combined analysis indicated that root weight was higher in *M. incognita* than *M. arenaria* infected plants (Table 2). *Meloidogyne incognita* and *M. javanica* formed significantly more (P < 0.05) EMs than *M. arenaria*, but *M. javanica* Pf was significantly

higher (P < 0.05) than *M. incognita* Pf (Table 2). However, the fecundity of the females did not differ among the three RKN isolates (Table 2). The hatching rate of *M. arenaria* (20 \pm 2) was lower (*P*<0.05) than that of *M. incognita* (32 ± 2) or *M. javanica* (30 ± 2) . Both Cucumis species (cucumber and melon) showed significantly higher (P=0.05) root weight than zucchini squash, followed by watermelon (Table 3). Significantly higher (P=0.05) EMs were observed on cucumber than on melon or zucchini squash followed by watermelon (Table 3). Statistical differences in Pf, and Rf among the cucurbits consistently corresponded with those observed for EMs. There was no difference in the fecundity of the females among the cucurbit hosts (Table 3). Hatching rates on cucumber (34 ± 2) , melon (30 ± 2) and watermelon (28+2) were comparable and significantly higher than on zucchini squash (22 ± 2) .

Plant species Common name (Cultivar)	Nematode species	Initial population density ^a	Egg masses	Final population density
<i>Cucurbita pepo</i> Zucchini squash (Amalthee)	M. arenaria	50±6 b C	18±9 a B	13857±6542 a AB
	M. incognita	104±10 a C	24±12 a B	5678±4013 b C
	M. javanica	47±6 c D	25±10 a B	15078±7249 a B
<i>Cucumis sativus</i> Cucumber (Dasher II)	M. arenaria	101±7 c A	46±16 b A	24762±14072 b A
	M. incognita	136±9 b A	73±21 a A	32918±10394 ab A
	M. javanica	155±4 a B	77±21 a A	48863±20384 a A
<i>Cucumis melo</i> Melon (Pistolero)	M. arenaria	87±5 c B	16±7 b B	7620±3791b BC
	M.incognita	134±10 b B	30±21 ab B	17481±8929 a B
	M. javanica	152±7 a A	46±22 a B	28355±15089 a AB
<i>Citrullus lanatus</i> Watermelon (Sugar Baby)	M. arenaria	42±10 c D	6±5 b C	4982±5077 b C
	M. incognita	54±7 b D	22±11 a B	17926±8769 a B
	M. javanica	82±13 a C	14±11 ab C	9474±11937 ab C

 Table 1
 Reproductive traits of Meloidogyne arenaria, M. incognita and M. javanica on four cucurbit species 45 days post inoculation with 200 second-stage juveniles (J2) in a growth chamber

Values are mean \pm standard deviation of ten replicates for initial population and 14 for reproductive traits. Values within cucurbit crop in the same column sharing the same lower-case letter are not significantly different. Values within nematode isolate in the same column sharing the same upper-case letter are not significantly different. Mean separation by Tukey HSD Test (P=0.05)

^a Number of J2 inside the roots 7 days post-inoculation

Discussion

All cucurbits were susceptible to the three *Meloidogyne* isolates but significant differences in root penetration and nematode reproduction that persisted through the experimental period were detected. Cucurbits also differed in root galling severity of *M. javanica* and *M. incognita* (Edelstein et al. 2010). The present study confirms that the genetic background of the host as well as that of the nematode affect the host-nematode interaction in good hosts and poor/resistant hosts (Ehwaeti et al. 1999; Fournet et al. 2012; Verdejo–Lucas et al.

2013). Root invasion and formation of egg masses were the primary factors explaining differences among cucurbits and the observed differences were thereafter consistently shown in final population densities and reproduction factors. Nevertheless, females that reached maturity laid similar numbers of eggs regardless the RKN isolate or the cucurbit host which corroborates that female fecundity is not a major factor in the response of the host to the nematode (Arens et al. 1981; Faske 2013). The exception to this trend was *M. incognita* on zucchini squash that showed reduced fecundity (236 eggs/female) in comparison to the other isolates (603 eggs/female). In

 Table 2
 Reproductive traits of three isolates of Meloidogyne spp.

 on cucurbit species (Cucurbita pepo cv Amalthee, Cucumis sativus cv Dasher II, C. melo cv Pistolero and Citrullus lanatus

cv Sugar Baby) grouped by nematode isolate 45 days post inoculation with 200 second-stage juveniles per plant in a growth chamber

Meloidogyne species	Root weight (g)	Egg masses	Final population density	Fecundity ^a	Rf ^b
M. arenaria	9.8±0.6 b	22±2 b	12997±1635 c	731±68 a	64±8 c
M. incognita	11.9±0.6 a	38±2 a	18734±1619 b	629±67 a	93±8 b
M. javanica	10.2±0.6 ab	40±2 a	25389±1619 a	710±67 a	127±8 a

Values are mean \pm standard deviation of 56 replicates (4 cucurbits \times 7 replicates \times 2 experiments). Values in the same column sharing the same letter do not differ according to Tukey HSD Test (P=0.05)

^a Pf/egg masses

^b Rf (Reproduction factor) = Pf/Pi

 Table 3
 Reproductive traits of three isolates of *Meloidogyne* spp., grouped according to cucurbit species 45 days post inoculation with 200 second-stage juveniles per plant in a growth chamber

Plant species Common name (Cultivar)	Root weight (g)	Egg masses	Final population density	Fecundity ^a	Rf ^b
<i>Cucurbita pepo</i> Zucchini squash (Amalthee)	9±0.7 b	22±2.5 bc	11680±1876 bc	708±77 a	58±9 bc
Cucumis sativus Cucumber (Dasher II)	15.3±0.7 a	65±2.4 a	35514±1853 a	569±77 a	178±9 a
<i>Cucumis melo</i> Melon (Pistolero)	15.4±0.7 a	31±2.5 b	18071±1923 b	716±79 a	90±10 b
Citrullus lanatus Watermelon (Sugar Baby)	3±0.7 c	14±2.4 c	10794±1853 c	768±77 a	53±9 c

Values are mean \pm standard deviation of 42 replicates (3 *Meloidogyne* isolates x 7 replicates x 2 experiments). Values in the same column followed by the same letter are not significantly different according to Tukey HDS Test (P=0.05)

^a Pf/egg masses

^b Rf (Reproduction factor) = Pf/Pi

general, *M. javanica* showed greater root penetration, faster development and reproduction than *M. arenaria* on the selected cucurbits. Seemingly, *M. javanica* invaded tobacco roots more rapidly and in greater numbers than *M. arenaria* or *M. incognita* (Arens et al. 1981). The lessen reproductive ability of *M. arenaria* in comparison to *M. javanica* on cucurbits is consistent with similar observations on tomato (Verdejo–Lucas et al. 2013) and could explain the restricted distribution and detection of *M. arenaria* in some vegetable areas (Giné et al. 2012; Talavera et al. 2012).

Both *Cucumis* species (cucumber and melon) were better hosts for nematode invasion and reproduction than zucchini squash followed by watermelon. *Meloidogyne incognita* and *M. javanica* showed similar root penetration pattern, infection and reproduction on cucumber and melon.

The *M. incognita* isolate showed a remarkable interaction with zucchini squash cv Amalthee with significantly greater numbers of penetrating J2, similar egg mass production and lower Pf than the other two RKN isolates. Accordingly, zucchini squash was a poorer host of *M. incognita* than *M. arenaria* or *M. javanica*. The specificity of this interaction for the crop cultivar or RKN isolate is not presently known but deserves further exploration since it resulted in increased root penetration and reduced Pf. This is an interesting combination of effects that might be of utility for the sustainable management of nematode infestations. Zucchini squash has been described as RKN susceptible (Thies et al. 2004) but whether squash cultivars differ in susceptibility levels is unknown. Species-specific and even race or population specific, non-host or resistance responses to *Meloidogyne* spp. were found on cultivars of cauliflower, tomato rootstocks and several wild plants (Khan and Khan 1991b; Ehwaeti et al. 1999; Cortada et al. 2009).

Apparently, the zucchini squash cv Amalthee hindered the development of the nematode from the J4stage to mature egg-laying-female with no effect on penetrating J2 since they progressively developed into immotile J3 and J4 stages. J4-stage juveniles may have died or stop developing as occurred on Cucumis sativus infected by M. hapla (Stephan and Trudgill 1982). Faske (2013) found empty galls on Cucumis melo var. texanus and suggested that juveniles developed to males and had left the roots. An increase in the male/female ratio on Cucumis myriocarpus, a non-host for M. incognita, was reported by Pofu and Mashela (2011). Egression of juveniles after penetration of the roots occurred on resistant Cucumis genotypes (Faske 2013) although it seems little likely to be the case here, since large numbers of penetrating J2 developed into J4-stages. Sub-optimal development of feeding sites unable to supply sufficient nutrients for the nematode could possibly explain the M. incognita-zucchini squash interaction. Such resistance mechanism has been suggested on RKN resistant Cucumis (Walters et al. 2006).

The response of watermelon cv Sugar Baby was differentiated by a great reduction in J2 penetration which suggested a pre-infectional mechanism that was not related to the RKN isolate. Allelochemicals released into the rizhosphere could affect nematode behavior, and thus modify the host recognition process (Dutta et al. 2011). Other mechanisms were retardation in juvenile development and low rates of penetrating J2 becoming egg-laying females. All these mechanisms together make watermelon cv Sugar Baby the less suitable host to the three RKN isolates among the tested cucurbits. Montalvo and Esnard (1994) found that Sugar baby supported the lowest M. incognita root galling and Rf among ten watermelon cultivars and all Rf values were significantly lower on watermelon than tomato. Other cucurbits also showed reduced Rf values in comparison to tomato or eggplants (Anwar and McKenry 2010). These results support the worth of searching for less suitable RKN host since resistance to M. arenaria, M. incognita and M. javanica is not commercially available on Cucumis, Cucurbita or Citrullus. Resistance to *M. hapla* has been reported on some cultivars of squash and melon (Carneiro et al. 2000).

In summary, the main results from this comparative study were: i) high root penetration and reproduction on cucumber and melon irrespective of the RKN isolate, ii) high penetration rates linked to low Pf values on M. incognita-infected zucchini squash, and iii) reduced invasion and delay in post-embryonic development on watermelon. These mechanisms operate on resistant germplasm (Khan and Khan 1991a) and poor host plants (Ehwaeti et al. 1999). Although the tested cucurbits were all susceptible to the three RKN isolates, differences in susceptibility levels were significant; cucumber cv Dasher II followed by melon cv Pistolero were the most susceptible hosts, and watermelon cv Sugar Baby the least. The host status affects not only the damage a crop is likely to suffer but also the residual populations left in the soil which are the inoculum for the next crop in the rotation. Thus, watermelon cv. Sugar Baby could be used for the sustainable management of the disease in double-cropping systems with Solanaceae since Rf were a third lower of that on cucumber. Similarly, zucchini squash cv. Amalthee could be used in M. incognita infested fields.

Acknowledgments The authors acknowledge the financial support provided by Intituto Nacional de Investigaciones Agrarias (INIA), project RTA2010-00017-C02-01 and the European Union through the Regional Development Funds. M. López-Gómez acknowledges INIA for support through a pre-doctoral grant. Thanks are given to Dr. Sorribas for statistical advice. The seeds were kindly provided by Gautier Seeds, Intersemillas S.A., Ramiro Arnedo S. A. and Sakata Vegetables Europe S.A.S.

References

- Anwar, S. A., & McKenry, M. V. (2010). Incidence and reproduction of *Meloidogyne incognita* on vegetable crops genotypes. *Pakistan Journal of Zoology*, 42, 135–141.
- Arens, M. L., Rich, J. R., & Dickson, D. W. (1981). Comparative studies on root invasion, root galling, and fecundity of three Meloidogyne spp. on a susceptible tobacco cultivar. *Journal* of Nematology, 13, 201–205.
- Bridge, J., & Page, L. J. (1982). The rice root-knot nematode, *Meloidogyne graminicola*, on deep water rice (*Oryza sativa* subsp. indica). *Revue Nematology*, 5, 225–232.
- Carneiro, R. M. D. G., Randig, O., Almeida, M., & Diniz Campos, A. (2000). Resistance of vegetable crops to *Meloidogyne* spp.: suggestion for a crop rotation system. *Nematologia Brasileira*, 24, 49–54.
- Cook, R., & Evans, K. (1987). Resistance and tolerance. In R. H. Brown & B. R. Kerry (Eds.), *Principles and Practice* of nematode control in crops (pp. 179–232). Sidney: Academic Press.
- Cortada, L., Sorribas, F. J., Ornat, C., Andrés, M. F., & Verdejo-Lucas, S. (2009). Response of tomato rootstocks carrying the *Mi*-resistance gene to populations of *Meloidogyne arenaria*, *M. incognita* and *M. javanica. European Journal of Plant Pathology*, 124, 337–343.
- Di Vito, M., Greco, N., & Carella, A. (1985). Population densities of *Meloidogyne incognita* and yield of *Capsicum annum*. *Journal of Nematology*, 17, 45–49.
- Dutta, T. K., Powers, S. J., Kerry, B. R., Gaur, H. S., & Curtis, R. H. C. (2011). Comparison of host recognition, invasion, development and reproduction of *Meloidogyne graminicola* and *M. incognita* on rice and tomato. *Nematology*, 13, 509–520.
- Edelstein, M., Oka, Y., Burger, Y., Eizenberg, H., & Cohen, R. (2010). Variation in the response of cucurbits to *Meloidogyne* incognita and *M. javanica. Israel Journal of Plant Sciences*, 58, 77–84.
- Ehwaeti, M. E., Fargette, M., Phillips, M. S., & Trudgill, D. L. (1999). Host status differences and their relevance by *Meloidogyne incognita. Nematology*, 1, 421–432.
- Faske, T. R. (2013). Penetration, post-penetration development and reproduction of *Meloidogyne incognita* on *Cucumis melo* var. texanus. Journal of Nematology, 45, 58–65.
- Fournet, S., Kerlan, M. C., Renault, J. P., Rouaux, C., & Montarry, J. (2012). Selection of nematodes by resistant plants has implications for local adaptation and cross-virulence. *Plant Pathology*, 62, 184–193.
- Giné, A., Bonmatí, M., Sarro, A., Stchigel, A., Valero, J., Ornat, C., Fernández, C., & Sorribas, F. J. (2012). Natural occurrence of fungal egg parasites of root-knot nematodes, Meloidogyne spp. in organic and integrated vegetable production systems in Spain. *BioControl*, 58, 407–416.
- Hussey, R. S., & Barker, K. R. (1973). A comparison of methods of collecting inocula of *Meloidogyne* spp., including a new technique. *Plant Disease*, 57, 1025–1028.
- Karssen, G. (2002). The plant- parasitic nematode genus Meloidogyne Göldi, 1892 (Tylenchida) in Europe. The Netherlands: Leiden.
- Khan, A. A., & Khan, M. W. (1991a). Penetration and development of *Meloidogyne incognita* race 1 and *Meloidogyne*

javanica in susceptible and resistant vegetables. *Nematropica*, 21, 71–77.

- Khan, A. A., & Khan, M. W. (1991b). Reaction of cauliflower cultivars to *Meloidogyne javanica* and races of *Meloidogyne incognita*. *Nematropica*, 21, 161–166.
- Montalvo, A. E., & Esnard, J. (1994). Reactions of ten cultivars of watermelon (*Citrullus lanatus*) to a Puerto Rican population of *Meloidogyne incognita*. Journal of Nematology, 26(Suppl.), 640–643.
- Omwega, C., Thomason, I. J., & Roberts, P. A. (1988). A nondestructive technique for screening bean germplasm for resistance to *Meloidogyne incognita*. *Plant Disease*, 72, 970– 972.
- Pofu, K. M., & Mashela, P. W. (2011). Using relative penetration and maleness indices in *Meloidogyne incognita* to establish resistance type in *Cucumis myriocarpus*. *African Journal of Biotechnology*, 10, 390–393.
- Roberts, P. A. (2002). Concepts and consequences of resistance. In J. L. Starr, R. Cook, & J. Bridge (Eds.), *Plant resistance to parasitic nematodes* (pp. 23–40). New York: CAB International UK.
- Roberts, P. A., & Thomason, I. J. (1989). A review of variability in four *Meloidogyne* spp. measured by reproduction on several hosts including *Lycopersicon*. *Agricultural Zoology Reviews*, 3, 225–252.
- Seinhorst, J. W. (1967). The relationships between population increase and population density in plant parasitic nematode. *Nematologica*, 13, 429–442.

- Stephan, Z. A., & Trudgill, D. L. (1982). Development of four populations of *Meloidogyne hapla* on two cultivars of cucumber at different temperatures. *Journal of Nematology*, 14, 545–549.
- Talavera, M., Sayadi, S., Chirosa-Rios, M., Salmerón, T., Flor-Peregrin, E., & Verdejo-Lucas, S. (2012). Perception of the impact of root-knot nematode induced diseases in horticultural protected crops of south-eastern Spain. *Nematology*, 14, 517–527.
- Taylor, A. L., & Sasser, J. N. (1978). Biology, identification, and control of root-knot nematodes (Meloidogyne species). North Carolina State University Graphics, The genus Meloidogyne (Root-Knot Nematodes) (pp. 1–13). NC: Raleigh.
- Thies, J. A., Davis, R. F., Mueller, J. D., Fery, R. L., Langston, D. B., & Miller, G. (2004). Double-cropping cucumbers and squash after bell pepper for root-knot nematode management. *Plant Disease*, 88, 589–593.
- Verdejo–Lucas, S., Blanco, M., Cortada, L., & Sorribas, F. J. (2013). Resistance of tomato rootstocks to *Meloidogyne* arenaria and *M. javanica* under intermittent elevated soil temperatures above 28°C. Crop Rotation, 46, 57–62.
- Walters, S. A., Wehner, T. C., Daykin, M. E., & Barker, K. R. (2006). Penetration rates of root-knot nematodes into *Cucumis sativus* and *C. metuliferus* roots and subsequent histological changes. *Nematropica*, 36, 231–242.
- Whitehead, A. G., & Hemming, J. R. (1965). A comparison of some quantitative methods of extracting small vermiform nematodes from soil. *Annals Applied Biology*, 55, 25–38.