Genetics of breast cancer bone metastasis: a sequential multistep pattern

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Abstract Bone metastasis accounts for the vast majority of breast cancer (BC) metastases, and is related to a high rate of morbidity and mortality. A number of seminal studies have uncovered gene expression signatures involved in BC development and bone metastasis; each of them points at a distinct step of the 'invasion-metastasis cascade'. In this review, we provide most recently discovered functions of sets of genes that are selected from widely accepted gene signatures that are implicate in BC progression and bone metastasis. We propose a possible sequential pattern of gene expression that may lead a benign primary breast tumor to get aggressiveness and progress toward bone metastasis. A panel of genes which primarily deal with features like DNA replication, survival, proliferation, then, angiogenesis, migration, and invasion has been identified. TGF-B, FGF, NFKB, WNT, PI3K, and JAK-STAT signaling pathways, as the key pathways involved in breast cancer development and metastasis, are evidently regulated by several genes in all three signatures. Epithelial to mesenchymal transition that is also an important mechanism in cancer stem cell generation and metastasis is evidently regulated by these genes. This review provides a comprehensive insight regarding breast cancer bone metastasis that may lead to a better understanding of the disease and take step toward better treatments.

Keywords Breast cancer metastasis · Bone metastasis · Gene signature · Signaling network

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Metastasis, a complex multistep event

Breast cancer (BC) metastasis is the leading cause of cancer related deaths among women (metastasis; the spread of tumor cells from primary site to distant organs) [1, 2]. More than 70 % of BC patients in advanced stage develop bone metastases which are related to high rate of morbidity and mortality [3, 4]. The vast majority of researchers have focused to identify the underlying molecular and cellular mechanisms of this complex phenomenon. Developing gene expression signatures, involved in certain stages of BC metastasis, are among seminal discoveries that may let to understand the basics involved in the 'invasion-metastasis cascade', and consequently the generation of effective therapies against metastasis.

'Invasion-metastasis cascade' represents a multistep process that consists a set of sequential events, in which tumor cells invade from primary site, disseminate through circulation, and reconstitute secondary tumors at distant tissue(s) [5]. It has been investigated that breast tumor cells encounter a plenty of changes when disseminating from primary to distant sites, including changes in gene expression pattern [6], and in the states of stemness [7], that define clonal evolution and cancer stem cell (CSC) theories, respectively. Genetic alterations and subsequent shift in cellular events and states must be considered in the context of different microenvironments in different steps of metastasis in order to uncover mysteries of the complex process of metastasis cascade.

Clonal evolution and CSC theories are two models for tumor heterogeneity, cancer development, and metastasis that are widely accepted [8, 9]. Clonal evolution theory indicates that different lineages of cancer cells are developed during multistep genetic and epigenetic alterations, which lead to acquiring different features required for tumor development and metastasis [10]. On the other hand, CSCs, which have features of self-renewal, tumorigenesis, multilineage differentiation, motility, invasiveness and apoptosis resistance, are believed to be required for the development and maintenance of several forms of human cancers, including BC. Based on the CSC theory, tumor cells are not naturally alike, based on the state of stemness, and are organized in a hierarchical pattern in which CSCs are considered to be at the top of the apex [11–13]. It is believed that both models of clonal evolution and CSC can be applied in cancer development and metastasis, and that tumors heterogeneity can be generated from both of them [8].

The pattern of gene expression is likely the most critical determinant of CSC state. A quite sophisticated program that leads to expression of a group of genes, and simultaneously suppression of others, directs all features of a tumor cell in a precise time. Gene expression signatures have been developed since more than a decade for a variety of diseases like cancer metastasis. Gene signatures that have been developed for BC metastasis are ostensibly behind-the-scene forces of cellular events. In this study, we provide most recently discovered functions of sets of genes that are selected from seminal widely accepted gene signatures. We then propose a possible sequential pattern of gene expression that may lead to a benign primary breast tumor to get aggressiveness and progress toward bone metastasis. We also discuss most prominent molecular mechanisms involved in BC bone metastasis.

Gene signatures associated to breast cancer progression and bone metastasis

A number of seminal studies have uncovered gene expression signatures involved in BC development and metastasis; each of them points at a distinct step of the 'invasion-metastasis cascade' (Tables 1, 2, 3, 4) [14-16]. Today, these findings have entered to the diagnosis as predictors of disease outcome in BC patients [17]. Particularly, such discoveries have heralded the new era of personalized medicine, while predicting the clinical outcome of patients based on a set of distinct gene expression patterns [18]. Although improving, our understanding of the exact molecular and, most importantly, cellular mechanisms of BC metastasis is poor, and therefore reliable treatments are lacking. Analysis of data resulting from high throughput genome wide assays, and translation of the molecular pattern to cellular mechanisms/pathways may provide novel perspective to understand the complex nature of metastasis, and subsequently develop new therapeutic strategies. Among studies that have provided gene signatures for BC progression and bone metastasis, the

Table 1 Gene signatures associated to breast cancer metastasis

Gene signature (authors)	Type of study	Year of study
Perou, C.M. et al.	In vitro, and in vivo (human primary breast tumor)	1999
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Zajchowski, D.A. et al.	In vitro	2001
Sorlie, T. et al.	In vivo (human primary breast tumor)	2001
West, M. et al.	In vivo (human primary breast tumor)	2001
van't Veer, L. J. et al.	In vivo (human primary breast tumor)	2002
Kang, Y. et al.	In vitro, and in vivo (mouse metastatic bone metastatic tumor)	2003
Smid, M. et al.	In vivo (human primary breast tumor)	2006

ones by van't Veer et al., Smid et al., and Kang et al. are of most seminal and widely accepted.

van't Veer's signature

Several studies regarding gene expression pattern in BC have been developed since more than a decade (Table 1). Earliest ones [19-23] were not quite sufficient to be utilized for predictive and therapeutic purposes. This may be because of the inconsistency in different studies (e.g., different kinds of primary tumors), and abundance of heterogeneity in tumors. The first highly applicable reported gene signature, by van't Veer and colleagues (van't Veer's signature), have established a 70-gene prognosis profile from primary tumors of young BC patients. (Table 2 comprises a panel of selected genes from van't Veer's signature, based on their significant expression in most poor prognosed patients, and the status of being well studied). They identified a set of genes strongly predicting distant metastasis in patients who were lymph node negative, called poor prognosis signature [14]. Their finding uncovered a pattern of gene expression required for primary tumor cells to become invasive; capable to evade from primary site.

van't Veer's signature comprises genes associated to a well-orchestrated program for the regulation of different features required for primary tumors to grow and escape from primary site, including cell cycle, DNA replication, proliferation, tumorigenesis, survival, angiogenesis, migration, and invasion (Table 2). In particular, among those, CCNB2, CCNE2, MCM6, TSPYL5, NUSAP1, CMC2, ECT2, ORC6, DTL, PRC1, MELK, EGLN1, SLC2A3, RAB6B, ESM1, RAD21, CDC25B, CDK16, CENPA, PGK1, MAD2L1, CKS2, BUB1, FGF18, WISP1, and IGFBP5 are known regulators of cell cycle, DNA

Table 2 van't Veer's signature of breast cancer poor/good prognosis genes

Gene symbol	Gene description	Functions and features
Cell cycle/DNA	replication/proliferation/tumorigenesis/survival	
CCNB2	Cyclin B2	Cell cycle control [24], Serves as oncogene, and potential biomarker of unfavorable prognosis [125], TGF-β-mediated cell cycle control [27]
CCNE2	Cyclin E2	Cell cycle control [24], resistance to both endocrine therapy and CDK4 inhibition [126], increase in pathogenesis and kinase activity [127]
МСМ6	Mini-chromosome maintenance proteins	DNA replication, and growth [25]
TSPYL5	TSPY-like 5	Suppression of p53 [128], cell growth via regulation of p21 and PTEN/AKT pathways [129]
NUSAP1	Nucleolar and spindle associated protein 1	Activation by E2F1 [130], spindle organization in mitosis thorough cdk1 activation [131, 132]
CMC2	COX assembly mitochondrial protein 2 homolog	Cytochrome C oxidase biogenesis [133]
ECT2	Epithelial cell transforming sequence 2 oncogene	Cell cycle progression and proliferation [134], mesenchymal amoeboid transition [135] and migration [136]
ORC6	Origin recognition complex, subunit 6	Proliferation [137]
DTL	Denticleless E3 ubiquitin protein ligase homolog	Tumor growth [138], proliferation, survival, and metastasis [139, 140]
PRC1	Protein regulator of cytokinesis 1	Proliferation [141]
MELK	Maternal embryonic leucine zipper kinase	Proliferation of tumor and CSCs, and inhibition of apoptosis [142-145]
EGLN1	egl-9 family hypoxia-inducible factor 1	Proliferation, survival [146], target of HIF [147], involved in the processing of TGF-β1 [28], angiogenesis [148], maintenance of epithelial differentiation [149]
SLC2A3	Solute carrier family 2 (facilitated glucose transporter), member 3	Preferential glucose uptake in TICs [150], induced by HIF [151]
IGFBP5	Insulin-like growth factor binding protein 5	Cell survival, cell adhesion through activation of integrin-linked kinase (ILK) and Akt, and reduces migration [152]
RAB6B	Member of RAS oncogene family	Tumor proliferation, malignant transformation [153]
ESM1	Endothelial cell specific molecule-1	Tumor growth, and metastasis through NFκB pathway [31], angiogenesis [154]
RAD21	Double-strand-break repair protein rad21 homolog	Sister chromatid cohesion in mitotic cells [155], DNA repair [156], and estrogen-mediated regulation of MYC [36]
CDK16	Cyclin dependent protein kinases	Cell proliferation [157]
CDC25B	Cell division cycle 25B	Cell proliferation [158], overexpression of c-myc [37]
CENPA	Centromere protein A	Tumor progression, cell cycle regulation, survival [159], chromosomal instability [160]
PGK1	Phosphoglycerate kinase 1	Increase in the expression of CXCR4, angiogenic switch, tumor growt h [45], target of PPARγ (cell proliferation) [161], and induce bone metastasis [44]
MAD2L1	Mitotic arrest-deficient 2 L1	Cell proliferation [162], tumorigenesis [163]
CKS2	Cdc28 kinase subunit 2	Tumorigenesis, proliferation, apoptosis resistance [164, 165]
BUB1	Mitotic checkpoint serine/threonine kinase	Cell proliferation [166, 167]
Angiogenesis/m	igration/invasion	
FLT1	Vascular endothelial growth factor receptor	Angiogenesis and vasculogenesis [168, 169], formation of premetastatic niche and bone directed metastasis of breast cancer [43]
EXT1	Exostosin glycosyltransferase 1	Bone development [170], migration [171]
DIAPH3	Diaphanous-related formin 3	Motility/migration and the formation of filopodium in tumor [172-174]
CDC42BPA	Serine/Threonine protein kinase	Tumor cell invasion [175], migration [176]
EXOC7	Exocyst complex component 7	Angiogenesis [177], migration [178]
NMU	Neuromedin U	Tumorigenicity, metastasis [179], cell migration, invasiveness, tumor cell dissemination [180]
VEGF	Vascular endothelial growth factor	Angiogenesis [181, 182], metastasis [183], EMT [41]
MMP9	Matrix metalloproteinase	Induction of VEGF [184, 185], EMT, invasion and metastasis [39, 40]
FGF18	Fibroblast growth factor 18	Cell survival [186], growth, migration, invasion through NFκB pathway, angiogenesis [32, 187]
WISP1	WNT1 inducible signaling pathway protein 1	WNT1/ β -catenin responsive oncogene [188], survival [189], induction of the expression of VCAM1 [190], increase the expression of MMP2 and migration through FAK, MEK, ERK, p65 and NF- κ B pathways [33], involved the regulation of TGF- β 1 to control osteoblast function [191], metastasis to bone [192]
TGFB3	Transforming growth factor, beta 3	Induction of breast CSCs [30], involved in the induction of MMPs, EMT, and invasion through MAPK pathway [29], induced by HIF-1 [193], induced by Snail/Slug/β-catenin-TCF4 pathway [194], induction of apoptosis [195]

Gene symbol	Gene description	Functions and features
TFF1	Trefoil factor-1	Oncogenicity [196], cell survival, anchorage-independent growth [52], angiogenesis, migration, and invasion [51], protect the mucosa from insults, stabilize the mucus layer and affect healing of the epithelium [197]
TFF3	Trefoil factor-3	Tumor progression and dissemination [198], regulation of VEGF expression induced by hypoxia [55], protect the mucosa from insults, stabilize the mucus layer and affect healing of the epithelium [197]
AGR2	A protein disulfide isomerase; anterior gradient homolog 2	Tumor growth, cell migration, cellular transformation [199], cancer cell survival [200], metastasis [201], and production of intestinal mucus [202]
NATI	Arylamine N-acetyltransferase-1	Cell growth, anchorage independent growth, E-cadherin (cell-cell contact) inhibition, and invasion [203, 204]
CRIP1	Cysteine-rich intestinal protein-1	Cellular growth and differentiation [205]
RND1	Member of the Rho GTPase family	Disassembly of actin filament structures, and loss of cell adhesion [206]
TSPAN1	Tetraspanin 1	Cell growth, migration, and invasion [207, 208]
FGFR3	Fibroblast growth factor receptor 3	Cellular proliferation, survival, migration, angiogenesis [97]
SCUBE2	Signal peptide CUB (complement proteins C1r/C1 s, Uegf, and Bmp 1)-EGF domain-containing protein2	Growth inhibitory effects, antagonizing bone morphogenetic protein, and suppressing the β -catenin pathway (interacting with E-cadherin) [63, 209], enhance the Sonic Hedgehog signaling activity [210]
CEACAM6	Carcinoembryonic antigen-related cell adhesion molecule 6	Tumorigenesis, disruption of cell polarity, and anoikis resistance [211, 212]
TOM1L1	Target of Myb-1 Like	Enhancement of IL2-Jak2-STAT3 signaling pathway [213], and involvement of EGF signaling [214]
KRT16	Keratin 16	Migration and invasion [215]
FGFBP1	Fibroblast growth factor binding protein1	Angiogenesis, enhancement in FGF-1- and FGF-2-dependent proliferation, FGF-2-induced ERK activation, and migration [59, 216, 217]
FOXO3A	Forkhead box class O3A transcription factor	Suppression of tumor cells; Regulation the expression of p27/Kip1, cyclin D1, and cyclin E, induction of apoptosis through Bim and FasL [218–222], regulation of glycolysis downstream of Akt through transcriptional control of Tsc1 [223], and repression of ER- α [224], and estrogen-dependent proliferation [225], repression of VEGF [226], promotion of invasive migration through MMP9 and MMP-13 upregulation [62]
KRT6B	Keratin 6B	Migration and invasion [215]
SNA11	Snail transcription factor	Induction of EMT, invasion, and metastasis [57] by suppressing E-cadherin [227], and cell cycle, and induction of apoptosis resistance [228]
TMSB15A	Thymosin beta 15a	Tumor progression and metastasis [229], motility [230]

Table 3 Smid's signature of breast cancer bone metastasis genes

replication, proliferation, and survival [24–26]. Upregulation of these genes can be considered as the first requirements of primary tumor for its growth in order to be prepared for dissemination. Afterward, *ECT2*, *EGLN1*, *ESM1*, *FLT1*, *EXT1*, *DIAPH3*, *EXOC7*, *NMU*, *CDC42BPA*, *VEGF*, *MMP9*, *FGF18*, *WISP1*, and *TGF-β3*, are wellknown pivotal elements that participate in angiogenesis, migration, and invasion. Tumor cells which express these genes seem to be adept for invasion from primary site (please see Table 2 for detail functions of gene products).

A variety of key molecular mechanisms are regulated by *van't Veer's signature* gene products. Remarkably, TGF- β

signaling pathway likely plays important roles in the regulation of cell cycle, proliferation, induction of EMT, CSCs, and MMPs [27–30] (Table 2). Furthermore, NFkB signaling is also involved in this level of tumor progression taking part in tumor growth and metastasis by the mediation of *ESM1*, *FGF18*, and *WISP1* [31–33]. In this step, the transcription factor MYC, which is well known for its association to breast tumor proliferation [34, 35], may also play important roles as is shown to be regulated by at least two of *van't Veer's signature* genes including *RAD21* and *CDC25B* [36, 37]. Notably, *'epithelial to mesenchymal transition'* (EMT), a process that is shown to be essential

Gene symbol	Gene description	Functions and features
Angiogenesis	, migration/invasion, EMT	
МСАМ	Melanoma cell adhesion molecule	Migration, invasion, tumorigenicity [231], motility [232], angiogenesis [233, 234], EMT [232, 235], and metastasis [236]
PTK7	Protein tyrosine kinase 7	Cell cycle regulation, EMT [237], motility [238], regulation of WNT/planar cell polarity pathway [239], invasion, angiogenesis [240], and target of MMP14 [241]
RGCC	Regulator of cell cycle	Induction of EMT, migration, and invasion via NF κ B signaling [242], interact with Smad3 to promote TGF- β 1 mediated EMT [65, 69]
CTGF	Connective Tissue Growth factor	Induction of EMT-like cell fate [243], protection of the vasculature [244], hypoxia induced [245] angiogenesis [246], migration and invasion [247] through MMP-13 upregulation, and FAK, ERK, NF κ B pathways [79], mediation of Smad-dependent TGF β signaling to regulate mesenchymal cell proliferation [71], involve in bone metastasis [80, 84], and upregulated by EGF [248]
FGF5	Fibroblast growth factor-5	Autocrine and paracrine dependent cell growth, enhance the MAPK signaling [78], angiogenic and mitogenic factor [249, 250], survival and migration [251]
ADAMTS1	A disintegrin and metalloprotease with thrombospondin motifs protein 1	Tumor growth and metastasis [252], migration [253, 254], endothelial invasion [255], inhibition of angiogenesis [256, 257], induce bone metastasis [81]
CXCR4	Chemokine (C-X-C motif) receptor 4	Modulating the trafficking of both cancer and normal stem cells [258], induction of bone metastasis [82], survival, proliferation, angiogenesis [259], migration, invasion through WNT/β-catenin pathway [260], upregulation/activation by TGF- β1 [72, 73], HIF [261], and Akt [262], and BMP4 [263], induces EMT [264], induces and maintains stemness in cancer stem cells [265]
IL11	Interleukin-11	TGF-β dependent [15, 74, 75] bone metastasis [74, 83–85], motility [266], and invasion via PI3K, Ras, STAT3, MAPK, and JNK mediation [267–269], induce production of RANKL [3]
MMP1	Matrix metalloproteinase-1	Tumorigenesis [270], induce bone metastasis [81], invasion, release IGF, FGF and TGF-β [76], vascular remodeling [271], induced by WNT signaling [272], MAPK [273], and BMP4 [263], activates PAR1 [270], roles in tumor extravasation [274]
Growth/angio	genesis inhibition	
FHL1	Four and a half LIM protein 1	Interaction with oestrogen receptors (ERs), breast cancer cell growth regulation [275], interaction with Smads 2,3,4 and suppress tumor growth and migration [276], suppression of VEGF [277]
DUSP1	Dual specificity phosphatase 1	Target of TGF-β1, inhibition of p38/MAPK and JNK [68, 278, 279]. Target of p53 and triggers apoptosis [280, 281], and involved in the auto-regulation of VEGF [282]
SOCS2	Suppressor of cytokine signaling 2	Suppression of proliferation and growth [283], prolactin-induced mammary gland development [284], enhance IL-2 and IL-3 signaling [285]
FST	Follistatin	Stimulation by TGF-β2 [70] and BRCA1 [286], Inhibition of BMPs [287–289], inhibition of activin [290], increase mTOR signaling via Smad3 [77], downstream of WNT4 signaling [291], inhibition of angiogenesis [292]
Survival and	microenvironmental factors	
SLC4A7	Solute carrier family 4, sodium bicarbonate cotransporter, member 7	Major determinant of pH(i) in breast cancer primary tumor and metastasis [293]
NCF2	Neutrophil cytosolic factor 2	Target of p53 in activation of ROS and inhibition of apoptosis [294]

for tumor dissemination and metastasis of breast carcinomas [38], is enhanced through at least two of genes in this signature including *MMP9* and *VEGF*. These two factors play determining roles in preparing a hospitable microenvironment in which emitted signals trigger EMT in order to induce/maintain CSCs [39–42]. On the other hand, two of genes in this panel may tend to trigger primary tumor cells to metastasize to bone, including FLT1 and PGK1 [43, 44]. PGK1 increases the expression of CXCR4, which is one of the most important bone metastasis factors [45]. FLT1 also provide a premetastatic niche in bone and direct bone metastasis of BC [43]. Together, above information suggest the involvement of key regulators like TGF- β , NF κ B, and MYC, as well as the process of EMT in first steps of BC metastasis within the primary tumor.

Biological functions of genes in *van't Veer's signature* define the hallmarks of cancer. Tian and colleagues have recently shown that *van't Veer's signature* gene products

functionally meet all the six *hallmarks of cancer* defined by Hanahan and Weinberg, including sustained proliferation, anti-growth signaling evasion, cell death resistance, immortality, angiogenesis, and invasion/metastasis [46]. They identified interconnected networks and showed that these genes are regulated by key tumorigenic factors like TP53, RB1, MYC, JUN and CDKN2A [47]. Interestingly, *van't Veer's signature* may also reflect the two additional hallmarks of next generation, including reprogramming of energy metabolism and evading immune destruction [1].

Adjustments of energy metabolism in order to fuel cell growth and division, is of most important features that lead to uncontrolled proliferation in neoplasms. Glycolytic fueling has been shown to be one of the most essential mechanisms in the reprogramming of energy, and associated with activated oncogenes like RAS, MYC, mutant tumor suppressors like TP53, certain signaling pathways like PI3K/Akt/PTEN, and hypoxia inducible factor 1(HIF1) [48, 49]. Now, as Tian and colleagues showed, and also from functions of certain genes including TSPYL5, CMC2, CDC25B, EGLN1, SLC2A3, RAB6B, and TGF- β 3 (see details of functions in Table 2), van't Veer's signature likely associates with the seventh hallmark of cancer, reprogramming of energy metabolism. As for the eighth hallmark, evading immune destruction, it has been shown that tumors that produce transforming growth factor (TGF)- β escape from immune surveillance, mainly by selective and direct suppression of the T cell cytotoxic gene responses [50]. Intriguingly, TGF- β signaling is undeniably of key factors in van't Veer's signature. Together, it seems that functions of gene products of van't Veer's signature also meet the two additional next generation hallmarks of cancer, in addition to the first six ones.

Smid's signature

Focusing on bone metastasis, Smid and colleagues have established a panel of genes in BC patients that are implicated to bone relapse (*Smid's signature*). They analyzed primary tumors of lymph node negative BC patients who, subsequently, had developed metastases. A set of 69 genes was identified to be differentially expressed in patients who had experienced bone metastasis versus patients with metastasis to other sites. (Table 3 comprises a panel of selected genes from *Smid's signature* that are significantly overexpressed and have been better studied). Notably, they developed classifier of tumors that metastasis is to bone that was applicable in clinic [16].

Smid's signature provides a pattern of gene expression in primary tumors that obligates them to metastasize to bone. Genes in *Smid's signature* participate in essential features of metastasis including tumor growth, proliferation, survival, angiogenesis, migration, and invasion (Table 3). Among those, *TFF1*, *TFF3*, *AGR2*, *NAT1*, *CRIP1*, *TSPAN1*, *FGFR3*, *CEACAM6*, and *TMSB15A* may be categorized to play roles in cell growth/proliferation, and survival. Importantly, a number of genes in this signature take part in angiogenesis that include *TFF1*, *TFF3*, *FGFR3*, and *FGFBP1*. Thereupon, the vast majority of the genes in this panel which have seminal roles in migration and invasion are reported to be *TFF1*, *TFF3*, *AGR2*, *NAT1*, *RND1*, *TSPAN1*, *FGFR3*, *CEACAM6*, *KRT16*, *FGFBP1*, *FOXO3A*, *KRT6B*, and *SNAI1*. Interestingly, *TFF1*, *TFF3*, and *FGFR3* are present in all above categories, and seemingly play pivotal roles in the process of BC metastasis to bone.

Well-characterized crucial molecular mechanisms are controlled by Smid's signature gene products. Trefoil factor-1 (TFF1), that was the most differentially expressed gene associated to bone metastasis in Smid's signature, is ascertained to play roles in cell survival, anchorage-independent growth, angiogenesis, migration, and invasion in breast (and also other) tumor cells [51-54]. TFF3 induces angiogenesis by the regulation of VEGF, under the control of hypoxia [55]. AGR2, NAT1, FGFR3, and TSPAN1 have shown to play roles in tumor growth/proliferation/survival and/or migration/invasion. CRIP1, CEACAM6, and TMSB15A regulate tumor growth/proliferation and survival. RND1, KRT16, KRT6B, FOXO3A, and SNAI1 induce migration/invasion. It should be noted that, SNAI1 (also SNAIL) has determined to play pivotal roles as a master regulator of EMT [38, 56-58]. On the contrary, SCUBE2, and FOXO3A have generally shown to encode suppressors of tumor growth/proliferation, inducers of apoptosis, and repressor of angiogenesis. SCUBE2, and FOXO3A, although, are distinguished to play roles in line with induction of invasiveness, and metastasis, by regulation of Hedgehog signaling and matrix metalloproteinases, respectively [59–62] (Table 3).

Critical signaling pathways are associated with the genes in the Smid's signature. Apparently, fibroblast growth factor (FGF) signaling takes fundamental parts in this circuit, as FGFR3 and FGFBP1 that are directly linked to this pathway are two important overexpressed genes in this panel. ERK signaling is likely involved through activation by FGFBP1 [59]. In addition, epidermal growth factor (EGF) and Janus kinase-signal transducers and activators of transcription (JAK-STAT) signaling pathways are demonstrated to be enhanced by TOM1L1 [63]. Together, from the above mentioned findings, it seems that the genes in Smid's signature direct a complex program in which tumor cells proliferate and survive, then further induce and maintain angiogenesis, and finally enhance migration and invasion. Passing these two steps of gene expression (van't Veer's and Smid's signatures), tumor cells are capable to invade, intravasate, and survive in

circulation, heading to bone. These two sets of genes may be regulated in an overlapped, or in a sequential pattern.

Kang's signature

Focused on the gene expression pattern of breast tumor cells heading to bone, Kang and colleagues investigated a multigenic program in highly aggressive osteolytic BC metastatic cells (*Kang's signature*) (Table 4 comprises a panel of selected genes from *Kang's signature* that are significantly overexpressed and have been better studied) [15]. Their work provided a framework for the identification of genes mediating metastasis to different organs. Although *Kang's signature* was established in animal model, it has recently been confirmed in human BC patients as well [64]. These genes mostly encode secreted and cell membrane proteins, and are associated to the preparation of a compatible metastatic niche.

Kang's signature provides a panel of genes that may be categorized into four groups including: angiogenesis, migration/invasion, EMT, and growth/angiogenesis inhibition. Of those, MCAM, PTK7, CTGF, FGF5, and CXCR4 play critical roles in angiogenesis. MCAM, PTK7, RGCC, CTGF, FGF5, ADAMTS1, CXCR4, IL-11, and MMP1 are well-known essential factors for tumor migration/invasion. Notably, a number of key genes in this signature encode important inducers of EMT and CSC features including: MCAM, PTK7, RGCC, CTGF, and CXCR4. RGCC and Smad3 direct the induction of EMT through regulation of SNAIL and SLUG EMT transcription factors [65]. CXCR4 activates several signaling pathways, including AKT [66], a process in which Src plays a critical role [67]. It should be noted that, some genes in Kang's signature controversially function against tumor growth or angiogenesis (fourth group). Those include FHL1, DUSP1, SOCS2, FST, and ADAMTS1 (see below). SLC4A7 and NCF2, which are not categorized in these groups, play roles in preparing the microenvironment and inhibition of apoptosis, respectively (Table 4). From the above mentioned information, MCAM, PTK7, CTGF, and CXCR4 are categorized into all first three groups, and likely play critical roles in the last steps of BC bone metastasis.

Genes in *Kang's signature* function toward controlling principal signaling pathways, and govern a sophisticated signaling network that leads to a successful metastasis. Table 4 demonstrates undeniable deviation in the regulation of key signaling pathways, like TGF- β , WNT, NF κ B, FGF, and MAPK, as underpinning functions of genes associated to bone metastasis. Almost half of the genes in this panel, including DUSP1, RGCC, FST, CTGF, CXCR4, IL-11, and MMP-1, are directly linked to the TGF- β signaling [65, 68–76]. FST also increase mTOR signaling via Smad3 [77]. PTK7, FST, CXCR4, and MMP-1 take part in the regulation of WNT signaling, and remarkably, RGCC and CTGF function via NF κ B pathway. FGF5 mediate FGF signaling, and enhance MAPK pathway [78]. CTGF also act through ERK and FAK pathways [79]. Importantly, distinct gene expression pattern of *Kang's signature* specifically direct disseminated tumor cells to overt bone metastasis. CTGF, ADAMTS1, CXCR4, IL-11, and MMP1 are considered crucial inducers of bone metastasis [80–85]. Together, it seems that for a successful bone metastasis such sophisticated signaling network is required, which is controlled by the power of gene expression regulation.

Breast cancer bone metastasis; a multistep cascade of events

Several studies have reported genes that are associated with BC bone metastasis, from which some prognostic tools are provided in order to obtain best available treatments for individual BC patients. However, a comprehensive understanding of the nature of metastasis is yet to be investigated. Accordingly, due to the short knowledge of this complex phenomenon, well-suited therapies are lacking. In this review, we aimed to use published gene signatures, which have been shown to be significantly linked to progression and metastasis of BC to bone, to unmask a sequential pattern of gene expression that leads to colonization of breast tumors in bone.

Primary breast tumors cells ought to arrange a programmed gene expression pattern for their growth, survival, and invasion. Therefore, programs related to cell cycle progression, proliferation, apoptosis resistant, angiogenesis, invasion, and distant colonization need to be directed by nucleus. Genes in van't Veer's and Smid's signature seem to provide such well-coordinated program for tumor cells, as both signatures are derived from primary tumors of BC patients, who developed metastasis (in general) and bone metastasis, respectively [14, 16]. We thought that genes in these two signatures likely express in a time period that primary tumor: first get committed to metastasize, and second committed to metastasize to bone. Then, as is obvious in Tables 2 and 3, primary tumor first get ready for metastasis, and then get features of invasion, and intravasation. Tumor cells, then, acquire bone specific directing features of homing, extravasation, micro-colonization, and eventually macro-colonization (metastatic colonization) from the genes in *Kang's signature* (Table 4) (Fig. 1). This is likely because Kang's signature is primarily derived from metastatic breast tumor cells from bone lesions [15]. Importantly, key genes in Kang's signature including MMP-1, CXCR4, FGF5 and CTGF, which was initially determined from study on metastatic MDA-MB-231 cells and mouse model, have been recently



Fig. 1 Sequential gene expression pattern, from primary tumor to metastatic colonization. *van't Veer's*, *Smid's*, and *Kang's* gene signatures shows a sequential pattern of expression that leads to breast cancer progression, metastasis. Genes in *van't Veer's* signature are mostly involved in the regulation of cell cycle, DNA replication, Proliferation, tumorigenesis, and survival, therefor, are essential for primary tumor cells to be prepared for invasion and metastasis. On the other hand some genes in this signature act toward angiogenesis, migration, and invasion. *ESM1*, *ECT2*, and *EGLN1* are present in both categories and likely play important roles in primary steps of tumor metastasis. Genes in *Smid's* signature also play essential roles in cell growth, proliferation, survival, angiogenesis, migration, and invasion.

confirmed in patients with BC and prostate cancer [64]. Figure 2 also shows deviation toward particular functions in each signature. In *van't Veer's* signature almost two out of third number of genes (24 out of 35) are in the category of proliferation. In *Smid's* signature 13 out of 17 genes are in the category of migration. In *Kang's* signature 9 out of 16 genes are involved in functions of migration and invasion. This may show the evolutionarily pattern of tumor progression and metastasis in BC bone metastasis.

A set of well-defined genes govern key molecular pathways, and are the behind-the-scene forces of BC bone metastasis. Genes from *van't Veer's*, *Smid's*, and *Kang's* signatures control signaling pathways that have been considered as pivotal driving forces of tumor progression and metastasis. TGF- β , FGF, NF κ B, WNT, PI3K, and JAK-STAT signaling pathways are induced/enhanced by genes of *van't Veer's*, and *Smid's* signatures in primary breast tumor cells. On the other hand, TGF- β , FGF, NF κ B, WNT, and PI3K pathways are also induced/enhanced by genes of *Kang's signature* in bone colonized tumor cells. In both primary and distant tumors EMT program is induced by these pathways, and also several genes such as SNAI1 and MMP9 in primary tumor, and MCAM, PTK7, CXCR4, through circulation. *TFF1*, *TFF3*, and *FGFR3* are present in all three categories. Genes in *Kang's* signature are mostly associated to angiogenesis, migration/invasion, EMT, and CSC factors. Many genes in this signature have been directly linked to bone metastasis of breast cancer. *MCAM*, *PTK7*, and *CTGF* are common genes in three categories. Genes in *Kang's* signature likely act toward homing, extravasation, formation of micrometastasis, and eventually metastatic colonization, in order to end this journey. Abbreviations: *EMT* epithelial to mesenchymal transition, *CSC* cancer stem cells

Tumor cells that overexpress genes in these two signatures seem to be

able to invade and intravasate from primary site and disseminate

RGCC, and CTGF in distant metastatic tumors (Fig. 3) (Tables 2, 3, 4).

Smad-dependent and Smad-independent TGF-B signaling pathways are essential for EMT and BC metastases [86–88]. Smad3 and Smad4 dependent TGF- β signaling have been shown to be indispensable for the induction of EMT and metastasis [89-91]. Smad transcription factors orchestrate overexpression of several important genes involved in EMT and metastasis, including SNAIL, TWIST, and ZEB families of transcription factor coding genes [92]. TGF- β also participates in the activation of several key signaling pathways such as Ras/ERK, and PI3K/Akt, called Smad-independent pathways, and regulates cell growth, survival, cytoskeletal reorganization, migration, and invasion [93]. For instance, MMP9 is shown to be induced by TGF- β -induced Akt-dependent ERK pathway [94]. TGF- β stimulation leads epithelial cells to obtain mesenchymallike features, and capability of migration, invasion, and dissemination through circulation to distant sites of metastasis, and features of stemness [95].

FGF signaling also acts as an important inducer of EMT and metastasis [96]. FGFs regulate a wide range of biological functions such as proliferation, survival, and



Fig. 2 Genes in *van't Veer's*, *Smid's*, and *Kang's* signatures are each deviated toward distinct functions. Certain numbers of genes in each signature are involved in certain functions categorized into: proliferation (*blue*), survival (*green*), angiogenesis (*red*), migration (*yellow*), and invasion (*brown*). Several genes have common

functions, which are located in interconnected territories of circles. *Red* highlighted genes have functions against the corresponding feature (*circle*). Size of the *circles* shows the deviation toward that function in each signature. (Color figure online)

migration [97], and likely play pivotal roles in bone metastasis of BC [16]. NFkB is also an important inducer of EMT, and act through direct activation of SNAIL and ZEB family of transcription factors [98, 99]. Interestingly, it is believed that the cooperation of NF κ B and TGF- β signaling pathways is critical for EMT and cancer metastasis [100, 101]. Wnt signaling is among the most important pathways involved in the induction of EMT and breast CSCs [86, 102, 103]. Notably, WNT and TGF- β signaling pathways likely induce an mutually reinforcing autocrine signaling network that is indispensable for constant expression of EMT associated transcription factors and CSC niche [87, 104]. PI3K signaling pathway is wellknown for its important roles in the induction of EMT and metastasis [105]. Intriguingly, it has been shown that PI3K signaling is essential for autocrine/paracrine TGF- β associated motility, invasiveness, and metastasis [106]. JAK-STAT signaling pathway has an essential regulatory role in growth and proliferation of breast CSCs [107]. JAK-STAT signaling is associated with essential features such as survival, cell cycle regulation, self-sufficiency in growth and metastasis [108, 109]. Importantly, JAK2 also interacts and activates PI3K and RAS signaling molecules [110].

Hypoxia and hypoxia inducible factors (HIFs) seem to have pivotal roles in BC bone metastasis. HIF directly regulates several genes from van't Veer's, Smid's, and Kang's signatures. TFF3, EGLN1, SNAI1, MMP9, TGFB3, SLC2A3, and CTGF are of genes that are directly regulated by hypoxia. In fact, HIFs are of the essential preliminary factors that trigger gene expression programs that lead to tumor progression and metastasis, and play critical roles in the induction of EMT and stemness state in CSCs [111]. Hypoxia and HIFs are likely essential factors in the regulation of on and off states of EMT between primary and secondary tumors. In primary tumors, localized hypoxia mediates HIFs to be activated, and therefore move toward EMT/CSC induction and metastasis. At the secondary sites, however, with likely no hypoxic environment, lack of hypoxia and other factors lead to the reversion of EMT and CSC features. This phenomenon is essential for metastasis of differentiated carcinomas [87, 112] (for a comprehensive review see Ref. [112]).

It seems that a comprehensive signaling network consisting of TGF- β , FGF, NF κ B, WNT, PI3K, and JAK-STAT is indispensable for breast tumor cells to progress to overt bone metastasis. Essential links between key bone metastatic factors, such as *vascular cell adhesion molecule 1* (VCAM1), *receptor activator of nuclear factor* κ *B ligand* (RNAKL), *parathyroid-hormone related peptide* (PTHrP), and BACH1 with these pathways further confirms this



Fig. 3 Well-orchestrated genes govern key molecular pathways, and are the behind-the-scene forces of breast cancer bone metastasis. Genes in van't Veer's, Smid's, and Kang's signatures govern a comprehensive signaling network in primary and secondary breast tumors. TGF-B, FGF, JAK-STAT, NFKB, WNT, and PI3K pathways in primary tumor, and TGF-B, FGF, NFkB, and PI3K pathways in secondary tumor are regulated by genes in these three signatures. In the primary tumor the six signaling pathways build a comprehensive signaling network that lead toward tumor growth, proliferation, survival, angiogenesis, migration, and invasion. In primary tumor (upleft), genes from van't Veer's, Smid's signatures and their related signaling molecules are showed. HIF seem to have profound effects in primary tumor development and dissemination. Notably, several genes and signaling cascades induce EMT, and therefor CSC associated features. Intravasated tumor cells form the population of circulating tumor cells that disseminate, home, and extravasate into the secondary organ (bone). Importantly, the majority of differentiated circulating tumor cells (yellow) cannot survive the inhospitable

signaling network. VCAM1 Promotes bone metastasis by attracting and tethering osteoclast progenitors that express $\alpha 4$ integrin and facilitating their maturation [113, 114], and

environment while in circulation, and a small proportion of CSCs (red) are able to reach distant sites and form metastasis. In metastatic tumor (down-right), genes from Kang's signature lead to activation of the five pathways, which build a comprehensive signaling network that governs features like invasion, migration, EMT, and CSC formation. It is important to mention that, tumors in primary and secondary sites can be different or identical regarding their state of differentiation. In most cases secondary tumors are at the same level of differentiation as primary tumor, or even more differentiated. But in some cases, like triple negative breast cancer, both primary and secondary tumors are mostly mesenchymal, and secondary tumors are even more mesenchymal (not shown in this figure). Abbreviations: TGF-B: transforming growth factor-beta, FGF: fibroblast growth factor, JAK-STAT: Janus kinase/signal transducers and activators of transcription, NFkB: nuclear factor kappa B, WNT, and PI3K: phosphatidylinositol 3 kinase, HIF: hypoxia inducible factor, EMT: epithelial to mesenchymal transition, CSC: cancer stem cells. (Color figure online)

induces PI3K-Akt signaling by the mediation of Ezrin [115]. RANKL regulates bone resorption [116], migration [117], invasion [117, 118], bone metastasis [3, 117, 119,

120], and induces tumorigenesis, EMT, stemness [121], and the upregulation of MMP1 [118]. PTHrP can be induced by TGF- β [122], and activates CTGF through protein kinase A/C and ERK pathways [80]. BACH1 is a common regulator of several bone metastasis genes, including MMP1 and CXCR4 [123], induced by TGF- β [124].

Genes play the central role in development and diseases. Controlling the cellular pathways is one of the most critical duties of genes, in which reciprocal feedbacks play essential roles. Gene expression pattern in a given cell likely relates the story of a journey in which the cell is born, grow, proliferate, and/or die. Regulating the cellular behavior is the most critical tasks of gene expression machinery. Malignant behavior in cancers is tightly controlled their by gene expression pattern. Connection of gene signatures discussed in this review may provide novel insight toward better understanding the journey in which tumor cells get features of malignancy and metastasize to distant sites, and therefore providing best fit treatments for any individual cancer.

Conflict of interest The authors declare that there is no conflict of interest.

References

- 1. Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. Cell 144(5):646–674
- Siegel R, Naishadham D (2013) Jemal A (2013) Cancer statistics. CA Cancer J Clin 63(1):11–30
- 3. Mundy GR (2002) Metastasis to bone: causes, consequences and therapeutic opportunities. Nat Rev Cancer 2(8):584–593
- Roodman GD (2004) Mechanisms of bone metastasis. N Engl J Med 350(16):1655–1664
- Valastyan S, Weinberg RA (2011) Tumor metastasis: molecular insights and evolving paradigms. Cell 147(2):275–292
- Ma XJ et al (2003) Gene expression profiles of human breast cancer progression. Proc Natl Acad Sci USA 100(10):5974–5979
- Yu M et al (2013) Circulating breast tumor cells exhibit dynamic changes in epithelial and mesenchymal composition. Science 339(6119):580–584
- Chaffer CL, Weinberg RA (2011) A perspective on cancer cell metastasis. Science 331(6024):1559–1564
- 9. Shackleton M et al (2009) Heterogeneity in cancer: cancer stem cells versus clonal evolution. Cell 138(5):822–829
- Greaves M, Maley CC (2012) Clonal evolution in cancer. Nature 481(7381):306–313
- Brabletz T et al (2005) Opinion: migrating cancer stem cells an integrated concept of malignant tumour progression. Nat Rev Cancer 5(9):744–749
- Charafe-Jauffret E et al (2009) Breast cancer cell lines contain functional cancer stem cells with metastatic capacity and a distinct molecular signature. Cancer Res 69(4):1302–1313
- Jordan CT, Guzman ML, Noble M (2006) Cancer stem cells. N Engl J Med 355(12):1253–1261
- van 't Veer LJ et al (2002) Gene expression profiling predicts clinical outcome of breast cancer. Nature 415(6871):530–536

- 15. Kang Y et al (2003) A multigenic program mediating breast cancer metastasis to bone. Cancer Cell 3(6):537–549
- Smid M et al (2006) Genes associated with breast cancer metastatic to bone. J Clin Oncol Off J Am Soc Clin Oncol 24(15):2261–2267
- Glas AM et al (2006) Converting a breast cancer microarray signature into a high-throughput diagnostic test. BMC Genom 7:278
- van't Veer LJ, Bernards R (2008) Enabling personalized cancer medicine through analysis of gene-expression patterns. Nature 452(7187):564–570
- Perou CM et al (1999) Distinctive gene expression patterns in human mammary epithelial cells and breast cancers. Proc Natl Acad Sci USA 96(16):9212–9217
- Perou CM et al (2000) Molecular portraits of human breast tumours. Nature 406(6797):747–752
- Zajchowski DA et al (2001) Identification of gene expression profiles that predict the aggressive behavior of breast cancer cells. Cancer Res 61(13):5168–5178
- 22. Sorlie T et al (2001) Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proc Natl Acad Sci USA 98(19):10869–10874
- West M et al (2001) Predicting the clinical status of human breast cancer by using gene expression profiles. Proc Natl Acad Sci USA 98(20):11462–11467
- 24. Grana X, Reddy EP (1995) Cell cycle control in mammalian cells: role of cyclins, cyclin dependent kinases (CDKs), growth suppressor genes and cyclin-dependent kinase inhibitors (CKIs). Oncogene 11(2):211–219
- 25. Ohtani K et al (1999) Cell growth-regulated expression of mammalian MCM5 and MCM6 genes mediated by the transcription factor E2F. Oncogene 18(14):2299–2309
- Mehdipour P et al (2009) Prognostic implication of CDC25A and cyclin E expression on primary breast cancer patients. Cell Biol Int 33(10):1050–1056
- 27. Liu JH et al (1999) Functional association of TGF-beta receptor II with cyclin B. Oncogene 18(1):269–275
- Wottawa M et al (2013) Knockdown of prolyl-4-hydroxylase domain 2 inhibits tumor growth of human breast cancer MDA-MB-231 cells by affecting TGF-beta1 processing. Int J Cancer J Int Cancer 132(12):2787–2798
- Petrella BL, Armstrong DA, Vincenti MP (2012) Interleukin-1 beta and transforming growth factor-beta 3 cooperate to activate matrix metalloproteinase expression and invasiveness in A549 lung adenocarcinoma cells. Cancer Lett 325(2):220–226
- Malanchi I et al (2012) Interactions between cancer stem cells and their niche govern metastatic colonization. Nature 481(7379):85–89
- Kang YH et al (2012) ESM-1 regulates cell growth and metastatic process through activation of NF-kappaB in colorectal cancer. Cell Signal 24(10):1940–1949
- 32. Sonvilla G et al (2008) FGF18 in colorectal tumour cells: autocrine and paracrine effects. Carcinogenesis 29(1):15–24
- Hou CH et al (2011) WISP-1 increases MMP-2 expression and cell motility in human chondrosarcoma cells. Biochem Pharmacol 81(11):1286–1295
- Dubik D, Dembinski TC, Shiu RP (1987) Stimulation of c-myc oncogene expression associated with estrogen-induced proliferation of human breast cancer cells. Cancer Res 47 24(Pt 1): 6517-21
- 35. Watson PH, Pon RT, Shiu RP (1991) Inhibition of c-myc expression by phosphorothioate antisense oligonucleotide identifies a critical role for c-myc in the growth of human breast cancer. Cancer Res 51(15):3996–4000
- McEwan MV, Eccles MR, Horsfield JA (2012) Cohesin is required for activation of MYC by estradiol. PLoS ONE 7(11):e49160

- 37. Wu W et al (1998) Overexpression of cdc25A and cdc25B is frequent in primary non-small cell lung cancer but is not associated with overexpression of c-myc. Cancer Res 58(18):4082–4085
- Yang J, Weinberg RA (2008) Epithelial-mesenchymal transition: at the crossroads of development and tumor metastasis. Dev Cell 14(6):818–829
- Deryugina EI, Quigley JP (2006) Matrix metalloproteinases and tumor metastasis. Cancer Metastasis Rev 25(1):9–34
- Orlichenko LS, Radisky DC (2008) Matrix metalloproteinases stimulate epithelial-mesenchymal transition during tumor development. Clin Exp Metastasis 25(6):593–600
- 41. Yang AD et al (2006) Vascular endothelial growth factor receptor-1 activation mediates epithelial to mesenchymal transition in human pancreatic carcinoma cells. Cancer Res 66(1):46–51
- Mani SA et al (2008) The epithelial-mesenchymal transition generates cells with properties of stem cells. Cell 133(4):704–715
- Kaplan RN et al (2005) VEGFR1-positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. Nature 438(7069):820–827
- 44. Jung Y et al (2009) Expression of PGK1 by prostate cancer cells induces bone formation. Mol Cancer Res MCR 7(10):1595–1604
- 45. Wang J et al (2007) A glycolytic mechanism regulating an angiogenic switch in prostate cancer. Cancer Res 67(1):149–159
- Hanahan D, Weinberg RA (2000) The hallmarks of cancer. Cell 100(1):57–70
- 47. Tian S et al (2010) Biological functions of the genes in the mammaprint breast cancer profile reflect the hallmarks of cancer. Biomark Insights 5:129–138
- DeBerardinis RJ et al (2008) The biology of cancer: metabolic reprogramming fuels cell growth and proliferation. Cell Metab 7(1):11–20
- Jones RG, Thompson CB (2009) Tumor suppressors and cell metabolism: a recipe for cancer growth. Genes Dev 23(5):537–548
- Thomas DA, Massague J (2005) TGF-beta directly targets cytotoxic T cell functions during tumor evasion of immune surveillance. Cancer Cell 8(5):369–380
- Buache E et al (2011) Deficiency in trefoil factor 1 (TFF1) increases tumorigenicity of human breast cancer cells and mammary tumor development in TFF1-knockout mice. Oncogene 30(29):3261–3273
- 52. Emami S et al (2001) Induction of scattering and cellular invasion by trefoil peptides in src- and RhoA-transformed kidney and colonic epithelial cells. FASEB J: official publication of the Federation of American Societies for Experimental Biology 15(2):351–361
- 53. Prest SJ, May FE, Westley BR (2002) The estrogen-regulated protein, TFF1, stimulates migration of human breast cancer cells. FASEB J: official publication of the Federation of American Societies for Experimental Biology 16(6):592–594
- 54. Rodrigues S et al (2003) Trefoil peptides as proangiogenic factors in vivo and in vitro: implication of cyclooxygenase-2 and EGF receptor signaling. FASEB J: official publication of the Federation of American Societies for Experimental Biology 17(1):7–16
- 55. Guleng B et al (2012) TFF3 mediated induction of VEGF via hypoxia in human gastric cancer SGC-7901 cells. Mol Biol Rep 39(4):4127–4134
- 56. Blanco MJ et al (2002) Correlation of Snail expression with histological grade and lymph node status in breast carcinomas. Oncogene 21(20):3241–3246
- Nieto MA (2002) The snail superfamily of zinc-finger transcription factors. Nat Rev Mol Cell Biol 3(3):155–166
- De Craene B, Berx G (2013) Regulatory networks defining EMT during cancer initiation and progression. Nat Rev Cancer 13(2):97–110

- 59. Liu R et al (2009) KLF5 promotes breast cell survival partially through fibroblast growth factor-binding protein 1-pERK-mediated dual specificity MKP-1 protein phosphorylation and stabilization. J Biol Chem 284(25):16791–16798
- 60. Harris LG et al (2012) Increased vascularity and spontaneous metastasis of breast cancer by hedgehog signaling mediated upregulation of cyr61. Oncogene 31(28):3370–3380
- Harris LG, Samant RS, Shevde LA (2011) Hedgehog signaling: networking to nurture a promalignant tumor microenvironment. Mol Cancer Res MCR 9(9):1165–1174
- 62. Storz P et al (2009) FOXO3a promotes tumor cell invasion through the induction of matrix metalloproteinases. Mol Cell Biol 29(18):4906–4917
- Lin YC et al (2011) Domain and functional analysis of a novel breast tumor suppressor protein, SCUBE2. J Biol Chem 286(30):27039–27047
- 64. Casimiro S et al (2012) Analysis of a bone metastasis gene expression signature in patients with bone metastasis from solid tumors. Clin Exp Metastasis 29(2):155–164
- 65. Guo X, Jose PA, Chen SY (2011) Response gene to complement 32 interacts with Smad3 to promote epithelial-mesenchymal transition of human renal tubular cells. Am J Physiol Cell Physiol 300(6):C1415–C1421
- 66. Epstein RJ (2004) The CXCL12-CXCR4 chemotactic pathway as a target of adjuvant breast cancer therapies. Nat Rev Cancer 4(11):901–909
- 67. Zhang XH et al (2009) Latent bone metastasis in breast cancer tied to Src-dependent survival signals. Cancer Cell 16(1):67–78
- 68. Mikami F et al (2006) The transforming growth factor-beta-Smad3/4 signaling pathway acts as a positive regulator for TLR2 induction by bacteria via a dual mechanism involving functional cooperation with NF-kappaB and MAPK phosphatase 1-dependent negative cross-talk with p38 MAPK. J Biol Chem 281(31):22397–22408
- 69. Huang WY et al (2009) RGC-32 mediates transforming growth factor-beta-induced epithelial-mesenchymal transition in human renal proximal tubular cells. J Biol Chem 284(14):9426–9432
- 70. Fitzgerald AM et al (2012) The effects of transforming growth factor-beta2 on the expression of follistatin and activin A in normal and glaucomatous human trabecular meshwork cells and tissues. Invest Ophthalmol Vis Sci 53(11):7358–7369
- Parada C, et al (2013) CTGF mediates Smad-dependent TGFbeta signaling to regulate mesenchymal cell proliferation during palate development. Mol Cell Biol 33(17):3482–3493
- 72. Chu CY, et al (2013) Induction of chemokine receptor CXCR4 expression by transforming growth factor-beta1 in human basal cell carcinoma cells. J Dermatol Sci 72(2):123–133
- Bertran E, et al (2013) Overactivation of the TGF-beta pathway confers a mesenchymal-like phenotype and CXCR4-dependent migratory properties to liver tumor cells. Hepatology 58(6):2032–2044
- 74. Gupta J et al (2011) TGFbeta-dependent induction of interleukin-11 and interleukin-8 involves SMAD and p38 MAPK pathways in breast tumor models with varied bone metastases potential. Cancer Biol Ther 11(3):311–316
- 75. Calon A et al (2012) Dependency of colorectal cancer on a TGFbeta-driven program in stromal cells for metastasis initiation. Cancer Cell 22(5):571–584
- Vu TH, Werb Z (2000) Matrix metalloproteinases: effectors of development and normal physiology. Genes Dev 14(17):2123–2133
- 77. Winbanks CE et al (2012) Follistatin-mediated skeletal muscle hypertrophy is regulated by Smad3 and mTOR independently of myostatin. J Cell Biol 197(7):997–1008
- 78. Kornmann M et al (1997) Fibroblast growth factor-5 stimulates mitogenic signaling and is overexpressed in human pancreatic cancer: evidence for autocrine and paracrine actions. Oncogene 15(12):1417–1424

- 79. Tan TW et al (2009) CTGF enhances migration and MMP-13 up-regulation via alphavbeta3 integrin, FAK, ERK, and NFkappaB-dependent pathway in human chondrosarcoma cells. J Cell Biochem 107(2):345–356
- Shimo T et al (2006) Pathogenic role of connective tissue growth factor (CTGF/CCN2) in osteolytic metastasis of breast cancer. J Bone Miner Res Off J Am Soc Bone Miner Res 21(7):1045–1059
- Lu X et al (2009) ADAMTS1 and MMP1 proteolytically engage EGF-like ligands in an osteolytic signaling cascade for bone metastasis. Genes Dev 23(16):1882–1894
- Muller A et al (2001) Involvement of chemokine receptors in breast cancer metastasis. Nature 410(6824):50–56
- McCoy EM et al (2013) IL-11 produced by breast cancer cells augments osteoclastogenesis by sustaining the pool of osteoclast progenitor cells. BMC Cancer 13:16
- 84. Gao YB et al (2013) Enhanced production of CTGF and IL-11 from highly metastatic hepatoma cells under hypoxic conditions: an implication of hepatocellular carcinoma metastasis to bone. J Cancer Res Clin Oncol 139(4):669–679
- Ren L et al (2013) Bone metastasis from breast cancer involves elevated IL-11 expression and the gp130/STAT3 pathway. Med Oncol 30(3):634
- 86. Jing Y et al (2011) Epithelial-mesenchymal transition in tumor microenvironment. Cell Biosci 1:29
- 87. Fazilaty H et al (2013) Crosstalk between breast cancer stem cells and metastatic niche: emerging molecular metastasis pathway? Tumour Biol J Int Soc Oncodev Biol Med 34(4):2019–2030
- Kalluri R, Weinberg RA (2009) The basics of epithelial-mesenchymal transition. J Clin Investig 119(6):1420–1428
- 89. Deckers M et al (2006) The tumor suppressor Smad4 is required for transforming growth factor beta-induced epithelial to mesenchymal transition and bone metastasis of breast cancer cells. Cancer Res 66(4):2202–2209
- 90. Kaimori A et al (2007) Transforming growth factor-betal induces an epithelial-to-mesenchymal transition state in mouse hepatocytes in vitro. J Biol Chem 282(30):22089–22101
- Ashcroft GS et al (1999) Mice lacking Smad3 show accelerated wound healing and an impaired local inflammatory response. Nat Cell Biol 1(5):260–266
- Xu J, Lamouille S, Derynck R (2009) TGF-beta-induced epithelial to mesenchymal transition. Cell Res 19(2):156–172
- Derynck R, Zhang YE (2003) Smad-dependent and Smadindependent pathways in TGF-beta family signalling. Nature 425(6958):577–584
- 94. Byun HJ et al (2006) A splice variant of CD99 increases motility and MMP-9 expression of human breast cancer cells through the AKT-, ERK-, and JNK-dependent AP-1 activation signaling pathways. J Biol Chem 281(46):34833–34847
- 95. Dang H et al (2011) Snail1 induces epithelial-to-mesenchymal transition and tumor initiating stem cell characteristics. BMC Cancer 11:396
- 96. Ciruna B, Rossant J (2001) FGF signaling regulates mesoderm cell fate specification and morphogenetic movement at the primitive streak. Dev Cell 1(1):37–49
- Turner N, Grose R (2010) Fibroblast growth factor signalling: from development to cancer. Nat Rev Cancer 10(2):116–129
- 98. Min C et al (2008) NF-kappaB and epithelial to mesenchymal transition of cancer. J Cell Biochem 104(3):733–744
- 99. Julien S et al (2007) Activation of NF-kappaB by Akt upregulates Snail expression and induces epithelium mesenchyme transition. Oncogene 26(53):7445–7456
- 100. Huber MA et al (2004) NF-kappaB is essential for epithelialmesenchymal transition and metastasis in a model of breast cancer progression. J Clin Investig 114(4):569–581

- 101. Maier HJ et al (2010) NF-kappaB promotes epithelial-mesenchymal transition, migration and invasion of pancreatic carcinoma cells. Cancer Lett 295(2):214–228
- 102. Velasco-Velazquez MA et al (2012) Breast cancer stem cells. Int J Biochem Cell Biol 44(4):573–577
- Malanchi I et al (2008) Cutaneous cancer stem cell maintenance is dependent on beta-catenin signalling. Nature 452(7187):650–653
- 104. Scheel C et al (2011) Paracrine and autocrine signals induce and maintain mesenchymal and stem cell states in the breast. Cell 145(6):926–940
- 105. Larue L, Bellacosa A (2005) Epithelial-mesenchymal transition in development and cancer: role of phosphatidylinositol 3' kinase/AKT pathways. Oncogene 24(50):7443–7454
- 106. Muraoka-Cook RS, Dumont N, Arteaga CL (2005) Dual role of transforming growth factor beta in mammary tumorigenesis and metastatic progression. Clin Cancer Res Off J Am Assoc Cancer Res 11(2 Pt 2):937s–943s
- 107. Marotta LL et al (2011) The JAK2/STAT3 signaling pathway is required for growth of CD44(+)CD24(-) stem cell-like breast cancer cells in human tumors. J Clin Investig 121(7):2723–2735
- Niu G et al (2002) Roles of activated Src and Stat3 signaling in melanoma tumor cell growth. Oncogene 21(46):7001–7010
- 109. Bowman T et al (2000) STATs in oncogenesis. Oncogene 19(21):2474–2488
- 110. Quintas-Cardama A, Verstovsek S (2013) Molecular pathways: Jak/STAT pathway: mutations, inhibitors, and resistance. Clin Cancer Res Off J Am Assoc Cancer Res 19(8):1933–1940
- 111. Lu X, Kang Y (2010) Hypoxia and hypoxia-inducible factors: master regulators of metastasis. Clin Cancer Res Off J Am Assoc Cancer Res 16(24):5928–5935
- 112. Brabletz T (2012) To differentiate or not-routes towards metastasis. Nat Rev Cancer 12(6):425–436
- 113. Lu X et al (2011) VCAM-1 promotes osteolytic expansion of indolent bone micrometastasis of breast cancer by engaging alpha4beta1-positive osteoclast progenitors. Cancer Cell 20(6):701–714
- 114. Chen Q, Massague J (2012) Molecular pathways: VCAM-1 as a potential therapeutic target in metastasis. Clin Cancer Res Off J Am Assoc Cancer Res 18(20):5520–5525
- 115. Chen Q, Zhang XH, Massague J (2011) Macrophage binding to receptor VCAM-1 transmits survival signals in breast cancer cells that invade the lungs. Cancer Cell 20(4):538–549
- 116. Boyce BF, Xing L (2007) Biology of RANK, RANKL, and osteoprotegerin. Arthr Res Ther 9(Suppl 1):S1
- 117. Jones DH et al (2006) Regulation of cancer cell migration and bone metastasis by RANKL. Nature 440(7084):692–696
- 118. Casimiro S et al (2013) RANKL/RANK/MMP-1 molecular triad contributes to the metastatic phenotype of breast and prostate cancer cells in vitro. PLoS ONE 8(5):e63153
- 119. Park HR et al (2003) Expression of osteoprotegerin and RANK ligand in breast cancer bone metastasis. J Korean Med Sci 18(4):541–546
- 120. Peng X et al (2013) Differential expression of the RANKL/ RANK/OPG system is associated with bone metastasis in human non-small cell lung cancer. PLoS ONE 8(3):e58361
- 121. Palafox M et al (2012) RANK induces epithelial-mesenchymal transition and stemness in human mammary epithelial cells and promotes tumorigenesis and metastasis. Cancer Res 72(11):2879–2888
- 122. Yin JJ et al (1999) TGF-beta signaling blockade inhibits PTHrP secretion by breast cancer cells and bone metastases development. J Clin Investig 103(2):197–206
- 123. Liang Y et al (2012) Transcriptional network analysis identifies BACH1 as a master regulator of breast cancer bone metastasis. J Biol Chem 287(40):33533–33544

- 124. Okita Y et al (2013) Transforming growth factor-beta induces transcription factors MafK and Bach1 to suppress expression of the heme oxygenase-1 gene. J Biol Chem 288(28):20658–20667
- 125. Shubbar E et al (2013) Elevated cyclin B2 expression in invasive breast carcinoma is associated with unfavorable clinical outcome. BMC Cancer 13:1
- 126. Caldon CE et al (2012) Cyclin E2 overexpression is associated with endocrine resistance but not insensitivity to CDK2 inhibition in human breast cancer cells. Mol Cancer Ther 11(7):1488–1499
- 127. Payton M et al (2002) Deregulation of cyclin E2 expression and associated kinase activity in primary breast tumors. Oncogene 21(55):8529–8534
- 128. Epping MT et al (2011) TSPYL5 suppresses p53 levels and function by physical interaction with USP7. Nat Cell Biol 13(1):102–108
- 129. Kim EJ et al (2010) TSPYL5 is involved in cell growth and the resistance to radiation in A549 cells via the regulation of p21(WAF1/Cip1) and PTEN/AKT pathway. Biochem Biophys Res Commun 392(3):448–453
- Gulzar ZG, McKenney JK, Brooks JD (2013) Increased expression of NuSAP in recurrent prostate cancer is mediated by E2F1. Oncogene 32(1):70–77
- 131. Chou HY et al (2011) Phosphorylation of NuSAP by Cdk1 regulates its interaction with microtubules in mitosis. Cell Cycle 10(23):4083–4089
- Raemaekers T et al (2003) NuSAP, a novel microtubule-associated protein involved in mitotic spindle organization. J Cell Biol 162(6):1017–1029
- 133. Horn D et al (2010) The conserved mitochondrial twin Cx9C protein Cmc2 Is a Cmc1 homologue essential for cytochrome c oxidase biogenesis. J Biol Chem 285(20):15088–15099
- 134. Xu J et al (2013) MiR-223/Ect2/p21 signaling regulates osteosarcoma cell cycle progression and proliferation. Biomed Pharmacother 67(5):381–386
- 135. Weeks A et al (2012) ECT2 and RASAL2 mediate mesenchymal-amoeboid transition in human astrocytoma cells. Am J Pathol 181(2):662–674
- 136. Cook DR et al (2011) The ect2 rho Guanine nucleotide exchange factor is essential for early mouse development and normal cell cytokinesis and migration. Genes Cancer 2(10):932–942
- 137. Thomae AW et al (2011) Different roles of the human Orc6 protein in the replication initiation process. Cell Mol Life Sci CMLS 68(22):3741–3756
- 138. Ueki T et al (2008) Involvement of elevated expression of multiple cell-cycle regulator, DTL/RAMP (denticleless/RAregulated nuclear matrix associated protein), in the growth of breast cancer cells. Oncogene 27(43):5672–5683
- 139. Pan HW et al (2006) Role of L2DTL, cell cycle-regulated nuclear and centrosome protein, in aggressive hepatocellular carcinoma. Cell Cycle 5(22):2676–2687
- 140. Liu CL et al (2007) L2dtl is essential for cell survival and nuclear division in early mouse embryonic development. J Biol Chem 282(2):1109–1118
- 141. Mollinari C et al (2002) PRC1 is a microtubule binding and bundling protein essential to maintain the mitotic spindle midzone. J Cell Biol 157(7):1175–1186
- 142. Joshi K et al (2013) MELK-dependent FOXM1 phosphorylation is essential for proliferation of glioma stem cells. Stem Cells 31(6):1051–1063
- 143. Gu C et al (2013) Tumor-specific activation of the C-JUN/ MELK pathway regulates glioma stem cell growth in a p53dependent manner. Stem Cells 31(5):870–881
- 144. Lin ML et al (2007) Involvement of maternal embryonic leucine zipper kinase (MELK) in mammary carcinogenesis through

is associated 146. Peurala E et al (2012) Expressions of individual PHDs associate with good prognostic factors and increased proliferation in

breast cancer patients. Breast Cancer Res Treat 133(1):179–188 147. Metzen E et al (2005) Regulation of the prolyl hydroxylase domain protein 2 (phd2/egln-1) gene: identification of a functional hypoxia-responsive element. Biochem J 387(Pt 3):711–717

interaction with Bcl-G, a pro-apoptotic member of the Bcl-2

is a key regulator of the proliferation of malignant brain tumors,

including brain tumor stem cells. J Neurosci Res 86(1):48-60

145. Nakano I et al (2008) Maternal embryonic leucine zipper kinase

family. Breast Cancer Res BCR 9(1):R17

- 148. Chan DA et al (2009) Tumor vasculature is regulated by PHD2mediated angiogenesis and bone marrow-derived cell recruitment. Cancer Cell 15(6):527–538
- 149. Mak P et al (2013) Estrogen receptor beta sustains epithelial differentiation by regulating prolyl hydroxylase 2 transcription. Proc Natl Acad Sci USA 110(12):4708–4713
- 150. Flavahan WA et al (2013) Brain tumor initiating cells adapt to restricted nutrition through preferential glucose uptake. Nat Neurosci 16(10):1373–1382
- 151. Mimura I et al (2012) Dynamic change of chromatin conformation in response to hypoxia enhances the expression of GLUT3 (SLC2A3) by cooperative interaction of hypoxia-inducible factor 1 and KDM3A. Mol Cell Biol 32(15):3018–3032
- 152. Sureshbabu A et al (2012) IGFBP5 induces cell adhesion, increases cell survival and inhibits cell migration in MCF-7 human breast cancer cells. J Cell Sci 125(Pt 7):1693–1705
- 153. Clark GJ, Der CJ (1995) Aberrant function of the Ras signal transduction pathway in human breast cancer. Breast Cancer Res Treat 35(1):133–144
- 154. Aitkenhead M et al (2002) Identification of endothelial cell genes expressed in an in vitro model of angiogenesis: induction of ESM-1, (beta)ig-h3, and NrCAM. Microvasc Res 63(2):159–171
- 155. Sonoda E et al (2001) Scc1/Rad21/Mcd1 is required for sister chromatid cohesion and kinetochore function in vertebrate cells. Dev Cell 1(6):759–770
- 156. Birkenbihl RP, Subramani S (1992) Cloning and characterization of rad21 an essential gene of Schizosaccharomyces pombe involved in DNA double-strand-break repair. Nucleic Acids Res 20(24):6605–6611
- 157. Wu G et al (2003) DeltaNp63alpha and TAp63alpha regulate transcription of genes with distinct biological functions in cancer and development. Cancer Res 63(10):2351–2357
- 158. Lammer C et al (1998) The cdc25B phosphatase is essential for the G2/M phase transition in human cells. J Cell Sci 111(Pt 16):2445–2453
- 159. Li Y et al (2011) ShRNA-targeted centromere protein A inhibits hepatocellular carcinoma growth. PLoS ONE 6(3):e17794
- 160. Tomonaga T et al (2003) Overexpression and mistargeting of centromere protein-A in human primary colorectal cancer. Cancer Res 63(13):3511–3516
- 161. Shashni B et al (2013) Glycolytic enzymes PGK1 and PKM2 as novel transcriptional targets of PPARgamma in breast cancer pathophysiology. J Drug Target 21(2):161–174
- 162. Fang G, Yu H, Kirschner MW (1998) The checkpoint protein MAD2 and the mitotic regulator CDC20 form a ternary complex with the anaphase-promoting complex to control anaphase initiation. Genes Dev 12(12):1871–1883
- 163. Sotillo R et al (2007) Mad2 overexpression promotes aneuploidy and tumorigenesis in mice. Cancer Cell 11(1):9–23
- 164. Lan Y et al (2008) Aberrant expression of Cks1 and Cks2 contributes to prostate tumorigenesis by promoting proliferation and inhibiting programmed cell death. Int J Cancer J Int Cancer 123(3):543–551

- 165. Kang MA et al (2009) Upregulation of the cycline kinase subunit CKS2 increases cell proliferation rate in gastric cancer. J Cancer Res Clin Oncol 135(6):761–769
- 166. Johnson VL et al (2004) Bub1 is required for kinetochore localization of BubR1, Cenp-E, Cenp-F and Mad2, and chromosome congression. J Cell Sci 117(Pt 8):1577–1589
- 167. Grabsch H et al (2003) Overexpression of the mitotic checkpoint genes BUB1, BUBR1, and BUB3 in gastric cancer: association with tumour cell proliferation. J Pathol 200(1):16–22
- 168. Waltenberger J et al (1994) Different signal transduction properties of KDR and Flt1, two receptors for vascular endothelial growth factor. J Biol Chem 269(43):26988–26995
- 169. Yoshiji H et al (1996) Expression of vascular endothelial growth factor, its receptor, and other angiogenic factors in human breast cancer. Cancer Res 56(9):2013–2016
- 170. Huegel J et al (2013) Perichondrium phenotype and border function are regulated by Ext1 and heparan sulfate in developing long bones: a mechanism likely deranged in Hereditary Multiple Exostoses. Dev Biol 377(1):100–112
- 171. Wang Y et al (2013) Involvement of Ext1 and heparanase in migration of mouse FBJ osteosarcoma cells. Mol Cell Biochem 373(1–2):63–72
- 172. Hager MH et al (2012) DIAPH3 governs the cellular transition to the amoeboid tumour phenotype. EMBO Mol Med 4(8):743–760
- 173. Gupton SL et al (2007) mDia2 regulates actin and focal adhesion dynamics and organization in the lamella for efficient epithelial cell migration. J Cell Sci 120(Pt 19):3475–3487
- 174. Block J et al (2008) Filopodia formation induced by active mDia2/Drf3. J Microsc 231(3):506–517
- 175. Wilkinson S, Paterson HF, Marshall CJ (2005) Cdc42-MRCK and Rho-ROCK signalling cooperate in myosin phosphorylation and cell invasion. Nat Cell Biol 7(3):255–261
- 176. Balasenthil S et al (2011) A migration signature and plasma biomarker panel for pancreatic adenocarcinoma. Cancer Prev Res (Phila) 4(1):137–149
- 177. Barkefors I et al (2011) Exocyst complex component 3-like 2 (EXOC3L2) associates with the exocyst complex and mediates directional migration of endothelial cells. J Biol Chem 286(27):24189–24199
- 178. Liu J et al (2012) Exo70 stimulates the Arp2/3 complex for lamellipodia formation and directional cell migration. Curr Biol CB 22(16):1510–1515
- 179. Wu Y et al (2007) Neuromedin U is regulated by the metastasis suppressor RhoGDI2 and is a novel promoter of tumor formation, lung metastasis and cancer cachexia. Oncogene 26(5):765–773
- 180. Ketterer K et al (2009) Neuromedin U is overexpressed in pancreatic cancer and increases invasiveness via the hepatocyte growth factor c-Met pathway. Cancer Lett 277(1):72–81
- Ferrara N, Gerber HP, LeCouter J (2003) The biology of VEGF and its receptors. Nat Med 9(6):669–676
- Gerhardt H et al (2003) VEGF guides angiogenic sprouting utilizing endothelial tip cell filopodia. J Cell Biol 161(6):1163–1177
- 183. Skobe M et al (2001) Induction of tumor lymphangiogenesis by VEGF-C promotes breast cancer metastasis. Nat Med 7(2):192–198
- 184. Hiratsuka S et al (2002) MMP9 induction by vascular endothelial growth factor receptor-1 is involved in lung-specific metastasis. Cancer Cell 2(4):289–300
- 185. Belotti D et al (2003) Matrix metalloproteinases (MMP9 and MMP2) induce the release of vascular endothelial growth factor (VEGF) by ovarian carcinoma cells: implications for ascites formation. Cancer Res 63(17):5224–5229
- 186. Gaughhofer C et al (2011) Up-regulation of the fibroblast growth factor 8 subfamily in human hepatocellular carcinoma for cell survival and neoangiogenesis. Hepatology 53(3):854–864

- 187. Wei W et al (2013) FGF18 as a prognostic and therapeutic biomarker in ovarian cancer. J Clin Investig 123(10):4435–4448
- 188. Xu L et al (2000) WISP-1 is a Wnt-1- and beta-cateninresponsive oncogene. Genes Dev 14(5):585-595
- 189. Su F et al (2002) WISP-1 attenuates p53-mediated apoptosis in response to DNA damage through activation of the Akt kinase. Genes Dev 16(1):46–57
- 190. Liu JF et al (2013) CCN4 induces vascular cell adhesion molecule-1 expression in human synovial fibroblasts and promotes monocyte adhesion. Biochim Biophys Acta 1833(5):966–975
- 191. Inkson CA et al (2008) TGF-beta1 and WISP-1/CCN-4 can regulate each other's activity to cooperatively control osteoblast function. J Cell Biochem 104(5):1865–1878
- 192. Ono M et al (2013) WISP1/CCN4: a potential target for inhibiting prostate cancer growth and spread to bone. PLoS ONE 8(8):e71709
- 193. Nishi H et al (2004) Hypoxia-inducible factor-1 transactivates transforming growth factor-beta3 in trophoblast. Endocrinology 145(9):4113–4118
- 194. Medici D, Hay ED, Olsen BR (2008) Snail and Slug promote epithelial-mesenchymal transition through beta-catenin-T-cell factor-4-dependent expression of transforming growth factorbeta3. Mol Biol Cell 19(11):4875–4887
- 195. Nguyen AV, Pollard JW (2000) Transforming growth factor beta3 induces cell death during the first stage of mammary gland involution. Development 127(14):3107–3118
- Amiry N et al (2009) Trefoil factor-1 (TFF1) enhances oncogenicity of mammary carcinoma cells. Endocrinology 150(10):4473–4483
- 197. Katoh M (2003) Trefoil factors and human gastric cancer (review). Int J Mol Med 12(1):3–9
- 198. Ahmed AR et al (2012) TFF3 is a normal breast epithelial protein and is associated with differentiated phenotype in early breast cancer but predisposes to invasion and metastasis in advanced disease. Am J Pathol 180(3):904–916
- 199. Wang Z, Hao Y, Lowe AW (2008) The adenocarcinoma-associated antigen, AGR2, promotes tumor growth, cell migration, and cellular transformation. Cancer Res 68(2):492–497
- 200. Hrstka R et al (2010) The pro-metastatic protein anterior gradient-2 predicts poor prognosis in tamoxifen-treated breast cancers. Oncogene 29(34):4838–4847
- 201. Innes HE et al (2006) Significance of the metastasis-inducing protein AGR2 for outcome in hormonally treated breast cancer patients. Br J Cancer 94(7):1057–1065
- 202. Park SW et al (2009) The protein disulfide isomerase AGR2 is essential for production of intestinal mucus. Proc Natl Acad Sci USA 106(17):6950–6955
- 203. Tiang JM, Butcher NJ, Minchin RF (2010) Small molecule inhibition of arylamine N-acetyltransferase Type I inhibits proliferation and invasiveness of MDA-MB-231 breast cancer cells. Biochem Biophys Res Commun 393(1):95–100
- 204. Tiang JM et al (2011) RNAi-mediated knock-down of arylamine N-acetyltransferase-1 expression induces E-cadherin up-regulation and cell-cell contact growth inhibition. PLoS ONE 6(2):e17031
- 205. Lanningham-Foster L et al (2002) Overexpression of CRIP in transgenic mice alters cytokine patterns and the immune response. Am J Physiol Endocrinol Metab 282(6):E1197–E1203
- 206. Nobes CD et al (1998) A new member of the Rho family, Rnd1, promotes disassembly of actin filament structures and loss of cell adhesion. J Cell Biol 141(1):187–197
- 207. Wang GL et al (2012) The effect of NET-1 on the proliferation, migration and endocytosis of the SMMC-7721 HCC cell line. Oncol Rep 27(6):1944–1952
- 208. Chen L et al (2010) Suppression of TSPAN1 by RNA interference inhibits proliferation and invasion of colon cancer cells in vitro. Tumori 96(5):744–750

- 209. Cheng CJ et al (2009) SCUBE2 suppresses breast tumor cell proliferation and confers a favorable prognosis in invasive breast cancer. Cancer Res 69(8):3634–3641
- 210. Tsai MT et al (2009) Isolation and characterization of a secreted, cell-surface glycoprotein SCUBE2 from humans. Biochem J 422(1):119–128
- 211. Ilantzis C et al (2002) Deregulated expression of the human tumor marker CEA and CEA family member CEACAM6 disrupts tissue architecture and blocks colonocyte differentiation. Neoplasia 4(2):151–163
- 212. Duxbury MS et al (2004) CEACAM6 gene silencing impairs anoikis resistance and in vivo metastatic ability of pancreatic adenocarcinoma cells. Oncogene 23(2):465–473
- 213. Elmarghani A, Abuabaid H, Kjellen P (2009) TOM1L is involved in a novel signaling pathway important for the IL-2 production in Jurkat T cells stimulated by CD3/CD28 co-ligation. Mediators Inflamm 2009:416298
- 214. Liu NS et al (2009) Participation of Tom1L1 in EGF-stimulated endocytosis of EGF receptor. The EMBO journal 28(22):3485–3499
- 215. Hendrix MJ et al (1996) Role of intermediate filaments in migration, invasion and metastasis. Cancer Metastasis Rev 15(4):507–525
- 216. Tassi E et al (2001) Enhancement of fibroblast growth factor (FGF) activity by an FGF-binding protein. J Biol Chem 276(43):40247–40253
- 217. Abuharbeid S, Czubayko F, Aigner A (2006) The fibroblast growth factor-binding protein FGF-BP. Int J Biochem Cell Biol 38(9):1463–1468
- Medema RH et al (2000) AFX-like Forkhead transcription factors mediate cell-cycle regulation by Ras and PKB through p27kip1. Nature 404(6779):782–787
- 219. Rena G et al (1999) Phosphorylation of the transcription factor forkhead family member FKHR by protein kinase B. J Biol Chem 274(24):17179–17183
- 220. Brunet A et al (1999) Akt promotes cell survival by phosphorylating and inhibiting a Forkhead transcription factor. Cell 96(6):857–868
- 221. Hu MC et al (2004) IkappaB kinase promotes tumorigenesis through inhibition of forkhead FOXO3a. Cell 117(2):225–237
- 222. Yang JY et al (2008) ERK promotes tumorigenesis by inhibiting FOXO3a via MDM2-mediated degradation. Nat Cell Biol 10(2):138–148
- 223. Khatri S et al (2010) FOXO3a regulates glycolysis via transcriptional control of tumor suppressor TSC1. J Biol Chem 285(21):15960–15965
- 224. Morelli C et al (2010) Akt2 inhibition enables the forkhead transcription factor FoxO3a to have a repressive role in estrogen receptor alpha transcriptional activity in breast cancer cells. Mol Cell Biol 30(3):857–870
- 225. Zou Y et al (2008) Forkhead box transcription factor FOXO3a suppresses estrogen-dependent breast cancer cell proliferation and tumorigenesis. Breast Cancer Res BCR 10(1):R21
- 226. Karadedou CT et al (2012) FOXO3a represses VEGF expression through FOXM1-dependent and -independent mechanisms in breast cancer. Oncogene 31(14):1845–1858
- 227. Cano A et al (2000) The transcription factor snail controls epithelial-mesenchymal transitions by repressing E-cadherin expression. Nat Cell Biol 2(2):76–83
- 228. Vega S et al (2004) Snail blocks the cell cycle and confers resistance to cell death. Genes Dev 18(10):1131–1143
- 229. Gu YM et al (2008) Elevated thymosin beta15 expression is associated with progression and metastasis of non-small cell lung cancer. APMIS Acta Pathol Microbiol Immunol Scand 116(6):484–490

- 230. Bao L et al (1996) Thymosin beta 15: a novel regulator of tumor cell motility upregulated in metastatic prostate cancer. Nat Med 2(12):1322–1328
- 231. Zeng G et al (2012) METCAM/MUC18 augments migration, invasion, and tumorigenicity of human breast cancer SK-BR-3 cells. Gene 492(1):229–238
- 232. Zabouo G et al (2009) CD146 expression is associated with a poor prognosis in human breast tumors and with enhanced motility in breast cancer cell lines. Breast Cancer Res BCR 11(1):R1
- 233. Jiang T et al (2012) CD146 is a coreceptor for VEGFR-2 in tumor angiogenesis. Blood 120(11):2330–2339
- 234. Stalin J et al (2013) Soluble melanoma cell adhesion molecule (sMCAM/sCD146) promotes angiogenic effects on endothelial progenitor cells through angiomotin. J Biol Chem 288(13):8991–9000
- 235. Imbert AM et al (2012) CD146 expression in human breast cancer cell lines induces phenotypic and functional changes observed in Epithelial to Mesenchymal Transition. PLoS ONE 7(8):e43752
- 236. Zhang X, et al. (2013) MCAM expression is associated with poor prognosis in non-small cell lung cancer. Clin Transl Oncol. Official publication of the Federation of Spanish Oncology Societies and of the National Cancer Institute of Mexico
- 237. Chan DN et al (2012) PTK7 marks the first human developmental EMT in vitro. PLoS ONE 7(11):e50432
- 238. Yen WW et al (2009) PTK7 is essential for polarized cell motility and convergent extension during mouse gastrulation. Development 136(12):2039–2048
- 239. Lu X et al (2004) PTK7/CCK-4 is a novel regulator of planar cell polarity in vertebrates. Nature 430(6995):93–98
- 240. Shin WS et al (2008) Soluble PTK7 inhibits tube formation, migration, and invasion of endothelial cells and angiogenesis. Biochem Biophys Res Commun 371(4):793–798
- 241. Golubkov VS et al (2010) The Wnt/planar cell polarity proteintyrosine kinase-7 (PTK7) is a highly efficient proteolytic target of membrane type-1 matrix metalloproteinase: implications in cancer and embryogenesis. J Biol Chem 285(46):35740–35749
- 242. Sun Q, et al. (2013) Overexpression of response gene to complement 32 (RGC32) promotes cell invasion and induces epithelial-mesenchymal transition in lung cancer cells via the NFkappaB signaling pathway. Tumour Biol J Int Soc Oncodev Biol Med
- 243. Sonnylal S et al (2013) Connective tissue growth factor causes EMT-like cell fate changes in vivo and in vitro. J Cell Sci 126(Pt 10):2164–2175
- 244. Muratoglu SC, et al. (2013) LRP1 protects the vasculature by regulating levels of connective tissue growth factor and HtrA1. Arterioscler Thromb Vasc Biol
- 245. Kondo S et al (2002) Connective tissue growth factor increased by hypoxia may initiate angiogenesis in collaboration with matrix metalloproteinases. Carcinogenesis 23(5):769–776
- 246. Lau LF, Lam SC (1999) The CCN family of angiogenic regulators: the integrin connection. Exp Cell Res 248(1):44–57
- 247. Chen PS et al (2007) CTGF enhances the motility of breast cancer cells via an integrin-alphavbeta3-ERK1/2-dependent S100A4-upregulated pathway. J Cell Sci 120(Pt 12):2053–2065
- 248. Hugo HJ et al (2009) Staurosporine augments EGF-mediated EMT in PMC42-LA cells through actin depolymerisation, focal contact size reduction and Snail1 induction: a model for crossmodulation. BMC Cancer 9:235
- 249. Basilico C, Moscatelli D (1992) The FGF family of growth factors and oncogenes. Adv Cancer Res 59:115–165
- 250. Giordano FJ et al (1996) Intracoronary gene transfer of fibroblast growth factor-5 increases blood flow and contractile

function in an ischemic region of the heart. Nat Med 2(5):534–539

- 251. Allerstorfer S et al (2008) FGF5 as an oncogenic factor in human glioblastoma multiforme: autocrine and paracrine activities. Oncogene 27(30):4180–4190
- 252. Ricciardelli C et al (2011) The ADAMTS1 protease gene is required for mammary tumor growth and metastasis. Am J Pathol 179(6):3075–3085
- 253. Krampert M et al (2005) ADAMTS1 proteinase is up-regulated in wounded skin and regulates migration of fibroblasts and endothelial cells. J Biol Chem 280(25):23844–23852
- 254. Esselens C et al (2010) The cleavage of semaphorin 3C induced by ADAMTS1 promotes cell migration. J Biol Chem 285(4):2463–2473
- 255. Su SC et al (2008) Molecular profile of endothelial invasion of three-dimensional collagen matrices: insights into angiogenic sprout induction in wound healing. Am J Physiol Cell Physiol 295(5):C1215–C1229
- 256. Luque A, Carpizo DR, Iruela-Arispe ML (2003) ADAMTS1/ METH1 inhibits endothelial cell proliferation by direct binding and sequestration of VEGF165. J Biol Chem 278(26):23656–23665
- 257. Vazquez F et al (1999) METH-1, a human ortholog of ADAMTS-1, and METH-2 are members of a new family of proteins with angioinhibitory activity. J Biol Chem 274(33):23349–23357
- 258. Kucia M et al (2005) Trafficking of normal stem cells and metastasis of cancer stem cells involve similar mechanisms: pivotal role of the SDF-1-CXCR4 axis. Stem Cells 23(7):879–894
- 259. Teicher BA, Fricker SP (2010) CXCL12 (SDF-1)/CXCR4 pathway in cancer. Clin Cancer Res Off J Am Assoc Cancer Res 16(11):2927–2931
- 260. Wang Z et al (2008) Blockade of SDF-1/CXCR4 signalling inhibits pancreatic cancer progression in vitro via inactivation of canonical Wnt pathway. Br J Cancer 99(10):1695–1703
- 261. Mimeault M, Batra SK (2013) Hypoxia-inducing factors as master regulators of stemness properties and altered metabolism of cancer- and metastasis-initiating cells. J Cell Mol Med 17(1):30–54
- 262. Conley-Lacomb MK et al (2013) PTEN loss mediated Akt activation promotes prostate tumor growth and metastasis via CXCL12/CXCR4 signaling. Mol Cancer 12(1):85
- 263. Guo D, Huang J, Gong J (2012) Bone morphogenetic protein 4 (BMP4) is required for migration and invasion of breast cancer. Mol Cell Biochem 363(1–2):179–190
- 264. Onoue T et al (2006) Epithelial-mesenchymal transition induced by the stromal cell-derived factor-1/CXCR4 system in oral squamous cell carcinoma cells. Int J Oncol 29(5):1133–1138
- 265. Jung MJ et al (2013) Upregulation of CXCR4 is functionally crucial for maintenance of stemness in drug-resistant non-small cell lung cancer cells. Oncogene 32(2):209–221
- 266. Li TM et al (2012) Interleukin-11 increases cell motility and upregulates intercellular adhesion molecule-1 expression in human chondrosarcoma cells. J Cell Biochem 113(11):3353–3362
- 267. Yoshizaki A et al (2006) Expression of interleukin (IL)-11 and IL-11 receptor in human colorectal adenocarcinoma: IL-11 upregulation of the invasive and proliferative activity of human colorectal carcinoma cells. Int J Oncol 29(4):869–876
- 268. Nakayama T et al (2007) Expression of interleukin-11 (IL-11) and IL-11 receptor alpha in human gastric carcinoma and IL-11 upregulates the invasive activity of human gastric carcinoma cells. Int J Oncol 30(4):825–833
- 269. Shin SY et al (2012) Transcriptional regulation of the interleukin-11 gene by oncogenic Ras. Carcinogenesis 33(12):2467–2476
- 270. Foley CJ et al (2012) Matrix metalloprotease-1a promotes tumorigenesis and metastasis. J Biol Chem 287(29):24330–24338
- 271. Gupta GP et al (2007) Mediators of vascular remodelling coopted for sequential steps in lung metastasis. Nature 446(7137):765–770

- 272. Masckauchan TN et al (2006) Wnt5a signaling induces proliferation and survival of endothelial cells in vitro and expression of MMP-1 and Tie-2. Mol Biol Cell 17(12):5163–5172
- 273. Reunanen N et al (2002) Activation of p38 alpha MAPK enhances collagenase-1 (matrix metalloproteinase (MMP)-1) and stromelysin-1 (MMP-3) expression by mRNA stabilization. J Biol Chem 277(35):32360–32368
- 274. Kim MY et al (2009) Tumor self-seeding by circulating cancer cells. Cell 139(7):1315–1326
- 275. Lin J et al (2009) Four and a half LIM domains 1 (FHL1) and receptor interacting protein of 140 kDa (RIP140) interact and cooperate in estrogen signaling. Int J Biochem Cell Biol 41(7):1613–1618
- 276. Ding L et al (2009) Human four-and-a-half LIM family members suppress tumor cell growth through a TGF-beta-like signaling pathway. J Clin Investig 119(2):349–361
- 277. Lin J et al (2012) FHL family members suppress vascular endothelial growth factor expression through blockade of dimerization of HIF1alpha and HIF1beta. IUBMB Life 64(11):921–930
- 278. Tong XK, Hamel E (2007) Transforming growth factor-beta 1 impairs endothelin-1-mediated contraction of brain vessels by inducing mitogen-activated protein (MAP) kinase phosphatase-1 and inhibiting p38 MAP kinase. Mol Pharmacol 72(6):1476–1483
- 279. Owens DM, Keyse SM (2007) Differential regulation of MAP kinase signalling by dual-specificity protein phosphatases. Oncogene 26(22):3203–3213
- 280. Li M et al (2003) The phosphatase MKP1 is a transcriptional target of p53 involved in cell cycle regulation. J Biol Chem 278(42):41059–41068
- 281. Liu YX et al (2008) DUSP1 is controlled by p53 during the cellular response to oxidative stress. Mol Cancer Res MCR 6(4):624–633
- 282. Bellou S et al (2009) VEGF autoregulates its proliferative and migratory ERK1/2 and p38 cascades by enhancing the expression of DUSP1 and DUSP5 phosphatases in endothelial cells. Am J Physiol Cell Physiol 297(6):C1477–C1489
- 283. Farabegoli F et al (2005) Suppressor of cytokine signalling 2 (SOCS-2) expression in breast carcinoma. J Clin Pathol 58(10):1046–1050
- Harris J et al (2006) Socs2 and elf5 mediate prolactin-induced mammary gland development. Mol Endocrinol 20(5):1177–1187
- 285. Tannahill GM et al (2005) SOCS2 can enhance interleukin-2 (IL-2) and IL-3 signaling by accelerating SOCS3 degradation. Mol Cell Biol 25(20):9115–9126
- 286. Karve TM et al (2012) BRCA1 regulates follistatin function in ovarian cancer and human ovarian surface epithelial cells. PLoS ONE 7(6):e37697
- 287. Wordinger RJ et al (2002) Expression of bone morphogenetic proteins (BMP), BMP receptors, and BMP associated proteins in human trabecular meshwork and optic nerve head cells and tissues. Mol Vis 8:241–250
- 288. Abe Y et al (2004) Follistatin restricts bone morphogenetic protein (BMP)-2 action on the differentiation of osteoblasts in fetal rat mandibular cells. J Bone Miner Res Off J Am Soc Bone Miner Res 19(8):1302–1307
- 289. Fainsod A et al (1997) The dorsalizing and neural inducing gene follistatin is an antagonist of BMP-4. Mech Dev 63(1):39–50
- 290. Shimonaka M et al (1991) Follistatin binds to both activin and inhibin through the common subunit. Endocrinology 128(6):3313–3315
- 291. Yao HH et al (2004) Follistatin operates downstream of Wnt4 in mammalian ovary organogenesis. Dev Dyn Off Pub Am Assoc Anat 230(2):210–215
- 292. Ogino H et al (2008) Follistatin suppresses the production of experimental multiple-organ metastasis by small cell lung

cancer cells in natural killer cell-depleted SCID mice. Clin Cancer Res Off J Am Assoc Cancer Res $14(3){:}660{-}667$

293. Boedtkjer E et al (2013) Contribution of Na+, HCO3(-)cotransport to cellular pH control in human breast cancer: a role for the breast cancer susceptibility locus NBCn1 (SLC4A7). Int J Cancer J Int Cancer 132(6):1288–1299

294. Italiano D et al (2012) Identification of NCF2/p67phox as a novel p53 target gene. Cell Cycle 11(24):4589–4596