

SYNTHESIS OF 3-METHYL DERIVATIVES FROM DIHYDROBETULONIC ACID METHYL ESTER

G. F. Krainova, Yu. A. Beloglazova, and V. V. Grishko*

A 2,3-secotriterpenoid with a methylketone was prepared from 1-hydroxyiminodihydrobetulonic acid methyl ester via a Grignard reaction followed by a Beckmann rearrangement. Functionalization of it led to the formation of 3-hydroxy- and 31-bromo-substituted derivatives; intramolecular cyclization, to the formation of a five-membered ring A with an alkene-nitrile. The synthetic products included compounds (5a and 9) with moderate cytotoxicity (IC₅₀ 25.22–46.66 μM) against MCF-7, HCT116, A549, and PC-3 cancer cells.

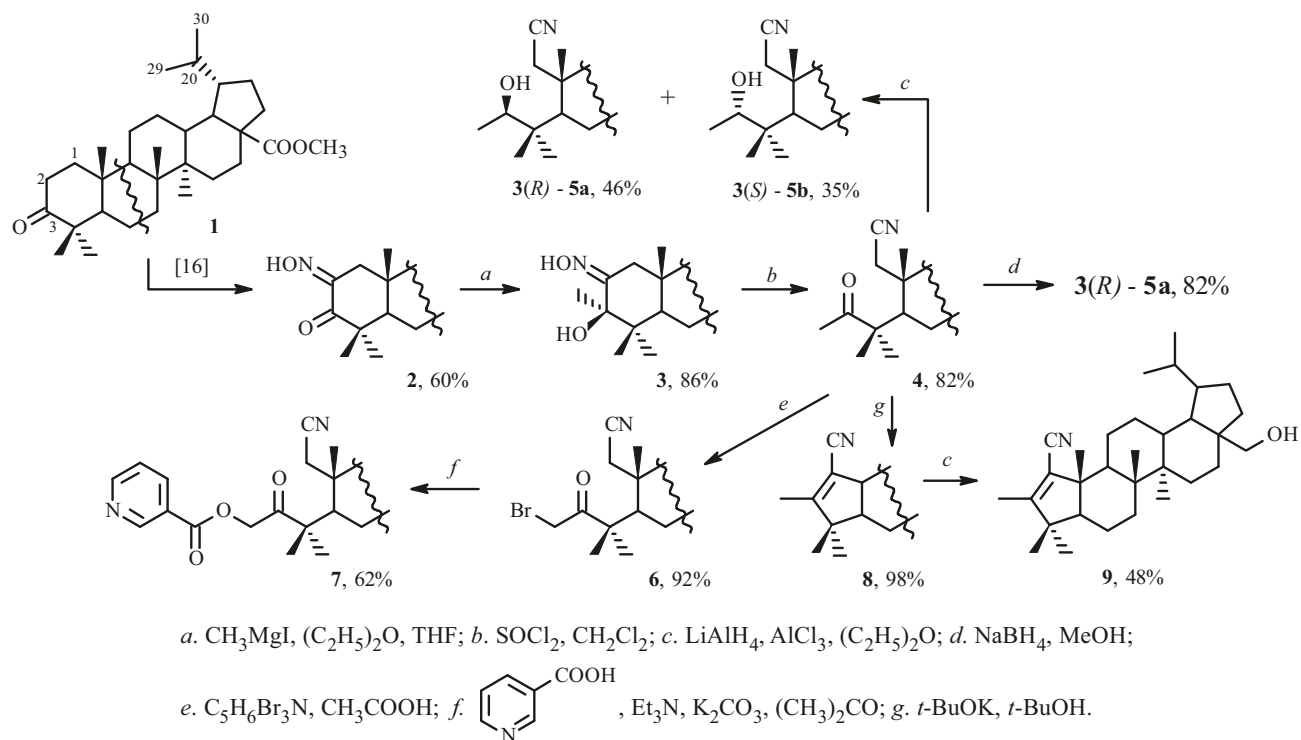
Keywords: A-secotriterpenoids, dihydrobetulonic acid, Beckmann rearrangement, nitrile-anionic cyclization, cytotoxic activity.

The significance of betulinic acid (3β-hydroxylup-20(29)-en-28-oic acid, BA) and semi-synthetic derivatives of BA for applications in medicinal chemistry derives mainly from their highly specific action against viruses and cancer cells with minimal harmful effects on healthy human cells [1–4]. The BA content in the extract of birch bark (*Betula pendula*) is usually less than 3–4% as compared to the elevated content (up to 80%) in the same extract of betulin, the biosynthetic precursor of BA [5]. BA can be produced in 86–92% yield by effective methods for chemical oxidation of betulin [6, 7]. Nevertheless, betulonic acid is most often used as starting material in laboratory syntheses. An effective method for preparative synthesis of it from betulin-containing birch-bark extract was recently published [8].

Previously, the ability to increase the antiviral and antitumor activity of pentacyclic triterpenoids by skeletal transformation of their C-3 alkylated derivatives was demonstrated by us [9–13]. In particular, a 2,3-secolupane α-bromo-substituted methylketone with pronounced cytotoxic activity (IC₅₀ 0.8–25.4 μM) against 11 tumor cell lines, including cancer cells with multi-drug resistance, was synthesized from betulonic acid methyl ester [14]. Also, introduction of a C-30 aldehyde into the isopropylidene fragment of lupane derivatives [10, 12, 14] most often increased the nonspecific cytotoxicity [15]. The present article involved the synthesis and screening of the cytotoxic properties of novel 3-methyl-substituted derivatives of dihydrobetulonic acid methyl ester (**1**) to assess the contribution of an oxidized isopropylidene moiety to the manifestation of cytotoxicity by the previously reported compounds [10, 12, 14].

The starting material for the work was α-hydroxyiminoketone **2**, the synthesis of which from **1** was recently reported by us [16]. A Grignard reaction [10–13, 17] of **2** with the alkylating agent CH₃MgI produced 3β-hydroxy-3α-methyl derivative **3** (Scheme 1). The IR spectrum of **3** exhibited bands for C=N–OH (1666 cm⁻¹) and OH stretching vibrations (3326, 3505 cm⁻¹). The ¹H NMR spectrum of **3** showed doublets for the enantiomeric 2H-1 methylene protons with centers at 2.22 and 3.37 ppm (AB-system, SSCC 12.0 Hz); a broad singlet at 5.18 ppm for the hydroxyl; and a characteristic singlet at 1.37 ppm for the methyl protons. The ¹³C NMR spectrum of **3** exhibited resonances for C atoms bonded to hydroxyl (76.84 ppm) and hydroxyimine (163.98 ppm). In the next step, alkylated derivative **3** underwent a Beckmann rearrangement using SOCl₂–CH₂Cl₂, which formed the corresponding methylketone **4**. The IR spectrum of **4** contained characteristic absorption bands for C=O (1703 cm⁻¹) and CN (2238 cm⁻¹). The PMR spectrum of **4** showed two doublets for the methylene 2H-1 protons with centers at 2.43 and 2.55 ppm (AB-system, SSCC 16.0 Hz) and a singlet for the C-3 methyl at 2.27 ppm. The ¹³C NMR spectrum of **4** included characteristic resonances for the CN and C=O C atoms at 118.65 and 214.28 ppm, respectively.

Institute of Technical Chemistry, Ural Branch, Russian Academy of Sciences, Perm, 614013, e-mail: grishko.v@itcras.ru. Translated from *Khimiya Prirodnykh Soedinenii*, No. 1, January–February, 2023, pp. 71–76. Original article submitted May 27, 2022.



Scheme 1

The nitrile of methylketone **4** remained inert upon reduction by LiAlH_4 in the presence of AlCl_3 . The reaction products were a mixture of diastereomeric alcohols **3(R)-5a** and **3(S)-5b** (Scheme 1), which were isolated pure in a 1.3:1.0 ratio by column chromatography over SiO_2 . ^{13}C NMR spectra of **5a** and **5b** had characteristic resonances for C-3 bonded to a hydroxyl at 73.11 or 72.92. Their IR spectra exhibited absorption bands for the OH at 3523 and 3521 cm^{-1} , respectively. The relative (*S*)-configuration of C-3 bonded to the hydroxyl in **5b** was confirmed by an X-ray crystal structure analysis (XSA) (Fig. 1). The resonances of the H-3 protons had practically the same shifts in ^1H NMR spectra of **3(R)-5a** and **3(S)-5b**. The quartets corresponding to them appeared at 3.825 and 3.83 ppm, respectively. The weak-field shift of the (*S*)-isomer protons was more noticeable for the 2H-1 protons, the doublets of which (AB-system, SSCC 18.0 Hz) were observed at 2.60 and 2.75 ppm for alcohol **3(R)-5a** or at 2.61 and 2.77 ppm for **3(S)-5b**, and the doublets for the 3H-32 protons (AB-system, SSCC 8.0 Hz) with centers at 1.14 or 1.24 ppm, respectively. Milder reduction of methylketone **4** by NaBH_4 occurred stereoselectively to form the **3(R)**-hydroxy derivative in 82% yield. This was consistent with data for the selective reduction by NaBH_4 of oleanane- and lupane-type methylketones [10].

The reaction of methylketone **4** with pyridinium tribromide in AcOH produced α -bromo-substituted derivative **6**. Its IR spectrum showed characteristic bands for stretching vibrations of carbonyl at 1722 cm^{-1} and cyano at 2238 cm^{-1} . The ^1H NMR spectrum of **6** contained doublets for the methylene 2H-1 protons with centers at 2.37 and 2.60 ppm (AB-system, SSCC 18.0 Hz) and resonances for the methylene protons of the bromomethyl moiety as doublets of an AB-system with centers at 4.29 and 4.35 ppm (SSCC 12.0 Hz). The ^{13}C NMR spectrum of **6** had characteristic resonances for the CN and C=O C atoms at 118.49 and 206.64 ppm, respectively.

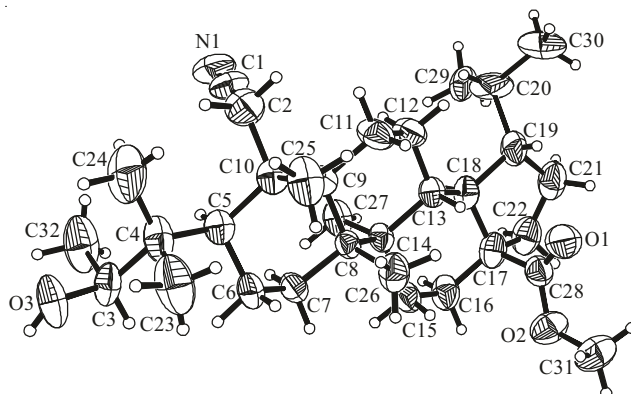


Fig. 1. Structure of **3(S)-5b** from XSA (N and O atoms refined as disordered).

TABLE 1. Cytotoxic Activity of **2**, **3**, **5a**, and **9** (IC₅₀, μM*)

Cell lines	2 [16]	3	5a	9	Doxorubicin
MCF-7	93.73 ± 8.69	62.40 ± 9.61	29.31 ± 1.11	25.22 ± 1.28	0.13 ± 0.02
HCT116	60.55 ± 5.42	> 100	42.69 ± 1.94	49.00 ± 1.77	1.88 ± 0.09
RD TE32	183.00 ± 49.43	> 100	> 100	63.80 ± 3.94	0.97 ± 0.06
MS	64.93 ± 4.58	66.67 ± 3.53	> 100	> 100	1.29 ± 0.16
A549	82.45 ± 0.51	64.9 ± 0.28	52.22 ± 1.10	46.66 ± 1.20	2.04 ± 0.22
PC-3	> 100	> 100	80.42 ± 3.23	37.75 ± 1.34	0.56 ± 0.08
HEpG2	> 100	97.15 ± 3.74	> 100	61.88 ± 3.15	1.78 ± 0.30
HEK293	109.10 ± 34.19	> 100	> 100	83.55 ± 3.74	0.44 ± 0.04

*IC₅₀, concentration of compound causing 50% death of cells.

Bromomethylketone **6** was *O*-alkylated by nicotinic acid with heating in an aprotic solvent in the presence of an excess of Et₃N and K₂CO₃ to afford the corresponding β-ketoester **7** in 62% yield [12, 18]. The ¹³C NMR spectrum of **7** exhibited typical resonances for the CN (118.53 ppm) and C=O C atoms (206.76 ppm), for two esters (163.51 and 176.74 ppm), and for the heteroaromatic ring (124.42, 126.86, 139.92, 148.47, and 150.60 ppm). The ¹H NMR spectrum included characteristic doublets of AB-systems for the two pairs of methylene protons 2H-1 and 2H-32 with centers at 2.49 and 2.66/5.27 and 5.35 ppm (SSCC 20.0/18.0 Hz) and resonances of the nicotinic acid aromatic protons in the range 7.57–9.30 ppm.

As expected [9–11, 13, 19], methylketone **4** underwent an intramolecular oxo-nitrile cyclization under basic conditions of *t*-BuOK–*t*-BuOH to form A-pentacyclic α,β-alkenenitrile **8** (Scheme 1). The IR spectrum of **8** exhibited characteristic bands for stretching vibrations at 1728 (COOCH₃) and 2207 cm⁻¹ (CN). The ¹H NMR spectrum of **8** showed the resonance for the C-3 methyl at 1.85 ppm. Its ¹³C NMR spectrum gave characteristic resonances of the alkenenitrile C atoms C-1 (120.45), C-3 (118.36), and C-3 (167.10 ppm). Compound **9**, the product of reduction of the C-28 carbomethoxyl group to hydroxyl, was obtained under reducing conditions (LiAlH₄, AlCl₃) from A-pentacyclic α,β-alkenenitrile **8**. The ¹H NMR spectrum of **9** exhibited a singlet at 1.86 ppm for the C-3 methyl protons and two doublets of an AB-system of the 2H-28 methylene protons at 3.32 and 3.75 ppm (SSCC 12.0 Hz). The ¹³C NMR spectrum of **9** gave characteristic resonances for the alkenenitrile moiety at 120.36 (C-1), 118.37 (C-2), and 167.19 ppm (C-3). The carbomethoxyl group of **4** remained inert under analogous conditions.

Compounds **2**, **4**, and **6–8** were nontoxic (IC₅₀ > 100 μM) according to a study of the cytotoxic activity of synthetic products **2–9** against seven tumor cell lines. The lack of cytotoxic properties for bromo-substituted methylketone **6** indicated that the specific activity of the previously reported lupane methylketone [10] was due largely to the presence in the triterpenoid of the C-30 aldehyde of the isopropylidene moiety or a combination in the triterpene structure of the aldehyde and a modified ring A. Compounds **5a** and **9** in the series of synthesized **2–9** were moderately active (IC₅₀ 25.22–46.66 μM) against cancer cell lines MCF-7, HCT116, A549, and PC-3. Alcohol **5a** appeared more selective, considering its low toxicity against normal HEK293 cells (Table 1).

EXPERIMENTAL

¹H NMR, ¹³C NMR, and DEPT spectra of the synthesized compounds in CDCl₃ with TMS internal standard were taken on a Bruker Avance II NMR spectrometer (400 and 100 MHz, respectively). IR spectra (ν, cm⁻¹) were recorded from thin films obtained by evaporation of CHCl₃ solutions of the compounds on a Bruker IFS 66/S FT-IR spectrometer (Germany). Optical rotation in CHCl₃ solution was measured at 589 nm on a PerkinElmer 341 polarimeter (USA). The threshold melting point at heating rate 1°C/min was determined on an OptiMelt MPA100 apparatus (USA). GC-MS spectra were analyzed by an Agilent Technologies 6890N instrument with a DB-35ms capillary column (30 m × 0.25 mm) at vaporizer temperature 240°C with programmed temperature of 20–40°C/min and He carrier gas. Elemental analyses (C, H, N) used a Vario EL cube analyzer (Germany). The course of reactions was monitored by TLC on Sorbfil plates (Russia) after treatment with H₂SO₄ (5%) followed by heating at 95–100°C for 2–3 min. Column chromatography used Macherey-Nagel silica gel (60–200 μm) and petroleum ether (40–70°)–EtOAc or petroleum ether (40–70°)–EtOAc–CHCl₃ eluent that was selected individually for each reaction product.

An X-ray crystal structure analysis (XSA) of **5b** used an Xcalibur Ruby single-crystal diffractometer (Agilent Technologies) and the standard method [Mo K α -radiation, 295(2) K, ω -scanning in 1° steps]. Absorption corrections were applied empirically using the SCALE3 ABSPACK algorithm [CrysAlisPro, Agilent Technologies, version 1.171.37.33 (release 27-03-2014 CrysAlis171.NET)]. The crystal (C₃₂H₅₃NO₃, MM 499.75) was monoclinic, space group *P*2₁, *a* = 7.0686(16), *b* = 17.595(4), *c* = 11.924(3) Å, β = 92.48(2)°, *V* = 1481.6(6) Å³, *Z* = 2, *d*_{calcd} = 1.12 g/cm³, μ = 0.070 mm⁻¹. The final refinement parameters were *R*₁ = 0.0598 [for 1935 reflections with *I* > 2 σ (*I*)], *wR*₂ = 0.1837 (for all 5470 independent reflections), *S* = 0.781, ratio of twinning components 0.7473(15):0.2527 (15). The structure was solved using the SHELXS program [20] and refined by anisotropic full-matrix least squares methods over *F*² for all nonhydrogen atoms using the SHELXL program [21] with an OLEX2 graphics interface [22]. H atoms were refined using a rider model. The crystal was refined using a dataset with reflection intensities in HKLF 5 format as a twin with two components. The XSA results for **5b** were deposited in the Cambridge Crystallographic Data Centre under No. CCDC 2174771 and can be requested at www.ccdc.cam.ac.uk/data_request/cif.

Anhydrous solvents were prepared by standard methods [23]. Dihydrobetulin was obtained in 70% yield by catalytic (Pd–C) hydrogenation of betulin [24]. Oxidation of dihydrobetulin by Jones reagent gave dihydrobetulonic acid, which was methylated by CH₃I in Me₂CO to the corresponding methyl ester [25].

Preparation of 3 β -Hydroxy-2-hydroxyimino-3 α -methylup-28-oic Acid (3). A freshly prepared solution of CH₃MgI (5.0 mmol) in Et₂O (5 mL) was treated dropwise with a solution (15 mL) of **2** (4.0 mmol) in a mixture (2:1) of Et₂O and THF. The reaction mixture was stirred for 2 h, chilled, treated dropwise with ice water (25 mL) and HCl solution (17–20%, 20 mL), and stirred until the precipitate dissolved completely. The reaction products were extracted with EtOAc (2 \times 50 mL). The organic layer was separated; washed with saturated Na₂S₂O₃ solution, NaHCO₃ solution (5%), and a small amount of H₂O; and dried over anhydrous MgSO₄. The solvent was distilled off. The solid was purified by column chromatography (CC) with elution by petroleum ether–EtOAc (15:1). Colorless crystals, yield 86%, *R*_f 0.46 (hexane–EtOAc, 7:3), mp 230.8°C (hexane–EtOAc), [α]_D²⁰ –32.6° (*c* 0.5, CHCl₃). IR (ν , cm⁻¹): 3505 (OH), 3326 (OH), 1726 (COOCH₃), 1666 (C=N–OH). ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 0.75, 0.86 (3H each, d, J = 8.0, CH₃), 0.76 (6H, s, CH₃), 0.91, 0.97, 0.98 (3H each, s, CH₃), 1.37 (3H, s, H-32), 2.22, 3.37 (1H each, d, J = 12.0, H-1, AB system), 2.21–2.28 (2H, m, H-19, 20), 3.64 (3H, s, H-31), 5.18 (1H, br.s, OH). ¹³C NMR (100 MHz, CDCl₃, δ , ppm): 14.67, 14.76, 15.79, 16.82, 18.61, 18.93, 21.36, 22.83, 22.89, 23.46, 24.01, 26.80, 29.77, 29.79, 32.16, 34.43, 36.38, 37.33, 38.23, 41.28, 41.55, 42.75, 44.33, 44.54, 49.10, 50.39, 51.05, 53.45, 57.09, 76.84 (C-3), 163.98 (C-2), 176.79 (C-28). Found, %: C, 74.52; H, 10.36; N, 2.72. C₃₂H₅₃NO₄. Calcd, %: C, 74.81; H, 10.39; N, 2.62. Mass spectrum *m/z* 515.4 [M]⁺. MM 515 g/mol.

Preparation of 3-Methyl-3-oxo-1-cyano-2,3-seco-2-norlup-28-oic Acid Methyl Ester (4). A solution of **3** (3.8 mmol) in CH₂Cl₂ (40 mL) was treated with SOCl₂ (7.1 mmol). The reaction mixture was stirred at room temperature for 30 min. Formation of products was monitored by TLC. The solvent was evaporated. The solid was rinsed with CH₂Cl₂ (2 \times 20 mL) and purified by CC with elution by petroleum ether–EtOAc (15:1). Colorless crystals, 82%, *R*_f 0.55 (hexane–EtOAc, 7:3), mp 211.7°C (hexane–EtOAc), [α]_D²⁰ +1.2° (*c* 0.5, CHCl₃). IR (ν , cm⁻¹): 2238 (C \equiv N), 1725 (COOCH₃), 1703 (C=O). ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 0.75, 0.85 (3H each, d, J = 8.0, CH₃), 0.91, 0.95, 1.01, 1.14, 1.21 (3H each, s, CH₃), 1.95–2.02 (2H, m, H-19, 20), 2.27 (3H, s, H-32), 2.43, 2.55 (1H each, d, J = 16.0, H-1, AB system), 3.63 (3H, s, H-31). ¹³C NMR (100 MHz, CDCl₃, δ , ppm): 14.62, 14.67, 15.74, 18.83, 21.38, 21.82, 22.75, 22.92, 23.70, 24.04, 25.74, 26.89, 29.46, 29.70, 29.71, 31.83, 33.45, 37.19, 38.14, 40.59, 42.49, 43.01, 44.12, 45.06, 48.76, 48.78, 51.14, 53.07, 57.01, 118.65 (C-2), 176.76 (C-28), 214.28 (C-3). Found, %: C, 77.22; H, 10.33; N, 2.81. C₃₂H₅₁NO₃. Calcd, %: C, 77.92; H, 10.23; N, 2.76. Mass spectrum *m/z* 497.4 [M]⁺. MM 497 g/mol.

Preparation of 3(R)-Hydroxy-3-methyl-1-cyano-2,3-seco-2-norlup-28-oic Acid Methyl Ester (5a). A solution of **4** (4.0 mmol) in MeOH (40 mL) was stirred and treated in portions with NaBH₄ (42 mmol). The reaction mixture was stirred at room temperature for 40 min and refluxed for 5 min. The formation of products was monitored by TLC. The solvent was evaporated. The resulting solid was dissolved in HCl solution (10%, 100 mL). The products were extracted with EtOAc (2 \times 50 mL). The organic layer was separated, washed with H₂O, and dried over anhydrous MgSO₄. The solvent was evaporated. The solid was purified by CC with elution by petroleum ether–EtOAc–CHCl₃ (20:1:1). Yield, 82%.

General Method for Preparing 5a,b and 9. A mixture of LiAlH₄ (12.5 eq) and AlCl₃ (4 eq) in anhydrous Et₂O was chilled in an ice bath, stirred for 15–20 min, treated with **4** (4.0 mmol, for **5a,b**) or **8** (4.2 mmol, for **9**), and stirred and heated for 2 h. The formation of products was monitored by TLC. The reaction mixture was diluted with aqueous NaOH solution (1%, 20 mL). The products were extracted by EtOAc (2 \times 50 mL). The organic layer was separated and dried over anhydrous MgSO₄. The solvent was evaporated. The solid was purified by CC with elution by petroleum ether–EtOAc–CHCl₃ (20:1:1).

3(R)-Hydroxy-3-methyl-1-cyano-2,3-seco-2-norlup-28-oic acid methyl ester (5a), colorless crystals, yield 46%, R_f 0.41 (hexane–EtOAc, 7:3), mp 176.1°C (hexane–EtOAc), $[\alpha]_D^{20}$ –3.0° (*c* 0.5, CHCl₃). IR (ν, cm⁻¹): 3523 (OH), 2239 (C≡N), 1726 (COOCH₃). ¹H NMR (400 MHz, CDCl₃, δ, ppm, J/Hz): 0.74, 0.84 (3H each, d, J = 8.0, CH₃), 0.83, 0.93, 0.98, 1.00, 1.08 (3H each, s, CH₃), 1.14 (3H, d, J = 8.0, H-32), 2.19–2.30 (2H, m, H-19, 20), 2.60, 2.75 (1H each, d, J = 18.0, H-1, AB system), 3.62 (3H, s, H-31), 3.825 (1H, q, J = 8.0, H-3). ¹³C NMR (100 MHz, CDCl₃, δ, ppm): 14.54, 14.66, 16.08, 17.63, 17.84, 21.18, 21.29, 22.19, 22.75, 22.92, 23.01, 27.17, 29.52, 29.68 (2C), 31.87, 33.61, 37.21, 38.31, 40.83, 42.39, 42.66, 43.05, 44.11, 45.50, 47.32, 48.74, 51.12, 57.06, 73.11 (C-3), 119.16 (C-2), 176.84 (C-28). Found, %: C, 76.90; H, 10.69; N, 2.80. C₃₂H₅₃NO₃. Calcd, %: C, 76.58; H, 10.89; N, 2.73. Mass spectrum *m/z* 499.4 [M]⁺. MM 499 g/mol.

3(S)-Hydroxy-3-methyl-1-cyano-2,3-seco-2-norlup-28-oic acid methyl ester (5b), colorless crystals, yield 35%, R_f 0.33 (hexane–EtOAc, 7:3), mp 180.7°C (hexane–EtOAc), $[\alpha]_D^{20}$ +3.0° (*c* 0.5, CHCl₃). IR (ν, cm⁻¹): 3521 (OH), 2239 (C≡N), 1726 (COOCH₃). ¹H NMR (400 MHz, CDCl₃, δ, ppm, J/Hz): 0.74, 0.85 (3H each, d, J = 8.0, CH₃), 0.93, 0.98 (6H, s, CH₃), 0.99, 1.05 (3H, s, CH₃), 1.24 (3H, d, J = 8.0, H-32), 2.19–2.30 (2H, m, H-19, 20), 2.61, 2.77 (1H each, d, J = 18.0, H-1, AB system), 3.63 (3H, s, H-31), 3.83 (1H, q, J = 8.0, H-3). ¹³C NMR (100 MHz, CDCl₃, δ, ppm): 14.56, 14.67, 16.06, 18.36, 18.79, 20.99, 21.23, 22.03, 22.17, 22.76, 22.93, 27.16, 29.70 (3C), 31.87, 33.58, 37.22, 38.31, 40.74, 42.71, 42.76, 43.04, 44.12, 45.34, 47.49, 48.75, 51.14, 57.06, 72.92 (C-3), 119.20 (C-2), 176.84 (C-28). Found, %: C, 76.90; H, 10.69; N, 2.80. C₃₂H₅₃NO₃. Calcd, %: C, 76.62; H, 10.81; N, 2.63. Mass spectrum *m/z* 499.5 [M]⁺. MM 499 g/mol.

3-Methyl-1-cyano-2-norlup-1,3-en-28-ol (9), colorless crystals, yield 48%, R_f 0.41 (hexane–EtOAc, 7:3), mp 123.8°C (hexane–EtOAc), $[\alpha]_D^{20}$ –4.8° (*c* 0.5, CHCl₃). IR (ν, cm⁻¹): 3319 (OH), 2208 (C≡N). ¹H NMR (400 MHz, CDCl₃, δ, ppm, J/Hz): 0.76, 0.84 (3H each, d, J = 4.0, CH₃), 0.92, 0.97, 1.02, 1.06, 1.10 (3H each, s, CH₃), 1.86 (3H, s, H-31), 3.32, 3.75 (1H each, d, J = 12.0, H-28, AB system). ¹³C NMR (100 MHz, CDCl₃, δ, ppm): 12.72, 14.80, 14.84, 17.41, 18.68, 19.60, 21.71, 22.24, 22.86, 26.42, 27.05, 27.09, 27.12, 29.38, 29.46, 34.07, 35.11, 36.69, 42.63, 43.35, 44.57, 46.79, 46.84, 47.84, 48.10, 50.85, 60.66, 62.17, 118.37 (C-2), 120.36 (C-1), 167.19 (C-3). Found, %: C, 82.42; H, 10.93; N, 3.10. C₃₁H₄₉NO. Calcd, %: C, 82.12; H, 10.99; N, 3.01. Mass spectrum *m/z* 451.1 [M]⁺. MM 451 g/mol.

Preparation of 3-Bromomethyl-3-oxo-1-cyano-2,3-seco-2-norlup-28-oic Acid Methyl Ester (6). A solution of **4** (4.0 mmol) in AcOH (40 mL) was treated with C₅H₆Br₃N (4.0 mmol). The reaction mixture was stirred at room temperature for 2 h. The formation of product was monitored by TLC. When the reaction was finished, the reaction mixture was diluted with H₂O. The product was extracted with EtOAc (2 × 50 mL). The organic layer was washed with aqueous NaHCO₃ solution (5%) and a small amount of H₂O and dried over anhydrous MgSO₄. The solvent was evaporated. The solid was purified by CC with elution by petroleum ether–EtOAc (10:1). Colorless crystals, yield 92%, R_f 0.46 (hexane–EtOAc, 7:3), mp 209.4°C (hexane–EtOAc), $[\alpha]_D^{20}$ +2.4° (*c* 0.5, CHCl₃). IR (ν, cm⁻¹): 2238 (C≡N), 1722 (COOCH₃, C=O). ¹H NMR (400 MHz, CDCl₃, δ, ppm, J/Hz): 0.75, 0.85 (3H each, d, J = 8.0, CH₃), 0.92, 0.96, 1.01, 1.25, 1.31 (3H each, s, CH₃), 1.93–2.00 (2H, m, H-19, 20), 2.37, 2.60 (1H each, d, J = 18.0, H-1, AB system), 3.63 (3H, s, H-31), 4.29, 4.35 (1H each, d, J = 12.0, H-32, AB system). ¹³C NMR (100 MHz, CDCl₃, δ, ppm): 14.56, 14.65, 15.80, 18.59, 21.72, 21.87, 22.73, 22.91, 23.95, 24.11, 26.89, 29.60, 29.65, 29.71, 31.78, 32.38, 33.27, 37.16, 38.12, 40.55, 42.50, 43.02, 44.09, 45.05, 48.72, 49.05, 51.14, 53.36, 56.99, 118.49 (C-2), 176.74 (C-28), 206.64 (C-3). Found, %: C, 66.65; H, 8.74; N, 2.43. C₃₂H₅₀BrNO₃. Calcd, %: C, 66.25; H, 8.63; N, 2.43. Mass spectrum *m/z* 576.1 [M]⁺. MM 576 g/mol.

Preparation of 31-O-Nicotinate-1-cyano-2,3-seco-2-norlup-28-oic Acid Methyl Ester (7). A solution of **6** (3.4 mmol) in anhydrous Me₂CO (10 mL) was treated with nicotinic acid (10.4 mmol), a 10-fold excess of Et₃N, and a four-fold excess of K₂CO₃. The reaction mixture was refluxed for 12 h. The course of the reaction was monitored by TLC. When the reaction was finished, the K₂CO₃ was filtered off. The reaction mixture was washed with HCl solution (10%) and with H₂O until neutral. The product was extracted with EtOAc (2 × 50 mL). The organic layer was separated, dried over anhydrous MgSO₄, and evaporated. The solid was purified by CC with elution by petroleum ether–EtOAc–CHCl₃ (5:1:1). Colorless crystals, yield 62%, R_f 0.15 (hexane–EtOAc, 7:3), mp 86.7°C (hexane–EtOAc), $[\alpha]_D^{20}$ +0.4° (*c* 0.5, CHCl₃). IR (ν, cm⁻¹): 2238 (C≡N), 1721 (COOCH₃, C=O). ¹H NMR (400 MHz, CDCl₃, δ, ppm, J/Hz): 0.75, 0.86 (3H each, d, J = 8.0, CH₃), 0.94, 1.00, 1.02, 1.26, 1.39 (3H each, s, CH₃), 2.49, 2.66 (1H each, d, J = 20.0, H-1, AB system), 3.64 (3H, s, H-31), 5.27, 5.35 (1H each, d, J = 18.0, H-32, AB system), 7.57–7.60 (1H, m, H-37), 8.52 (1H, d, J = 8.0, H-38), 8.84 (1H, d, J = 4.0, H-36), 9.30 (1H, br.s, H-35). ¹³C NMR (100 MHz, CDCl₃, δ, ppm): 14.56, 14.64, 15.83, 18.85, 21.86 (2C), 22.73 (2C), 22.89, 24.40, 26.90, 29.64, 29.70, 29.75, 31.78, 33.33, 37.15, 38.12, 40.57, 42.52, 43.02, 44.09, 45.12, 48.71, 48.77, 51.13, 52.02, 56.98, 65.91 (C-32), 118.53 (C-2), 124.42, 126.86, 139.92, 148.47, 150.60, 163.51, 176.74 (C-28), 206.76 (C-3). Found, %: C, 73.75; H, 8.80; N, 4.53. C₃₈H₅₄N₂O₅. Calcd, %: C, 73.95; H, 8.65; N, 4.30. Mass spectrum *m/z* 618.4 [M]⁺. MM 618 g/mol.

Preparation of 3-Methyl-1-cyano-2-norlup-1(3)-en-28-oic Acid Methyl Ester (8). A solution of **4** (4.0 mmol) in *t*-BuOH (15 mL) was treated with *t*-BuOK (12.0 mmol). The reaction mixture was refluxed for 2 h. The course of the reaction was monitored by TLC. When the reaction was finished, the mixture was diluted with HCl solution (5%) until neutral. The product was extracted by EtOAc (2 × 50 mL). The organic layer was separated, dried over anhydrous MgSO₄, and evaporated. The solid was purified by CC with elution by petroleum ether–EtOAc (15:1). Colorless crystals, yield 98%, *R_f* 0.43 (hexane–EtOAc, 5:1), mp 89.3°C (hexane–EtOAc), $[\alpha]_D^{20} -5.2^\circ$ (*c* 0.5, CHCl₃). IR (ν , cm⁻¹): 2207 (C≡N), 1728 (COOCH₃). ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 0.74, 0.85 (3H each, d, J = 4.0, CH₃), 0.92, 0.94, 0.96, 1.01, 1.09 (3H each, s, CH₃), 1.85 (3H, s, H-32), 2.18–2.25 (2H, m, H-19, 20), 3.64 (3H, s, H-31). ¹³C NMR (100 MHz, CDCl₃, δ , ppm): 12.70, 14.60, 14.74, 17.40, 17.41, 18.69, 19.57, 22.32, 22.78, 22.88, 26.43, 27.04, 29.62, 29.75, 32.27, 35.14, 37.39, 38.08, 42.37, 43.03, 44.28, 46.81, 47.00, 49.01, 50.91, 51.11, 56.88, 62.32, 118.36 (C-2), 120.45 (C-1), 167.10 (C-3), 176.81 (C-28). Found, %: C, 80.12; H, 10.30; N, 2.92. C₃₂H₄₉NO₂. Calcd, %: C, 80.92; H, 10.18; N, 2.82. Mass spectrum *m/z* 479.4 [M]⁺. MM 479 g/mol.

Screening for Cytotoxic Activity of 3–9. Cytotoxic activity of the synthesized compounds was determined by the classical MTT assay [26] using HEPG2, HCT116, MS, RD-TE32, MCF-7, A549, and PC-3 tumor cell lines and normal HEK293 cells. Cells were cultivated in DMEM (HEPG2, HCT116, MCF-7) and RPMI-1640 medium (RD TE32, MS, A549, PC-3) with added fetal bovine serum (10%), L-glutamine (2 mM), and penicillin-streptomycin solution (1%) at 37°C and 5% CO₂ in a humid atmosphere for 24 h. Then, the cell cultures were treated with the tested compounds in DMSO at concentrations of 3.125–100 μM. The controls were wells with added DMSO, the final concentration of which was ≤ 1% and was nontoxic to the cells. Cell survival was assessed after incubation for 72 h with the tested compounds by adding to each well a solution of MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide, 5 mg/mL] (20 μL) with subsequent determination of the optical density of the formed formazan at 544 nm on a FLUOstar Optima spectrophotometer (BMG Labtech, Germany). The 50% inhibitory concentrations (IC₅₀) of the tested compounds were determined using dose-dependent curves and GraphPad Prism 6.0 software. Results were given as means of three independent tests ± SD (Table 1).

ACKNOWLEDGMENT

The work was financially supported by Russian Science Foundation grant No. 21-13-00161. We thank the Center for Collective Use of PFRC, UrB, RAS, Research on Materials and Compounds, for facilitating the spectral and analytical studies and Candidate M. V. Dmitriev, Department of Organic Chemistry, Perm State Natl. Res. Univ., for performing the XSA of **5b**.

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