CHEMICAL CONSTITUENTS OF THE LEAVES OF *Michelia figo*

H. C. Chen,¹ C. L. Kao,² C. T. Chen,³ H. T. Li,^{4*} and C. Y. Chen^{5*}

Species belonging to the genus *Michelia* are arboreous plants, growing in temperate zones of oriental India, southern China, Malaysia, and Indonesia. The species most utilized is *Michelia champaca*: its cortex and seeds are used as febrifuge and tonic-aromatic; the roots are employed as emmenagogue, the leaves as astringent, the gemmae in the treatment of hemorrhage, and the flowers and fruits are believed to possess curative properties in enteritis [1]. The less known species, *M. figo*, is used as ornamental plants and to obtain essences [1]. *M. figo* is an evergreen medium shrub, commonly called banana shrub, because of the heavy, sweet fragrant banana scent of its purple flowers. The plant is also known in Indian folk medicine as a remedy against hypertension [2]. To further understand the chemotaxonomy of the *Michelia* species [3–7], *M. figo* was chosen for phytochemical investigation. The chemical constituents of the leaves of this plant have not yet been reported. The compounds derived from the leaves include three alkaloids, (–)-nuciferine (1) [8], (–)-anonaine (2) [9], and *N*-methylcorydaldine (3) [10]; two steroids, β -sitostenone (4) [9] and stigmasta-4,22-dien-3-one (5) [9]; four benzenoids, *p*-hydroxybenzaldehyde (6) [10], *p*-hydroxybenzoic acid (7) [11], methylparaben (8) [11], and vanillin (9) [11]; six chlorophylls, pheophytin a (10) [12], pheophorbide a (11) [12], pheophytin b (12) [13], pheophorbide b (13) [13], aristophyll-C (14) [14], 13²-hydroxy-(13²-*S*)pheophytin a (15) [15]; and one sesquiterpene lactone, 11,13-dehydrolanuginolide (16) [16]. All of these known compounds were obtained for the first time from the leaves of this plant and were identified by direct comparison with authentic samples (TLC, UV, IR, ESI-MS and NMR) and the literature [8–16].

The leaves of M. figo (Lour.) Spreng. were collected from Chiayi County, Taiwan, May 2011. Plant material was identified by Prof. Fu-Yuan Lu (Department of Forestry and Natural Resources, College of Agriculture, National Chiayi University). A voucher specimen (Michelia 5) was deposited in the School of Medicinal and Health Sciences, Fooyin University, Kaohsiung City, Taiwan. The air-dried leaves of M. figo (4.8 kg) were extracted with MeOH (6 L×4) at room temperature, and a MeOH extract (121.6 g) was obtained upon concentration under reduced pressure. The MeOH extract, suspended in H₂O (1 L), was partitioned with CH_2Cl_2 (3 L × 5) to give fractions soluble in CH_2Cl_2 (67.9 g) and H_2O . The CH_2Cl_2 -soluble fraction was chromatographed over silica gel (950 g, 70-230 mesh) using n-hexane-EtOAc-MeOH mixtures as eluents to give five fractions. Part of fraction 1 (8.24 g) was subjected to silica gel chromatography by eluting with *n*-hexane–EtOAc (60:1) and enriched gradually with EtOAc to furnish five fractions (1-1-1-5). Fraction 1-2 (3.11 g) was further purified on a silica gel column using *n*-hexane–EtOAc mixtures to obtain 4 (12 mg) and 5 (6 mg). Part of fraction 2 (11.78 g) was subjected to silica gel chromatography by eluting with *n*-hexane–EtOAc (60:1) and enriched gradually with EtOAc to furnish five fractions (2-1–2-5). Fraction 2-1 (2.78 g) was further purified on a silica gel column using *n*-hexane–EtOAc mixtures to obtain 10 (3 mg) and 11 (4 mg). Fraction 2-2 (2.17 g) was further purified on a silica gel column using *n*-hexane–EtOAc mixtures to obtain 12 (1 mg) and 13 (2 mg). Fraction 2-3 (2.86 g) was further purified on a silica gel column using n-hexane-EtOAc mixtures to obtain 14 (13 mg) and 15 (9 mg). Fraction 2-4 (1.55 g) was further purified on a silica gel column using *n*-hexane–EtOAc mixtures to obtain 16 (18 mg). Part of fraction 3 (16.97 g) was subjected to silica gel chromatography by eluting with n-hexane–EtOAc (40:1) and enriched with EtOAc to furnish six further fractions (3-1–3-6). Fraction 3-3 (4.24 g) was further purified on a silica

1) Department of Nutrition and Health Science, Fooyin University, Ta-Liao, 83102, Kaohsiung, Taiwan; 2) Tzu Hui Institute of Technology, Pingtung County, Taiwan; 3) Department of Medical Sciences Industry, College of Health Sciences, Chang Jung Christian University, Tainan, Taiwan; 4) Department of Medical Laboratory Science and Biotechnology, Fooyin University, Ta-Liao, 83102, Kaohsiung, Taiwan, e-mail: mt085@fy.edu.tw; 5) School of Medical and Health Sciences, Fooyin University, Ta-Liao, 83102, Kaohsiung, Taiwan, e-mail: xx377@fy.edu.tw: Published in *Khimiya Prirodnykh Soedinenii*, No. 2, March–April, 2018, pp. 343–345. Original article submitted June 24, 2016.

gel column using CH_2Cl_2 –MeOH mixtures to obtain **3** (7 mg). Fraction 3-4 (3.24 g) was further purified on a silica gel column using CH_2Cl_2 –MeOH mixtures to obtain **2** (19 mg) and (–)-nuciferine (2 mg). Part of fraction 4 (21.23 g) was subjected to silica gel chromatography by eluting with CH_2Cl_2 –MeOH (40:1) and enriched with MeOH to furnish four fractions (4-1–4-4). Fraction 4-2 (16.72 g) eluted with CH_2Cl_2 –MeOH (40:1) was further separated using silica gel column chromatography and preparative TLC (CH_2Cl_2 –MeOH (50:1)) to give **6** (17 mg),**7** (22 mg), and **8** (13 mg). Part of fraction 5 (16.12 g) was subjected to silica gel chromatography by eluting with CH_2Cl_2 –MeOH (40:1) and enriched with MeOH to furnish four fractions (5-1–5-4). Fraction 5-3 (5.31 g) eluted with CH_2Cl_2 –MeOH (40:1) was further separated using silica gel column chromatography and preparative TLC (CH_2Cl_2 –MeOH (45:1)) to give **9** (14 mg).

(-)-Nuciferine (1). $C_{19}H_{21}NO_2$, brown powder (MeOH), mp 164–166°C. UV (λ_{max} , nm): 230, 274, 312. IR (ν_{max} , cm⁻¹): 1250, 1375, 1425, 1500, 1605. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 2.71–3.15 (4H, m, H-4, 5), 3.65 (3H, s, 1-OCH₃), 3.88 (3H, s, 2-OCH₃), 6.62 (1H, s, H-3), 7.23–7.26 (3H, m, H-8, 9, 10), 8.35 (1H, d, J = 7.6, H-11). ESI-MS *m/z* 319 [M + Na + H]⁺ [8].

(-)-Anonaine (2). $C_{17}H_{15}NO_2$, pale yellow powder (MeOH), mp 121–123°C. UV (λ_{max} , nm): 230, 272, 310. IR (v_{max} , cm⁻¹): 950, 1040. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 2.65 (1H, t, J = 13.4, H-7a), 2.85 (1H, dd, J = 13.4, 5.2, H-7b), 3.11–3.29 (3H, m, H-4a, 4b, 5a), 3.53 (1H, m, H-5b), 3.98 (1H, dd, J = 13.4, 5.2, H-6a), 5.92, 6.06 (each 1H, d, J = 1.6, OCH₂O), 6.55 (1H, s, H-3), 7.21–7.30 (3H, m, H-8, 9, 10), 8.06 (1H, d, J = 7.6, H-11). ESI-MS *m/z* 289 [M + Na + H]⁺ [9].

N-Methylcorydaldine (3). $C_{11}H_{15}NO_3$, colorless crystals (CHCl₃), mp 112–113°C. UV (λ_{max} , nm): 200, 219, 263, 303. IR (v_{max} , cm⁻¹): 1612, 1656. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 2.93 (2H, t, J = 6.8, H-4), 3.12 (3H, s, N-CH₃), 3.56 (2H, t, J = 6.8, H-3), 3.92 (3H, s, 6-OCH₃), 3.93 (3H, s, 7-OCH₃), 6.58 (1H, s, H-5), 7.41 (1H, s, H-8). ESI-MS *m/z* 245 [M + Na + H]⁺ [10].

β-Sitostenone (4). $C_{29}H_{48}O$, white needles (CHCl₃), mp 85–86°C. IR (v_{max} , cm⁻¹): 1375, 1385, 1460, 1620, 1675. ¹H NMR (400 MHz, CDCl₃, δ, ppm, J/Hz): 0.68 (3H, s, H-18), 0.81 (3H, d, J = 6.7, H-26), 0.84 (3H, s, H-27), 0.86 (3H, t, J = 7.1, H-29), 0.92 (3H, d, J = 6.0, H-21), 1.02 (3H, s, H-19), 5.72 (1H, d, J = 1.4, H-3). EI-MS *m/z* 414 [M]⁺ [9].

Stigmasta-4,22-dien-3-one (5). $C_{29}H_{46}O$, white needles (CHCl₃), mp 135–136°C. IR (v_{max} , cm⁻¹): 1375, 1385, 1460, 1620, 1675. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 0.68 (3H, s, H-18), 0.81 (3H, d, J = 6.7, H-26), 0.84 (3H, s, H-27), 0.86 (3H, t, J = 7.1, H-29), 0.93 (3H, d, J = 6.0, H-21), 1.02 (3H, s, H-19), 5.02 (1H, dd, J = 16.1, 8.3, H-22), 5.12 (1H, dd, J = 16.1, 8.3, H-23), 5.72 (1H, d, J = 1.4, H-3). EI-MS *m/z* 412 [M]⁺ [9].

p-Hydroxybenzaldehyde (6). $C_7H_6O_2$, brown powder (CHCl₃). UV (λ_{max} , nm): 223, 285, 290. IR (ν_{max} , cm⁻¹): 3200, 1660, 1600, 1155, 826. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 6.92 (2H, d, J = 8.4, H-3, 5), 7.80 (2H, d, J = 8.4, H-2, 6), 9.87 (1H, s, C<u>H</u>O). ESI-MS *m/z* 146 [M + Na + H]⁺ [10].

p-Hydroxybenzoic Acid (7). $C_7H_6O_3$, brown powder (CHCl₃). UV (λ_{max} , nm): 250, 285, 290. IR (ν_{max} , cm⁻¹): 3500, 1660, 1590, 1165, 845. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 6.85 (2H, d, J = 8.6, H-3, 5), 7.96 (2H, d, J = 8.6, H-2, 6). ESI-MS *m/z* 162 [M + Na + H]⁺ [11].

Methylparaben (8). $C_8H_8O_3$, colorless needles (CHCl₃), mp 130–131°C. UV (λ_{max} , nm): 225, 256, 310. IR (ν_{max} , cm⁻¹): 3400, 2950, 1695, 1610. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 3.88 (3H, s, COOCH₃), 5.46 (1H, br.s, OH), 6.87 (2H, d, J = 8.8, H-3, 5), 7.96 (2H, d, J = 8.8, H-2, 6). ESI-MS *m/z* 176 [M + Na + H]⁺ [11].

Vanillin (9). $C_8H_8O_3$, yellow powder (CHCl₃). UV (λ_{max} , nm): 220, 280, 310. IR (ν_{max} , cm⁻¹): 3400, 1670, 1595, 1025. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 3.95 (3H, s, 3-OCH₃), 6.20 (1H, br.s, OH), 7.09 (1H, d, J = 8.0, H-5), 7.31 (1H, d, J = 2.0, H-2), 7.42 (1H, dd, J = 8.0, 2.0, H-6), 9.77 (1H, s, CHO). ESI-MS *m*/z 176 [M + Na + H]⁺ [11].

Pheophytin a (10). $C_{55}H_{74}N_4O_5$, deep green needles (CHCl₃), mp 113–114°C. UV (λ_{max} , nm): 229, 274, 330, 372, 406, 508, 540, 610, 665. IR (ν_{max} , cm⁻¹): 3400, 1740, 1700, 1620. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): -1.62 (1H, br.s, NH, D₂O exchangeable), 0.81, 0.83 (each 3H, d, J = 6.6, H-38, 39), 0.87 (6H, d, J = 6.6, H-36, 37), 1.60 (3H, s, H-40), 1.01–1.10 (21H, m, H-24–35), 1.64 (3H, t, J = 7.6, H-8²), 2.22 (1H, m), 2.32 (1H, m), 2.54 (1H, m), 2.65 (1H, m), 3.22, 3.42 (each 3H, s, H-7¹, 2¹), 3.54 (2H, q, J = 7.6, H-8¹), 3.72, 3.88 (each 3H, s, H-12¹, OCH₃), 4.22 (1H, m, H-17), 4.48 (2H, m, H-18), 4.49 (1H, d, J = 7.2, H-21), 5.16 (1H, t, J = 7.4, H-22), 6.19 (1H, d, J = 11.6, H-3²), 6.30 (1H, d, J = 17.8, H-3²), 6.28 (1H, s, H-13²), 8.00 (1H, dd, J = 17.8, 11.4, H-3¹), 8.57, 9.39, 9.52 (each 1H, s, H-20, 5, 10). FAB-MS *m/z* 871 [M + H]⁺ [12].

Pheophorbide a (11). $C_{35}H_{36}N_4O_5$, deep green needles (CHCl₃), mp 115–116°C. UV (λ_{max} , nm): 229, 274, 330, 372, 408, 506, 536, 608, 665. IR (ν_{max} , cm⁻¹): 3400, 1740, 1700, 1620. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): –1.63 (1H, br.s, NH, D₂O exchangeable), 1.64 (3H, t, J = 7.6, H-8²), 2.22 (1H, m), 2.35 (1H, m), 2.50 (1H, m), 2.65 (1H, m), 3.22, 3.40 (each 3H, s, H-7¹, 2¹), 3.52 (2H, q, J = 7.6, H-8¹), 3.70, 3.88 (each 3H, s, H-12¹, OCH₃), 4.22 (1H, m, H-17), 4.47 (2H, m, H-18),

6.17 (1H, d, J = 11.4, H-3²), 6.28 (1H, s, H-13²), 6.30 (1H, d, J = 17.8, H-3²), 8.00 (1H, dd, J = 17.8, 11.4, H-3¹), 8.57, 9.39, 9.52 (each 1H, s, H-20, 5, 10). FAB-MS m/z 593 [M + H]⁺ [12].

Pheophytin b (12). $C_{55}H_{72}N_4O_6$, deep green needles (CHCl₃), mp 118–119°C. UV (λ_{max} , nm): 233, 280, 330, 411, 435, 536, 608, 665. IR (ν_{max} , cm⁻¹): 3500, 1730, 1700, 1665, 1616. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): -1.49 (1H, br.s, NH, D₂O exchangeable), 0.77 (3H, d, J = 6.4, H-38), 0.78 (3H, d, J = 6.4, H-39), 0.83 (6H, d, J = 6.6, H-36, 37), 1.58 (3H, s, H-40), 1.00–1.90 (21H, m, H-24–35), 1.63 (3H, t, J = 7.6, H-8²), 2.21 (1H, m), 2.35 (1H, m), 2.50 (1H, m), 2.64 (1H, m), 3.38 (3H, s, H-2¹), 3.69 (3H, s, H-7¹), 3.88 (2H, q, J = 7.6, H-8¹), 3.90 (3H, s, OCH₃), 4.07 (1H, m, H-17), 4.45 (2H, m, H-18), 4.53 (2H, m, H-21), 5.14 (1H, t, J = 7.2, H-22), 6.21 (1H, d, J = 11.6, H-3²), 6.23 (1H, s, H-13²), 6.37 (1H, d, J = 17.8, H-3²), 8.01 (1H, dd, J = 17.8, 11.6, H-3¹), 8.54, 9.66, 10.38 (each 1H, s, H-20, 5, 10), 11.01 (1H, s, CHO). FAB-MS *m/z* 885 [M + H]⁺ [13].

Pheophorbide b (13). $C_{35}H_{34}N_4O_6$, deep green needles (CHCl₃), mp 118–119°C. UV (λ_{max} , nm): 233, 280, 330, 411, 435, 536, 608, 665. IR (ν_{max} , cm⁻¹): 3500, 1730, 1700, 1665, 1616. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): –1.49 (1H, br.s, NH, D₂O exchangeable), 1.63 (3H, t, J = 7.6, H-8²), 2.21 (1H, m), 2.35 (1H, m), 2.50 (1H, m), 2.64 (1H, m), 3.38 (3H, s, H-2¹), 3.69 (3H, s, H-7¹), 3.88 (2H, q, J = 7.6, H-8¹), 3.90 (3H, s, OCH₃), 4.07 (1H, m, H-17), 4.45 (2H, m, H-18), 6.21 (1H, d, J = 11.6, H-3²), 6.23 (1H, s, H-13²), 6.37 (1H, d, J = 17.8, H-3²), 8.01 (1H, dd, J = 17.8, 11.6, H-3¹), 8.54, 9.66, 10.38 (each 1H, s, H-20, 5, 10), 11.01 (1H, s, C<u>H</u>O). FAB-MS *m/z* 607 [M + H]⁺ [13].

Aristophyll-C (14). $C_{53}H_{70}N_4O_5$, deep green needles (CHCl₃), mp 247–248°C. UV (λ_{max} , nm): 282, 360, 412, 480, 512, 550, 642, 702. IR (ν_{max} , cm⁻¹): 1740, 1725. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): -0.18, 0.12 (each 1H, br.s, NH, D₂O exchangeable), 0.78, 0.80 (each 3H, d, J = 7.0, H-38, 39), 0.83 (6H, d, J = 6.8, H-36, 37), 1.01–1.62 (21H, m), 1.62 (3H, s, H-40), 1.66 (3H, t, J = 7.6, H-8²), 1.75 (3H, d, J = 7.0, H-18¹), 2.04 (1H, m, H-17¹), 2.45 (2H, m, H-17¹, 17²), 2.73 (1H, m, H-17²), 3.15 (3H, s, CH₃-7), 3.37 (3H, s, CH₃-2), 3.62 (2H, q, J = 7.6, H-8¹), 3.75 (3H, s, CH₃-12), 4.36 (1H, q, J = 7.0, H-18), 4.52 (2H, m, H-21), 5.22 (2H, m, H-17, 22), 6.22 (1H, d, J = 11.4, H-3²), 6.30 (1H, d, J = 18.0, H-3²), 7.89 (1H, dd, J = 18.0, 11.5, H-3¹), 8.56, 9.40, 9.56 (each 1H, s, H-20, 5, 10). FAB-MS *m/z* 843 [M + H]⁺ [14].

13²-Hydroxy-(13²-S)-pheophytin a (15). $C_{55}H_{74}N_4O_6$, deep green needles (CHCl₃), mp 205–206°C. UV (λ_{max} , nm): 225, 410, 505, 611, 665. IR (ν_{max} , cm⁻¹): 3400, 1740, 1700, 1620. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): -1.82 (1H, br.s, NH, D₂O exchangeable), 0.77, 0.78 (each 3H, d, J = 6.4, H-38, 39), 0.83 (3H, d, J = 6.4, H-36), 0.86 (3H, d, J = 6.4, H-37), 1.60 (3H, s, H-40), 1.62 (3H, d, J = 7.4, H-18¹), 2.21–2.34 (2H, m), 2.56 (1H, m), 2.93 (1H, m), 3.27, 3.43 (each 3H, s, H-7¹, 2¹), 3.61 (3H, s, H-12¹, OCH₃), 3.75 (3H, s, H-13⁴, OCH₃), 4.16 (1H, m, H-17), 4.52 (2H, m, H-18), 4.57 (1H, d, J = 7.2, H-21), 5.20 (1H, t, J = 7.2, H-22), 6.20 (1H, d, J = 11.6, H-3²), 6.30 (1H, d, J = 17.8, H-3²), 8.03 (1H, dd, J = 17.8, 11.6, H-3¹), 8.64, 9.50, 9.63 (each 1H, s, H-20, 5, 10). FAB-MS *m/z* 887 [M + H]⁺ [15].

11,13-Dehydrolanuginolide (16). $C_{17}H_{22}O_5$, colorless needles (MeOH), mp 168–170°C. UV (λ_{max} , nm): 210. IR (ν_{max} , cm⁻¹): 1770, 1655. ¹H NMR (400 MHz, CDCl₃, δ , ppm, J/Hz): 1.29 (3H, s, H-15), 1.84 (3H, s, H-14), 3.30 (1H, dd, J = 7.2, 3.6, H-7), 4.32 (1H, dd, J = 9.2, 6.8, H-6), 4.60 (1H, m, H-8), 5.33 (1H, m, H-1), 5.80, 6.43 (each 1H, d, J = 3.4, H-13). ESI-MS *m/z* 330 [M + Na + H]⁺ [16].

ACKNOWLEDGMENT

This investigation was supported by a grant from the Ministry of Science and Technology of the Republic of China (MOST-104-2320-B-0242- 001-MY3) awarded to C. Y. Chen.

REFERENCES

- 1. C. T. Huang, S. J. Chen, H. M. Wu, Y. F. Kang, H. L. Chen, W. J. Li, H. T. Li, and C. Y. Chen, *Chem. Nat. Compd.*, **50**, 1047 (2014).
- 2. S. Chericoni, L. Testai, E. Campeol, V. Calderone, I. Morelli, and E. Martinotti, J. Ethnopharmacol., 91, 263 (2004).
- 3. W. L. Lo, L. Y. Huang, H. M. Wang, and C. Y. Chen, *Chem. Nat. Compd.*, **46**, 664 (2010).
- 4. H. M. Wang, W. L. Yang, S. C. Yang, and C. Y. Chen, Chem. Nat. Compd., 46, 327 (2010).
- M. J. Cheng, W. L. Lo, J. C. Huang, Y. T. Yeh, Z. L. Hong, Y. C. Lu, M. S. Chang, and C. Y. Chen, *Nat. Prod. Res.*, 24, 682 (2010).

- 6. W. L. Lo, J. C. Huang, L. Y. Huang, and C. Y. Chen, *Nat. Prod. Res.*, 24, 326 (2010).
- C. Y. Chen, L. Y. Huang, L. J. Chen, W. L. Lo, S. Y. Kuo, Y. D. Wang, S. H. Kuo, and T. J. Hsieh, *Chem. Nat. Compd.*, 44, 137 (2008).
- 8. H. Guinaudeau, A. Cave, and R. R. Paris, *Phytochemistry*, **10**, 1963 (1971).
- 9. C. Y. Chen, F. R. Chang, and Y. C. Wu, J. Chin. Chem. Soc., 44, 313 (1997).
- 10. A. K. Sinha, S. C. Verma, and U. K. Sharma, J. Sep. Sci., 30, 15 (2007).
- 11. C. Y. Chen, F. R. Chang, C. M. Teng, and Y. C. Wu, J. Chin. Chem. Soc., 46, 77 (1999).
- 12. J. S. Walkera, A. H. Squiera, D. A. Hodgsonb, and B. J. Keelya, Org. Geochem., 33, 1667 (2002).
- 13. Y. Nakatani, G. Ourisson, and J. P. Beck, *Chem. Pharm. Bull.*, **29**, 2261 (1981).
- 14. Y. Y. Chan, Y. L. Leu, and T. S. Wu, Chem. Pharm. Bull., 47, 887 (1999).
- 15. L. Ma and D. Dolphin, J. Org. Chem., 61, 2501 (1996).
- 16. S. K. Talapatra, A. Patra, and B. Talapatra, *Phytochemistry*, **12**, 1827 (1973).