NEW FLAVONOIDS FROM Nonea rossica AND Tournefortia sibirica

D. N. Olennikov,^{1*} M. E. Kartashova,² V. V. Velichko,² and D. S. Kruglov²

The chemical composition of two species in the family Boraginaceae, Nonea rossica Steven and Tournefortia sibirica L., in which alkaloids, hydroxycinnamates, and flavonoids, including two new acylglycosides 1 and 2 were observed, was studied. UV and NMR spectroscopy and mass spectrometry found that the new compounds were quercetin $3-O-(2''-O-caffeoyl-6''-O-acetyl)-\beta-D-glucopyranoside (noneaside, 1; from N. rossica) and kaempferol <math>3-O-(2''-O-caffeoyl-6''-O-acetyl)-\beta-D-glucopyranoside (tournefoside, 2; from T. sibirica). Both flavonoids possessed antiradical activity.$

Keywords: Nonea rossica, Tournefortia sibirica, Boraginaceae, flavonoids, quercetin, kaempferol, antioxidants.

The family Boraginaceae is represented in Siberia by 28 genera including >100 species. Despite the broad distribution, the chemical compositions of most representatives have not been reported or are incomplete [1]. In continuation of research on this family [2, 3], two broadly distributed species in the region, *Nonea rossica* Steven [*N. pulla* subsp. *pulla*, *N. pulla* subsp. *rossica* (Steven) Soo] and *Tournefortia sibirica* L. [*Arguzia rosmarinifolia* Steven, *Messerschmidia sibirica* (L.) L.], were studied. The chemical composition of *N. rossica* is unknown while essential oil [4], flavones [5], the alkaloid tournesibirin [6], and cembrane diterpenoids [7] were observed in *T. sibirica* of Chinese origin. Both species were used in traditional Buryat medicine under the name gyer-shing-pa as antipyretic and antibacterial agents [8]. Herein, results of a chemical study of the aerial parts of *N. rossica* and *T. sibirica* growing in Baikal District are reported.

The EtOH extract of *N. rossica* was separated by column chromatography (CC) over polyamide, Al₂O₃, normal and reversed-phase silica gel, and Sephadex LH-20 and by preparative HPLC to afford 27 compounds including the alkaloids intermedine (**3**) [9], lycopsamine (**4**) [9], intermedine *N*-oxide (**5**) [9], and lycopsamine *N*-oxide (**6**) [9]; the flavonoids kaempferol 3-*O*-galactoside (**7**) [10], kaempferol 3-*O*-glucoside (*8*) [10], kaempferol 3-*O*-rutinoside (**9**) [10], kaempferol 3-*O*-neohesperidoside (**10**) [10], quercetin 3-*O*-glucoside (*11*) [10], quercetin 3-*O*-galactoside (**12**) [10], quercetin 3-*O*-rutinoside (**13**) [10], quercetin 3-*O*-neohesperidoside (**14**) [11], kaempferol 3-*O*-gentiobioside (**15**) [10], quercetin 3-*O*-gentiobioside (**16**) [10], quercetin 3-*O*-(2"-*O*-acetyl)-glucoside (**22**) [12], quercetin 3-*O*-(6"-*O*-acetyl)-glucoside (**23**) [12], and quercetin 3-*O*-(2",6"-di-*O*-acetyl)-glucoside (**24**) [12]; hydroxycinnamates 2-*O*-caffeoylthreonic acid (**17**) [13], 3-*O*-caffeoylthreonic acid (**18**) [13], 2-*O*-caffeoylglyceric acid (**19**) [13], 3-*O*-caffeoylglyceric acid (**20**) [13], globoidnan B (**21**) [13], rosmarinic acid (**25**) [13], and salvianolic acids B (**26**) [13] and L (**27**) [14]; in addition to new compound **1**.

The molecular formula $C_{32}H_{28}O_{16}$ for 1 was determined using mass spectrometry (HR-ESI-MS, *m/z* 669.4083 $[M + H]^+$, calcd for $C_{32}H_{29}O_{16}$, 669.5174) and ¹³C NMR spectroscopy. The shape of the absorption spectrum of 1 was characteristic of flavonoids acylated by a caffeic acid moiety, which was observed after hydrolysis of 1 with trifluoroacetic acid (TFA, 2 M) together with quercetin and D-glucose. Daughter ions of the protonated ion in the mass spectrum appeared at *m/z* 507, 465, and 303, indicating loss of fragments with masses of 162 (caffeoyl), 42 (acetyl), and 162 amu (glucose), respectively.

¹⁾ Institute of General and Experimental Biology, Siberian Branch, Russian Academy of Sciences, 6 Sakh'yanovoi St., Ulan-Ude, 670047, fax: 7 (3012) 43 47 43, e-mail: olennikovdn@mail.ru; 2) Novosibirsk State Medical University, Ministry of Health of the Russian Federation, 52 Krasnyi Prosp., Novosibirsk, 630091, e-mail: velichkvik@rambler.ru. Translated from *Khimiya Prirodnykh Soedinenii*, No. 6, November–December, 2022, pp. 859–862. Original article submitted April 25, 2022.

C atom	1		2	
	$\delta_{ m H}$	δ _C	$\delta_{ m H}$	δ _C
2	_	157.0	_	157.2
3	_	134.8	_	134.5
4	_	178.0	_	177.8
5	_	161.1	_	161.0
6	6.14 (1H, d, J = 2.0)	99.0	6.42 (1H, d, J = 2.0)	99.3
7	_	163.4	-	163.0
8	6.40 (1H, d, J = 2.0)	94.5	6.84 (1H, d, J = 2.0)	94.2
9	_	156.1	-	155.8
10	_	104.2	-	104.1
1′	_	120.3	-	119.4
2'	7.31 (1H, d, J = 2.1)	116.5	8.02 (2H, d, J = 9.0)	130.5
3'	_	145.4	6.99 (2H, d, J = 9.0)	115.1
4'	_	148.3	_	160.3
5'	6.91 (1H, d, J = 8.0)	115.9	6.99 (2H, d, J = 9.0)	115.1
6'	7.24 (1H, dd, J = 2.1, 8.0)	122.3	8.02 (2H, d, J = 9.0)	130.5
1″	5.01 (1H, d, J = 7.2)	102.1	4.95 (1H, d, J = 7.0)	101.8
2″	5.14 (1H, m)	74.9	5.16 (1H, m)	74.6
3″	3.68 (1H, m)	76.3	3.70 (1H, m)	76.5
4″	3.45 (1H, m)	71.6	3.47 (1H, m)	71.9
5″	3.61 (1H, m)	75.4	3.60 (1H, m)	75.2
6″	4.54 (1H, dd, J = 2.1, 11.8).	65.3	4.51 (1H, dd, J = 2.0, 12.0).	65.2
Ū.	4.29 (1H, dd, J = 5.4, 11.8)		4.33 (1H, dd, J = 5.2, 12.0)	
1‴	_	127.3	_	127.2
2.""	7.03 (1H, d, $J = 2.0$)	115.3	7.00 (1H, d, J = 1.9)	114.7
3'''	_	148.8	_	148.6
4'''	_	146.0	_	145.7
5'''	6.73(1H, d, J = 8.1)	117.2	6.70 (1H, d, J = 8.0)	117.1
5 6'''	6.84 (1H dd I = 2.0.81)	123.2	6.85(1H dd I = 1.9.80)	123.0
0 7‴′	7.55(1H d I = 15.8)	147.2	7.52 (1H d I = 16.0)	147.4
0""	6.30 (1H, d, J = 15.8)	11/.2	6.25(1H, d, J = 16.0)	11/.1
o 0‴	0.50 (111, 4, 5 – 15.0)	168 1	0.25 (111, d, 5 – 10.0)	168.0
۶ ۲ CU CO	- 2 03 (3H s)	21.2	- 2 01 (3H s)	21.0
$0 - \underline{C}H_3CO$	2.03 (311, 8)	21.3 160.2	2.01 (311, 8)	21.0
6 -СН <u>3С</u> О	_	109.5	—	109.1

TABLE 1. ¹H (500 MHz) and ¹³C NMR (125 MHz) Spectra of 1 and 2 (DMSO-d₆, 298 K, δ , ppm, J/Hz)

Work up of **1** with NaOH solution (0.5%) formed quercetin-3-*O*-glucoside (**11**) [10], which indicated acyl fragments were present in the carbohydrate part of the compound. A comparative analysis of ¹H NMR spectra of **1** and **11** revealed weak-field shifts of resonances in **1** for C-2" (δ_C 73.4 \rightarrow 74.9) and C-6" (δ_C 61.4 \rightarrow 65.3) (Table 1). The HMBC spectrum showed correlations between resonances of H-2" and the carbonyl C atom of caffeic acid (δ_H/δ_C 5.14/168.1) and H-6" and the carbonyl C atom of acetyl (δ_H/δ_C 4.29, 4.54/169.3). As a result, compound **1** was elucidated as quercetin 3-*O*-(2"-*O*-caffeoyl-6"-*O*-acetyl)- β -D-glucopyranoside, which we called noneaside.

Chromatographic separation of the EtOH extract of *T. sibirica* isolated **1**, **4**, **6**, **8**, **9**, **11**, **13**, **23**–**25**, 7-*O*-acetyllycopsamine (**28**) [15], 7-*O*-acetyllycopsamine *N*-oxide (**29**) [15], piperonal (**30**) [16], lithospermic acid (**31**) [13], rosmarinic acid 9-*O*-methyl ester (**32**) [17], kaempferol 3-*O*-(6"-*O*-acetyl)-glucoside (**33**) [10], and a new flavonoid **2**. All compounds were observed in *T. sibirica* for the first time.

Compound 2 ($C_{32}H_{28}O_{15}$, HR-ESI-MS, *m/z* 653.3004 [M + H]⁺, calcd for $C_{32}H_{29}O_{15}$, 653.5184) gave kaempferol, D-glucose, and caffeic acid after hydrolysis by TFA (2 M). Work up with NaOH solution (0.5%) gave kaempferol-3-*O*-glucoside (8) [10]. UV and NMR spectroscopic (Table 1) and mass spectrometric data indicated that 2 was an analog of 1 containing kaempferol as the aglycon or kaempferol 3-*O*-(2"-*O*-caffeoyl-6"-*O*-acetyl)- β -D-glucopyranoside, which was named tournefoside.



A study of the antiradical activity against DPPH radical showed the greatest activity (IC₅₀ 7.43 μ M) for 1, which exceeded those of the reference compound Trolox (IC₅₀ 30.08 μ M) and 2 (IC₅₀ 35.71 μ M).

According to the results, *N. rossica* and *T. sibirica* contained dehydropyrrolizidine alkaloids, caffeic acid esters, rosmarinic acid and its derivatives, and quercetin and kaempferol *O*-glycosides. These compound groups were encountered in other *Boraginaceae* species [18, 19]. However, flavonoid acylglycosides were found for the first time in the family.

EXPERIMENTAL

The aerial part of the flowering plants was collected in the Republic of Buryatia and air-dried in the shade (humidity < 5%): *N. rossica*, near Mukhorshibir (Mukhorshibirsky District, Jun. 20, 2020; 51°01′58.9″ N, 107°49′15.1″ E, 650 m above sea level); *T. sibirica*, near Gusinozersk (Selenginsky District, Jul. 15, 2019; 51°12′39.2″ N, 106°31′47.3″ E, 420 m above sea level). The species were determined by Dr. N. K. Chirikova. Specimens of raw material are preserved in the herbarium of the IGEB, SB, RAS (No. BU/BOR-0620/51-062, BU/BOR-0719/92-114). Column chromatography (CC) used polyamide, Al₂O₃, normal (SiO₂) and reversed-phase silica gel (RP-SiO₂), and Sephadex LH-20 (Sigma-Aldrich, St. Louis, MO, USA). Spectrophotometric studies used an SF-2000 spectrophotometer (OKB Spectr, St. Petersburg, Russia). Mass spectra were recorded in an LCMS-8050 TQ-mass spectrometer (Shimadzu, Columbia, MD, USA) [20]. NMR spectra were taken on a VXR 500S spectrometer (Varian, Palo Alto, CA, USA). Preparative HPLC used an LC-20 Prominence liquid chromatograph (Shimadzu) equipped with a Shim-pak PREP-ODS column (20×250 mm, d 15 µm) and an SPD-M30A diode array detector (Shimadzu) using flow rate 1.0 mL/min and column temperature 20°C. Antiradical activity of the compounds against 2,2-diphenyl-1-picrylhydrazyl radicals (DPPH, Sigma-Aldrich) was determined by a microplate spectrophotometric method [21].

Extraction and Isolation of 1 and 3–27 from *N. rossica*. Ground raw material (3.5 kg) was extracted with EtOH [70%, 1:10, 50°C, 3 ×, ultrasound (US) bath]. The EtOH extract was concentrated to dryness. The dry solid was exhaustively extracted with hexane and water-saturated BuOH. The BuOH fraction (630 g) was separated by CC (1:10) over polyamide with elution by H_2O (fraction A), EtOH (60%, faction B), and NH_3 (0.5% in 95% EtOH, fraction C). Fraction A (320 g) was separated by CC over Al_2O_3 (eluent $CHCl_3$ –EtOH– NH_3 , 99:0:1→59:40:1) and RP-SiO₂ (1 × 30 cm, eluent H_2O –MeCN, 90:10→50:50) and by preparative HPLC (eluent A, 0.2% HCOOH in H_2O ; eluent B, 0.2% HCOOH in MeCN; isocratic mode, 15% B) to isolate intermedine (10 mg, 3) [9], lycopsamine (5 mg, 4) [9], intermedine *N*-oxide (35 mg, 5) [9], and lycopsamine *N*-oxide (30 mg, 6) [9].

Fraction B (80 g) was chromatographed over SiO₂ (3×40 cm, eluent hexane–EtOAc, $100:0 \rightarrow 70:30$, EtOAc–Me₂CO, $100:0 \rightarrow 80:20$), Sephadex LH-20 (2×80 cm, eluent EtOH–H₂O, $90:10 \rightarrow 50:50$), and RP-SiO₂ (1×30 cm, eluent H₂O–MeCN, $95:5 \rightarrow 20:80$) and by preparative HPLC (eluent A, H₂O; eluent B, MeCN; gradient mode, %B: 0–60 min, 5–50%, 60–80 min, 50-60%) to afford kaempferol 3-*O*-galactoside (trifolin, 10 mg, 7) [10], kaempferol 3-*O*-glucoside (astragalin, 50 mg, **8**) [10], kaempferol 3-*O*-rutinoside (nicotiflorin, 60 mg, **9**) [10], kaempferol 3-*O*-neohesperidoside (30 mg, **10**) [10], quercetin 3-*O*-glucoside (isoquercitrin, 25 mg, **11**) [10], quercetin 3-*O*-galactoside (hyperoside, 20 mg, **12**) [10], quercetin 3-*O*-gentiobioside (rutin, 15 mg, **13**) [10], quercetin 3-*O*-gentiobioside (9 mg, **16**) [10]. Fraction C (200 g) was separated over Sephadex LH-20 ($2 \times 80 \text{ cm}$, eluent EtOH–H₂O, $50:50 \rightarrow 0:100$) and RP-SiO₂ ($1 \times 30 \text{ cm}$, eluent H₂O–MeCN, $95:5 \rightarrow 50:50$) and by preparative HPLC

(eluent A, 0.5% AcOH in H_2O ; eluent B, 0.5% AcOH in MeCN; gradient mode, %B: 0–20 min, 5–10%, 20–90 min, 10–40%, 90–150 min, 40–50%) to isolate 12 compounds including **1** (30 mg), 2-*O*-caffeoylthreonic acid (20 mg, **17**) [13], 3-*O*-caffeoylthreonic acid (25 mg, **18**) [13], 2-*O*-caffeoylglyceric acid (55 mg, **19**) [13], 3-*O*-caffeoylglyceric acid (70 mg, **20**) [13], globoidnan B (25 mg, **21**) [13], quercetin 3-*O*-(2"-*O*-acetyl)-glucoside (30 mg, **22**) [12], quercetin 3-*O*-(6"-*O*-acetyl)-glucoside (40 mg, **23**) [12], quercetin 3-*O*-(2",6"-di-*O*-acetyl)-glucoside (65 mg, **24**) [12], rosmarinic acid (1.4 g, **25**) [13], salvianolic acid B (30 mg, **26**) [13], and salvianolic acid L (45 mg, **27**) [14].

Noneaside (1), $C_{32}H_{28}O_{16}$. UV (MeOH, λ_{max} , nm): 255, 267 sh., 300 sh., 341; +AlCl₃ 271, 296 sh., 348 sh., 422; +AlCl₃ + HCl 265, 296 sh., 340, 400; +NaOAc 260, 296 sh., 388; +NaOAc + H₃BO₃ 260, 360. HR-ESI-MS, *m/z* 669.4083 [M + H]⁺ (calcd for $C_{32}H_{29}O_{16}$, 669.5174). ESI-MS, *m/z* (%): 669 [M + H]⁺ (100). ESI-MS² [669]: 507 [(M + H) – 162]⁺ (73), 465 [(M + H) – 162 – 42]⁺ (14), 303 [(M + H) – 2 × 162 – 42]⁺ (100). ¹H NMR (500 MHz, DMSO-d₆, 298 K) and ¹³C NMR (125 MHz, DMSO-d₆, 298 K), see Table 1.

Extraction and Isolation of 1, 2, 4, 6, 8, 9, 11, 13, 23–25, and 28–33 from *T. sibirica*. The above scheme was used to isolate from *T. sibirica* herb (1 kg) 1 (10 mg), 2 (35), 4 (15), 6 (40), 8 (10), 9 (25), 11 (20), 13 (10), 23 (140), 24 (35), 25 (220), 7-O-acetyllycopsamine (20 mg, 28) [15], 7-O-acetyllycopsamine *N*-oxide (5 mg, 29) [15], piperonal (heliotropin, 15 mg, 30) [16], lithospermic acid (40 mg, 31) [13], rosmarinic acid 9-O-methyl ester (35 mg, 32) [17], and kaempferol 3-O-(6"-O-acetyl)-glucoside (25 mg, 33) [10].

Tournefoside (2). $C_{32}H_{28}O_{15}$. UV (MeOH, λ_{max} , nm): 270, 328; +AlCl₃ 272, 298, 340, 387 sh.; +AlCl₃ + HCl 275, 298, 333, 388 sh.; +NaOAc 269, 332; +NaOAc + H₃BO₃ 261, 350. HR-ESI-MS, *m/z* 653.3004 [M + H]⁺ (calcd for $C_{32}H_{29}O_{15}$, 653.5184). ESI-MS, *m/z* (%): 653 [M + H]⁺ (100). ESI-MS² [653]: 491 [(M + H) – 162]⁺ (70), 449 [(M + H) – 162 – 42]⁺ (10), 287 [(M + H) – 2 × 162 – 42]⁺ (100).¹H NMR (500 MHz, DMSO-d₆, 298 K) and ¹³C NMR (125 MHz, DMSO-d₆, 298 K), see Table 1.

Hydrolysis. Acid hydrolysis was performed in TFA (2 M) [22] followed by analysis of the monosaccharide composition (HPLC after derivatization with 3-methyl-1-phenyl-2-pyrazolin-5-one) [23], determination of the D/L-series type (HPLC after reductive amination with L-tryptophan) [24], and determination of the aglycons by HPLC-MS [25]. The hydrolysis products contained quercetin (1), kaempferol (2), D-glucose (1, 2), and caffeic acid (1, 2). Alkaline hydrolysis used NaOH solution (0.5%) as before [22]. The hydrolysis products of 1 and 2 were identified using UV, NMR, and mass spectrometry as quercetin 3-*O*-glucoside (11) [10] and kaempferol 3-*O*-glucoside (8) [10], respectively.

ACKNOWLEDGMENT

The research was supported by the Ministry of Education and Science of the Russian Federation (Project No. 121030100227-7).

REFERENCES

- 1. R. A. Sharma, B. Singh, D. Singh, and P. Chandrawat, J. Med. Plants Res., 3, 1153 (2009).
- 2. D. N. Olennikov, Z. V. Daironas, and I. N. Zilfikarov, Chem. Nat. Compd., 53, 953 (2017).
- 3. D. N. Olennikov, D. S. Kruglov, Z. V. Daironas, and I. N. Zilfikarov, Chem. Nat. Compd., 56, 713 (2020).
- 4. K. Morteza-Semnani, M. Saeedi, and M. Akbarzadeh, J. Essent. Oil Res., 20, 207 (2008).
- 5. S. Diao, M. Jin, C. S. Jin, C.-X. Wei, J. Sun, W. Zhou, and G. Li, Nat. Prod. Res., 33, 3021 (2019).
- 6. H. Hu, A. Bao, S. Pan, J. Hao, and Y. Xin, Nat. Prod. Res., 36 (8), 2028 (2022).
- 7. S. Diao, M. Jin, J. Sun, C. Jin, R. Wang, W. Zhou, and G. Li, *Tetrahedron Lett.*, 61 (4), 151413 (2020).
- 8. S. M. Batorova, G. P. Yakovlev, and T. A. Aseeva, *Guide to Medicinal Plants of Traditional Tibetan Medicine* [in Russian], Nauka, Novosibirsk, 2003, 291 pp.
- 9. S. M. Colegate, D. R. Gardner, J. M. Betz, and K. E. Panter, *Phytochem. Anal.*, 25, 429 (2014).
- S. S. Azimova and V. I. Vinogradova, Natural Compounds. Flavonoids: Plant Sources, Structure and Properties, Springer, New York, 2013.
- 11. N. F. Komissarenko, V. T. Chernobai, and A. I. Derkach, Chem. Nat. Compd., 24, 675 (1988).

- Y. B. Wang, J. X. Pu, H. Y. Ren, J. F. Zhao, S. X. Mei, Z. Y. Li, H. B. Zhang, and L. Li, Chin, *Chem. Lett.*, 14, 1268 (2003).
- 13. J. Krzyzanowska-Kowalczyk, L. Pecio, J. Moldoch, A. Ludwiczuk, and M. Kowalczyk, *Molecules*, 23, 2277 (2018).
- 14. Y. Lu and L.Y. Foo, *Tetrahedron Lett.*, **42**, 8223 (2001).
- 15. J. Roitman, Aust. J. Chem., 36, 769 (1983).
- 16. B. Meriga, B. Parim, V. R. Chunduri, R. R. Naik, H. Nemani, P. Suresh, S. Ganapathy, and V. V. S. Uddandrao, *Nutr. Metab.*, **14**, 72 (2017).
- 17. A. Abedini, V. Roumy, S. Mahieux, M. Biabiany, A. Standaert-Vitse, C. Riviere, S. Sahpaz, F. Bailleul, C. Neut, and T. Hennebelle, *Evidence-Based Complementary Altern. Med.*, **2013**, 604536 (2013).
- 18. E. Wollenweber, R. Wehde, M. Dorr, and J. F. Stevens, Z. Naturforsch., C: J. Biosci., 57, 445 (2002).
- 19. M. Petersen and M. S. J. Simmonds, *Phytochemistry*, **62**, 121 (2003).
- 20. D. N. Olennikov, V. V. Chemposov, and N. K. Chirikova, *Plants*, 10, 2525 (2021).
- 21. D. N. Olennikov, C. S. Kirillina, and N. K. Chirikova, Antioxidants, 10, 1300 (2021).
- 22. D. N. Olennikov and N. K. Chirikova, Chem. Nat. Compd., 55, 1032 (2019).
- 23. D. N. Olennikov, N. K. Chirikova, N. I. Kashchenko, T. G. Gornostai, I. Y. Selyutina, and I. N. Zilfikarov, *Int. J. Mol. Sci.*, **18**, 2579 (2017).
- 24. M. Akabane, A. Yamamoto, S. Aizawa, A. Taga, and S. Kodama, Anal. Sci., 30, 739 (2014).
- 25. D. N. Olennikov, N. K. Chirikova, A. G. Vasilieva, and I. A. Fedorov, Antioxidants, 9, 526 (2020).