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Quantification of genetic variability and stable genotype selection over the years in the germplasm of critically endangered Prishanparni (*Uraria picta* Desv.)

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Abstract Prishanparni (Uraria picta Desv.), a critically endangered annual shrub belonging to the family 'Papillionaceae'. It is widely distributed throughout India, Sri Lanka, Bangladesh, Tropical Africa, Malay Islands, and the Philippines. The consistent performances of U. picta accessions based on ten economic traits studied were identified as P-12, P-16, P-21, P-22, P-31, P-47, and P-48. These accessions could be used for commercial cultivation in northern Indian plains. Among the total twentythree studied accessions P-50, P-21, P-48, and P-47 were found superior for rhoifolin content in their aerial as well as root part, which may have various therapeutic potentials used in traditional and modern systems of medicines. These accessions can be exploited for commercial cultivation or in a hybridization program for further crop improvement. Wide range cultivation of the selected accessions in the Indo-Gangetic plains will fit in the existing cropping systems of this region, resulting in comparatively

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better supplementation of herb to the pharmaceutical and herbal drug industries and reducing the pressure on the wild populations.

Keywords Bioactive compounds \cdot Dashmoola \cdot G \times E interactions \cdot Prishanparni \cdot Stability

Introduction

Prishanparni is the popular name for Uraria picta Desv. (2n = 22), a severely endangered annual plant in the Papillionaceae family. It can be found in India, Sri Lanka, Bangladesh, Tropical Africa, the Malay Islands, and the Philippines (Kirtikar et al., 1993; Yusuf et al., 1994; Thong-on et al., 2013). It is one of the constituents in Dashmula, which is utilized in the indigenous system of medicine. The Dashmula drug is a combination of the root of ten medicinal plants, involving Bilva (Aegle marmelos), Agnimantha (Premna integrifolia), Shoynaka (Oroxylum indicum), Gambhari (Gmelina arborea), Patla (Sterospermum suaveolens), Shalaparni (Desmodium gangeticum), Prishnaparni (Uraria picta), Brahati (Solanum indicum), Kantkari (Solanum xanthocarpum), and Gokshuran (Tribulus terrestris), utilized in the treatment of general fatigue, oral sores, and several gynecological disorder. Almost every part of U. picta has medicinal value; the fruits and pods are used to cure mouth sores, while the roots are used to treat chills, cough, and fever

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(Kirtikar et al., 1993; Yusuf et al., 1994). The major biologically active flavonoid, rhoifolin exhibits a wide range of activities exhibiting effectiveness against vasorelaxing (Yadav et al., 2009), hypodynamic (Occuhiuto et al., 1990), pulmonary hypertensive cases (Occuhiuto and Limardi, 1994). It also exhibits anti-inflammatory, anti-thrombotic, anti-hepatic properties and is used in treatment of Alzheimer's disease due to their free radical scavenging activity (Yadav et al., 2009).

It is classified as threatened shrub by the IUCN Red Data Book because of unplanned extensive collection from natural sources for pharmaceutical/herbal medication businesses. The quality of ayurvedic formulations such as Dashmoolarisht, Sudarshanchuran, and others has been harmed by an uneven quality profile reported in the chemical ingredient of the plant. Hence, there is a need for organized commercial cultivation of this shrub to meet its high market demand (Prasad et al., 2012; Rakotondralambo et al., 2013). The quality and efficacy of the herb have been established based on biologically active marker compound rhoifolin (Apigenin-7-O-neohesperidoside) analyzed through liquid chromatographic analysis (Yadav et al., 2009). The ultrasonic extraction (30 min at 40 °C) with methanol solvent is a highly optimized extraction method for screening of rhoifolin in raw material of U. picta and in the herbal products (Yadav et al., 2009).

Since biomass production and biosynthesis of secondary metabolites in medicinal herbs are significantly influenced by genotype (G), environment (E), and genotype (G) \times environmental (E) interactions. Thus, a multi-environment trial was conducted to study the performance of genotypes across a range of environmental conditions, genotype \times environment interaction (GE), and genotype stability. The result of study would be recommending stable genotypes having relatively consistent performance for commercial cultivation in a particular environmental niche (Rahajanirina et al., 2012; Randriamampionona et al., 2007; Thomas et al., 2010).

No breeding strategies on stability of genotype \times environment interaction have been conducted on *U. picta*. The goal of this study is to analyze the Indian Prishnaparni germplasm based on field experiments and identify those accessions (s) which are widely accepted for cultivation in the Indo-Gangetic plains (Fig. 1).

Materials and methods

Planting and experimental lay-out

The twenty-three accessions of U. picta used in this study were originally collected from different geographical regions of Uttar Pradesh and Uttarakhand i.e., Lucknow, Padrauna, Basti, Ambedkar Nagar districts of Uttar Pradesh and Dehradun and Nainital (Haldwani) districts of Uttarakhand, respectively (Table 4). The experiments were conducted at the experimental field station of CSIR-Central Institute of Medicinal and Aromatic Plants (CIMAP), Lucknow, India (northern Indo-Gangetic plains; 25.5°N latitude, 80.5° E longitude, an altitude of 128 m above mean sea level (amsl)). The seeds were exposed to presowing conditions to break the dormancy mechanism imposed by keeping the seeds at room temperature in a glass bottle. Seeds were sown at a depth of 3 cm, during the month of July in each year 2017 and 2018. Seedling emergence and germination in the field were recorded at regular intervals from 7th day of sowing. Four-week-old seedlings with 6-8 leaflets were transplanted during the last week of August each year 2017 and 2018 in the experimental plot (20 m² plots). With the row to row and plant to plant spacing of 15×15 cm² in a randomized block design (RBD) for each consecutive year (2017 and 2018) with three replications per accession. Before transplanting, the soil was fertilized with a standard fertilization regime of 50 kg N + 80 kg P_2O_5 + 80 kg K_2O/ha , respectively. The texture of soil at the experimental location is sandy-loam with pH 7.8. The climate of the experimental site is demonstrated as semi-arid subtropical with severe hot summer and cool winter. The mean maximum (32.9, 32.3) and minimum (23.8, 23.0) temperature (Fig. 2a), mean maximum (83.3, 79.6) and minimum (59.2, 60.3) relative humidity (Fig. 2b) in 2017 and 2018, respectively. The onset of monsoon in this region occurs from the month of June to the month of September. The average rainfall for the planting season is 5.4 and 5.1 mm for each year 2017 and 2018, respectively (Fig. 2c). The soil was subsequently irrigated at 7-10 days intervals as per requirement of the field.

On full maturation, five plants were uprooted from each accession for recording morpho-metric and yield attributing data in November (150 days from sowing) in both years: 2017 and 2018 involving eight



Fig. 1 I. Leaf morphological variation at nursery stage (a: P-2, b: P-47, c: P-48, d: P-51, e: P-52, f: P-50); II. Leaf morphological variation at mature stage (a: P-21, b: P-16, c:

economical traits viz. plant height (cm), root length (cm), inflorescence length (cm), number of leaves per plant, leaf length (cm), leaf width (cm), fresh aerial part yield per plant (gm), and fresh root yield per plant (gm).

HPLC analysis for Rhoifolin

The dried uprooted plants were used to determine the percentage of rhoifolin content in aerial and root parts for the aforementioned accessions using the HPLC method following the protocol of Yadav et al. (2009) for both succeeding years (2017 and 2018).

Statistical analysis

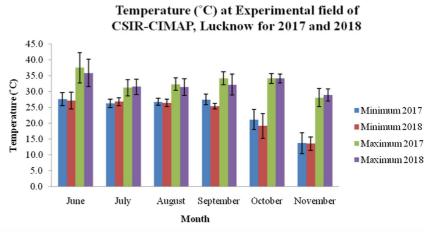
The year-wise analysis of variance (ANOVA), pooled mean analysis of variance (ANOVA), and mean

P-25, **d**: P-50), III. Mature leaf of P-50; IV. Seed colour variation (**a**: P-31, **b**: P-50, **c**: P-10, **d**: P-52, **e**: P-22, **f**: P-51); V. Plant height variation (**a**: P-50, **b**: P-47, **c**: P-4, **d**: P-22)

performance of ten traits of the twenty-three accessions for the two consecutive years (2017 and 2018) were done using statistical analysis software ver. 3.0 available at the Department of Genetics and Plant Breeding of CSIR-CIMAP, Lucknow based on Panse and Sukhatme (1967) and Singh and Chaudhary (1985). All the values at p < 0.01 and p < 0.05 were considered significant.

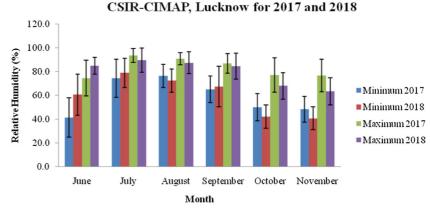
Results and discussion

Analysis of variance revealed that the mean sum of squares due to genotype showed highly significant for all the quantitative and qualitative characters in both years (Tables 1 and 2). The pooled ANOVA for genotype, year, and genotype × year interactions were found to be significant (p < 0.01) for all the

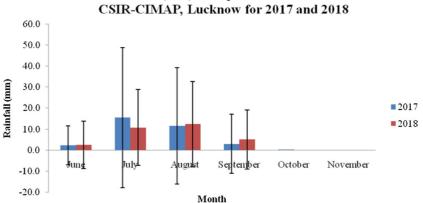


(a) Temperature (°C) at Experimental field of CSIR-CIMAP, Lucknow from the month of June to November of 2017 and 2018.

Relative Humidity (%) at Experimental field of



(b) Relative Humidity (%) at Experimental field of CSIR-CIMAP, Lucknow from the month of June to November of 2017 and 2018.



Rainfall (mm) at Experimental field of

(c) Rainfall (mm) at Experimental field of CSIRCIMAP, Lucknow from the month of June to November - of 2017 and 2018.

Fig. 2 a Temperature (°C) at Experimental field of CSIR-CIMAP, Lucknow from the month of June to November of 2017 and 2018. b Relative Humidity (%) at Experimental field of CSIR-CIMAP, Lucknow from the month of June to November of 2017 and 2018. c Rainfall (mm) at Experimental field of CSIR-CIMAP, Lucknow from the month of June to November of 2017 and 2018

Source of	df	Mean sum	Mean sum of squares (MSS)	(ISS)							
Variation		Plant height	Root length	Inflorescence length	No. of Leaves	Leaf length	Leaf width	Aerial part weight	Root weight	Rhoifolin in aerial Rhoifolin in root part	Rhoifolin in root part
Replication	2	17.9258	4.0537	7.242	3.5801	8.749	0.0632	2.5147	0.558	0.0172	0.0001
Treatment	22	884.65**	22 884.65** 67.7519**	100.9008^{**}	248.715**	23.216^{**}	0.0751^{**}	57.34**	3.982**	0.1557^{**}	0.0074^{**}
Error	44	9.7003	6.1165	22.1974	20.458	1.6721	0.0289	1.7599	0.2885	0.0126	0.0008

r (2018) at CSIR-CIMAP, Lucknow
U. picta grown for IIst year
on 23 genotypes of U .
morpho-chemical traits
(ANOVA) for different
Analysis of variance
Table 2

•				•))	•		
Source of	df	df Mean sum of squares (MSS)	of squares (N	ASS)							
Variation		Plant height	Root length	Inflorescence length	No. of Leaves	Leaf length	Leaf width	Aerial part weight	Root weight	Rhoifolin in aerial Rhoifolin in root part part	Rhoifolin in root part
Replication	2	34.281	23.869	8.0146	0.8828	3.178	0.036	0.193	1.647	0.0007	0.0001
Treatment	52	492.407** 49.775**	49.775**	73.666**	215.333**	16.723^{**}	0.121^{**}	8.668**	4.747**	0.1124^{**}	0.0006^{**}
Error	4	44 50.517 16.97	16.97	8.2417	18.596	2.327	0.03	0.709	0.599	0.007	0.0001

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Source of	d.f	d.f Mean sum of squares (MSS)	f squares (M	SS)							
Variation		Plant height	Root length	Inflorescence length	No. of Leaves	Leaf length	Leaf width	Aerial part weight	Root weight	Rhoifolin in aerial Rhoifolin in roo part part	Rhoifolin in root part
Replication	2	2 21.531	22.355	4.225	0.609	0.689	0.03	0.885	1.681	0.008	0.0001
Treatment	45	774.565**	77.682**	125.84^{**}	228.175**	24.263**	0.094^{**}	35.039**	4.296**	0.129^{**}	0.005^{**}
Years	1	5334.516**	987.746**	1948.133^{**}	286.961**	237.397**	0.019	159.584^{**}	5.544**	0.057*	0.059^{**}
Genotype	22	637.65**	67.602**	1504.91^{**}	359.1**	20.638^{**}	0.146^{**}	40.27^{**}	6.897^{**}	0.162^{**}	0.004^{**}
Genotype × Year 22	22	739.416**	49.926**	2335.578**	104.949^{**}	19.302**	0.051	25.739**	1.833 **		0.004^{**}
Error	90	30.188	11.413	1361.383	19.179	2.205	0.031	1.248	0.446	0.01	0.00041

Fable 3 Pooled Analysis of variance (ANOVA) of $G \times Y$ interaction for different morpho-chemical traits on 23 genotypes of U. *picta* grown for two consecutive years (2017)

studied economic traits except for leaf width which showed non-significance at year and genotype \times year interaction (Table 3). Interaction of rhoifolin content in aerial part at year level was significant at p < 0.05. Evaluation trials were conducted in two years to study the effect of $G \times E$ interaction. For the same reason, the data about environmental variations in the morpho-metric traits and rhoifolin content in aerial and root part (%) of the twenty-three accessions under Lucknow agro-climatic conditions are summarized in Tables 4. The data suggest that significance among treatments at genotype level could be due to collection of genotypes from different geo-climatic zonal distribution and genotype by environment interaction (Gupta, 2018a, 2018b; Lal, 2013; Lal et al., 2019; Shukla et al., 2018). While the significant difference at year and genotype \times year interactions could be attributed to change in humidity, precipitation, climatic conditions, soil condition, or other cultivation practices pursued during the cropping season (Prasad et al., 2014, 2016; Yadav et al., 2018a, 2018b).

Yield and its components are polygenic traits strongly influenced by the environment in U. picta. Summarizing the mean data of the morpho-metric traits and rhoifolin content of twenty-three accessions for each consequent year 2017 and 2018 are presented in Table 4. It was prominently elucidated the significant variation among the genotype, year, and genotype \times year interactions studied. Among the genotypes studied, the most stable plant height was obtained by P-31 (27.66 cm in both years) followed by P-21 (31.33 cm in 2017 and 31.66 in 2018); while the highest plant height was recorded for P-50 (90 cm in 2018), followed by P-51 (87.66 cm in 2017). With regard to root length, the most superior and stable performance was attained by the genotype P-50 (28.33 cm) followed by P-5 (21.66 cm), P-31 (19.33 cm), and P-4 (21.66 cm in 2017 and 22.00 cm in 2018). Plant height and root length are the most sensitive to environmental fluctuations. It is indicated that the relative inconsistent performance of other genotypes was marked due to genotype and environment interaction. For the inflorescence length, the genotype P-50 (35.00 cm in 2018) followed by P-44 (34.33 cm in 2017) projected the highest inflorescence length which could be directly correlated with larger seed formation. The most stable inflorescence length was achieved by P-16 (16.66 cm)

Table 4 M Lucknow	Table 4 Morphological and bioactive compound Lucknow	d bioact	ive com		comparis	comparison of U . picta accessions for two consecutive years	picta E	accession	is for tv	vo cons	ecutive	years (I	I = 2017	and	II = 20	18) und	2018) under agro-climatic conditions	-climat	ic con	litions	of
Accession no	Origin/places of collection	Plant height (cm)	height	Root le (cm)	length	Inflorescence length (cm)	cence (cm)	No. of Leaves		Leaf length (cm)	ngth	Leaf width (cm)		Aerial part weight (gm)	part (gm)	Root weight (gm)	я п	Rhoifolin in aerial part (%)	ц	Rhoifolin in root part (%)	art
Year		1	Π	I	Π	I	II	I	II	I	Π	I	п		п	I	п 1	Π		Π	
P- 1	Lucknow	22.00	44.33	17.33	21.66	25.00	13.33	25.66	25.00	10.66	5.93	1.40	1.50	5.91	4.09	1.47 3	3.08 0	0.37 0	0.40 0.	0.06 0.	0.00
P- 2	Dehradun	25.00	32.00	18.33	26.66	20.33	13.33	20.00	23.66	17.16	6.36	1.36	1.43	5.70	1.63	1.47	.08 0	0.20 0	0.36 0.	0.01 0.	0.01
P- 4	Haldwani	27.00	39.66	21.66	22.00	31.33	11.66	27.00	19.66	12.66	6.10	1.50	1.66	3.76	2.31	1.36	.92 0	0.61 0	0.33 0.	0.03 0.	0.02
P- 5	Haldwani	25.33	44.33	21.66	21.66	31.00	13.66	32.33	22.33	11.33	6.33	1.33	1.50	2.17	1.78	2.42	8.4 0	0.35 0	0.20 0.	0.06 0.	0.01
P-11	Haldwani	23.33	42.33	19.83	16.66	24.66	13.00	33.66	40.00	11.33	6.40	1.50	1.66	2.43	1.81	1.23	.34 0	0.38 0	.47 0.	0.18 0.	0.02
P-12	Haldwani	21.33	40.00	20.17	17.33	25.00	12.33	29.00	22.66	14.00	8.00	1.56	1.33	3.73	2.37	2.10	.61 0	0.43 0	0.57 0.	0.04 0.	0.03
P-16	Haldwani	20.00	52.00	17.83	20.66	16.66	16.66	24.33	27.33	9.09	9.00	1.33	1.33	3.53	2.82	2.20	.66 0	0.36 0	0.24 0.	0.03 0.	0.02
P-17	Haldwani	25.66	41.66	19.33	28.00	24.66	16.00	30.00	23.00	7.63	9.00	1.16	1.46	5.79	2.38	2.54 2	2.59 0	0.09 0	0.23 0.	0.01 0.	0.01
P-21	Haldwani	31.33	31.66	20.16	24.33	29.00	15.66	32.33	31.00	12.16	9.33	1.46	1.33	3.71	1.94	2.52 2	2.43 0	0.27 0	0.73 0.	0.14 0.	0.03
P-22	Haldwani	23.00	56.66	16.33	21.00	21.66	18.33	30.00	30.00	12.66	13.00	1.43	1.36	5.73	1.62	1.37	.52 0	0.46 0	0.69.0	0.05 0.	0.05
P-25	Haldwani	21.00	38.33	12.33	21.66	19.66	15.66	30.00	17.66	10.50	10.00	1.30	1.56	3.55	1.62	1.32	.73 0	0.75 0	0.48 0.	0.03 0.	0.02
P-29	Haldwani	22.66	50.33	17.83	24.33	19.00	13.66	24.33	29.00	9.59	10.66	1.66	1.50	4.69	3.15	1.70	2.00 0	0.33 0	0.20 0.	0.03 0.	0.05
P-31	Haldwani	27.66	27.66	19.33	19.33	17.00	17.00	31.66	31.66	10.66	10.66	1.66	1.16	4.93	4.92	2.54 2	2.53 0	0.34 0	0.27 0.	0.03 0.	0.02
P-34	Haldwani	31.66	47.33	28.33	31.33	26.00	13.00	31.33	26.33	15.66	8.33	1.46	1.40	6.33	4.23	2.72	2.96 0	0.16 0	0.51 0.	0.06 0.	0.04
P-39	Haldwani	27.66	46.66	17.33	22.00	24.33	11.00	37.66	26.00	9.50	10.33	1.26	1.33	2.93	3.29	1.72	2.08 0	0.60 0	0.49 0.	0.10 0.	0.01
P-41	Haldwani	28.00	40.66	20.33	30.33	22.00	13.00	30.00	26.33	11.83	10.33	1.36	1.36	3.16	3.05	1.66	2.63 0	0.22 0	0.45 0.	0.03 0.	0.01
P-44	Haldwani	64.66	45.00	20.33	26.66	34.33	17.00	60.00	35.00	14.66	12.00	1.56	1.66	5.07	2.95	2.31 2	2.32 0	0.69.0	0.34 0.	0.03 0.	0.01
P-47	Haldwani	25.33	42.33	20.83	19.33	21.33	21.00	28.33	24.00	12.86	10.66	1.63	1.40	2.22	2.53	1.16	.98 0	0.67 0	0.39 0.	0.19 0.	0.02
P-48	Haldwani	22.33	53.33	16.33	27.66	25.33	15.66	30.00	31.66	12.50	13.00	1.46	1.36	2.60	4.21	1.52 (0.87 0	0.78 0	0.12 0.	0.09 0.0	0.01
P-49	Haldwani	18.66	51.66	9.43	28.33	11.16	19.00	16.66	31.66	8.83	9.00	1.13	1.43	1.24	5.94	0.73 3	3.62 0	0.24 0	0.18 0.	0.05 0.	0.01
P-50	Padrauna	52.33	90.00	28.33	28.33	22.66	35.00	50.33	58.33	14.43	14.66	1.73	2.10	17.28	8.87	5.75	-	0 10.	0.75 0.	0.07 0.	0.00
P-51	Basti	87.6	28.33	12.33	22.33	29.00	10.66	36.33	22.00	19.33	9.60	1.50	1.13	19.42	1.87	4.53 2	2.67 0	0.43 0	0.67 0.	0.01 0.	0.02
P-52	Ambedkar Nagar	57.66	31.00	9.00	25.33	12.00	14.66	21.66	22.00	10.66	10.66	1.33	1.26	5.43	2.47	3.43	1.61 0	0.39 0	0.09 0.	0.02 0.	0.01

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I = Ist Year = 2017; II = IInd Year = 2018

followed by P-31 (17.00 cm) and P-47 (21.33 cm for 2017 and 21.00 for 2018).

Plant height, leaf, and seed morphological variation are the primary descriptor for characterization of germplasm. However, very few attempts have been conducted on characterizing the germplasm/accessions based on descriptor data in U. picta. In the present study plant, seed, and leaf descriptor data for diverged accessions in the collection is represented in Fig. 1. These descriptor data are important for the identification and categorization of new or existing clones, identification of duplicate accessions, determining relative relationships among accessions, establishment of core collections, and selecting parental materials in breeding programs (Huaman, 1992). Leaf morphological characteristics reflected wide range of diversity among collected germplasm of U. picta. In relation to the number of leaves, it was observed that P-22 (30.00 cm) and P-31 (31.66 cm) demonstrated the most stable trait for the two years studied followed by P-21 (32.33 cm for 2017 and 31.00 for 2018). While in reference to leaf length, the stable performance was presented by P-31 (10.66 cm) followed by P-16 (9.09 cm in 2017 and 9.00 cm in 2018) and in case of leaf width, genotype P-16 (1.33 cm) followed by P-41 (1.36 cm) was recorded stable in the study.

Aerial part weight and root part weight are the major economic traits and are important for successful agronomic practices and industrial demand. Thus, exploiting the highest aerial part weight was observed in genotype P-51 (19.42 gm in 2017) followed by P-50 (17.28 gm in 2017 and 8.87 gm in 2018), whereas for

root part weight, genotype P-50 (5.75 gm in 2017 and 7.27 gm in 2018) was recorded highest. While the most stable genotype for aerial part weight was recorded in P-31 (4.93 gm in 2017 and 4.92 gm in 2018) followed by P-41 (3.16 gm in 2017 and 3.05 gm in 2018), and root part weight was revealed by genotype P-44 (2.31gm and 2.32 gm) followed by P-31 (2.52 gm and 2.53 gm) and P-17 (2.54 gm and 2.59 gm) in 2017 and 2018, respectively.

WHO and modern pharmacopeia give strong emphasis on the content (%) of biologically active compounds for determining the quality and efficacy of medicinal plants (Saxena et al., 2016; Sharma et al., 2010; Vasudevan, 2009). The pooled mean percentage of rhoifolin content in aerial parts and root parts of the twenty-three accessions ranged from 0.16% to 1.01% and 0.12% to 0.75% in aerial parts and from 0.01% to 0.19% and 0.00% to 0.05% in root parts for 2017 and 2018, respectively (Fig. 3). The accession P-50 (1.01%) was recorded to establish the highest rhoifolin content (%) in aerial parts followed by accession P-48 (0.78%) and P-25 (0.75%), while the highest rhoifolin content in the root part was depicted by P-47 (0.19%) followed by P-11 (0.18%) and P-21 (0.14%) (Table 4 and Fig. 4). A superior trend of rhoifolin content in both aerial and root part were observed in P-50 (1.01% and 0.07% in 2017), P-21 (0.73% in 2018 and 0.14% in 2017), P-48 (0.78% and 0.09% in 2017) and P-47 (0.67% and 0.19% in 2017), respectively. Rhiofolin content is higher in the aerial part than in the root part suggesting that the harvesting of aerial parts would be helpful for sustainable exploitation of herb and natural

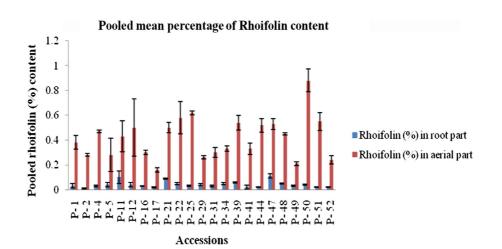
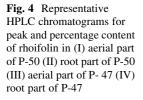
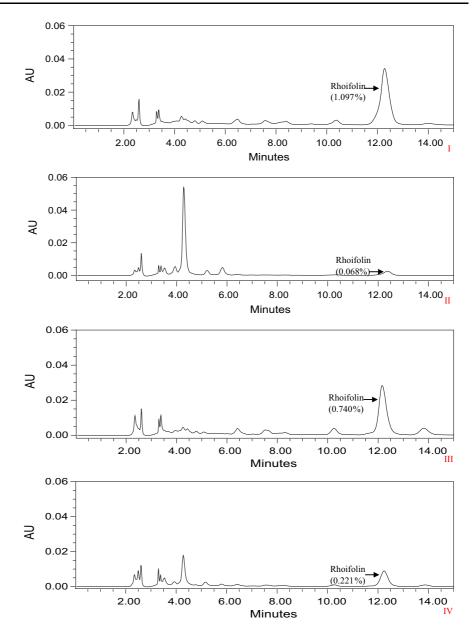


Fig. 3 Pooled mean percentage of rhoifolin content in aerial parts and root parts for two consecutive years (2017 and 2018)





conservation. Few accessions, on the other hand, demonstrated inconsistent rhoifolin content throughout two years, which was most likely due to the consequence of polygenic control, environmental or nongenetic influences, and their combined interaction ($G \times E$ interaction). (Kang, 1998; Lal et al., 2018; Prasad et al., 2014; Sarkar & Lal, 2018).

When the performance of the traits was compared between the two years, it was clear that the accessions performed superior for different morpho-metric and yield attributing traits viz., inflorescence length, leaf length, aerial part weight, rhoifolin content in aerial parts, and rhoifolin content in root part in the first year (2017). The accessions performed superior for plant height, root length, and root weight characters in the second consecutive year (2018), while leaf width exhibited non-significant performance for both years.

Overall based on the ten morpho-chemical attributes, the accessions P-12, P-16, P-21, P-22, P-31, P-47, and P-48 displayed a comparable consistent performances pattern in the agro-climatic conditions of the Indo-Gangetic plains for the two consecutive years studied (Table 4). P-50, P-21, P-48, and P-47 had higher rhoifolin content in both aerial and root parts. Other genotypes responded differently in terms of yield-attributing traits in both the years. The result revealed that the genotype-by-environment interaction complicates the process of crop variety generation, reduces the effectiveness of a breeding program aiming at yield improvement (Ahmad et al., 2011). Nonetheless, eight accessions, P-12, P-16, P-21, P-22, P-31, P-47, P-48, and P-50, may be utilized for continued commercial cultivation on a large scale based on mean performances and stability.

Conclusion

To conserve resources, broad adaptation and performance reliability across a variety of situations are commendable goals. To achieve greater success, researchers have focused on the phenomena of genotype environments ($G \times E$) interaction, which allows them to distinguish genotype performance in various settings and to selectively target appropriate genotypes for commercial cultivation to certain environmental niches. P-12, P-16, P-21, P-22, P-31, P-47, and P-48 were found as the consistently performing U. picta accessions based on ten morpho-chemical attributes studied. These accessions might be employed for commercial production under the agroclimatic conditions of the Indo-Gangetic plains. Accession P-50, P-21, P-48, and P-47, which have higher rhoifolin content in aerial and root parts, have therapeutic potential and could be used in traditional and modern medicine to treat weariness, oral sores, anti-thrombotic, hepatoprotective, and a variety of gynecological problems.

These accessions can help in commercial cultivation, hybridization, and agricultural enhancement. Wide-scale cultivation of selected accessions in the Indo-Gangetic plains will fit well within the region's managed agricultural cropping systems, ensuring a sustainable and far better supplementation of herb to the industry while reducing the impact of unregulated exploitation of wild populations.

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Data availability The datasets used or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Conflict of interest The authors have no conflict of interest.

Consent to participate All authors have read and agreed with their participation.

Consent to publish All authors have read and agreed to the published version of the manuscript.

Human and animal rights No animals were harmed in this research.

References

- Ahmad, B., Lal, H. A., Shahid, R. M., Asif, M., Muhammad, I., & Ramatullah, Q. (2011). Grain yield stability in chickpea (*Cicer arietinum* L.) Across environments. *Pakistan Journal of Botany*, 43(5), 2947–2951.
- Gupta, V. (2018a). Smart materials in dentistry: A review. International Journal of Advanced Research, 3(6), 89–96.
- Gupta, V. (2018b). Endodontic and esthetic rehabilitation of a discoloured non-vital young permanent tooth—A case report. *International Journal of Advanced Research*, 3(6), 84–88.
- Huaman, Z. (1992). Morphologic identification of duplicates in collections of Ipomoea batatas. CIP Res. Guide 36, International Potato Center (CIP), Lima, Peru.
- Kang, M. S. (1998). Using genotype-by-environment interaction for crop cultivar development. Advances in Agronomy, 62, 199–253.
- Kirtikar, K. R., & Basu, B. D. (1993). 'Indian Medicinal Plant' Orient Enterprises, Dehradun, India. 1:496–498.
- Lal, R. K. (2013). On genetic diversity in germplasm of vetiver 'Veteveria zizanioides (L.) Nash'. Industrial Crops and Products, 43, 93–98.
- Lal, R. K., Chanotiya, C. S., Singh, V. R., Dhawan, S. S., Gupta, P., Shukla, S., & Mishra, A. (2019). Induced polygenic variations through γ- rays irradiation and selection of novel genotype in chamomile (*Chamomilla recutita* [L.] Rauschert}. *International Journal of Radiation Biology*, 95(9), 1242–1250.
- Lal, R. K., Gupta, P., & Sarkar, S. (2018). Phylogenetic Relationships, path and principal component analysis for genetic variability and high oil yielding clone selection in vetiver (*Vetiveria zizanioides* L.) nash. *Journal of Plant Breeding and Genetics*, 2(1), 105–113.

- Occuhiuto, F., Circosta, C., De Pasquale, A., & Briguglio, F. (1990). Comparative hemodynamic effects of the flavonoids rhoifolin and vitexin in the dog. *Phytotherapy Research*, 4(3), 118–120.
- Occuhiuto, F., & Limardi, F. (1994). Comparative effects of the flavonoids luteolin, apiin and rhoifolin on experimental pulmonary hypertension in the dog. *Phytotherapy Research*, 8(3), 153–156.
- Panse, V. G., & Sukhatme, P. V. (1967). Statistical methods for agricultural workers (2nd ed.). Indian Council of Agricultural Research.
- Prasad, A., Dhawan, S. S., Mathur, A. K., Prakash, O., Gupta, M. M., Verma, R. K., Lal, R. K., & Mathur, A. (2014). Morphological, chemical and molecular characterization of *Centella asiatica* germplasms for commercial cultivation in the indo-gangetic plains. *Natural Product Communications*, 9(6), 779–784.
- Prasad, A., Yadav, K. S., Yadav, N. P., Mathur, A., Sreedhar, R. V., Lal, R. K., & Mathur, A. K. (2016). Biomass and centellosides production in two elite *Centella asiatica* germplasms from India in response to seasonal variation. *Industrial Crops and Products*, 94, 711–720.
- Prasad, A., Pragadheesh, V. S., Mathur, A., Srivastava, N. K., Singh, M., & Mathur, A. K. (2012). Growth and centelloside production in hydroponically established medicinal plant-*Centella asiatica* (L.). *Industrial Crops and Products*, 35, 309–312.
- Rakotondralambo, S. O. R., Goud, M. R., Rivallan, R., Lussert, A., Danthu, P., Lamotte, F., Ralambofetra, E., Ramavovololona, P., Noyer, J. L., & Baurens, F. C. (2013). Insight into the biology, genetics and evolution of the *Centella* asiatica polyploidy complex in Madagascar. *Industrial Crops and Products*, 47, 118–125.
- Rahajanirina, V., Raoseta, S. O. R., Roger, E., Razafindrazaka, H., Pirotais, S., Boucher, M., & Danthu, P. (2012). The influence of certain taxonomic and environmental parameters on biomass production and triterpenoid content in the leaves of *Centella asiatica* (L.) Urb. from Madagascar. *Chemistry & Biodiversity*, 9, 298–308.
- Randriamampionona, D., Diallo, B., Rakotoniriana, F., Rabemanantsoa, C., Cheuk, K., Corbisier, A. M., Mahillion, J., Ratsimamanga, S., & Jaziri, M. E. J. (2007). Comparative analysis of active constituents in *Centella asiatica* samples from Madagascar: Application for *ex-situ* conservation and clonal propagation. *Fitoterapia*, 78, 482–489.
- Sarkar, S., & Lal, R. K. (2018). Genetics, inheritance pattern and genotype selection in Isabgol (*Plantago Ovata* Forsk). *SCIOL Genetic Science*, 1, 68–79.
- Saxena, H. O., Mohan, B., & Kakkar, A. (2016). Assessment of variation in rhoifolin content in aerial parts of *Uraria picta*

Desv. From different location of Madhya Pradesh. *Journal* of Pharmacy Research, 10(5), 185–190.

- Sharma, P., Kaushik, S., Jain, A., & Sikarwar, S. M. (2010). Preliminary Phytochemical screening and HPLC fingerprinting of *Nicotiana tabacum* leaf. *Journal of Pharmaceutical Research*, 3(5), 1114–1145.
- Shukla, S., Lal, R. K., Shanker, K., & Singh, S. (2018). Evolutionary pattern on genetic resources and diversity in Indian landraces of Opium Poppy (*Papaver somniferum* L.). Acta Scientific Agriculture, 2(9), 116–122.
- Singh, R. K., & Chaudhary, B. D. (1985). Variance and covariance analysis. Biometrical methods in quantitative genetic analysis (p. 57). Kalyani Publisher.
- Thomas, M. T., Kurup, R., Johnson, A. J., Chandrika, S. P., Mathew, P. J., Dan, M., & Baby, S. (2010). Elite genotypes/chemotypes, with high contents of madecassoside and asiaticoside, from sixty accessions of *Centella asiatica* of south India and the Andaman Islands: For cultivation and utility in cosmetic and herbal drug applications. *Industrial Crops and Products*, 32, 545–550.
- Thong-on, W., Arimatsu, P., Pitiporn, S., Soonthornchareonnon, N., & Prathanturarug, S. (2013). Field evaluation of *in-vitro*-induced tetraploid and diploid *Centella asiatica* (L.) Urban. *Journal of Natural Medicines*, 68, 267–273.
- Vasudevan, H. (2009). DNA fingerprinting in the standardization of herbs and neutraceuticals. *The Science Creative Quarterly* 4.
- Yadav, A. K., Yadav, D., Shanker, K., Verma, R. K., Saxena, A. K., & Gupta, M. M. (2009). Flavone glycoside based validated RP-LC method for quality evaluation of Prishnaparni (*Uraria picta*). *Chromatographia*, 69, 653–658.
- Yadav, R., Lal, R. K., & Gupta, M. M. (2018). Prediction of morpho-genetic variability at the intra-specific level and elite line selection in Black Henbane (*Hyoscyamus niger* L.). Acta Scientific Agriculture, 2(9), 94–103.
- Yadav, R., Lal, R. K., & Singh, V. R. (2018). Intra-specific differentiation, genetic variability and their prospect for exploitation in medicinally important plant Black henbane (*Hyoscyamus niger* L.). *Trends in Phytochemical Research*, 2(3), 147–154.
- Yusuf, M., Chowdhury, J. U., Wahab, M. A., & Begum, J. (1994). The Essential Oil Is Used to Treat Cholera and as an Antiseptic and Disinfectant. Medicinal Plants of Bangladesh. BCSIR Laboratories, Chittagong, Bangladesh.

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