

CHEMICAL CONSTITUENTS OF *Penicillium chrysogenum*, AN ENDOPHYTIC FUNGUS FROM *Strychnos toxifera*

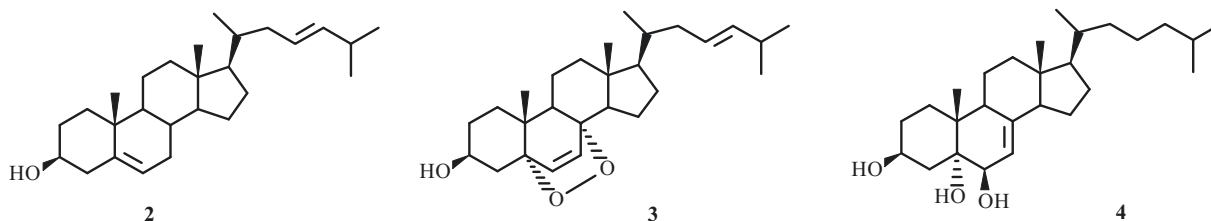
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UDC 547.918

Strychnos toxifera is a poisonous plant used by Amazon indians in the manufacture of curare poison [1]. Its toxicity is due to indole alkaloids, which are the major constituents of this poison [2]. Endophytes are considered outstanding and underexplored sources of novel chemically diverse and bioactive compounds. These microorganisms can be detected at a particular moment within the tissues of apparently healthy plant hosts, and they have been found in all plant species examined to date [3]. As they occupy unique biological niches, the complex web of interactions with other endophytes and with the host might give rise to new chemical diversity and bioactive compounds [4]. In the course of screening for biologically active agents from Amazonian endophytic fungi [5], *Penicillium chrysogenum*, an endophytic fungus from the trunk of *S. toxifera*, was chosen for further phytochemical investigation.

The MeOH extract of the mycelial mass from *P. chrysogenum* was subjected to chromatographic separation to afford four steroids [6] [ergosterol (1), brassicasterol (2), ergosterol peroxide (3), and cerevisterol (4)], two aromatic acids [7] [cinnamic acid (5) and *p*-hydroxybenzoic acid (6)], and three mycotoxins [8] [kojic acid (7), penicillic acid (8), and patulin (9)]. These compounds were obtained and characterized by a comparison of their physical and spectral data (UV, IR, NMR and MS), with values in the literature.

S. toxifera was collected from Manaus, Brazil in July, 2008. The strain of *P. chrysogenum* was obtained, purified, and identified from the healthy tissues of the trunk using a previous methodology [9]. After cultivation in liquid media [5], the mycelial mass (1.1 kg) was extracted with MeOH (3 × 500 mL) at room temperature, and a MeOH extract (30.4 g) was obtained upon concentration under reduced pressure. The MeOH extract was chromatographed over silica gel (600 g, 70–230 mesh) using *n*-hexane containing increasing amounts of EtOAc to obtain nine fractions. Fraction 1 (321.7 mg) was purified on a silica gel column with *n*-hexane–CHCl₃ with increasing gradient, yielding 1 (22 mg), 2 (7 mg), 3 (12 mg), and 4 (9 mg). Fraction 5 (507 mg) was subjected to silica gel chromatography with CHCl₃–MeOH with increasing gradient, yielding 5 (4 mg), 6 (11 mg), 7 (89 mg), 8 (9 mg), and 9 (5 mg).



ACKNOWLEDGMENT

This investigation was supported by a Grant from the Fundacao de Amparo a Pesquisa do Estado do Amazonas (FAPEAM).

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REFERENCES

1. M. A. Silva, A. R. M. Souza-Brito, C. A. H. Lima, S. Sannomiya, and W. Vilegas, *Rev. Bras. Farmacogn.*, **15**, 256 (2005).
2. G. Phillipe, L. Angenot, T. Monique, and F. Michel, *Toxicon*, **44**, 405 (2004).
3. J. Clardy and C. Walsh, *Nature*, **432**, 829 (2004).
4. B. Schulz and C. Boyle, *Mycol. Res.*, **109**, 661 (2005).
5. H. H. F. Koolen, E. R. Soares, F. M. A. da Silva, R. A. Almeida, A. D. L. de Souza, L. S. de Medeiros, E. Rodrigues-Filho, and A. Q. L. de Souza, *Quim Nova*, **35**, 771 (2012).
6. A. M. R. Marinho, P. S. B. Marinho, and E. Rodrigues-Filho, *Quim. Nova*, **32**, 1710 (2009).
7. W. A. Ayer, L. M. Browne, C. M. Feng, H. Orszanska, and H. S. Ghomi, *Can. J. Chem.*, **64**, 904 (1986).
8. B. Andersen, J. Smedsgaard, and J. Frisvad, *J. Agric. Food Chem.*, **52**, 2421 (2004).
9. A. Q. L. de Souza, A. D. L. de Souza, S. Astolfi-Filho, M. L. B. Pinheiro, M. I. M. Sarquis, and J. O. Pereira, *Acta Amazon.*, **34**, 185 (2004).