

Diet of finfish targeted by fishers in North West Australia and the implications for trophic cascades

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Abstract Detailed information of fish diets is required if we are to understand complex interactions between species and successfully manage resources at an ecosystem level. We compiled diet information from 76 species of fish targeted by recreational and commercial fishers in North West Australia. Based on 81 independent studies we demonstrate that species targeted by the fishery are all carnivores, however the type of prey they consume and their trophic level is variable (3.31–4.49) and trophic range of some species spans different trophic levels (e.g. *Lethrinus nebulosus*, 3.46–4.35). These findings infer that in highly diverse systems, such as coral reefs, trophic cascades instigated by fishing must be investigated at the species, rather than functional or trophic level. Moreover, as prey availability is likely to vary spatially and temporally, diet must be quantified locally to assess ecosystem level impacts of fishing.

Keywords Trophic cascades · Predator prey relationships · Coral reefs · Ecosystem based management

Introduction

Owing to their high commercial value and catchability, large predatory fishes are often selectively harvested in the recreational and commercial fisheries of the world (Pauly et al. 1998; Jennings et al. 2001). The suite of life history characteristics that typifies the majority of these large predatory fish, i.e. being long-lived, slow-growing and with advanced ages at maturity (Reynolds et al. 2005), infers low rates of intrinsic population growth. As a consequence these species are extremely vulnerable to over exploitation (Myers et al. 1997; Denney et al. 2002). In evidence of this inherent vulnerability, recent estimates have shown that populations of large predatory fish have been declining at a rapid pace worldwide (Friedlander and DeMartini 2002; Myers and Worm 2003; Baum et al. 2003; Myers et al. 2007).

While the consequences of removing predators from marine ecosystems are not well understood (Hughes et al. 2003; Hawkins and Roberts 2004), their depletion in fish assemblages has been linked to a lowering of mean trophic level (Pauly et al. 1998; Jennings et al. 2002; Pinnegar et al. 2002; Nicholson and Jennings 2004; Essington et al. 2006), and the alteration of trophic guild structure (Link and Garrison 2002; Allen and Clarke 2007; Watson et al. 2007). The removal of top predatory fish is also likely to have significant indirect effects on marine ecosystems, influencing a range of ecological processes that might include predation, competition and herbivory

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(Babcock et al. 1999; Pinnegar et al. 2000; Willis and Anderson 2003).

Understanding the potential impacts of declines in biomass of large predatory fish is essential for the management of marine ecosystems and the fisheries that they sustain. Ecosystem models are a common method for exploring the wider effects of such top-predator declines (Pauly et al. 2000a), but these rely on the model creator having a detailed knowledge of trophic linkages within the particular system being examined. Dietary composition data is an important component of this required knowledge, yet these data are often only available for a very limited range of species in a given area or ecosystem. The strong push towards the modeling of marine ecosystems in recent times, particularly in data-poor regions, has often led to a heavy reliance on data obtained from outside the local area. For highly specialized feeders whose diet does not vary spatially information for such models may be gathered from several different locations. However, for species with a generalist diet the trophic level will reflect availability of prey and is likely to vary both spatially and temporally. Identifying generalists and specialists and using appropriate dietary information is therefore an important facet of building credible ecosystem models.

In this paper, we summarize all available dietary data for a range of recreationally and commercially important fish species from the inshore waters of North West Australia. Trawl and trap fisheries along this coast target lethrinids, lutjanids and serranids, producing ~3,000 t of finfish, worth 12 million AUD, annually (Fletcher and Santoro 2008). There are also significant fisheries for mackerel (predominantly *Scomberomorus* spp.), a gillnet fishery that targets barramundi (*Lates calcarifer*) and threadfin salmon (e.g. *Eleutheronema tetradactylum*), a shark fishery (predominantly *Carcharhinus* spp.) and an expanding recreational fishery. Clearly the focus in this region is on fish from higher trophic levels and, unlike many other tropical fisheries, herbivorous fish species are generally not targeted. Indirect effects of fishing are therefore not confounded by removal of herbivorous prey species and may be more apparent.

Based on the compilation and comparison of multiple studies, conducted in locations around the world, we assess how diet of fisheries targeted finfish may vary spatially. We also identify the fish families most commonly preyed upon, providing an indication

of what species may benefit from removal of predators and how this may affect the ecosystem. This information will facilitate a better understanding of the indirect effects of fishing, highlighting gaps in our current knowledge of fish diets and provide direction for future research in this region.

Methods

Diet of fish recreationally and/or commercially targeted in the inshore waters of N W Australia was assessed using gut content information collected from locations around the world. A paucity of local data precluded exclusive use of data collected from N W Australia, however by taking this global approach we were able to use variation in trophic level as an indicator of plasticity in species diet. A list of key targeted fish species was formed using data obtained from recreational catch surveys (Sumner et al. 2002; Williamson et al. 2006), and from several years of commercial catch statistics (Fletcher and Head 2006; Fletcher and Santoro 2007, 2008), for the northern Gascoyne (including Ningaloo Marine Park), Pilbara and Kimberley bioregions. This list was expanded to include predatory fish species, which may be competitors to the main target species of the aforementioned fisheries and could thus benefit from their removal.

Systematic, computerized literature searches were conducted to find all available dietary data on these species, irrespective of where the study was conducted. Abstracting and indexing services, such as Aquatic Sciences and Fisheries Abstracts (ASFA), Biological Abstracts (BIOSYS), Web of Science (WoS) and Google Scholar were searched for peer-reviewed articles, and grey literature, including books, reports and unpublished theses. Unpublished raw data was also utilized for some fish species.

Information provided from the literature was tabulated by species, study area, sample size (number of stomachs containing food items), length range of specimens, method used for diet expression, and dietary composition. Dietary information was summarized using 11 food item categories: teleosts, chondrichthyans, cephalopods, molluscs (other than cephalopods), crustaceans, echinoderms, annelids, other invertebrates, birds, reptiles, and mammals. These represented the major taxa ingested. Overall

dietary composition for each species was calculated using a weighted average that takes into account the number of fish sampled by each study. The proportion of each prey item category P_j to the overall diet was calculated as;

$$P_j = \frac{\sum_{i=1}^n P_{ij}N_i}{\sum_{j=1}^{11} \left(\sum_{i=1}^n P_{ij}N_i \right)}, \quad (1)$$

where P_{ij} is the proportion of prey category j in study i , N_i is the number of stomachs with food used to calculate P_{ij} in study i , n is the number of studies, j is the number of prey categories, and $\sum P_j = 1$.

Fractional trophic levels were calculated for each individual study using the 11 dietary categories and by substituting into the following equation the average values for each dietary category as described in the TrophLab manual (Eq. 2, Pauly et al. 2000b);

$$TROPH_i = 1 + \sum_{j=1}^n DC_{ij} \times TROPH_j, \quad (2)$$

where $TROPH_j$ is the fractional trophic level of prey j as defined by TrophLab (Pauly et al. 2000b), DC_{ij} represents the fraction of j in the diet of i , and n is the total number of prey species. Trophic levels for species were calculated for each study, overall TROPHs were then calculated for a species using a weighted average that takes into account the sample size in each study.

The examination of trophic level variation within and between fish species was limited to those species for which there were at least three independent studies, each involving the examination of 10 or more fish. Trophic level of species was only assessed from studies where weight or volume of dietary items were expressed as a proportion of total gut contents, as these values were typically similar within species. We excluded from calculations those studies where only presence or percentage occurrence of dietary items was recorded. Studies involving the examination of juvenile diet only, or where juveniles may have contributed substantially to the sample, were also excluded. We also examined if the number of studies on a particular species diet influence trophic breadth by assessing the strength of the relationship between the number of independent studies and TROPH range. The number of studies was not

strongly correlated to diet breadth ($F_{1,14}=0.24$, $P=0.63$, $r^2=0.01$), suggesting sample size did not unduly influence our results. Similarly, diet breadth was not related to the mean distance between study locations from which dietary information was collated ($F_{1,14}=0.49$, $P=0.49$, $r^2<0.03$), suggesting geographic range of studies did not influence results.

To assess which fish are susceptible to predation by fisheries targeted species the teleost component of piscivorous fish diets was examined separately. Only those species for which teleosts accounted for >50% of the overall diet were considered piscivores, as prevalence of fish in guts suggests these are the predators most likely to remove fish prey and affect top down control of community structure. Prey information was collated at the family level and the frequency of occurrence of teleost families in the diets of major piscivores was determined. Pelagic and demersal predators and prey were identified to provide a rough gauge of how well these two systems are linked. The percentage of pelagic and benthic prey in the diet of target species was used as an indicator of where predators predominantly feed. Pelagic species were defined as those belonging to groups 35–38 of the FAO International Standard Statistical Classification of Aquatic Animals and Plants (ISSCAAP). This includes the sharks, scombrids, carangids and species which spend most of their time swimming in open water. Demersal species were defined as those belonging to ISSCAAP groups 31 and 34. These included serranids, lutjanids and lethrins and species that live in close association with the benthos, typically swimming near to, or sheltering within, benthic structures.

Results

Eighty-one dietary studies were included in the present study to calculate standardized dietary compositions of 76 fish species from the inshore waters of N W Australia (Table 1). Although most of these species are an important component of the recreational and commercial catch in Western Australia, there has been very little study into the diets of these species locally. Only five studies were found to have been conducted in Western Australian waters, with these containing quantitative (volumetric or weight) dietary data on 10 of the 76 species examined. A

Table 1 Standardised dietary composition and mean fractional trophic level (TROPH) of fish species targeted by recreational and commercial fishers in North West Australia

	Standardised dietary composition (%)	TROPH	Sources
Carcharhinidae			
<i>Carcharhinus amblyrhynchos</i>	tel (80), crus (11), ceph (5), oth (4)	4.25	1, 2, 3*, 4*, 5*, 6*
<i>Carcharhinus limbatus</i>	tel (98), chon (1), oth (1)	4.48	3, 7*, 8*, 9, 10*, 11, 12, 13*
<i>Carcharhinus melanopterus</i>	tel (89), ceph (10), oth (1)	4.47	1, 14*, 15*
<i>Carcharhinus sorrah</i>	tel (86), ceph (6), crus (7), oth (1)	4.38	1, 2, 3*, 16*, 17*#
<i>Carcharhinus tilstoni</i>	tel (98), crus (2), oth (<1)	4.47	1, 2, 16*, 17*#
<i>Galeocerdo cuvier</i>			4*, 5*, 7*, 8*, 18*, 19*, 20*, 21*, 22*, 23*, 24*, 25*#, 26*#
<i>Negaprion acutidens</i>	tel (100), oth (<1)	4.49	2, 3*, 14*
Sphymidae			
<i>Sphyrna lewini</i>	tel (65), ceph (30), crus (5)	4.45	5*, 7*, 16*, 27, 28*#
<i>Sphyrna mokarran</i>	chon (94), tel (6), oth (<0.1)	4.64	7*, 28*#, 29
Synodontidae			
<i>Saurida undosquamis</i>	tel (92), crus (7), ceph (2)	4.44	30, 31, 32, 33*#
<i>Synodus variegatus</i>	tel (100)	4.50	34*, 35
Ariidae			
<i>Neoarius graeffei</i>	tel (25), ann (23), crus (19), insect (6), oth (28)	2.95	36, 37, 38
<i>Netuma thalassina</i>	tel (54), crus (34), mol (3), echin (3), ceph (2), ann (1), oth (2)	4.01	36, 39
Platycephalidae			
<i>Platycephalus arenarius</i>	tel (89), crus (10), oth (1)	4.36	37
<i>Platycephalus indicus</i>	tel (95), crus (5), oth (<1)	4.44	3, 40, 41
Latidae			
<i>Lates calcarifer</i>	tel (82), crus (16), oth (2)	4.30	1, 3, 37, 42*
<i>Psammoderus waigiensis</i>	crus (87), tel (9), mol (2), oth (2)	3.55	1
Serranidae			
<i>Cephalopholis argus</i>	tel (96), crus (4)	4.46	34*, 43, 44*, 45*
<i>Cephalopholis boenak</i>	tel (70), crus (29), oth (1)	4.18	30, 46, 47
<i>Cephalopholis cyanostigma</i>	tel (97), crus (3), oth (<1)	4.47	46
<i>Cephalopholis miniata</i>	tel (92), crus (8)	4.42	30, 44*
<i>Epinephelus coioides</i>	crus (50), tel (37), mol (13)	3.91	30
<i>Epinephelus fasciatus</i>	crus (71), tel (23), echin (6), oth (<1)	3.72	30, 43
<i>Epinephelus malabaricus</i>	crus (70), tel (24), mol (6)	3.76	30
<i>Epinephelus polyphekadion</i>	crus (50), tel (43), mol (5), ann (2)	3.94	30
<i>Plectropomus leopardus</i>	tel (93), crus (5), ceph (2)	4.45	30, 47, 48*, 49*, 50*
<i>Variola louti</i>	tel (83), crus (17)	4.33	30
Terapontidae			
<i>Amniataba caudavittata</i>	crus (57), ann (24), oth (19)	2.92	3, 40
Apogonidae			
<i>Cheilodipterus artus</i>	tel (88), crus (12)	4.38	51
<i>Cheilodipterus macrodon</i>	tel (100)	4.50	34, 52, 53
<i>Cheilodipterus quiquelineatus</i>	crus (56), tel (34), mol (8), oth (2)	3.84	51, 53, 54
Sillaginidae			
<i>Sillago analis</i>	mol (61), crus (32), ann (8)	3.65	55, 56*
<i>Sillago sihama</i>	ann (57), crus (29), mol (4), tel (2), oth (9)	3.06	30, 55, 57

Table 1 (continued)

	Standardised dietary composition (%)	TROPH	Sources
Rachycentridae			
<i>Rachycentron canadum</i>	tel (55), crus (37), ceph (4), mol (3), chon (1)	4.10	39, 58, 59, 60, 61, 62*
Carangidae			
<i>Carangoides fulvoguttatus</i>	tel (91), crus (9), oth (<0.1)	4.41	30, 39
<i>Caranx ignobilis</i>	tel (73), crus (26), oth (1)	4.23	1, 30, 63, 64
<i>Caranx sexfasciatus</i>	tel (77), crus (22), oth (1)	4.27	3, 39, 58, 64
<i>Gnathanodon speciosus</i>	crus (58), tel (22), oth (20)	3.22	1, 3, 39, 40
<i>Scomberoides commersonnianus</i>	tel (88), crus (8), ceph (2), oth (2)	4.35	1, 3, 37, 40
<i>Trachinotus bailloni</i>	tel (100)	4.50	34
Lutjanidae			
<i>Lutjanus argentimaculatus</i>	crus (89), tel (11)	3.61	3, 30
<i>Lutjanus bohar</i>	tel (66), ceph (15), crus (12), ann (3), mol (2), oth (2)	4.24	30, 65
<i>Lutjanus carponotatus</i>	tel (58), crus (32), mol (10), oth (<1)	4.11	1, 66
<i>Lutjanus fulviflamma</i>	crus (78), tel (18), mol (1), oth (3)	3.61	30, 53, 66, 67
<i>Lutjanus malabaricus</i>	tel (95), mol (2), ann (1), crus (1), oth (.1)	4.46	39, 58, 68#
<i>Lutjanus quinquelineatus</i>	crus (49), tel (24), ann (11), ceph (9), mol (4), echin (3)	3.79	30, 69
<i>Lutjanus russellii</i>	tel (54), crus (45), oth (1)	4.01	3, 30, 39, 40, 53
<i>Lutjanus sebae</i>	tel (67), crus (23), ann (5), ceph (3), mol (1), oth (2)	4.13	30, 39
<i>Lutjanus vitta</i>	tel (53), crus (43), ann (2), mol (1)	4.03	30, 34, 39, 47
<i>Symphorus nematophorus</i>	tel (55), crus (35), echin (6), ceph (4)	4.08	30
Haemulidae			
<i>Diagramma labiosum</i>	crus (48), tel (15), ann (12), ceph (1), mol (1), oth (23)	3.04	39
Lethrinidae			
<i>Lethrinus atkinsoni</i>	crus (22), mol (22), echin (20), ceph (16), tel (13), ann (7), oth (1)	3.78	30, 54, 70#
<i>Lethrinus laticaudis</i>	tel (66), mol (15), crus (9), echin (5), ann (2), oth (4)	4.08	1, 39, 70#
<i>Lethrinus lentjan</i>	crus (40), tel (22), ceph (12), mol (11), echin (6), ann (4), oth (4)	3.75	1, 30, 39, 40, 70#
<i>Lethrinus miniatus</i>	tel (60), crus (20), echin (20)	4.08	30
<i>Lethrinus nebulosus</i>	mol (55), crus (17), ceph (9), echin (7), tel (7), ann (4), oth (<1)	3.79	30, 39, 70#, 71*
<i>Lethrinus rubrioperculatus</i>	crus (58), tel (23), ceph (11), ann (3), mol (2), echin (2), oth (1)	3.81	30
<i>Lethrinus variegatus</i>	crus (53), tel (43), ceph (3), mol (1)	3.96	30, 71*
Sparidae			
<i>Acanthopagrus latus</i>	mol (55), crus (23), tel (4), oth (18)	3.27	72#
Sciaenidae			
<i>Protonibea diacanthus</i>			73*
Mullidae			
<i>Parupeneus cyclostomus</i>	crus (58), tel (42)	3.92	34*, 53
Sphyraenidae			
<i>Sphyraena barracuda</i>	tel (97), ceph (1), crus (1), oth (1)	4.49	30, 47, 61, 74*
<i>Sphyraena jello</i>	tel (100)	4.50	58
Polynemidae			
<i>Eleutheronema tetradactylum</i>	tel (79), crus (18), ceph (2), oth (2)	4.26	3, 37, 40, 75
Labridae			
<i>Bodianus bilunulatus</i>	mol (100)	3.80	35
<i>Bodianus perditio</i>	mol (86), crus (7), echin (6), tel (1)	3.76	30

Table 1 (continued)

	Standardised dietary composition (%)	TROPH	Sources
<i>Cheilinus trilobatus</i>	mol (33), crus (33), echin (26), tel (8)	3.65	53
<i>Cheilinus undulatus</i>	mol (38), echin (25), tel (22), crus (14), oth (1)	3.78	76
<i>Choerodon rubescens</i>	mol (58), echin (22), crus (14), oth (7)	3.53	77#
<i>Choerodon schoenleinii</i>	mol (83), crus (9), oth (7)	3.58	77#
<i>Thalassoma lunare</i>	crus (48), mol (43), tel (5), ann (2), oth (2)	3.65	53, 66
<i>Thalassoma lutescens</i>	crus (65), echin (19), mol (14), tel (2)	3.54	34*, 53
Scombridae			
<i>Euthynnus affinis</i>	tel (98), crus (1), oth (1)	4.48	47, 58, 78, 79
<i>Scomberomorus commerson</i>	tel (97), crus (2), mol (1)	4.47	1, 47, 58, 80*
<i>Scomberomorus queenslandicus</i>	tel (98), ceph (2)	4.50	39, 80*, 81*

Asterisk denotes studies that presented dietary data as% occurrence only and which were not used to calculate overall dietary composition

indicates studies carried out in N W Australia

Diet abbreviations: ann *annelids*, bird *birds*, chon *chondrichthyans*, ceph *cephalopods*, crus *crustaceans*, echin *echinoderms*, insect *insects*, mam *mammals*, oth *other invertebrates*, rept *reptiles*, tel *teleosts*

Species from the families Synodontidae, Terapontidae and Apogonidae are presented as examples of meso-predators that are not targeted by fishers in N WAustralia

(1) Brewer et al. 1995, (2) Salini et al. 1992, (3) Salini et al. 1990, (4) DeCrosta et al. 1984, (5) Wass 1971, (6) Wetherbee et al. 1997, (7) Dodrill 1977, (8) Bass et al. 1973, (9) Tavares 1997, (10) Castro 1996, (11) Hueter 1994, (12) Dudley and Cliff 1993, (13) Castillo et al. 1992, (14) Stevens 1984b, (15) Lyle 1987, (16) Simpfendorfer and Milward 1993, (17) Stevens and Wiley 1986, (18) Lowe et al. 1996, (19) Randall 1992, (20) Simpfendorfer 1992, (21) Stevens and McLoughlin 1991, (22) Stevens 1984a, (23) Taylor and Naftel 1978, (24) Bell and Nichols 1921, (25) Heithaus 2001, (26) Simpfendorfer et al. 2001, (27) Galván-Magana et al. 1989, (28) Stevens and Lyle 1989, (29) Cliff 1995, (30) Kulbicki et al. 2005, (31) Yamashita et al. 1991, (32) Rao 1981, (33) Sainsbury unpubl., (34) Hiatt and Strasburg 1960, (35) Hobson 1974, (36) Blaber et al. 1994, (37) Salini et al. 1998, (38) Sumpton and Greenwood 1990, (39) Salini et al. 1994, (40) Haywood et al. 1998, (41) Marais 1984, (42) Davis 1985, (43) Harmelin-Vivien and Bouchon 1976, (44) Shpigel and Fishelson 1989, (45) Randall and Brock 1960, (46) Beukers-Stewart and Jones 2004, (47) Blaber et al. 1990, (48) St John et al. 2001, (49) Kingsford 1992, (50) Goeden 1978, (51) Marnane and Bellwood 2002, (52) Barnett et al. 2006, (53) Sano et al. 1984, (54) Nakamura et al. 2003, (55) Gunn and Milward 1985, (56) Brewer and Warburton 1992, (57) Hajisamae et al. 2006, (58) Bachok et al. 2004, (59) Arendt et al. 2001, (60) Franks et al. 1996, (61) Randall 1967, (62) Knapp 1951, (63) Meyer et al. 2001, (64) Blaber and Cyrus 1983, (65) Wright et al. 1986, (66) Connell 1998, (67) Kamukuru and Mgaya 2004, (68) Bulmer unpubl., (69) Sweatman 1993, (70) Westera 2003, (71) Walker 1978, (72) Platell et al. 2007, (73) Phelan et al. 2008, (74) Wilson unpubl., (75) Stanger 1974, (76) Randall et al. 1978, (77) Lek 2004, (78) Griffiths et al. 2009, (79) Al-Zibdah and Odat 2007, (80) Jenkins et al. 1984, (81) Begg and Hopper 1997

further 20 studies, conducted in Queensland and the Northern Territory, provided quantitative dietary data on 44 species. Globally, 28 studies were found to contribute quantitative dietary data on 51 of the 78 species included in the present study. While quantitative dietary data were available for the majority of fishery target species in N W Australia, there were some recreationally and/or commercially important species for which data was not found. These included: *Epinephelus areolatus*, *E. bilobatus*, *E. microdon*, *E. multinotatus*, *E. rivulatus*, *Carangoides gymnotethus*, *Lutjanus lemniscatus*, *L. lutjanus*, *Lethrinus hutchinsi*, *Grammatocynus bicarinatus* and *Acanthopagrus palmaris*. For some species dietary information is available, however these studies have only recorded prey presence in guts rather

than providing a quantitative assessment of how this prey contributes to total weight or volume of ingested matter. Most notable are the 13 studies on tiger shark (*Galeocerdo cuvier*) diet, none of which provide data on volume or weight of prey ingested (Table 1).

All of the 76 species for which dietary data was obtained were carnivorous, the mean fractional trophic levels ranging from 2.92 for *Amniataba caudavittata* up to 4.64 for *Sphyrna mokarran* (Table 1). The majority of these fishes, (44 species), could be classified as piscivores, as teleosts represented >50% of volume or weight ingested. Piscivorous families targeted by fishers included the Platycephalidae, Serranidae, Rachycentridae, Carangidae and Lutjanidae, whilst non-targeted piscivores

included Muraenidae and Synodontidae. The Sillaginidae, Haemulidae, Lethrinidae, Sparidae, Sciaenidae and Mullidae are also targeted by many fishers, but are mostly invertivores, predominantly feeding on crustaceans and molluscs.

Pelagic species had a slightly higher mean (\pm standard error) fractional trophic level (4.39 ± 0.05) than demersal species (3.91 ± 0.10). A *T*-test finding the difference to be significant ($t_{18} = 4.16$, $p < 0.01$). Pelagic fish such as barracuda, tuna and mackerel tended to feed on pelagic prey such as carangids, clupeids and mugilids. However, sharks that forage over both benthic and pelagic environments fed on both pelagic and benthic prey. Predators such as grouper, lethrinids and some of the lutjanids, that are typically associated with the benthos, fed mainly on fish that are associated with the benthos (Table 2). A notable exception to this pattern was a high percentage of clupeids in the diet of the *Lutjanus bohar*, which, although classified as a benthic predator is often observed swimming well above the substratum. The lizardfish, *Saurida undosquamis*, which is typically associated with the benthos, also had a high percentage of clupeids in the diet.

Members of the family Clupeidae were the most frequently consumed teleost prey, occurring in the diet of 21/27 species (Fig. 1). Clupeids were present in both demersal and pelagic piscivores, representing a prominent component of the diet of eight demersal and 13 pelagic species. Mugilidae, Carangidae, Monacanthidae and Antherinidae were all prey of five or more pelagic predators, whilst Nemipteridae, Mullidae and Pomacentridae were typically preyed upon by benthic predators. Fish from the families Leiognathidae, Engraulidae, Gerriidae, Labridae and Haemulidae were a prominent component of both pelagic and benthic piscivore diets (Fig. 1).

Dietary assessment of fish varied considerably between studies. The smallest range in mean fractional trophic levels were observed for the pelagic species *S. barracuda*, *Euthynnus affinis* and *Carcharhinus tilstoni*, while the largest amount of variation was observed for the demersal *Lethrinus nebulosus* and the reef-associated pelagic *Caranx ignobilis* (Fig. 2). Dietary assessment of both *L. nebulosus* and *C. ignobilis* spanned trophic levels 3 and 4, emphasizing the extent of plasticity in their diet. Moreover, comparison of species within genera revealed that trophic ranges do not always overlap (Fig. 2). For

example there was no overlap in the trophic values of *Lutjanus vitta* and *L. fulviflamma* inferring that these congeners are trophically distinct.

Discussion

Effective ecosystem based management is founded upon a sound knowledge of how ecosystem components interact (Levin and Lubchenco 2008). In particular, dietary information is critical for modeling trophic pathways and assessing the implications of predator removal from a system. For example, fishing typically removes large bodied predators from the system, but details of predator diet are required before cascading consequences can be investigated. We have demonstrated that prey consumed by fisheries targeted species varies between species and that not all fish targeted by N W Australian fishers are piscivores. For example, the blacktip shark, *Carcharhinus tilstoni*, an important component of the northern shark fishery (Fletcher and Santoro 2008), feed almost exclusively on other fish, yet the whiting *Silago sihama*, which is targeted by recreational fishers (Williamson et al. 2006), feeds exclusively on invertebrates. Moreover, there is considerable intra-specific variation in the diet of some species. This plasticity in diet is best demonstrated by the Spangled Emperor, *Lethrinus nebulosus*, which is targeted by both recreational and commercial fishers (Sumner et al. 2002; Fletcher and Santoro 2008). At two locations *L. nebulosus* fed predominantly on invertebrates and only small volumes (<5%) of fish (Westera 2003; Kulbicki et al. 2005). But at another location fish represented >80% of the weight of ingested matter (Salini et al. 1994). Differences in density of prey items may account for a small amount of variation when diet is assessed by weight and volume, however the large disparity in diet from these studies suggests *L. nebulosus* will feed on a variety of prey resources. Importantly, diet clearly varies among and within species targeted by N W Australian fishers and the impact of fishing on prey will depend on the species and quantity of fish removed.

Furthermore, for species whose diet is spatially variable it may be inappropriate to assess indirect effects of fishing based on dietary information collected from other locations. Diet for many fish may also change ontogenetically (e.g., Davis 1985;

Table 2 Prevalence of fish prey in the diet of fish species targeted by fishers in North West Australia. Only species where teleost account for >50% of the diet are included. Information compiled at the family level for prey

Taxa	% Tel	Teleost families consumed	%P	%B	Sources
Carcharhinidae					
<i>Carcharhinus amblyrhynchos</i>	80.2	Car 18, Mug 7, Leiog 7, Clup 4, Arii 3, Engr 2, Lab 2, Mull 2, Gerr 1	31	15	1, 2
<i>Carcharhinus melanopterus</i>	88.9	Car 13, Leiog 5, Mona 5, Lab 4, Clup 3, Leth 3, Sigan 3, Hemi 2, Mug 2, Gerr 1	20	21	1
<i>Carcharhinus sorrah</i>	85.6	Clup 13, Mug 12, Leiog 4, Sigan 3, Tera 3, Haem 1, Mona 1	26	11	1, 2
<i>Carcharhinus tilstoni</i>	97.7	Mug 8, Clup 8, Arii 5, Leiog 4, Cara 3, Lab 3, Gerr 2, Haem 2, Mona 2, Scian 2	26	13	1, 2
<i>Negaprion acutidens</i>	99.7	Mug 28, Clup 18, Car 3	49	0	2
Synodontidae					
<i>Saurida undosquamis</i>	88.8	Clup 25, Leiog 20, Engr 18, Syno 10, Scom 3, Caes 2, Nem 2	46	34	3, 4
Ariidae					
<i>Netuma thalassina</i>	54.2	Nem 15, Leiog 10, Syno 3	0	28	5
Platycephalidae					
<i>Platycephalus arenarius</i>	89.1	Gob 51, Scian 19	19	51	6
Latidae					
<i>Lates calcarifer</i>	82.0	Engr 17, Mug 13, Mull 10, Gerr 8, Clup 6, Arii 2, Ball 2, Leth 2	38	22	1, 6
Serranidae					
<i>Cephalopholis boenak</i>	70.1	Apog 42, Pom 32, Ser 7, Clup 6, Ather 3, Pseud 3, Gob 2,	9	87	7, 8
<i>Cephalopholis cyanostigma</i>	96.6	Caes 26, Apog 26, Pom 19, Lutj 13, Clup 5, Hol 3, Syno 3, Leth 2	5	92	7
Rachycentridae					
<i>Rachycentron canadum</i>	52.4	Engr 15, Syno 13, Pleur 10, Scian 9, Clup 9, Car 7, Ang 6, Bat 3, Gerr 3, Pomt 3, Spar 3	43	35	5, 9, 10
Carangidae					
<i>Carangoides fulvoguttatus</i>	91.3	Nem 25, Haem 9	0	34	5
<i>Caranx ignobilis</i>	73.1	Spar 43, Gob 27, Amb 9, Clup 8 Lab 5	8	84	1, 11
<i>Caranx sexfasciatus</i>	76.8	Amb 41, Mug 17, Gob 16, Spar 7, Engr 7, Ather 1, Clup 1,	26	64	11
<i>Scomberoides commersonianus</i>	87.5	Clup 10, Lab 10, Hemi 8, Leth 7, Car 5, Gerr 4, Sigan 3, Ather 2, Mona 2, Mug 2, Leigo 1	28	27	1, 6
Lutjanidae					
<i>Lutjanus bohar</i>	66.1	Clup 61, Opi 13, Engr 8, Caes 7, Pom 5, Acan 4, Ser 2	69	31	12
<i>Lutjanus carponotatus</i>	57.9	Blen 20, Leth 9, Clup 5, Lab 4, Pom 4, Apog 2, Gob 2, Mona 2	5	43	1, 13
<i>Lutjanus russellii</i>	53.5	Leiog 30, Nem 29	0	59	5
<i>Lutjanus sebae</i>	66.5	Leiog 35, Mull 28, Gerr 9, Haem 8, Ang 4, Nem 3, Syno 2	0	89	5
<i>Lutjanus vitta</i>	53.2	Clup 27, Leiog 25, Car 3, Nem 2, Haem 1, Pleur 1	30	29	5, 8
Lethrinidae					
<i>Lethrinus laticaudis</i>	65.6	Gerr 74, Haem 15, Ang 4,	0	93	5
Sphyraenidae					
<i>Sphyraena barracuda</i>	96.1	Car 17, Clup 14, Ather 9, Haem 7, Lutj 7, Sphy 7, Mona 5, Acan 3, Bel 3, Scar 3, Syno 3	50	28	8, 14
Polynemidae					
<i>Eleutheronema tetradactylum</i>	78.8	Leiog 25, Clup 11, Mull 6, Tera 5, Scat 5, Pol 4, Mug 3, Sil 3, Syno 3, Engr 2, Haem 2	16	53	6, 15

Table 2 (continued)

Taxa	% Tel	Teleost families consumed	%P	%B	Sources
Scombridae					
<i>Scomberomorus commerson</i>	97.3	Mug 13, Clup 12, Car 6, Hemi 4, Engr 2, Ather 1	38	0	1, 8
<i>Scomberomorus queenslandicus</i>	98.0	Clup 62	62	0	5
<i>Euthynnus affinis</i>	98.3	Engr 42, Car 25, Clup 22, Bel 4, Mona 2	93	2	8, 16

Values are the percentage prey families in the teleost component in predator’s diet. Where this does not sum to 100% the remaining teleosts are unidentified. Each prey family classified as either predominantly from pelagic (P) or benthic (B) environment and summed to provide an estimate of pelagic and benthic fish in predators diet

Family abbreviations: Acan *Acanthuridae*^B, Amb *Ambassidae*^B, Ang *Anguilliformes*^B, Apog *Apogonidae*^B, Arii *Ariidae*^B, Ather *Atherinidae*^P, Ball *Ballistidae*^B, Bat *Batrachoididae*^B, Bel *Belonidae*^P, Blen *Blenniidae*^B, Caes *Caesionidae*^B, Car *Carangidae*^P, Clup *Clupeidae*^P, Engr *Engraulidae*^P, Gerr *Gerreidae*^B, Gob *Gobiidae*^B, Haem *Haemulidae*^B, Hemi *Hemiramphidae*^P, Hol *Holocentridae*^B, Leiog *Leiognathidae*^B, Lab *Labridae*^B, Leth *Lethrinidae*^B, Lutj *Lutjanidae*^B, Mona *Monacanthidae*^B, Mug *Mugilidae*^P, Mull *Mullidae*^B, Mur *Muraenidae*^B, Nem *Nemipteridae*^B, Opi *Ophichthidae*^B, Pem *Pempheridae*^B, Pleur *Pleuronectiformes*^B, Pol *Polynemidae*^B, Pom *Pomacentridae*^B, Pomt *Pomatomidae*^B, Pseud *Pseudochromidae*^B, Scar *Scaridae*^B, Scat *Scatophagidae*^B, Scian *Sciaenidae*^P, Scom *Scombridae*^P, Sigan *Siganidae*^B, Ser *Serranidae*^B, Sil *Sillaginidae*^B, Spar *Sparidae*^B, Sphy *Sphyracidae*^P, Syno *Synodontidae*^B, Tera *Terapontidae*^B

(1) Brewer et al. 1995, (2) Salini et al. 1992, (3) Yamashita et al. 1991, (4) Rao 1981, (5) Salini et al. 1994, (6) Salini et al. 1998, (7) Beukers-Stewart and Jones 2004, (8) Blaber et al. 1990, (9) Arendt et al. 2001, (10) Franks et al. 1996, (11) Blaber and Cyrus 1983, (12) Wright et al. 1986, (13) Connell 1998, (14) Randall 1967, (15) Stanger 1974, (16) Griffiths et al. 2009

St John 1999; Cocheret de la Moriniere et al. 2003) or seasonally (e.g., Kingsford 1992; Arendt et al. 2001; Griffiths et al. 2009). These spatial, ontogenetic and temporal changes reflect shifts in fish feeding capacity, energetic demands and availability

of prey. As a consequence, any trophic cascade instigated by fishing will be dependant on the size, age and type of fish species removed. This is especially true in ecosystems as complex as coral reefs, where there is a greater diversity of species

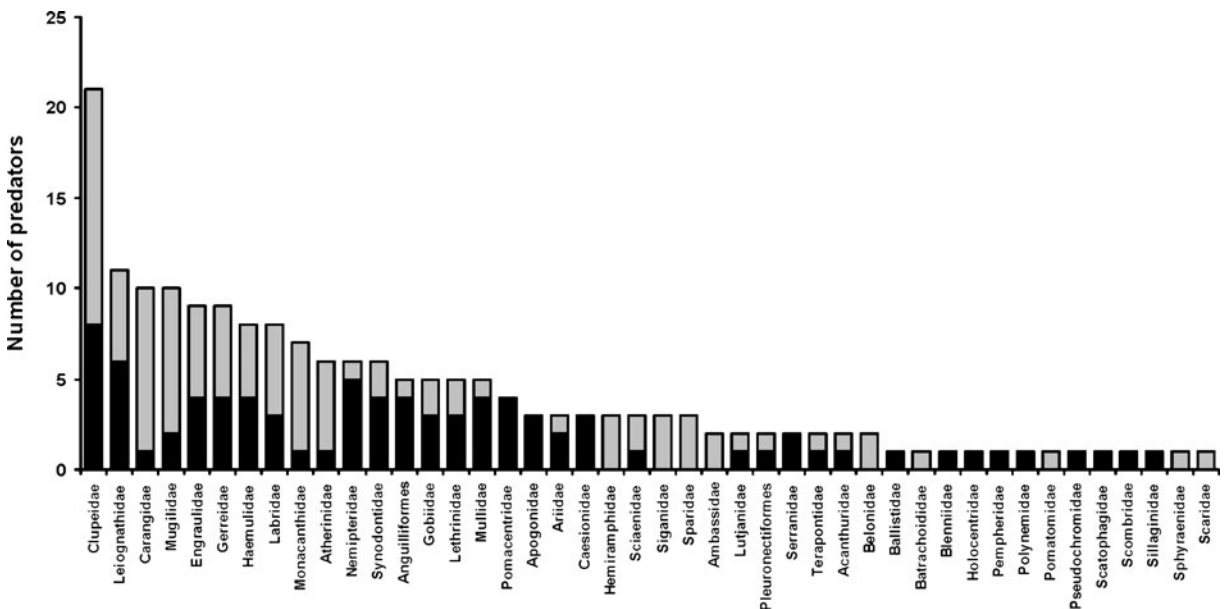


Fig. 1 Fish families most frequently preyed on by fisheries targeted species. Histogram shows the frequency at which teleost prey families contributed to the teleost component of the diets of species for which teleosts accounted for >50% of the

overall diet. Black and grey shading represents the proportion of demersal or pelagic piscivores, respectively, preying upon a particular teleost family

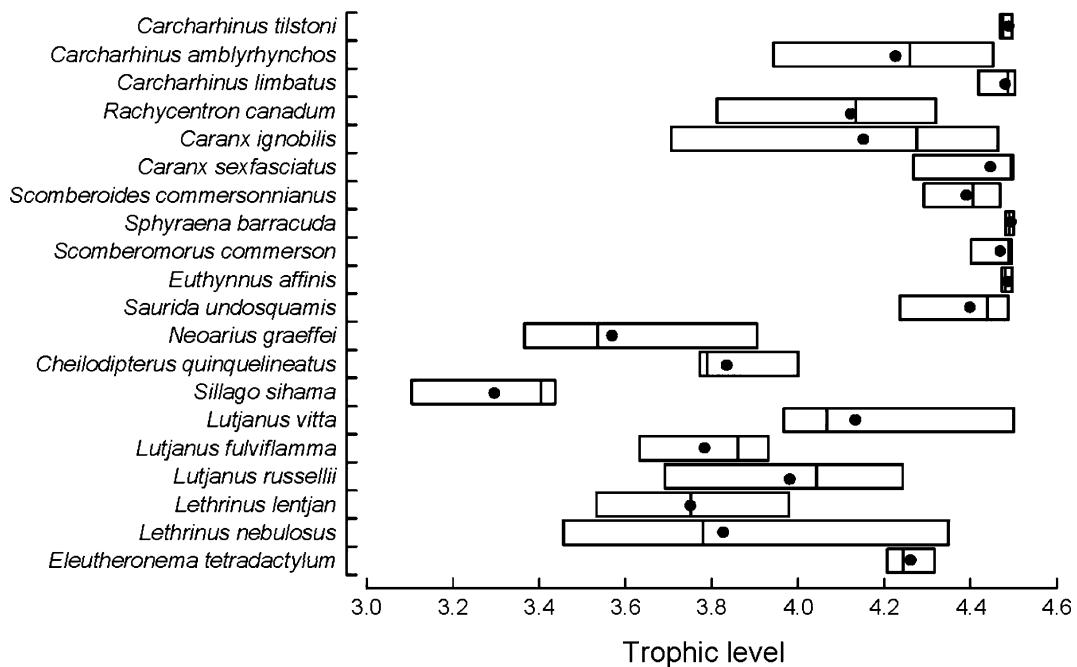


Fig. 2 Trophic range of species targeted by fishers in North West Australia. Box-whisker plots show fractional trophic levels (TROPHs) for those species for which there were between three and five independent studies, each involving the examination of 10 or more samples, and that expressed dietary composition

and carnivores are often opportunistic generalists (Russ 1991).

There is a general paucity of local dietary information for fisheries target species in N W Australia. There are only five studies containing quantitative dietary data on 10 recreationally and/or commercially important fish species in this region. Furthermore, while information on many other fisheries target species could be obtained from studies conducted outside of this region, there are a few species for which no quantitative dietary data is currently available. These include: *Lethrinus hutchinsi*, *Epinephelus multinotatus* and *Epinephelus rivulatus*, which form an important component of the recreational and commercial fisheries of Western Australia (Sumner et al. 2002; Williamson et al. 2006; Fletcher and Santoro 2008).

The dietary data obtained during the present study indicates that the majority of recreationally and commercially targeted fishes in N W Australia are carnivorous, with many of these being large predatory fishes with high mean fractional trophic levels. The vulnerability of top predatory fishes to fishing is well documented in the scientific literature, with many

volumetrically or by weight as a proportion of total gut contents. Boxes extend out to the minimum and maximum values of the data. The mean and median are displayed within the box as a black dot and a vertical line, respectively

Northern Hemisphere fisheries reporting significant declines in the mean trophic level of their catches (Pauly et al. 1998; Pinnegar et al. 2002; Essington et al. 2006). While there has been a much shorter history of fishing in Western Australian waters, the expansive nature of the coastline of this state means that any regional trophic declines may not be observed in the catch data while fishers are still finding and fishing new grounds. Monitoring and management of fish assemblages in N W Australia will therefore require comprehensive spatial and temporal assessments of both fish populations and fishing effort.

The removal of predators from marine ecosystems through exploitation is thought to result in elevated prey abundance, which, in turn, can influence primary productivity (Pace et al. 1999; Pinnegar et al. 2000; Shurin et al. 2002). While there is some evidence for trophic cascades on coral reefs due to the removal of predatory fish (McClanahan et al. 2002; Graham et al. 2003; Dulvy et al. 2004), such events have been more commonly reported for less diverse ecosystems, such as lakes and rocky reefs (Pace et al. 1999). High species diversity on coral reefs is thought to confer a

measure of redundancy that maintains ecological processes when individual species decline (McNaughton 1977; King and Pimm 1983; Tilman et al. 1996; Naeem 1998). Indeed variation in response to environmental drivers within trophic groups can often mask trophic cascades that occur at species level (Wilson et al. 2008). Consequently, even in complex systems flow on effects from exploitation of marine resources may exist and it is vital to examine trophic interactions at a species level.

Importantly there may be some key species that have relatively distinct ecological roles and may not always be fully redundant, interchangeable or replaceable (Rowan et al. 1997). For example, it is often hypothesized that removal of predators could increase the abundance of herbivores, reducing the standing biomass of algae. However on the reefs of N W Australia herbivorous fish such as kyphosids, scarids, siganids and acanthurids were not identified as an important component in the diet of predatory fish targeted by fishers. These large bodied herbivores were present in only five of the species targeted by fishers and represented <5% of ingested material in any one species. Consequently trophic cascades are unlikely to be instigated by increased predation of large bodied herbivores. Similarly, links between large herbivorous fish and predators in the Caribbean are weak (Mumby et al. 2006) and increased predation of herbivores in marine protected areas does not overwhelm the ecological benefits of increased herbivory afforded by these reserves (Mumby et al. 2007). Indirect effects of fishing on herbivory on N W Australian reefs may however be altered by removal of fish that feed on juveniles, increasing the survival rates of post-settlement fish. Mortality during this early life history stage is high and changes in predation may have a profound impact on adult populations and associated processes (Webster 2003). Moreover, some predators target invertebrates, including herbivorous echinoderms, which may alter rates of herbivory and erosion. On the reefs of N W Australia, fishers target lethrinids and labrids, which often feed on echinoderms. If predation by these fish controls echinoderm populations fishing may reduce predator abundance resulting in the release of echinoderm prey. For example, where fishing is prohibited at Ningaloo reef, abundance of some lethrinids is greater, density of herbivorous urchins is lower and algal communities differ,

implying fishing has instigated trophic cascades (Westera 2003). Similarly, reduced fishing pressure in Kenyan reserves increased the abundance of the triggerfish, *Balistapus undulatus*, and reduced the abundance of urchins (McClanahan 2000). Removal of invertebrate feeding species by fishers has also been associated with increased abundance and outbreaks of the coral feeding starfish, *Acanthaster planci* (Dulvy et al. 2004; Sweatman 2008), which have caused extensive declines in coral cover (Chesher 1969; Randall 1973; Pratchett et al. 2009). These studies indicate trophic cascades can occur on tropical reefs and further investigations are required to validate and understand trophic links and possible cascades caused by fishing in NW Australia.

Clupeids, carangids, engraulids and mugilids, which spend most of their time in the water column, were the primary prey of scombrids which were classified as pelagic predators. This suggests pelagic fisheries are not tightly linked to benthic prey. Several benthic predators (e.g., *Saurida undosquamis*, *Lutjanus bohar*) did however ingest pelagic prey, and carchar-inids that move between these two habitats fed on both pelagic and benthic prey, suggesting there are some links between these environments. Moreover, most benthic marine animals have a bipartite life history, where early life stages are spent in the pelagic environment. Predation of “larval” fish by pelagic species may be an important determinant of juvenile abundance, as large pelagic species, such as greater barracuda, (*Sphyraena barracuda*), can have large numbers of larval fish in their stomachs (Wilson *pers. obs.*). Careful assessment of predation on pre-settlement stages of demersal species by pelagic fish is therefore required.

Overall N W Australian fishers currently target carnivores which feed on both fish and invertebrates. Large herbivorous fish are not a major component of these fishes diet and there is little evidence to suggest fishing will alter herbivory by fish. Fishing may however result in the release of invertebrate prey, which could alter herbivory and other processes attributable to invertebrates. To fully appreciate the indirect effects of fishing further work is required to elucidate functional roles and trophic links of fish and invertebrates on N W Australian reefs. Clearly assessing and predicting the cascading effects of fishing on coral reefs requires collection of dietary information from relevant species at appropriate

temporal and spatial scales. This information must then be combined with information on predator/prey abundance and consumption rates of predators to assess the magnitude of trophic fluxes. Current trophic knowledge of reef fish diet is largely based on a handful of studies that collected large numbers of species at specific locations and carried out gut content analyses on these fish (e.g., Hiatt and Strasburg 1960; Randall 1967; Kulbicki et al. 2005). A concerted effort to build on the information collected in these studies is required if we are to understand trophic interactions on reefs. This should involve a combination of stable isotope, fatty acid and gut content analyses to provide diet assessments of high taxonomic resolution over extended time frames.

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