## **NEW FLAVONOIDS FROM** *Nonea rossica* **AND** *Tournefortia sibirica*

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*The chemical composition of two species in the family Boraginaceae,* Nonea rossica *Steven and* Tournefortia sibirica *L., in which alkaloids, hydroxycinnamates, and flavonoids, including two new acylglycosides 1 and 2 were observed, was studied. UV and NMR spectroscopy and mass spectrometry found that the new compounds were quercetin 3-*O*-(2*′′*-*O*-caffeoyl-6*′′*-*O*-acetyl)-*β*-D-glucopyranoside (noneaside, 1; from* N. rossica*) and kaempferol 3-*O*-(2*′′*-*O*-caffeoyl-6*′′*-*O*-acetyl)-*β*-D-glucopyranoside (tournefoside, 2; from* T. sibirica*). Both flavonoids possessed antiradical activity.*

**Keywords**: *Nonea rossica*, *Tournefortia sibirica*, Boraginaceae, flavonoids, quercetin, kaempferol, antioxidants.

The family Boraginaceae is represented in Siberia by 28 genera including >100 species. Despite the broad distribution, the chemical compositions of most representatives have not been reported or are incomplete [1]. In continuation of research on this family [2, 3], two broadly distributed species in the region, *Nonea rossica* Steven [*N. pulla* subsp. *pulla*, *N. pulla* subsp. *rossica* (Steven) Soo] and *Tournefortia sibirica* L. [*Arguzia rosmarinifolia* Steven, *Messerschmidia sibirica* (L.) L.], were studied. The chemical composition of *N. rossica* is unknown while essential oil [4], flavones [5], the alkaloid tournesibirin [6], and cembrane diterpenoids [7] were observed in *T. sibirica* of Chinese origin. Both species were used in traditional Buryat medicine under the name gyer-shing-pa as antipyretic and antibacterial agents [8]. Herein, results of a chemical study of the aerial parts of *N. rossica* and *T. sibirica* growing in Baikal District are reported.

The EtOH extract of *N. rossica* was separated by column chromatography (CC) over polyamide,  $Al_2O_3$ , normal and reversed-phase silica gel, and Sephadex LH-20 and by preparative HPLC to afford 27 compounds including the alkaloids intermedine (**3**) [9], lycopsamine (**4**) [9], intermedine *N*-oxide (**5**) [9], and lycopsamine *N*-oxide (**6**) [9]; the flavonoids kaempferol 3-*O*-galactoside (**7**) [10], kaempferol 3-*O*-glucoside (*8*) [10], kaempferol 3-*O*-rutinoside (**9**) [10], kaempferol 3-*O*-neohesperidoside (**10**) [10], quercetin 3-*O*-glucoside (**11**) [10], quercetin 3-*O*-galactoside (**12**) [10], quercetin 3-*O*-rutinoside (**13**) [10], quercetin 3-*O*-neohesperidoside (**14**) [11], kaempferol 3-*O*-gentiobioside (**15**) [10], quercetin 3-*O*-gentiobioside (**16**) [10], quercetin 3-*O*-(2′′-*O*-acetyl)-glucoside (**22**) [12], quercetin 3-*O*-(6′′-*O*-acetyl)-glucoside (**23**) [12], and quercetin 3-*O*-(2′′,6′′-di-*O*-acetyl) glucoside (**24**) [12]; hydroxycinnamates 2-*O*-caffeoylthreonic acid (**17**) [13], 3-*O*-caffeoylthreonic acid (**18**) [13], 2-*O*-caffeoylglyceric acid (**19**) [13], 3-*O*-caffeoylglyceric acid (**20**) [13], globoidnan B (**21**) [13], rosmarinic acid (**25**) [13], and salvianolic acids B (**26**) [13] and L (**27**) [14]; in addition to new compound **1**.

The molecular formula  $C_{32}H_{28}O_{16}$  for 1 was determined using mass spectrometry (HR-ESI-MS,  $m/z$  669.4083  $[M + H]^+$ , calcd for  $C_{32}H_{29}O_{16}$ , 669.5174) and <sup>13</sup>C NMR spectroscopy. The shape of the absorption spectrum of 1 was characteristic of flavonoids acylated by a caffeic acid moiety, which was observed after hydrolysis of **1** with trifluoroacetic acid (TFA, 2 M) together with quercetin and D-glucose. Daughter ions of the protonated ion in the mass spectrum appeared at *m/z* 507, 465, and 303, indicating loss of fragments with masses of 162 (caffeoyl), 42 (acetyl), and 162 amu (glucose), respectively.

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C atom	$\mathbf{1}$		$\overline{2}$	
	$\delta_{\rm H}$	$\delta_{\rm C}$	$\delta_{\rm H}$	$\delta_{\rm C}$
$\sqrt{2}$		157.0		157.2
$\sqrt{3}$		134.8		134.5
$\overline{4}$		178.0		177.8
$\mathfrak s$		161.1		161.0
$\sqrt{6}$	6.14 (1H, d, $J = 2.0$ )	99.0	6.42 (1H, d, $J = 2.0$ )	99.3
$\boldsymbol{7}$		163.4		163.0
$\,$ 8 $\,$	6.40 (1H, d, $J = 2.0$ )	94.5	6.84 (1H, d, $J = 2.0$ )	94.2
9		156.1		155.8
10		104.2		104.1
1'		120.3		119.4
$2^\prime$	7.31 (1H, d, $J = 2.1$ )	116.5	$8.02$ (2H, d, J = 9.0)	130.5
3'		145.4	6.99 (2H, d, $J = 9.0$ )	115.1
$4'$		148.3		160.3
$5^{\prime}$	6.91 (1H, d, $J = 8.0$ )	115.9	6.99 (2H, d, $J = 9.0$ )	115.1
$6^{\prime}$	7.24 (1H, dd, $J = 2.1$ , 8.0)	122.3	$8.02$ (2H, d, J = 9.0)	130.5
$1^{\prime\prime}$	5.01 (1H, d, $J = 7.2$ )	102.1	4.95 (1H, d, $J = 7.0$ )	101.8
$2^{\prime\prime}$	$5.14$ (1H, m)	74.9	$5.16$ (1H, m)	74.6
$3^{\prime\prime}$	$3.68$ (1H, m)	76.3	3.70(1H, m)	76.5
$4^{\prime\prime}$	$3.45$ (1H, m)	71.6	$3.47$ (1H, m)	71.9
$5^{\prime\prime}$	$3.61$ (1H, m)	75.4	$3.60$ (1H, m)	75.2
6''	4.54 (1H, dd, $J = 2.1$ , 11.8),	65.3	4.51 (1H, dd, $J = 2.0, 12.0$ ),	65.2
	4.29 (1H, dd, $J = 5.4$ , 11.8)		4.33 (1H, dd, $J = 5.2$ , 12.0)	
$1^{\prime\prime\prime}$		127.3		127.2
$2^{\prime\prime\prime}$	7.03 (1H, d, $J = 2.0$ )	115.3	7.00 (1H, d, $J = 1.9$ )	114.7
$3^{\prime\prime\prime}$		148.8		148.6
$4^{\prime\prime\prime}$		146.0		145.7
$5^{\prime\prime\prime}$	6.73 (1H, d, $J = 8.1$ )	117.2	6.70 (1H, d, $J = 8.0$ )	117.1
$6^{\prime\prime\prime}$	6.84 (1H, dd, $J = 2.0, 8.1$ )	123.2	6.85 (1H, dd, $J = 1.9$ , 8.0)	123.0
$7^{\prime\prime\prime}$	7.55 (1H, d, $J = 15.8$ )	147.2	7.52 (1H, d, $J = 16.0$ )	147.4
$8^{\prime\prime\prime}$	6.30 (1H, d, $J = 15.8$ )	114.8	6.25 (1H, d, $J = 16.0$ )	114.5
9'''		168.1		168.0
$6''$ -CH <sub>3</sub> CO	$2.03$ (3H, s)	21.3	$2.01$ (3H, s)	21.0
		169.3		169.1
$6''$ -CH <sub>3</sub> CO				

TABLE 1. <sup>1</sup>H (500 MHz) and <sup>13</sup>C NMR (125 MHz) Spectra of 1 and 2 (DMSO-d<sub>6</sub>, 298 K,  $\delta$ , ppm, J/Hz)

Work up of **1** with NaOH solution (0.5%) formed quercetin-3-*O*-glucoside (**11**) [10], which indicated acyl fragments were present in the carbohydrate part of the compound. A comparative analysis of 1H NMR spectra of **1** and **11** revealed weak-field shifts of resonances in 1 for C-2'' ( $\delta_C$  73.4→74.9) and C-6'' ( $\delta_C$  61.4→65.3) (Table 1). The HMBC spectrum showed correlations between resonances of H-2" and the carbonyl C atom of caffeic acid ( $\delta_H/\delta_C$  5.14/168.1) and H-6" and the carbonyl C atom of acetyl (δ<sub>H</sub>/δ<sub>C</sub> 4.29, 4.54/169.3). As a result, compound 1 was elucidated as quercetin 3-*O*-(2"-*O*-caffeoyl-6"-*O*-acetyl)-β-Dglucopyranoside, which we called noneaside.

Chromatographic separation of the EtOH extract of *T. sibirica* isolated **1**, **4**, **6**, **8**, **9**, **11**, **13**, **23–25**, 7-*O*-acetyllycopsamine (**28**) [15], 7-*O*-acetyllycopsamine *N*-oxide (**29**) [15], piperonal (**30**) [16], lithospermic acid (**31**) [13], rosmarinic acid 9-*O*-methyl ester (**32**) [17], kaempferol 3-*O*-(6′′-*O*-acetyl)-glucoside (**33**) [10], and a new flavonoid **2**. All compounds were observed in *T. sibirica* for the first time.

Compound **2** (C<sub>32</sub>H<sub>28</sub>O<sub>15</sub>, HR-ESI-MS,  $m/z$  653.3004 [M + H]<sup>+</sup>, calcd for C<sub>32</sub>H<sub>29</sub>O<sub>15</sub>, 653.5184) gave kaempferol, D-glucose, and caffeic acid after hydrolysis by TFA (2 M). Work up with NaOH solution (0.5%) gave kaempferol-3-*O*-glucoside (**8**) [10]. UV and NMR spectroscopic (Table 1) and mass spectrometric data indicated that **2** was an analog of **1** containing kaempferol as the aglycon or kaempferol 3-*O*-(2′′-*O*-caffeoyl-6′′-*O*-acetyl)-β-D-glucopyranoside, which was named tournefoside.



A study of the antiradical activity against DPPH radical showed the greatest activity  $(IC_{50} 7.43 \mu M)$  for 1, which exceeded those of the reference compound Trolox (IC<sub>50</sub> 30.08  $\mu$ M) and **2** (IC<sub>50</sub> 35.71  $\mu$ M).

According to the results, *N. rossica* and *T. sibirica* contained dehydropyrrolizidine alkaloids, caffeic acid esters, rosmarinic acid and its derivatives, and quercetin and kaempferol *O*-glycosides. These compound groups were encountered in other *Boraginaceae* species [18, 19]. However, flavonoid acylglycosides were found for the first time in the family.

## **EXPERIMENTAL**

The aerial part of the flowering plants was collected in the Republic of Buryatia and air-dried in the shade (humidity < 5%): *N. rossica*, near Mukhorshibir (Mukhorshibirsky District, Jun. 20, 2020; 51°01′58.9′′ N, 107°49′15.1′′ E, 650 m above sea level); *T. sibirica*, near Gusinozersk (Selenginsky District, Jul. 15, 2019; 51°12′39.2′′ N, 106°31′47.3′′ E, 420 m above sea level). The species were determined by Dr. N. K. Chirikova. Specimens of raw material are preserved in the herbarium of the IGEB, SB, RAS (No. BU/BOR-0620/51-062, BU/BOR-0719/92-114). Column chromatography (CC) used polyamide,  $\text{Al}_2\text{O}_3$ , normal (SiO<sub>2</sub>) and reversed-phase silica gel (RP-SiO<sub>2</sub>), and Sephadex LH-20 (Sigma-Aldrich, St. Louis, MO, USA). Spectrophotometric studies used an SF-2000 spectrophotometer (OKB Spectr, St. Petersburg, Russia). Mass spectra were recorded in an LCMS-8050 TQ-mass spectrometer (Shimadzu, Columbia, MD, USA) [20]. NMR spectra were taken on a VXR 500S spectrometer (Varian, Palo Alto, CA, USA). Preparative HPLC used an LC-20 Prominence liquid chromatograph (Shimadzu) equipped with a Shim-pak PREP-ODS column ( $20 \times 250$  mm, d 15  $\mu$ m) and an SPD-M30A diode array detector (Shimadzu) using flow rate 1.0 mL/min and column temperature 20°C. Antiradical activity of the compounds against 2,2-diphenyl-1-picrylhydrazyl radicals (DPPH, Sigma-Aldrich) was determined by a microplate spectrophotometric method [21].

**Extraction and Isolation of 1 and 3–27 from** *N. rossica*. Ground raw material (3.5 kg) was extracted with EtOH [70%, 1:10, 50 $\degree$ C, 3  $\times$ , ultrasound (US) bath]. The EtOH extract was concentrated to dryness. The dry solid was exhaustively extracted with hexane and water-saturated BuOH. The BuOH fraction (630 g) was separated by CC (1:10) over polyamide with elution by  $H_2O$  (fraction A), EtOH (60%, faction B), and NH<sub>3</sub> (0.5% in 95% EtOH, fraction C). Fraction A (320 g) was separated by CC over  $Al_2O_3$  (eluent CHCl<sub>3</sub>–EtOH–NH<sub>3</sub>, 99:0:1→59:40:1) and RP-SiO<sub>2</sub> (1 × 30 cm, eluent H<sub>2</sub>O–MeCN, 90:10→50:50) and by preparative HPLC (eluent A, 0.2% HCOOH in H<sub>2</sub>O; eluent B, 0.2% HCOOH in MeCN; isocratic mode, 15% B) to isolate intermedine (10 mg, **3**) [9], lycopsamine (5 mg, **4**) [9], intermedine *N*-oxide (35 mg, **5**) [9], and lycopsamine *N*-oxide (30 mg, **6**) [9].

Fraction B (80 g) was chromatographed over SiO<sub>2</sub> (3 × 40 cm, eluent hexane–EtOAc, 100:0→70:30, EtOAc–Me<sub>2</sub>CO, 100:0→80:20), Sephadex LH-20 (2 × 80 cm, eluent EtOH–H<sub>2</sub>O, 90:10→50:50), and RP-SiO<sub>2</sub> (1 × 30 cm, eluent H<sub>2</sub>O–MeCN, 95:5→20:80) and by preparative HPLC (eluent A, H<sub>2</sub>O; eluent B, MeCN; gradient mode, %B: 0–60 min, 5–50%, 60–80 min, 50–60%) to afford kaempferol 3-*O*-galactoside (trifolin, 10 mg, **7**) [10], kaempferol 3-*O*-glucoside (astragalin, 50 mg, **8**) [10], kaempferol 3-*O*-rutinoside (nicotiflorin, 60 mg, **9**) [10], kaempferol 3-*O*-neohesperidoside (30 mg, **10**) [10], quercetin 3-*O*-glucoside (isoquercitrin, 25 mg, **11**) [10], quercetin 3-*O*-galactoside (hyperoside, 20 mg, **12**) [10], quercetin 3-*O*-rutinoside (rutin, 15 mg, **13**) [10], quercetin 3-*O*-neoherperidoside (calendoflavobioside, 8 mg, **14**) [11], kaempferol 3-*O*-gentiobioside (5 mg, **15**) [10], and quercetin 3-*O*-gentiobioside (9 mg, **16**) [10]. Fraction C (200 g) was separated over Sephadex LH-20 (2 × 80 cm, eluent EtOH–H<sub>2</sub>O, 50:50→0:100) and RP-SiO<sub>2</sub> (1 × 30 cm, eluent H<sub>2</sub>O–MeCN, 95:5→50:50) and by preparative HPLC

(eluent A,  $0.5\%$  AcOH in H<sub>2</sub>O; eluent B,  $0.5\%$  AcOH in MeCN; gradient mode,  $%B: 0-20$  min,  $5-10\%$ ,  $20-90$  min,  $10-40\%$ , 90–150 min, 40–50%) to isolate 12 compounds including **1** (30 mg), 2-*O*-caffeoylthreonic acid (20 mg, **17**) [13], 3-*O*-caffeoylthreonic acid (25 mg, **18**) [13], 2-*O*-caffeoylglyceric acid (55 mg, **19**) [13], 3-*O*-caffeoylglyceric acid (70 mg, **20**) [13], globoidnan B (25 mg, **21**) [13], quercetin 3-*O*-(2′′-*O*-acetyl)-glucoside (30 mg, **22**) [12], quercetin 3-*O*-(6′′-*O*-acetyl)-glucoside (40 mg, **23**) [12], quercetin 3-*O*-(2′′,6′′-di-*O*-acetyl)-glucoside (65 mg, **24**) [12], rosmarinic acid (1.4 g, **25**) [13], salvianolic acid B (30 mg, **26**) [13], and salvianolic acid L (45 mg, **27**) [14].

**Noneaside (1)**, C<sub>32</sub>H<sub>28</sub>O<sub>16</sub>. UV (MeOH, λ<sub>max</sub>, nm): 255, 267 sh., 300 sh., 341; +AlCl<sub>3</sub> 271, 296 sh., 348 sh., 422;  $+AICI<sub>3</sub> + HCl$  265, 296 sh., 340, 400;  $+AaOAc$  260, 296 sh., 388;  $+AaOAc + H<sub>3</sub>BO<sub>3</sub>$  260, 360. HR-ESI-MS,  $m/z$  669.4083  $[M + H]$ <sup>+</sup> (calcd for C<sub>32</sub>H<sub>29</sub>O<sub>16</sub>, 669.5174). ESI-MS,  $m/z$  (%): 669 [M + H]<sup>+</sup> (100). ESI-MS<sup>2</sup> [669]: 507 [(M + H) – 162]<sup>+</sup> (73), 465  $[(M + H) - 162 - 42]^+$  (14), 303  $[(M + H) - 2 \times 162 - 42]^+$  (100). <sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>, 298 K) and <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ , 298 K), see Table 1.

**Extraction and Isolation of 1, 2, 4, 6, 8, 9, 11, 13, 23–25, and 28–33 from** *T. sibirica*. The above scheme was used to isolate from *T. sibirica* herb (1 kg) **1** (10 mg), **2** (35), **4** (15), **6** (40), **8** (10), **9** (25), **11** (20), **13** (10), **23** (140), **24** (35), **25** (220), 7-*O*-acetyllycopsamine (20 mg, **28**) [15], 7-*O*-acetyllycopsamine *N*-oxide (5 mg, **29**) [15], piperonal (heliotropin, 15 mg, **30**) [16], lithospermic acid (40 mg, **31**) [13], rosmarinic acid 9-*O*-methyl ester (35 mg, **32**) [17], and kaempferol 3-*O*-(6′′-*O*acetyl)-glucoside (25 mg, **33**) [10].

**Tournefoside (2)**.  $C_{32}H_{28}O_{15}$ . UV (MeOH,  $\lambda_{max}$ , nm): 270, 328; +AlCl<sub>3</sub> 272, 298, 340, 387 sh.; +AlCl<sub>3</sub> + HCl 275, 298, 333, 388 sh.; +NaOAc 269, 332; +NaOAc + H<sub>3</sub>BO<sub>3</sub> 261, 350. HR-ESI-MS,  $m/z$  653.3004 [M + H]<sup>+</sup> (calcd for C<sub>32</sub>H<sub>29</sub>O<sub>15</sub>, 653.5184). ESI-MS,  $m/z$  (%): 653 [M + H]<sup>+</sup> (100). ESI-MS<sup>2</sup> [653]: 491 [(M + H) – 162]<sup>+</sup> (70), 449 [(M + H) – 162 – 42]<sup>+</sup> (10), 287 [(M + H) – 2 × 162 – 42]<sup>+</sup> (100).<sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>, 298 K) and <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>, 298 K), see Table 1.

**Hydrolysis**. Acid hydrolysis was performed in TFA (2 M) [22] followed by analysis of the monosaccharide composition (HPLC after derivatization with 3-methyl-1-phenyl-2-pyrazolin-5-one) [23], determination of the D/L-series type (HPLC after reductive amination with L-tryptophan) [24], and determination of the aglycons by HPLC-MS [25]. The hydrolysis products contained quercetin (**1**), kaempferol (**2**), D-glucose (**1**, **2**), and caffeic acid (**1**, **2**). Alkaline hydrolysis used NaOH solution (0.5%) as before [22]. The hydrolysis products of **1** and **2** were identified using UV, NMR, and mass spectrometry as quercetin 3-*O*-glucoside (**11**) [10] and kaempferol 3-*O*-glucoside (**8**) [10], respectively.

## **ACKNOWLEDGMENT**

The research was supported by the Ministry of Education and Science of the Russian Federation (Project No. 121030100227-7).

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