REVIEW



Molecular panorama of therapy resistance in prostate cancer: a pre-clinical and bioinformatics analysis for clinical translation

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Abstract

Prostate cancer (PCa) is a malignant disorder of prostate gland being asymptomatic in early stages and high metastatic potential in advanced stages. The chemotherapy and surgical resection have provided favourable prognosis of PCa patients, but advanced and aggressive forms of PCa including CRPC and AVPC lack response to therapy properly, and therefore, prognosis of patients is deteriorated. At the advanced stages, PCa cells do not respond to chemotherapy and radiotherapy in a satisfactory level, and therefore, therapy resistance is emerged. Molecular profile analysis of PCa cells reveals the apoptosis suppression, pro-survival autophagy induction, and EMT induction as factors in escalating malignant of cancer cells and development of therapy resistance. The dysregulation in molecular profile of PCa including upregulation of STAT3 and PI3K/Akt, downregulation of STAT3, and aberrant expression of non-coding RNAs are determining factor for response of cancer cells to chemotherapy. Because of prevalence of drug resistance in PCa, combination therapy including co-utilization of anti-cancer drugs and nanotherapeutic approaches has been suggested in PCa therapy. As a result of increase in DNA damage repair, PCa cells induce radioresistance and RelB overexpression prevents irradiation-mediated cell death. Similar to chemotherapy, nanomaterials are promising for promoting radiosensitivity through delivery of cargo, improving accumulation in PCa cells, and targeting survival-related pathways. In respect to emergence of immunotherapy as a new tool in PCa suppression, tumour cells are able to increase PD-L1 expression and inactivate NK cells in mediating immune evasion. The bioinformatics analysis for evaluation of drug resistance-related genes has been performed.

Keywords Prostate cancer \cdot Chemoresistance \cdot Radioresistance \cdot Immune evasion \cdot Bioinformatics

Highlights

- The drug resistance development in prostate cancer (PCa) has resulted in therapy failure in patients.
- Abnormal biological mechanisms including apoptosis, autophagy, and EMT mediate chemoresistance.
- Dysregulation of molecular pathways induces both chemoresistance and radioresistance in PCa.
- Pharmacological compounds and nanostructures increase therapy sensitivity.
- Immune evasion compromises function of immunotherapy in PCa suppression.
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1 Introduction

A significant health concern in the world results from development of prostate cancer (PCa) in males that in 2020, this malignancy caused approximately 400,000 deaths in patients that is in warning stage [1]. Due to malignant transformation of cells present in the prostate gland, the development

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of PCa occurs, and when disease is at first and localized stage, the tumour cells only proliferate in the tissue, while conditions are different at advanced stages with metastasis into surrounding tissue and disseminate in the body [2]. The prognosis of PCa appears to be promising and desirable at early stages with intervention of chemotherapy and surgical resection [3]. However, the prognosis is deteriorated; when PCa cells begin their spread in the body and after dissemination, the ability of aforementioned therapeutic approaches is diminished significantly. Among various factors, the survival and proliferation of malignant PCa cells depend on androgen signalling, and therefore, androgen-deprivation therapy (ADT) has been rationally developed for therapy of PCa [4]. In spite of bringing hope in the treatment of cancer patients, after PCa therapy with ADT for 2-3 years, castration-resistant prostate cancer (CRPC) is emerged that compromises the efficacy of ADT and results in resistance to this strategy [4, 5]. The CRPC is defined as a condition in which androgen signalling is stimulated in spite of androgen deprivation [6]. However, CRPC is not the end of PCa progression, and after abnormal progression of CRPC, it is developed into a new form that is known as aggressive variant prostate cancer (AVPC) that its most prominent characteristic is high metastatic potential [7]. The AVPC is also known as neuroendocrine prostate cancer (NEPC), since neuroendocrine markers display high expression in this subtype [7]. The treatment of AVPC is more difficult than CRPC, since CRPC still requires androgen receptor (AR) signalling for malignant progression, while alternative survival and proliferative pathways are stimulated in AVPC, and therefore, current ADT-based interventions are not applicable for AVPC, and therefore, it is completely lethal with average survival of less than 1 year [8]. Figure 1 depicts the stages of PCa with short description of PCa pathogenesis.

One of the prominent feature of PCa cells is the chromatin remodelling and the dysregulation of related pathways. Along with progression of PCa stem cells, the MUC1-C plays a significant role in process of tumourigenesis. MUC1-C increases E2F1 expression to upregulate PBAF, BRG1, and esBAF. Then, overexpression of ARID1A and PBRM1 occurs to mediate progression of CSCs. Therefore, the stimulation of chromatin remodelling complex PBAF by MUC1-C can accelerate tumourigenesis [11]. Furthermore, the chromatin remodelling pathways can determine the immune reactions in PCa. The upregulation of MUC1-C can promote BAF, PBAF, and NuRD levels. FBXW7 is suppressed by NuRD to upregulate IFNGR1, while BAF directly increases IFNGR1 expression. Then, upregulation of STAT1 occurs to promote IRF1 levels, mediating immunosuppression [12]. The proteins related to the chromatin remodelling can also enhance the progression and metastasis of PCa. BRG1 as a chromatin remodelling protein has ability of ELOVL3 transactivation in enhancing the invasion and migration of PCa [13]. Interestingly, overexpression of CHD6 in PCa can increase the detachment of nucleosomes from the promoters to stimulate the tumour-promoting factors in PCa [14]. The changes in expression levels of chromatin remodelling ATPase BRG1 and PTEN can affect tumourigenesis and clinical outcome. The upregulation of BRG1 and downregulation of PTEN can cause the poor prognosis in PCa [15].

The accumulation of mutations in PCa progression from early stages that symptoms are not specific to advanced and metastatic stages with clinically detectable symptoms results in changes in biological behaviour of tumour cells. The genomic background of PCa appears to be unstable [16], and most commonly studied germline mutations are BRCA1 and BRCA2 genes as tumour-suppressor factor [17–20]. However, molecular profile of PCa is not ended to dysregulation of BRCA1 and BRCA2, thanks to advances in field of biology that has made it possible to highlight the molecular profile of PCa. An integrative analysis has revealed that changes in levels of ASCL1, FOXA2, NKX2-2, POU3F2, and SOX2 can occur in NEPC [21]. More importantly, FOXA1 and Ku70/Ku80 are considered as potential therapeutic targets of ivermectin through integrated analysis for treatment of PCa [22]. Such alterations in genomic landscape of PCa can result in heterogeneous nature and plasticity of tumour cells [23]. More importantly, AR mutations and changes can be observed in 75% of metastatic CRPC patients [24], confirming its function as potential carcinogenic factor. The multiomics analysis of NSUN2 highlights its upregulation in PCa and its function in unfavourable prognosis of patients [25]. Even the immune system function in PCa and the infiltration of immune cells in the tumour microenvironment (TME) are changed by the molecular landscape [26, 27]. However, the story is beyond simple function of genes in control of PCa progression and epigenetic factors [28] and epigenetic silencing of genes [29] play a vital role of in tumourigenesis process. It is widely accepted that increase in malignancy of tumour cells mediated by genetic and epigenetic factors can result in chemoresistance. Therefore, current review aims to provide a detailed discussion on the function of molecular networks in progression-related chemoresistance in PCa, the interaction of signalling pathways, and the molecular mechanisms regulating drug resistance. This molecular landscape is evaluated for determining response of PCa cells to chemotherapy, radiotherapy, and immunotherapy. Moreover, a bioinformatics analysis is provided to shed more light on the genes involved in therapy resistance in PCa. The current therapeutic approaches including application of combination therapy and nanostructures for impairing tumourigenesis in PCa are discussed.

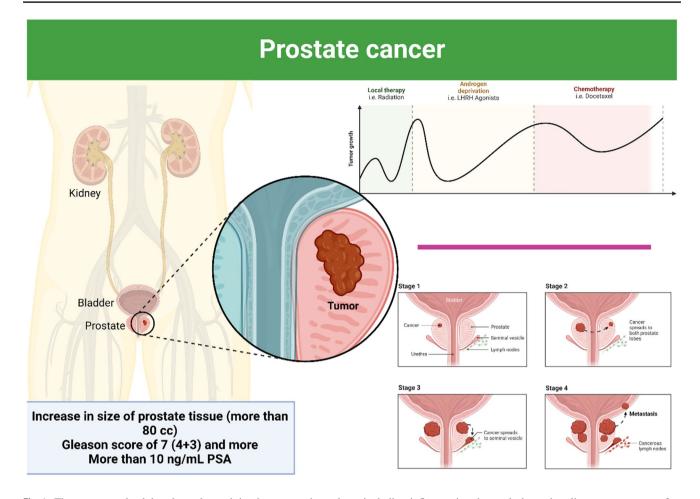


Fig. 1 The prostate gland has been located in the retroperitoneal space of body, and this male-specific gland responds to hormones. The prostate gland has a physical connection with urethra and bladder tissue, and in terms of anatomical view, it is divided into four categories and zones including peripheral zone, central zone, transitional zone, and periurethral zone. In view of histological analysis, prostate gland has a basal layer of epithelial cells that are covered by columnar secretory cells and fibromuscular stroma separates the cells. The growth and viability of cells present in the prostate tissue are controlled by androgens, and prostate atrophy occurs as a result of androgen removal. Three pathological events can occur in prostate tissue

2 Search strategy

The data and studies for the current review were selected from the major databases including PubMed, Google Scholar, and ScienceDirect using the keywords including "prostate cancer," "cancer drug resistance," "prostate cancer + drug resistance," "prostate cancer + radioresistance," "immune evasion + prostate cancer," "hormone resistance + prostate cancer," "pharmacological intervention," and "nanoparticles + prostate cancer." Moreover, the bioinformatics analysis was performed by the GEO database and using the RStudio software.

including inflammation, hyperplasia, and malignant tumour transformation [9]. After the development of PCa, a number of characteristics are found including increase in size of prostate tissue to more than 80 cc, enhanced level of PSA to more than 10 ng/mL, and Gleason score of 7 (4+3) [10]. Radiation is used as a local therapy, and then, it is followed by application of hormone therapy and chemotherapy with docetaxel. Four stages for metastasis of PCa are considered that in stage I, the tumour is localized, and in stage II, cancer cells disseminate in both lobes of tumour tissue. Then, spread of cancer cells to seminal vesicles in stage III occurs and followed by dissemination to surrounding tissues in stage IV

3 The mechanisms of drug resistance: a biological standpoint

3.1 Cancer mass and growth factors

The curability of a certain cancer mainly depends on its size [30, 31], and the size of tumour is used to determine the prognosis of patients. The risk of metastasis and invasion to surrounding tissues could be escalated with enhancement in size of tumours [32]. Although the correlation between tumour size, curability, and chemotherapy response has not been fully predicted, it is quite obvious that application of

various classes of anti-cancer drugs that individually suppress distinct pathways and cells in the TME can result in full eradication of cancer that has been approved by early mathematical methods including "log kill" hypothesis [33]. This has been shown to be correct for a number of cancers including lymphomas and germ cell cancers, but it is not valid for all cancer types. For better understanding, other kinds of models and hypotheses including Goldie-Coldman hypothesis were published that was informed by seminal microbiology experiments [34, 35] and it is responsible for understanding the relationship between tumour burden and therapy resistance risk. This hypothesis brings into attention that tumour size and mutation rates are two critical factors in determining resistance [34]. Therefore, if a same mutation rate is considered, tumour size can determine resistance to chemotherapy. As a result, this notion has been developed that if therapy regimens with non-cross-resistant probabilities are used, the chance of resistance is lower than when all the treatment strategies are utilized at once. However, such hypothesis has not been applied in clinic, showing that there are still complications and complexities to be revealed [36].

3.2 Heterogeneous nature of cancers

The easiest understanding of the chemoresistance in human cancers is role of tumour heterogeneity [37]. Throughout their progression, genomic changes accumulate in cancer cells that form a spatial and temporal genetic diversity and promote cancer heterogeneity [38]. The evolutionary speed of such heterogeneity is unique, and it can be slow such as mutations related to age, and it can be frequent occurring by APOBEC enzymes. Moreover, such changes can be rapid and quick, similar to what happens by genomic instability [39], chromothripsis [40], and chromosomal instability [41].

3.3 Physical barriers

The distinct feature of TME of cancers is hypoxia that is defined by low oxygenation. The spatial gradients are generated in the tumours that can prevent the normal blood flow in TME, and therefore, a hypoxic condition with tumourigenesis function is created, and moreover, less anti-cancer drugs are reached to tumour site. It may be a question that anti-angiogenic strategies have been applied in cancer therapy and reducing blood flow to cancer site is a therapeutic approach. However, this is not the complete story and there are evidences showing that anti-angiogenic therapies may result in normalization of vascular structure and function [42]. Therefore, chemotherapy drugs and targeted therapy are delivered to tumour site in an appropriate way [43]. That is why tyrosine kinase inhibitors with anti-angiogenic function have been used along with anti-PD-1/PD-L1 therapies in synergistic tumour suppression [44, 45]. Moreover, blood-brain barrier (BBB) is considered as another impediment towards effective cancer chemotherapy that by reducing chemical exposure of brain tumours, the risk of resistance enhances, urging to develop targeted systems for crossing over BBB [46].

3.4 Drug efflux transporters

The drug efflux transporters presence on the surface of cancer cells are connected to the phenomenon of chemoresistance that enhance efflux of chemotherapy drugs from cancer cells. ATP-binding cassette (ABC) transporters have been localized across cell membrane with different structures and have been related to cancer drug resistance [47, 48]. Although there are 49 members of this protein family, only three of them have been under attention and widely analysed including MDR1 (also known as P-glycoprotein (P-gp) or ABCB1), MRP1 (ABCC1), and BCRP (ABCG2) [49]. The first drug transporter that was identified is MDR1, and the glycoprotein is bounded to cell membrane and is low expressed in approximately all tissues with higher expression and activity in epithelial cells with excretory functions including those lining colon, intestine, pancreatic tubules, and bile ducts [50, 51]. The upregulation of MDR1 is found on various tumours, and interestingly, chemotherapy drugs have ability of increasing its expression [52]. As a result, its inhibitors including zosuquidar and tariquidar have been introduced and developed, but their clinical efficacy is comprised. For instance, co-application of tariquidar with anthracycline or taxanes has restricted potential in females with breast cancer at stages III-IV [53].

3.5 Activation and inactivation of chemotherapy drug

Owing to specific changes occurring in drugs, they may be inactivated or their activation is suppressed that is observed in platinum compounds after inactivation by thiol glutathione [54]. Moreover, when there is absence of cellular enzymes, the transformation of 5-flourouracil (5-FU) and methotrexate (MTX) to their active form does not occur [55, 56]. The conversion of capacitabine to 5-FU occurs by function of phosphorylase [57]. Therefore, capacitabine resistance occurs when methylation of gene encoding thymidine phosphorylase occurs [58]. The inactivation of irinotecan is observed by function of UGT1A1, and when DNA methylation of UTG1A1 promoter occurs, it can escalate activity of irinotecan [59, 60].

3.6 Decrease in drug uptake

Although main focus is on the role of dug efflux transporters, there are also evidences showing that decrease in drug uptake can result in development of chemoresistance in cancers [61]. However, the mechanism of action is similar to drug efflux to decrease accumulation of chemotherapy drug in cancer cells and restrict its potential. A number of anti-cancer drugs including 5-FU, cisplatin (CP), and 8-aza-guanine use solute carriers for cancer cell internalization and their cellular uptake may be diminished [62]. In order to solve such problem, it is suggested to use active transport pathways or focus on passive permeability of anti-cancer drug.

3.7 Changes in drug targets

The mutations and abnormal changes in the target of drugs have been considered as another way in the development of resistance to anti-cancer drugs [63]. The Cancer Genome Atlas (TCGA) has detected the mutations of genes in 12 cancer classes [64], and a number of tumour-promoting factors including EGFR, RAS, RAF, and PI3K are considered as most common mutations in tumours. Furthermore, the mutations and changes can result in downregulating or silencing factors including PTEN, Rb, and p16INK4a that are common in human cancers [65]. In addition to changes, mutations and amplifications of genes, such changes in amino acids, can also mediate resistance of tumours to chemotherapy.

3.8 Apoptotic machinery

The aberrant proliferation of tumour cells along with apoptosis suppression can result in development of a condition in which promote tumourigenesis [66]. The alterations in molecular factors such as MYC that participate in regulation of proliferation can also modulate apoptosis. Apoptosis is the final aim of anti-cancer drugs in reducing tumourigenesis, and alterations in apoptotic machinery can lead to development of resistance [67]. Drug resistance has been considered as a result of such defects and abnormal changes in apoptotic machinery, and hence, effective removal of cancer cells depends on caspase-dependent and -independent mechanisms [68]. The anti-apoptotic factors such as Bcl-2 (upregulation) and pro-apoptotic factors such as Bax (downregulation) can be controlled in development of chemoresistance in human cancers [69].

3.9 DNA damage repair

Another major mechanism in the development of chemoresistance is DNA damage repair. In addition to apoptosis, anti-cancer drugs plan in triggering damage into DNA of tumour cells in mediating cell death. However, the stimulation of DNA damage repair mechanisms can induce chemoresistance in human cancers [70]. The anti-cancer drugs stimulate double-strand break (DSB) in tumour cells, and when DNA repair pathways and homologous recombination are stimulated, such activity is reduced or suppressed [71]. The role of DNA damage repair is not certain to a single type of cancer, and it can result in chemoresistance in various tumours including breast tumour [72], hepatocellular carcinoma [73], and ovarian cancer [74]. BMAL1 has been suggested to collaborate with CLOCK in enhancing DNA damage repair and the development of drug resistance [75]. The function of copper is related to increased DNA damage repair through ATOX1 in cancer drug resistance [76].

3.10 Cancer stem cells and drug resistance

Cancer stem cells (CSCs) are a rare population in the tumour colonies that are correlated with the malignant biological features of cancers including drug resistance, radioresistance, cancer recurrence, and capacity of G0 phase arrest in the development of new tumours [77, 78]. The first characterization of CSCs was performed in 1990s in which they were recognized in leukaemia and their isolation occurred through CD34⁺ and CD38⁻ surface marker expression [79, 80]. Then, the CSCs were detected in the different solid and haematological tumours and they express a number of surface makers including CD133, nestin, and CD44 [81, 82]. The CSCs have self-renewal capacity and ability of differentiation into several cellular subtypes [83]. The function of CSCs can be modulated by a number of intracellular and extracellular factors, providing new insights and promising targets in cancer therapy [84]. The process of carcinogenesis can be regulated by CSCs in the different tumour classes. The CSC-like properties can be increased in ovarian tumour through function of actin-like protein 6A, and this stimulates unfavourable prognosis [85]. The suppression of AKT/ GSK3β/β-catenin axis by MASM can impair the CSC-like features in EpCAM⁺ cells [86]. Furthermore, the proanthocyanidins have ability of suppressing Wnt/β-catenin axis to impair the CSC features in colorectal tumour [87]. The extracellular vesicles can be derived from CSCs and possess the ability of targeting MHC-II-macrophages and $PD^{1+}T$ cells [88], showing the ability of CSCs in the tumour microenvironment remodelling. The several molecular pathways have ability of regulating CSCs including Notch, Hedgehog, PI3K/Akt, Wnt, MAPK, and JAK/STAT. These related pathways of CSCs can be targeted by the natural products for cancer therapy and impairing tumourigenesis and therapy resistance [89].

3.11 Pro-survival autophagy

A programmed cell death (PCD) mechanism with potential functions in the regulation of cancer drug resistance is known as autophagy. The aberrant activation of autophagy has been well-documented in human cancer [90], and it is considered as a potent regulator of drug resistance [91], capable of decreasing/increasing cancer progression, and therapy resistance based on its dual function. Adaptor SH3BGRL elevates stability of ATG12 and increases translation of PIK3C3 to mediate autophagy in the development of drug resistance [92]. The sequestration of miR-488-3p by circPOFUT1 results in stimulation of PLAG1/ATG12 axis in autophagy induction and enhancing chemoresistance [93]. SMC4 is another factor in the development of autophagy to mediate drug resistance [94]. Other types of autophagy also play a significant role of drug resistance such as mitophagy in cancer drug resistance [95]. Figure 2 displays the role of factors in development of drug resistance in human cancers. Table 1 represents the mechanisms causing drug resistance in PCa.

4 The molecular landscape of drug resistance in prostate cancer

4.1 microRNAs

The small RNA molecules lacking ability in protein translation are known as microRNAs (miRNAs) that their length is less than 24 nts and their aberrant expression

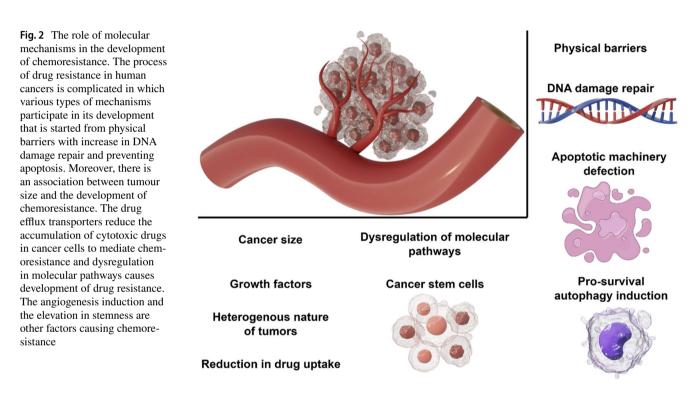


 Table 1
 An overview of drug resistance mechanisms in PCa

Molecular profile	Highlights	Ref
CircFoxo3	CircFoxo3 impairs EMT and reduces proliferation and survival of tumour cells in increasing docetaxel sensitivity	[<mark>96</mark>]
WDR5	WDR5 upregulates PD-L1 expression and stimulates drug resistance	[<mark>97</mark>]
DRD2	Bromocriptine promotes DRD2 expression and reverses drug resistance	[<mark>98</mark>]
BMP	The downregulation of BMP enhances drug sensitivity	[<mark>99</mark>]
Akt	The downregulation of Akt isoforms, especially Akt1 and Akt2 significantly suppress chemoresistance	[100]
ZEB1	ZEB1 increases MRP4 levels to induce drug resistance	[<mark>10</mark> 1]
E-cadherin	E-cadherin downregulation causes Notch upregulation to mediate chemoresistance	[102]
EED/EZH2	The non-canonical EED-EZH2 axis suppression by pharmacological compounds impairs drug resistance	[103]
NPRL2	NPRL2 impairs mTOR axis to induce autophagy and inhibit apoptosis, causing drug resistance	[104]
NOTCH1	NOTCH1 increases ABCC1 expression in causing drug resistance	[105]
IGFBP2	Hyperglycaemia increases IGFBP2 expression to mediate drug resistance	[106]
ETS1	ETS1 increases MDR1 and MMP0 levels in causing drug resistance and increasing cancer metastasis	[107]

has been confirmed in various diseases, especially cancer [108]. The recent studies have emphasized on the potential role of miRNAs in regulation of apoptosis [109], metastasis [110], and other biological behaviours. The miR-375 has been suggested to involve in the development of DTX resistance in PCa. SEC23A and YAP1 play a significant role in development of chemosensitivity. However, miR-375 reduces levels of YAP1 and SEC23A to diminish response of PCa cells to DTX chemotherapy [111]. On the other hand, miR-34a enhances chemosensitivity through stimulation of apoptosis and cell cycle arrest by decreasing SIRT1 expression [112]. miR-99b-5p suppression and mTOR enhancement can jointly participate in the development of chemoresistance in African American PCa [113]. Interestingly, miRNAs can be enriched in exosomes as small extracellular vesicles to regulate tumourigenesis and exosomes function as cell-cell communication tools [114]. The enrichment of miR-27a in exosomes is observed in PCa that by reducing p53 expression, they participate in the drug resistance development in tumour cells [115]. However, a few studies have focused on this potential of exosomal miRNAs in the regulation of chemoresistance and more studies should be conducted in this case. According to the studies, miRNAs can participate in both reducing and increasing response of PCa cells to chemotherapy. For instance, miR-21 participates in the development of chemoresistance in PCa through downregulation of PDCD4, and therefore, manipulating expression level of miR-21 can contribute to modulation of tumourigenesis and therapy response in PCa [116]. In order to increase potential in increasing drug sensitivity, studies have focused on regulating more than one factor. The upregulation of miR-145 and downregulation of TR4 can result in DTX sensitivity of PCa cells [117]. miR-17-92 cluster has been considered as a factor involved in drug resistance that stimulates Akt axis in promoting ERK1/2 expression [118]. miR-199a is a critical player in regulating tumourigenesis and miR-199a-5p inhibition results in upregulation of HIF-1a to increase tumourigenesis in PCa [119]. ROR2 reduces expression level of miR-199a-5p to suppress metastasis of PCa cells [120]. Hence, miR-199a disrupts pathogenesis and tumourigenesis of PCa; miR-199a diminishesYES1 expression to enhance DTX sensitivity [121]. miR-34a is able to increase drug sensitivity; miR-34a diminishes JAG1/Notch1axis to increase paclitaxel (PTX) sensitivity [122]. On the other hand, miR-129-5p stimulates DTX resistance through downregulation of CAMK2N1 to trigger DTX resistance [123]. Hence, regulating expression level of miRNAs, especially silencing miRNAs can enhance drug sensitivity. miR-193a-5p diminishes Bach2 expression to stimulate DTX resistance, and therefore, silencing miR-193a-5p can increase drug sensitivity [124].

4.2 LncRNAs

The long non-coding RNAs (lncRNAs) are other factors with significant contribution in regulating PCa tumourigenesis. The function of lncRNAs in PCa is versatile, and they are promising modulators of tumourigenesis that at cellular and molecular level; they can control a number of pathways and mechanisms including proliferation, metastasis, apoptosis, autophagy, cell cycle arrest, and related molecular pathways [125]. The recent studies have revealed function of lncRNAs in the regulation of chemoresistance in PCa. The function of chemotherapy drugs in reducing tumourigenesis is attributed to the potential in preventing cell death. The ferroptosis is an iron-dependent cell death that its regulation in PCa is mediated by a number of molecular pathways. CEMIP is capable of preventing ferroptosis to promote viability of PCa cells [126]. Moreover, SGK2 escalates expression level of GPX4 to suppress ferroptosis in increasing invasion of PCa cells [127]. The expression level of IncRNA PCAT1 can be increased by function of TFAP2C that subsequently enhances stability of c-Myc. Moreover, PCAT1 increases SLC7A11 expression through miR-25-3p suppression in decreasing iron accumulation and avoiding ferroptosis in cancer cells [128]. The function of lncRNAs in drug resistance regulation in PCa is more than simple control of molecular mechanisms such as ferroptosis. LncRNA HOTTIP has been involved in the development of cisplatin resistance in PCa; to this end, HOTTIP stimulates Wnt/βcatenin axis to trigger cisplatin resistance [129]. Silencing such lncRNAs can contribute to increased toxicity of anticancer drugs. Silencing SNHG6 leads to the suppression of growth and metastasis of PCa cells and increases PCa sensitivity to PTX chemotherapy [130]. Moreover, knock-out of LINC01963 results in DTX sensitivity in PCa cells through promoting expression level of its target miR-216b-5p and suppressing carcinogenesis and lung invasion [131]. The function of lncRNAs in the regulation of drug sensitivity in PCa has been widely evaluated in regard to regulation of miRNAs including miR-183 [132], miR-24-3p [133], miR-149-5p [134], miR-497-5p [135], and miR-33b-5p [136]. However, function of lncRNAs in the modulation of drug resistance in PCa can be distinct from miRNA sponging like function of HOXD-AS1 that recruits WDR5 for drug resistance development [137].

4.3 CircRNAs

The circular RNAs (circRNAs) are similar to lncRNAs and miRNA from the standpoint that none of them are able to encode proteins and their functions in cells is regulatory. The circRNAs have a covalently closed loop structure, and they are promising candidates in the regulation of tumourigenesis in PCa, mainly through sponging and controlling miRNAs [138–141]. However, there are studies showing that circRNA function in PCa can be mediated in a miRNAindependent manner such as circ-0006156 that prevents ubiquitination of S100A9 in reducing PCa invasion [142]. A few experiments have evaluated the potential of circRNAs in the modulation of drug resistance in PCa; circ-0004087 is at class of oncogenic circRNAs that a result of its interaction with SND1 is to escalate mitotic error in promoting DTX resistance [143]. However, the role of circRNAs in the regulation of chemoresistance has been mainly evaluated in terms of regulation of miRNAs. miR-7 has ability of increasing DTX sensitivity in PCa; however, circ-0000735 reduces miR-7 expression to escalate DTX resistance [144]. Hence, circRNA-miRNA circuit regulates response of PCa cells to chemotherapy [145]. Recently, circRNAs enriched in exosomes have obtained much attention in field of tumour diagnosis and acting as biomarker [146]. The function of exosomal circRNAs in human cancers is versatile, and they are potent regulators of proliferation, invasion, and metabolism [147]. The exosomal circ-SFMBT2 has been considered as a regulator of tumourigenesis in PCa and response to DTX chemotherapy. Exosomal circ-SFMBT2 positively

regulates TRIB1 expression through miR-136-5psponging to escalate tumourigenesis for DTX resistance development [148]. Figure 3 depicts the role of non-coding RNAs in regulation of chemotherapy response in PCa cells.

4.4 PI3K/Akt

The PI3K/Akt axis is another factor in the development of chemoresistance in PCa. Increasing evidence has shown potential of this axis in enhancing carcinogenesis. The upregulation of PI3K/Akt enhances tumourigenesis, induces EMT, and stimulates DTX resistance. The function of INPP4B is against tumourigenesis in PCa, and INPP4B reduces PI3K/Akt expression to suppress EMT and DTX resistance [149]. In fact, PI3K/Akt participates in both increasing invasions of PCa cells and reducing the response to chemotherapy. The progression and malignancy of PCa cells enhance by the function of EpCAM, and it stimulates PI3K/Akt/mTOR axis in triggering resistance to chemotherapy and radiotherapy [150]. The role of PI3K/Akt in the development of chemoresistance in human cancers is related to preventing apoptosis. In fact,

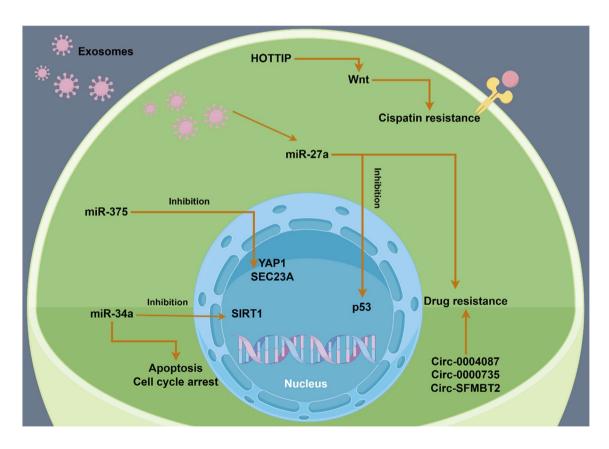


Fig. 3 The non-coding RNA-mediated regulation of chemotherapy response. In case of circRNAs, only oncogenic circRNAs with ability of inducing drug resistance have been described including circ-0004087, circ-0000735, and circ-SFMBT2. Moreover, miRNAs and their exosomal forms can suppress apoptosis and induce chemoresist-

ance through regulation of p53, SIRT1, and YAP1, among others. In terms of lncRNAs, they can sponge miRNAs for ferroptosis inhibition. Moreover, HOTTIP stimulates Wnt axis to mediate cisplatin resistance CCR9 stimulates PI3K/Akt axis to enhance and evoke anti-apoptotic signal in decreasing response of PCa cells to etoposide chemotherapy [151]. In addition to apoptosis, the expression level of MRP-1 as a potential factor involved in the chemoresistance is affected by PI3K. The upregulation of MRP-1 by PI3K results in reduction in response of PCa cells to chemotherapy [152]. Moreover, the drug efflux transporters including MDR1 can be stimulated by EGFR through upregulation of AKt to induce DTX resistance [153]. However, PI3K/Akt is not the only pathway and an oncogenic factor can regulate different downstream targets. CD44v6 increases lymph node metastasis of PCa cells and increases sphere formation. Interestingly, CD44v6 enhances expression level of PI3K/Akt/ mTOR, EMT, and Wnt/ β -catenin in the development of chemoresistance and radioresistance in PCa cells [154]. As a result of silencing such carcinogenic factors in cancers, the drug sensitivity enhances. The interesting point is the involvement of endothelial cells in development of DTX resistance in PCa cells through stimulation of Akt/mTOR axis [155].

4.4.1 PTEN

The previous section focused on the role of PI3K/Akt axis in the development of drug resistance in PCa cells. The function of PI3K/Akt in PCa is oncogenic, and it mediates chemoresistance. However, PTEN is upstream mediator of PI3K/Akt and suppresses its activation. The PTEN deficiency escalates the progression of PCa cells, and when loss of PTEN occurs, it causes degradation of FBP1 to induce Warburg impact in increasing tumourigenesis [156]. Furthermore, loss of PTEN mediates the poor prognosis of PCa patients [157]. The therapy response of Pca cells is regulated by PTEN signalling. BRD4 escalates expression level of KDM5C through transcriptional level to suppress PTEN expression in enhancing tumourigenesis and escalating cancer growth [158]. When expression level of PTEN enhances, it promotes potential of drugs in apoptosis induction and enhances sensitivity of Pca cells to DR-induced cell death [159]. Furthermore, PTEN reduces expression level of Bcl-2 as an anti-apoptotic protein in increasing drug sensitivity [160]. The activity of drug transporters is regulated by PTEN. Low expression level of miR-21 results in PTEN overexpression to downregulate P-gp in increasing drug sensitivity in Pca [161]. Notably, low expression of PTEN can result in the development of resistance to more than one anticancer drug. TUBB3 reduces PTEN expression, and when TUBB3 silencing is performed, it causes PTEN upregulation and sensitivity to DTX and cabazitaxel promotes [162]. A clinical study has revealed that upregulation of TUBB3 and downregulation of PTEN result in poor prognosis [163].

4.4.2 STAT3

STAT3 is considered as another factor involved in tumourigenesis in PCa. Increasing evidence highlights the function of STAT3 in favour of tumourigenesis in PCa and its downregulation by Pinus mugo essential oil leads to apoptosis and oxidative damage in tumour cells [164]. The upregulation of JAK/STAT3 by HSPB8 leads to increase in PCa progression [165] and STAT3 downregulation mediates apoptosis [166]. Interestingly, the strategy of drug repurposing has focused on regulation of STAT3 in cancer therapy [167]. The dysregulation and abnormal alterations in gut microbiota mediate the DTX resistance in PCa that is due to stimulation of NF-kB axis that promotes IL-6 levels in STAT3 induction [168]. Therefore, IL-6 can function as upstream mediator of STAT3 in cancer drug resistance. Interestingly, cancerassociated fibroblasts are able to secrete IL-6 in stimulation of JAK/STAT3 axis to stimulate doxorubicin resistance in PCa [169]. The suppression of STAT3 promotes enzalutamide sensitivity of PCa cells [170]

5 Molecular mechanisms of drug resistance in prostate cancer

5.1 Apoptosis

Apoptosis was mentioned before as a factor in cancer chemotherapy that defects in this system can result in the development of drug resistance. The stimuli including ROS overgeneration and endoplasmic reticulum stress can result in apoptosis. Several caspases participate in process of apoptosis and finally, irreversible alterations occur in cells. The cell shrinkage, chromatin condensation, blebbing, and generation of apoptotic bodies are morphological characteristics of apoptosis [171]. The lack of response of PCa cells to apoptosis can pave the way in development of chemoresistance. The molecular interactions that prevent apoptosis in Pca play a critical role in drug resistance development. Calcitonin has ability of increasing expression level of Akt to induce survivin expression. Then, apoptosis resistance is observed against the anti-cancer drugs in Pca [172]. The apoptotic rate and proliferation of Pca cells have close interaction. The exposure of Pca cells to chemotherapy results in apoptosis induction and reduction in proliferation rate. Such association has been evaluated in the function of WD repeat domain 5 (WDR5) in which regulates apoptosis and proliferation of Pca cells. Moreover, apoptosis and DNA damage repair can be regulated together in determining the response of Pca cells to the therapy. The apoptosis inhibition and DNA damage repair can be mediated by WDR5 in triggering cisplatin resistance in PCa, and a number of factors such as AURKA, CCNB1, E2F1, PLK1, BIRC5, XRCC2,

and PD-L1 are modulated [97]. The DTX is another chemotherapy drug in PCa suppression that is potential is based on modulating depolymarization of microtubules. The DTXmediated apoptosis can be suppressed by molecular interactions; miR-204 upregulation results in ZEB1 downregulation to stimulate apoptosis and prevent drug resistance [173]. Furthermore, there is correlation between drug efflux transporters and apoptosis in PCa cells [174]. Cofilin-1 is considered as a factor relating drug efflux and apoptosis. Exosomal transfer RPS3 results in stimulation of PI3K/Akt axis to enhance cofilin-1 levels in drug resistance development [175]. The phosphorylation of cofilin-1 can be stimulated by ERK1/2 and phatycodin D can suppress drug resistance by suppressing cofilin-1 [176]. The cofilin-1 increases levels of p38 MAPK to upregulate activity of MDR1 to prevent apoptosis in drug resistance development [174].

CXCR4 increases HMGA2 expression to induce chemoresistance [177] and HMGA2 escalates FOXL2 expression to stimulate EMT in increasing drug resistance [178]. HMGA2 is able to increase expression level of Wnt in promoting 5-FU resistance [122]. Upregulation of HMGA2 prevents apoptosis in PCa [179], and this may result in chemoresistance in tumour cells in the future. USP9x reduces apoptosis in PCa cells and can develop drug resistance. Reduction USP9x can lead to the degradation to PBX1 to trigger apoptosis and reverse drug resistance [180]. The high expression level of Notch-1 PCa can decrease potential of DTX in cancer therapy. Therefore, silencing Notch-1 participates in apoptosis induction and mediating mitotic arrest mediated by DTX [181]. The cleavage of PARP1 is vital for triggering apoptosis in PCa cells. Interestingly, cisplatin stimulates DNA damage and promotes PARP1 claevage to induce apoptosis. Moreover, NRF2 downregulation leads to ROS overgeneration and sensitivity of PCa cells to cisplatin [182]. The regulation of anti- and pro-apoptotic proteins can control apoptosis in PCa cells. The TLR4 upregulation prevents apoptosis to induce DTX resistance in PCa, while silencing TLR4 promotes Bax expression and reduces Bcl-2 levels in apoptosis induction and reversing DTX insensitivity [183]. However, the regulation of apoptosis is more than simple regulation of apoptotic proteins. PDIA-4 is able to induce phosphorylation of Akt in apoptosis inhibition and mediating DTX resistance [184]. Therefore, apoptosis is a determining factor of therapy response in PCa cells.

5.2 Autophagy

Autophagy is considered as a regulated cell death similar to apoptosis in which degrades toxic macromolecules and aged organelles in cell homeostasis, while its function is transformed in tumour cells and it can reduce/increase carcinogenesis. The autophagy is a highly regulated mechanism in which autophagy-related genes (ATGs), ULK1, Beclin-1, and AMPK stimulate autophagy, while mTOR suppresses autophagy. Autophagy function in PCa has been fully investigated showing that proliferation and metastasis of PCa cells can be dually induced/suppressed by autophagy. This mechanism can regulate apoptosis in PCa cells and therefore, understanding its function in therapy response of PCa cells is of importance that is aim of current section.

The role of autophagy in chemotherapy response of PCa cells can be protective that upon upregulation of autophagy by Ambra1 in PCa cells, apoptosis is reduced and therefore, sensitivity of tumour cells to chemotherapy diminishes [185]. STAT3 is a regulator of tumourigenesis in human cancers and evaluation of its function in PCa displays that STAT3 controls the biological behaviour of PCa cells and its phosphorylation escalates tumourigenesis [186]. The miR-125a upregulation by curcumol results in STAT3 suppression to impair carcinogenesis in PCa [187]. Furthermore, the expression level of JAK2/STAT3 in PCa is regulated by SOCS3 [188]. In CRPC, the tumour cells lack response to DTX chemotherapy that is attributed to pro-survival autophagy stimulation. Notably, STAT3 overexpression is vital for autophagy induction and lack of response of PCa cells to DTX chemotherapy [189]. Interestingly, regulation of autophagy can sensitize tumour cells to apoptosis and improve chemosensitivity. NPRL2 has been suggested to reduce expression level of mTOR in autophagy induction, while silencing NPRL2 suppresses autophagy and escalates apoptosis in mediating DTX sensitivity [104].

The NEPC is a malignant and progressive type of PCa that IL-6 is critical for NEPC. Interestingly, autophagy is a pre-requisite for IL-6-medaited NEPC and stimulates drug resistance. IL-6 increases AMPK phosphorylation and reduces mTOR expression in triggering drug resistance and advancing progression into NEPC. Autophagy suppression promotes apoptosis in PCa cells [190]. Furthermore, AMPK/ mTOR axis is vital for regulation of ATG4B expression in PCa. The methylation of miR-34a results in downregulation of this miRNA, and then, AMPK upregulation occurs. Then, mTOR expression reduce to increase ATG4B expression in autophagy induction and accelerating drug resistance [191]. Owing to this critical function of autophagy in cancer drug resistance, its suppression can lead to reduction in resistance. However, function of autophagy is lethal in PCa sometimes, and in this case, autophagy suppression induces chemoresistance. It has been reported that PrLZ is an oncogenic factor in PCa that reduces LKB1 expression to suppress AMPK/autophagy axis in triggering DTX resistance [192]. The AMPK/mTOR axis is not the only factor regulating autophagy in PCa and upregulation of Beclin-1 results in autophagy stimulation to induce drug resistance [193]. After the downregulation and inhibition of PLIN3, autophagy intensification occurs that mediates DTX resistance in PCa and such drug resistance is suppressed by chloroquine as inhibitor of autophagy [194].

Cisplatin resistance also commonly occurs in PCa; the glycolysis dysfunction can be induced by RSL3 in accelerating potential of cisplatin in PCa suppression [195]. DUSP1 upregulation can be provided by resveratrol in apoptosis induction and increasing cisplatin sensitivity of PCa [196]. However, stimulation of pro-survival autophagy can mediate cisplatin resistance in PCa. Silencing CFTR promotes cisplatin sensitivity of PCa cells through autophagy inhibition [197]. At the time that autophagy exerts protective role, its inhibition enhances apoptosis and reverses drug resistance [198]. In case that autophagy stimulates drug resistance, the autophagy-related proteins can be directly targeted. For instance, FOXM1 stimulates AMPK/mTOR axis in autophagy induction and mediating DTX resistance. Silencing ATG7, Beclin-1, or application of chloroquine can suppress DTX resistance [199]. Furthermore, enhanced autophagy through NPRL2 stimulates Everolimus resistance in PCa [200]. Interestingly, the progression of PCa can be provided by HMGB1 through AR upregulation [201]. HMGB1 stimulates Akt axis in increasing carcinogenesis in PCa [202], and it is a regulator of NF-kB in enhancing cancer metastasis [203]. The gemcitabine resistance can be facilitated through HMGB1-induced autophagy [204]. Therefore, autophagy plays a critical role in regulating chemotherapy response of PCa cells [205-209].

5.3 Cancer stem cells in prostate cancer and therapy resistance

The regulation of CSC features and phenotype in PCa occurs through different molecular mechanisms. SFRP1 has ability of elevating the CSC features and phenotype in prostate cancer through upregulation of Wnt/β-catenin axis and increasing levels of SOX2, NANOG, and OCT4 [210]. Both SFRP1 and GSK-3ß can be regulated by miR-1301-3p. SFRP1 and GSK-3β are suppressed by miR-1301-3p that are Wnt inhibitor, causing upregulation of OCT4, SOX2, NANOG, CD44, KLF4, c-MYC, and MMP2 and improving CSC features in PCa [211]. The upregulation of ZEB1 accelerates the CSC features in PCa, and downregulation of ZEB1 reduces levels of CD44, CD133, and SOX2 as markers of stemness [212]. The stimulation of Jagged1/Notch1 axis by bone marrow mesenchymal stem cells can enhance the stemness of PCa [213]. The increasing evidences have shown that stemness and CSCs participate in the development of therapy resistance in PCa. The CSCs appear to be resistant into radiotherapy and a combination of fractionated irradiation and B7-H3 can significantly suppress the growth of PCa [214]. An interesting part is the relationship of CSCs and EMT in the regulation of cancer metastasis. The upregulation of CSC markers in PCa including SOX2, NANOG, KIF4,

OCT3/4, and c-Myc can accelerate the invasion and metastasis through overexpression of Slug, ZEB1, and Twist1 [215]. The increase in CSC features and cancer invasion can cause the development of castration resistance [216]. The proliferation of CSCs occurs slowly, and they stimulate DNA damage repair, and upregulation of γ H2AX is another factor in the reduced anti-cancer activity of etoposide against PCa [217]. The overexpression of NANOG promotes levels of CXCR4, CD133, ALDH1, and IGFBP5 in the expansion of CSCs and the development of androgen deprivation resistance [218]. Therefore, it can be perceived that CSCs contribute to the development of therapy resistance in PCa.

5.4 Epithelial-mesenchymal transition

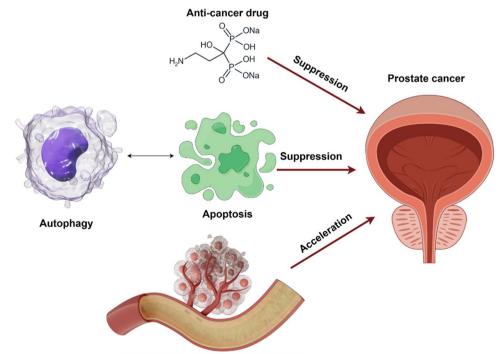
The epithelial-mesenchymal transition (EMT) has been suggested to be one of the leaders in the regulation of cancer metastasis with growing contribution to low therapy response of tumour cells. With improvement in understanding the molecular and structural changes of EMT, it is now obvious that morphological aspect of EMT is mesenchymal transformation of epithelial cells, and at molecular level, E-cadherin as epithelial marker is suppressed, and then, N-cadherin and vimentin are escalated. The increasing evidence highlights the function of EMT in the development of resistance to various chemotherapy drugs [219, 220]. The recent experiments have highlighted the fact that EMT plays a significant role in the progression of PCa cells [221]. The loss of vitamin D results in EMT stimulation in escalating cancer metastasis [222], and EMT suppression by atractylenolide I increases anti-cancer function of cabozantinib [223]. The increase in progression of PCa cells can be accompanied with the development of chemoresistance. The EMT and drug resistance occur simultaneously in PCa. SPP1 is able to stimulate PI3K/Akt and ERK1/2 pathways in mediating EMT and development of enzalutamide resistance [224]. CNTN-1 stimulates PI3K/Akt axis and induces EMT in evoking DTX resistance [225]. The wealth evidence displays that EMT induction by oncogenic factors can mediate chemoresistance in PCa cells. The EMT mechanism can participate in the chemoresistance in Pca through both intrinsic and extrinsic pathways. In intrinsic mechanism, EGFR promotes Rad51 expression to stimulate EMT and increase DNA damage repair in intrinsic resistance [226]. Furthermore, the ability of CRPC cells in developing resistance to enzalutamide is based on progression acceleration that is mediated by function of ISL1 in inducing EMT [227]. There is essential association between metastasis of Pca cells to bone and their ability of developing resistance to DTX chemotherapy. TGF- β escalates acetylation of KLF5 in promoting bone metastasis of Pca cells, and acetylated form of KLF5 promotes osteoclastogenesis and increases bone metastatic lesions through stimulation of CXCR4. Moreover,

acetylated KLF-5 has been associated with enhancement in carcinogenesis and increasing DTX resistance (Fig. 4) [228].

6 Combination therapies in prostate cancer: application of small molecule inhibitors and natural products

Both natural compounds and synthetic small molecules have been introduced in regulating the therapy response of PCa cells regarding molecular view. Each of these compounds have ability of affecting a certain molecular pathway in increasing chemosensitivity of PCa cells. There are differences about using natural products or small molecules in the treatment of PCa that their efficacy is various. For instance, small molecules are designed for a specific target in PCa therapy, while natural compounds are able to affect various pathways. Furthermore, phytochemicals are more available compared to small molecules, they are affordable, and their biocompatibility is higher compared to small molecules. The quercetin is a natural compound commonly utilized in the treatment of PCa; guercetin is capable of escalating radiosensitivity of PCa cells [229], and by increasing ER stress and ROS generation, it promotes paclitaxel sensitivity of PCa cells [230]. The upregulation of c-met can induce doxorubicin resistance in PCa cells; quercetin decreases c-met expression to suppress PI3K/ Akt axis in enhancing doxorubicin sensitivity of PCa cells [231]. Moreover, the response of PCa cells to docetaxel can be improved. Quercetin is able to suppress AR and PI3K/ Akt pathways and increase apoptosis to promote docetaxel sensitivity of PCa cells [232]. The important molecular pathways regulating tumourigenesis in PCa are affected by natural compounds. For instance, eupatilin is able to suppress NF-kB axis and elevate PTEN expression in disrupting growth and metastasis of PCa cells [233]. The combination of natural compounds in exerting more inhibitory impact on PCa cells than monotherapy is suggested. A combination of green tea and quercetin can be utilized for downregulation of PI3K/Akt to escalate DTX sensitivity of PCa cells [234]. Owing to multi-targeting feature of green sources, they are capable of regulating more than one molecular pathway in PCa removal. The β -elemonic acid is capable of stimulating apoptosis in PCa cells, and for this purpose, it diminishes expression level of JAK2/STAT3 to downregulate MCL-1, and it suppresses NF-KB axis in impairing tumourigenesis [235]. Moreover, natural products are beneficial in disrupting tumourigenesis in drug resistant-tumour cells. Acetyl-11-keto-β-boswellic acid has been shown to downregulate Akt and STAT3 pathways and inhibits stem-like features in PCa to impair tumourigenesis [236]. Even components of tumour microenvironment can be regulated in affecting chemoresistance in PCa cells. Qi Ling is capable of suppressing IL-6/STAT3 axis, and it reduces M2 polarization of macrophages in elevating PTX sensitivity of PCa cells [237]. STAT3/AR downregulation by galiellalactone can disrupt enzalutamide insensitivity in PCa [238]. Notably, reduction in metastatic potential of PCa cells can participate in chemosensitivity; metformin has been suggested to suppress EMT through STAT3 downregulation in escalating enzalutamide

Fig. 4 The role of biological mechanisms in development of chemoresistance. In the development of drug resistance, several biological mechanisms demonstrate interactions with each other in which protective autophagy inhibits apoptosis, while lethal autophagy stimulates apoptosis to increase drug sensitivity in prostate cancer. Moreover, the increase in the EMT can accelerate drug resistance. The association of autophagy and EMT in prostate cancer drug resistance requires more investigation



Cancer metastasis and EMT induction

sensitivity [239]. When a molecular pathways such as Akt is suppressed by phytochemicals, it causes reduction in the proliferation of PCa cells and promotes their chemosensitivity [240]. Hence, phytochemicals are promising candidates in the regulation of drug sensitivity in PCa [241, 242].

In addition to natural products, the small molecules have been widely utilized in regulating response of PCa cells to chemotherapy. GBP730 is a small molecule inhibitor of STAT3 that has ability of increasing sensitivity of PCa cells to enzalutamide chemotherapy [243]. The suppression of STAT33/AR axis by niclosamide results in reduction in metastasis of PCa cells to reverse enzalutamide resistance [244]. Upon regulation of molecular pathways, a plan is provided for regulating apoptosis and drug efflux transporters in PCa cells. BKM1972 is considered as a small molecule in which reduces survivin expression and downregulates P-gp activity in enhancing DTX sensitivity in PCa cells [245]. Rapamycin is a regulator of autophagy that diminishes expression level of cyclin D1 in increasing potential of cisplatin in PCa suppression [246]. Therefore, combination therapy can bring new hopes in the treatment of PCa and the reason is regulation of underlying molecular pathways in suppressing tumourigenesis. Everolimus (RAD001) can be utilized for reducing levels of HIF-1 α and sphingosine kinase 1 to increase DTX sensitivity of PCa cells [247]. Accordingly, both drugs, small molecules and phytochemicals, are promising candidates in regulation of drug sensitivity in PCa.

7 Radioresistance in prostate cancer

7.1 Molecular interactions

Although the main focus of the studies is on evaluation the underlying molecular pathways regulating chemoresistance, there are evidences showing that molecular interactions can also regulate response of PCa cells to radiotherapy. The complicated networks among pathways can regulate radioresistance in PCa. HIF-1 α is able to escalate nuclear transfer of β-catenin in apoptosis inhibition, increasing growth and metastasis, and promoting DNA repair to mediate radioresistance [248]. Interestingly, chemoresistance and radioresistance in PCa can occur simultaneously. Keap1 deficiency leads to stimulation of chemoresistance and radioresistance in PCa cells. This is due to the fact that loss of Keap1 results in upregulation of Nrf2 to mediate therapy resistance [249]. The factors regulating radioresistance have been shown to induce therapy resistance in both in vitro and in vivo. EHMT2 is an inducer of radioresistance in PCa that decreases ERP29 expression in mediating resistance in both in vitro and in vivo [250]. Similar to chemoresistance, non-coding RNAs have been considered as factors regulating radioresistance. As it was mentioned in introduction section, enhancement in DNA damage repair can cause development of radioresistance in PCa. The miR-205 is able to escalate radiosensitivity in PCa cells, and to this end, miR-205 diminishes expression levels of ZEB1 and PKCe to preventing DNA damage repair and enhancing radiosensitivity [251]. Hence, regulation of DNA damage repair can lead to changes in radiosensitivity of PCa cells. Downregulation of cyclin D1 is vital for disrupting DNA break repair [252].

The progression of PCa cells is highly dependent on the hypoxia in TME that is due to insufficient oxygenation of cancer cells. The stimulation of HIF-1α/Notch1 can occur in PCa to increase stem cell phenotype [253], and hypoxia is capable of triggering radioresistance [254]. The expression level of miRNAs is regulated by hypoxia. The hypoxic TME reduces expression levels of miR-124 and miR-144 as a way to upregulate PIM1 to evoke autophagy in development of radioresistance [255]. Similar to chemotherapy, function of autophagy in radiotherapy can be oncogenic or onco-suppressor. Downregulation of PC1/PrLZ results in reduction in DNA damage repair and stimulation of autophagic cell death in enhancing radiosensitivity [256]. Interestingly, plasticity is a determining factor for radiotherapy response of PCa cells. Upon irradiation, the number of ALDH + cells exceeds ALDH- to mediate radioresistance [257]. The increase in antioxidant defense system can lead to development of radioresistance, while miR-17-3p has ability of impairing antioxidant defense in PCa cells to enhance radiosensitivity [258]. Hence, radiotherapy response of PCa cells is regulated by a number of underlying molecular interactions and mechanisms including apoptosis, autophagy, and DNA damage repair participate in this condition.

7.2 Therapeutic approaches

The therapeutic approaches have been displayed to be beneficial in enhancing radiosensitivity in PCa cells. The application of PARP1 inhibitor along with irradiation can result in increase in radiosensitivity of PCa cells [259]. For overcoming radioresistance, intermittent radiotherapy has been suggested. However, fractionated irradiation can lead to neuroendocrine differentiation of PCa cells. Therefore, JNJ-64619178 has been utilized as a factor to avoid DNA damage repair and increase radiosensitivity of PCa cells [260]. Both in vitro and in vivo experiments have revealed that combination therapy can elevate drug sensitivity of PCa cells. Atorvastatin is able to enhance response of PCa cells and mice to radiotherapy, and for this purpose, it stimulates interaction of Bcl-2 and MSH2 in accelerating potential of irradiation in PCa removal [261]. Besides, the DNA damage repair can be regulated by therapeutic compounds. Silibinin as anti-cancer agent is able to diminish DNA damage repair in PCa cells, and this is beneficial for increasing radiosensitivity [262].

Recently, resveratrol has been suggested as a new therapy for PCa. The TRAF6/PTCH/SMO axis can be suppressed by resveratrol in impairing PCa progression [263]. The liposomes for co-delivery of resveratrol and docetaxel have been fabricated to suppress PCa malignancy [264]. Moreover, vasculogenic mimicry in PCa can be inhibited by function of resveratrol through downregulation of Akt [265]. Interestingly, resveratrol is beneficial in increasing radiosensitivity of PCa cells. Administration of resveratrol escalates apoptosis and cell senescence in PCa and disrupt growth through increasing p15, p21, and mutant p53; reducing cyclin B, cyclin D, and cdk2; and enhancing Fas and TRAILR1 to accelerate radiation-mediated PCa removal [266]. The control of DNA-PKcs is another factor in regulation of irradiation response of PCa cells. Interestingly, metformin is able to reduce levels of EGFR/PI3K/Akt to prevent phosphorylation of DNA-PKcs in elevating radiosensitivity [267]. The reason of using complementary therapies with PCa is that when tumour cells are exposed to radiation, they may develop relapse and metastasis in next steps. Therefore, application of BEZ235 as suppressor of PI3K and mTOR is suggested to impair tumourigenesis and mediate cell cycle arrest at G2/M phase in enhancing radiosensitivity [268]. In fact, the anti-cancer drugs and small molecules are able to disrupt to barriers towards effective radiotherapy of PCa cells. HZ08 has shown potential in increasing ROS production, decreasing mitochondrial respiration, reducing level of MnSOD as antioxidant enzymes, suppressing PI3K/Akt/ IKKα, and impairing nuclear transfer of RelB that all of these impacts result in impairment in PCa progression and increased radiosensitivity [269]. The high expression level of both PI3K/Akt and MAPK can result in radioresistance in PCa that antrocin has ability of downregulating MAPK and PI3K/Akt in enhancing radiosensitivity [270]. Table 2 and Fig. 5 summary the radioresistance mechanism in PCa cells.

8 Immune evasion in prostate cancer

Immunotherapy has been emerged as a new tool in treatment of PCa that its potential is based on stimulation of immune cells, increasing their infiltration in TME and preventing the activation of immunosuppressive pathways including PD-1/ PD-L1 axis. Upregulation of PD-L1 results in immune evasion of PCa cells. Interestingly, there is close association between immune evasion and chemoresistance in PCa cells. The endocrine therapy results in low expression level of FLII to suppress YBX1/PD-L1 axis in immune evasion. However, FLII has ability of suppressing enzalutamide resistance through regulating PD-L1-mediated immune evasion [286]. Moreover, there is association between metastasis and immune evasion of PCa cells. N-cadherin is a regulator of EMT and its upregulation stimulates EMT. The overexpression of N-cadherin mediates immune evasion in PCa cells through reducing CD8+T cells and enhancing number of CD4+/DOXP3+cells [287]. Regarding the function of PD-L1 in immune evasion, its related molecular interactions in PCa cells have been understood. The lncRNA

Table 2 The underlying mechanisms involved in radioresistance of PCa cells

Molecular network	Remark	Ref
GAS5/miR-320a/RAB21	GAS5 reduces miR-320a expression to promote RAB21 levels in mediating radiosensitivity	[271]
RelB	RelB upregulation prevents apoptosis	[272]
JMJD1A	Increase in DNA damage repair	[273]
-	The PI3K/Akt/mTOR inhibitors reduce radioresistance	[274]
LPAR3/miR-513b-5p/JPT1	CircRNA LPAR3 increases JPT1 expression through miR-513b-5p suppression in glycolysis induction and increasing radioresistance	[275]
miR-106a	miR-106a reduces LITAF expression in increasing tumourigenesis	[276]
Glucocorticoid receptor	Overexpression of GR induces radioresistance	[277]
Circ-ABCC4/miR-1253/SOX4	Circ-ABCC4 escalates SOX4 expression through miR-1253 sponging in radioresistance development	[141]
miR-301a/b/NDRG2	The miR-301a/b upregulation by hypoxia occurs to reduce NDRG2 expression in radioresistance devel- opment	[278]
AP-1	AP-1 increases proliferation and radioresistance	[279]
Circ-ZNF609/miR-501-3p/HK2	Circ-ZNF609 modulates miR-501-3p/HK2 axis to enhance glycolysis in radioresistance development	[280]
αvβ3 Integrin/survivin	$\alpha\nu\beta3$ Integrin escalates survivin expression to induce radioresistance	[281]
MEK5	Downregulation of MEK5 impairs NHEJ mechanism and increases DNA damage	[282]
PAFR/Beclin-1	PAFR reduces Beclin-1 expression in autophagy inhibition and enhancing radiosensitivity	[283]
MEK/ERK/c-Myc	Suppressing MEK/ERK/c-Myc axis increases radiosensitivity	[284]
IL-6	IL-6 increases activity of DNA repair factors to induce radioresistance	[285]

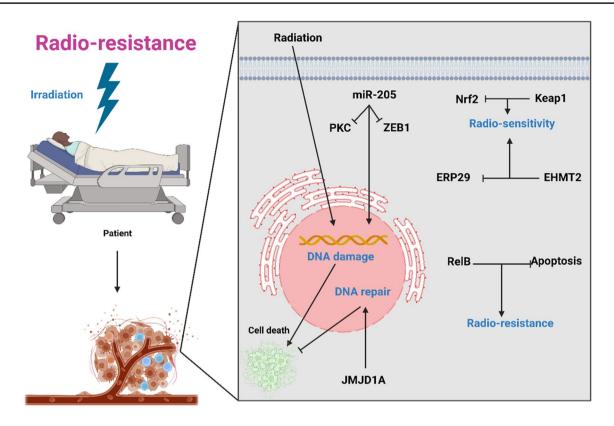


Fig. 5 The radioresistance development in PCa cells. The application of irradiation is mainly related to the DNA damage induction, while JMJD1A causes DNA damage repair to increase tumourigenesis and

KCNQ10T1 has ability of sponging miR-15a as a mechanism to increase PD-L1 expression for facilitating immune evasion in PCa [288]. In PCa patients, there is loss of JUN and ATF3 that can cause immune evasion. Interestingly, application of lipid nanostructures for delivery of JUN and ATF3 results in increase in immune surveillance for improving immunotherapy potential in PCa suppression [289].

Inflammation is considered as a factor involved in PCa pathogenesis and nitric oxide stimulates inflammation by affecting macrophages to escalate tumourigenesis [290]. The pro-inflammatory cytokines have been identified as factors involved in immune evasion. Upregulation of TLR9 and LIF secretion by PCa cells can upregulate STAT3 to increase infiltration of PMN-MDSCs in TME and mediating immunosuppression [291]. The inflammation and immune evasion have association in PCa; Porphyromonas gingivalis can cause chronic inflammation, and this is vital for promoting PD-L1 expression in triggering immune evasion in PCa [292]. The pathway in which inflammation causes immune evasion is that inflammation can promote IKKβ activation for purpose of ARID1A phosphorylation. Then, downregulation of ARID1A occurs to stimulate NF-kB axis in providing myeloid-derived suppressor cell chemotaxis [293]. Due to important function of PD-L1 in immune evasion,

prevent cell death, causing radioresistance. Moreover, apoptosis inhibition by RelB stimulates radioresistance. The upregulation of Nrf2 as redox regulator mediates radioresistance

multiple regulators including C5a receptor [294] and RelB [295] have ability of enhancing PD-L1 expression in tumour cells. More importantly, upregulation of Dickkopf-1 in PCa can prevent activation of natural killer cells in mediating immune evasion [296]. Notably, PRC1 participates in providing stemness, immune evasion, and neoangiogenesis in increasing metastasis of PCa cells [297]. Furthermore, APOE has ability of increasing TREM2 expression to increase senescence of neutrophils [298].

9 Hormone, castration, and ADT resistance in prostate cancer

Although main focus is on radio- and chemo-resistance in PCa, there are studies evaluating ability of PCa cells in development of resistance to hormone therapy and other strategies. In CRPC, HSP90 chaperone function is related to hormone resistance; HSP90 has ability of interaction with AR-FL/AR-V7 to develop hormone resistance, while bruceantin targets HSP90 in suppressing hormone resistance [299]. In addition, Hsp60 has been considered as a factor in enhancing lymph node metastasis and development of hormone resistance in PCa [300]. Even under the androgen deprivation, the PCa cells have ability of developing hormone resistance. There is close association between lipid synthesis and hormone resistance. Notably, caveolin-1 is considered as a factor in enhancing ACC1/FASIN expression and elevates lipid synthesis in development of hormone resistance [301]. On the other hand, decrease in EBP1 expression results in enhancement in carcinogenesis and mediates hormone resistance [302]. Moreover, the upregulation of Bcl-xL is observed in hormone-resistant PCa cells [303]. Even PCa cells can develop resistance to intensive hormone therapy due to loss of chromosome 10g and alterations to TP53 [304]. The interesting point is that the presence of castration-resistant PCa-like cells can cause development of hormone resistance [305]. Another challenge is castration resistance that can be caused by dysregulation of PI3K/Akt/mTOR axis [306] and interference in Hippo/YAP axis by MYBL2 [307].

The castration resistance has been a challenge into the treatment of PCa. In the advanced stages of mCRPC, there are mutations in RB1, TP53, and AR and such genomic alterations can be utilized for the diagnosis and prediction of castration resistance [308]. GREM1 has been considered as a factor causing castration resistance in PCa causing linear plasticity and mediating castration resistance. Furthermore, AR suppresses GREM1 transcription, while it is activated during ADT. Moreover, GREM1 stimulates MAPK axis through interaction with FGFR1 [309]. OBP2A is considered as a small extracellular protein that participates in transporting small and volatile molecules or the odorants via the nasal mucus to olfactory receptors. Moreover, OBP2A may function as a scavenger of toxic odours [310]. Recently, the function of OBP2A in the regulation castration resistance in PCa has been mentioned. The application of ADT causes the release of OBP2A from the tumour cells to upregulate the expression level of CXCL15/IL-8 as pro-survival factors and promoting the MDSC infiltration into the tumour microenvironment, accelerating the castration resistance in PCa [311]. A number of factors demonstrate high expression during the progression of PCa and the development of castration resistance. The AR-V7 expression is poor in primary PCa, while in almost 75% of cases, the expression of AR-V7 enhances after AD therapy and upon the exposure to abiraterone acetate or enzalutamide, its expression remarkably promotes and can be considered as a factor in the stimulation of castration resistance [312]. The stability of AR-V7 enhances due to its interaction with USP14 and USP22. However, application of nobiletin can prevent the interaction of AR-V7 with USP14 and USP22 to enhance the proteasomal degradation of AR-V7 and overcoming castration resistance [313]. The overexpression of SLC12A5 during the progression of PCa can cause the castration resistance in which SLC12A5 interacts with YTHDC1 in the nucleus to enhance HOXB13 expression, increasing the proliferation and metastasis of cancer cells as well as triggering castration resistance [314]. Moreover, the CRISPR screening has recognized RNF19A as a new factor in the development of castration resistance in PCa. AR upregulates HIF1A to increase RNF19A expression, upregulating TRIP13 expression and mediating castration resistance in PCa [315].

10 Nanotherapeutic approaches in prostate cancer

The nanotechnological approaches have been shown to be promising in PCa therapy. The nanostructures can be utilized for gene and drug delivery, immunotherapy, bioimaging, and preventing chemoresistance in PCa cells. The application of nanoparticles for delivery of cargo can remarkably disrupt tumourigenesis. The cyclodextrin-based nanostructures can be functionalized with sialic acid to deliver CSF-1R siRNA. Such nano-scale delivery results in targeted delivery of cargo and switching M2 macrophages to M1 macrophages in improving cancer immunotherapy [316]. Even in PCa cells that are resistant to DTX chemotherapy, application of curcumin-loaded nanostructures can cause cytotoxicity against tumour cells [317]. Besides, the polymeric nanostructures developed from carboxymethylcellulose are able to deliver cabazitaxel in suppressing PCa cells resistant to DTX [318]. The efficacy of current therapeutics can be significantly improved using nanostructures. The GSH-responsive nanoarchitectures are capable of co-delivery of cisplatin and AZD7762 in improving potential in PCa removal [319]. Interestingly, the hybrid nanostructures developed from lipid and polymer can be utilized for co-delivery of DTX and curcumin in effective PCa elimination [320]. It was mentioned that PCa cells have ability of developing resistance to enzalutamide; however, targeted delivery of this drug by nanostructures increases PCa removal [321]. Such nanocarriers display high biocompatibility in treatment of PCa [322, 323] and the interesting point is their ability in co-delivery of drugs and genes in cancer therapy [324].

The wealth evidence has highlighted the fact that nanostructures can improve response of PCa cells to therapy. Although GSH can be exploited as a factor in development of smart and multifunctional nanocarriers, the high levels of GSH in the tumour cells stimulate chemoresistance. For overcoming such condition, small molecule–based nanodrugs loaded with carboplatin have been developed that have 48% drug loading and great plasma stability. Such nanocarriers are able to diminish GSH levels in the tumour cells, and therefore, the ability of platinum compounds as anti-cancer drugs in chelation with DNA increases, overcoming chemoresistance [325]. The drug resistance in PCa cells can be reversed as a result of using nanostructures for co-delivery of drugs and genes. The core–shell

nanostructures have been functionalized with aptamer to deliver PTX and siRNAs. After cytoplasmic delivery of cargo, they suppressed EMT and enhanced PTX sensitivity of cancer cells [326]. Moreover, epigenetic factors such as miR-205 can be loaded in nanostructures comprised of iron oxide core decorated with PEI/PEG layers to increase its cellular uptake, stimulate apoptosis and promote drug sensitivity [327]. More importantly, targeted delivery of two anti-cancer drugs including resveratrol and DTX can be provided by PBM nanostructures in overcoming chemoresistance [328]. More importantly, nanostructures have ability of being used for radiosensitivity of PCa cells [329]. High expression of AR results in radioresistance and delivery of AR-shRNA by nanostructures suppresses this condition. For improving targeting of PCa cells, the functionalization of nanoparticles with folate has been mediated [330]. More interestingly, chemotherapy drug (DTX)-loaded titanate nanotubes have ability of increasing PCa radiosensitivity [331]. Figure 6 highlights the application of nanomaterials in therapy sensitivity.

11 Bioinformatic analysis of molecular pathways-related to carcinogenesis

We analysed GSE46602 datasets (https://www.ncbi.nlm.nih. gov/geo/) [332] and identified 135 differentially expressed genes between prostate cancer tissues and normal specimens (Fig. 7A and B). Then, we performed GO assays and found that 135 differentially expressed genes were mainly enriched in cell junction assembly, cell-substrate junction assembly, cell-substrate junction organization, collagen-containing extracellular matrix, neuron projection terminus, presynaptic active zone membrane, receptor ligand activity, signalling receptor activator activity, and sulphur compound binding (Fig. 7C). The results of KEGG assays suggested that 135 differentially expressed genes were mainly related to glutathione metabolism, malaria, drug metabolism-cytochrome P450, and ECM-receptor interaction (Fig. 7D). In addition, DO analysis revealed that 135 differentially expressed genes were mainly associated with prostate cancer, male reproductive organ cancer, cell type benign neoplasm, urinary system cancer, renal carcinoma, and kidney cancer (Fig. 7E). Then,

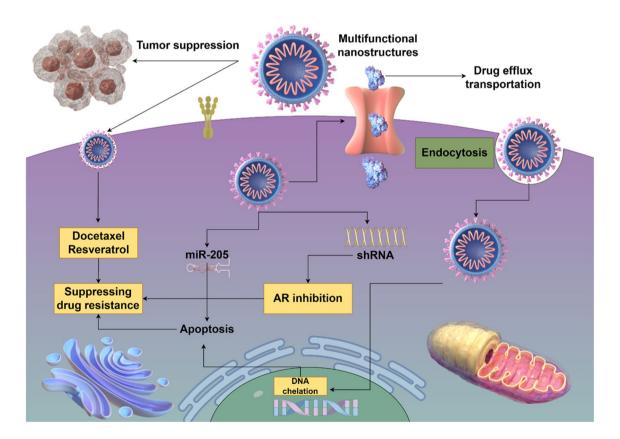
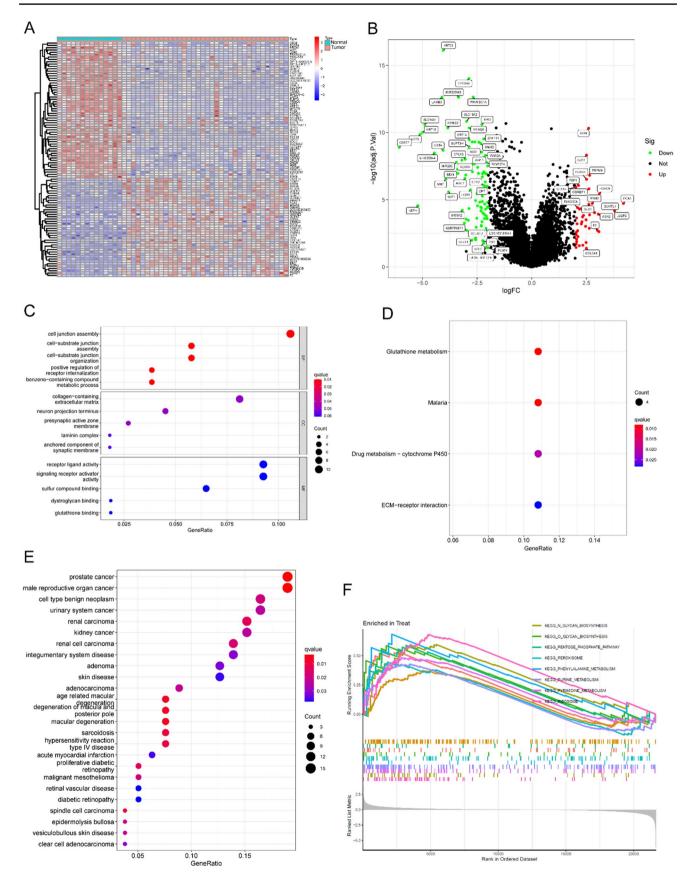


Fig. 6 The role of nanostructures for therapy sensitivity. The most important application of nanostructures is related to increasing the internalization of cargos (drugs and genes) in the tumour cells to improve potential in cancer suppression. The nanoparticles can mediate DNA chelation to cause cell death. Furthermore, the co-delivery

of docetaxel and resveratrol by nanostructures can impair the chemoresistance. The miR-205 delivery by nanoparticles can induce apoptosis in enhancing chemosensitivity. Furthermore, the nanoparticles can bypass drug efflux transporters and reduce the export of the drugs out of cancer cells



◄Fig. 7 Bioinformatics analysis of the datasets for GSE46602 with prostate cancer. A and B Differentially expressed genes between prostate cancer tissues and normal specimens, shown in Heat Map and volcano plot. C GO function assays of differentially expressed genes. D KEGG pathway assays of differentially expressed genes. E Disease assays of differentially expressed genes. F GSEA analysis for prostate cancer specimens

we performed GSEA analysis and found that the N GLY-CAN BIOSYNTHESIS, O GLYCAN BIOSYNTHESIS, PENTOSE PHOSPHATE PATHWAY, PEROXISOME, PHENYLALANINE METABOLISM, PURINE METAB-OLISM, YRIMIDINE_METABOLISM, and RIBOSOME were significantly enriched in the tumour group (Fig. 7F). Importantly, several studies have confirmed that abnormalities in glutathione metabolism may be associated with tumour development, drug resistance, and treatment response. Tumour cells are often in a state of heightened oxidative stress and require increased antioxidant capacity to counteract free radicals and oxidative damage. Glutathione, as a major intracellular antioxidant, plays a crucial role in tumour cells. By protecting cells from oxidative stress damage, glutathione may influence the sensitivity of tumour cells to treatment. In addition, some enzymes in the glutathione metabolism pathway, such as glutathione S-transferase (GST), are involved in drug metabolism and detoxification processes. Elevated activity of GST in tumour cells may enhance the metabolism and clearance of anti-cancer drugs, thereby affecting drug efficacy. Moreover, certain genes related to glutathione metabolism play a significant role in tumour drug resistance. For instance, the expression levels of genes such as glutathione reductase and glutathione peroxidase may be associated with tumour resistance to chemotherapy drugs. On the other hand, cytochrome P450 is a class of enzymes involved in drug metabolism and plays a crucial role in the transformation and clearance of drugs. The association of differentially expressed genes with the drug metabolism-cytochrome P450 pathway suggests that changes may have occurred in drug metabolism and drug responsiveness in the study subjects. This could potentially have implications for drug efficacy, drug safety, and drugdrug interactions. Our findings suggested the 135 differentially expressed genes may influence drug resistance via regulating glutathione metabolism and drug metabolismcytochrome P450.

12 Conclusion and remarks

The urological cancers are responsible for high death, even in developed countries, and among them, PCa is a threatening disease for men. The molecular profile of PC in terms of development of resistance to therapy is interesting and this resistance can occur to chemotherapy, radiotherapy, immunotherapy, and hormone therapy. The major focus of studies is on evaluating radioresistance and chemoresistance in PCa. However, underlying mechanisms involved in immune evasion and hormone resistance have been highlighted, showing that the genomic alterations can stimulate therapy failure. The changes in molecular interactions can affect a wide variety of mechanisms related to drug resistance including activity of drug transporters, DNA damage, apoptosis, and autophagy machinery. The molecular interactions are not specified to nucleus and due to changes in both cytoplasm and nucleus, and interactions at transcriptional, translational, and post-translational levels can affect chemoresistance. The major focus in radiotherapy is effect on the DNA damage in tumour cells, and when DNA damage repair occurs, the response to radiotherapy decreases. About hormonal therapy resistance, the interesting point is ability of castration-resistant PC cells in hormone resistance, showing that there are some crosslinkings between them. The bioinformatics analysis revealed some of the most dysregulated genes in therapy resistance in PC, and therefore, future studies can focus on such genes to improve ability in treatment of PCa at clinical level.

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Data availability Not applicable.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication Not applicable.

Competing interests The authors declare no conflict of interest.

References

- Silvestri, R., Nicolì, V., Gangadharannambiar, P., Crea, F., & Bootman, M. D. (2023). Calcium signalling pathways in prostate cancer initiation and progression. *Nature Reviews Urology*. https://doi.org/10.1038/s41585-023-00738-x
- Rebello, R. J., Pearson, R. B., Hannan, R. D., & Furic, L. (2017). Therapeutic approaches targeting MYC-driven prostate cancer. *Genes*, 8, 71.
- Taichman, R. S., Loberg, R. D., Mehra, R., & Pienta, K. J. (2007). The evolving biology and treatment of prostate cancer. *The Journal of clinical investigation*, 117, 2351–2361. https:// doi.org/10.1172/jci31791
- Crawford, E. D., Heidenreich, A., Lawrentschuk, N., Tombal, B., Pompeo, A. C. L., Mendoza-Valdes, A., Miller, K., Debruyne,

F. M. J., & Klotz, L. (2019). Androgen-targeted therapy in men with prostate cancer: Evolving practice and future considerations. *Prostate cancer and prostatic diseases*, 22, 24–38. https://doi.org/10.1038/s41391-018-0079-0

- Ehsani, M., David, F. O., Baniahmad, A. (2021). Androgen receptor-dependent mechanisms mediating drug resistance in prostate cancer. *Cancers*. 13. https://doi.org/10.3390/cancers130 71534
- Hoang, D. T., Iczkowski, K. A., Kilari, D., See, W., & Nevalainen, M. T. (2017). Androgen receptor-dependent and -independent mechanisms driving prostate cancer progression: Opportunities for therapeutic targeting from multiple angles. *Oncotarget*, 8, 3724–3745. https://doi.org/10.18632/oncotarget. 12554
- Merkens, L., Sailer, V., Lessel, D., Janzen, E., Greimeier, S., Kirfel, J., Perner, S., Pantel, K., Werner, S., & von Amsberg, G. (2022). Aggressive variants of prostate cancer: underlying mechanisms of neuroendocrine transdifferentiation. *Journal of experimental & clinical cancer research : CR, 41*, 46. https://doi. org/10.1186/s13046-022-02255-y
- Montironi, R., Cimadamore, A., Lopez-Beltran, A., Scarpelli, M., Aurilio, G., Santoni, M., Massari, F., Cheng, L. (2020). Morphologic, molecular and clinical features of aggressive variant prostate cancer. *Cells* 9. https://doi.org/10.3390/cells9051073
- Coleman, W. B. (2018). Chapter 25 Molecular pathogenesis of prostate cancer. In Molecular pathology (Second Edition), Coleman, W.B., Tsongalis, G.J., Eds.; Academic Press. pp. 555–568.
- Avkshtol, V., Ruth, K. J., Ross, E. A., Hallman, M. A., Greenberg, R. E., Price, R. A., Jr., Leachman, B., Uzzo, R. G., Ma, C., Chen, D., et al. (2020). Ten-year update of a randomized, prospective trial of conventional fractionated versus moderate hypofractionated radiation therapy for localized prostate cancer. *Journal of clinical oncology : Official journal of the American Society of Clinical Oncology, 38*, 1676–1684. https://doi.org/10. 1200/jco.19.01485
- Hagiwara, M., Fushimi, A., Yamashita, N., Bhattacharya, A., Rajabi, H., Long, M. D., Yasumizu, Y., Oya, M., Liu, S., & Kufe, D. (2021). MUC1-C activates the PBAF chromatin remodeling complex in integrating redox balance with progression of human prostate cancer stem cells. *Oncogene*, 40, 4930–4940. https://doi. org/10.1038/s41388-021-01899-y
- Hagiwara, M., Fushimi, A., Bhattacharya, A., Yamashita, N., Morimoto, Y., Oya, M., Withers, H. G., Hu, Q., Liu, T., Liu, S., et al. (2022). MUC1-C integrates type II interferon and chromatin remodeling pathways in immunosuppression of prostate cancer. *Oncoimmunology*, *11*, 2029298. https://doi.org/10.1080/ 2162402x.2022.2029298
- 13 Yang, Y., Liu, L., Li, M., Cheng, X., Fang, M., Zeng, Q., & Xu, Y. (2019). The chromatin remodeling protein BRG1 links ELOVL3 trans-activation to prostate cancer metastasis. *Biochimica et biophysica acta. Gene Regulatory Mechanisms, 1862*, 834–845. https://doi.org/10.1016/j.bbagrm.2019.05.005
- Zhao, D., Zhang, M., Huang, S., Liu, Q., Zhu, S., Li, Y., Jiang, W., Kiss, D. L., Cao, Q., Zhang, L., et al. (2022). CHD6 promotes broad nucleosome eviction for transcriptional activation in prostate cancer cells. *Nucleic Acids Research*, 50, 12186–12201. https://doi.org/10.1093/nar/gkac1090
- Ding, Y., Li, N., Dong, B., Guo, W., Wei, H., Chen, Q., Yuan, H., Han, Y., Chang, H., Kan, S., et al. (2019). Chromatin remodeling ATPase BRG1 and PTEN are synthetic lethal in prostate cancer. *The Journal of clinical investigation*, *129*, 759–773. https://doi. org/10.1172/jci123557
- Rajwa, P., Quhal, F., Pradere, B., Gandaglia, G., Ploussard, G., Leapman, M. S., Gore, J. L., Paradysz, A., Tilki, D., Merseburger, A. S., et al. (2023). Prostate cancer risk, screening and management in patients with germline BRCA1/2 mutations.

Nature Reviews Urology, 20, 205–216. https://doi.org/10.1038/ s41585-022-00680-4

- Loeb, S., & Giri, V. N. (2021). Clinical implications of germline testing in newly diagnosed prostate cancer. *European urology oncology*, 4, 1–9. https://doi.org/10.1016/j.euo.2020. 11.011
- Oh, M., Alkhushaym, N., Fallatah, S., Althagafi, A., Aljadeed, R., Alsowaida, Y., Jeter, J., Martin, J. R., Babiker, H. M., McBride, A., et al. (2019). The association of BRCA1 and BRCA2 mutations with prostate cancer risk, frequency, and mortality: A metaanalysis. *The Prostate*, 79, 880–895. https://doi.org/10.1002/pros. 23795
- Pritchard, C. C., Mateo, J., Walsh, M. F., De Sarkar, N., Abida, W., Beltran, H., Garofalo, A., Gulati, R., Carreira, S., Eeles, R., et al. (2016). Inherited DNA-repair gene mutations in men with metastatic prostate cancer. *The New England Journal of Medicine*, 375, 443–453. https://doi.org/10.1056/NEJMoa1603144
- Shore, N., Oliver, L., Shui, I., Gayle, A., Wong, O. Y., Kim, J., Payne, S., Amin, S., & Ghate, S. (2021). Systematic literature review of the epidemiology of advanced prostate cancer and associated homologous recombination repair gene alterations. *The Journal of urology*, 205, 977–986. https://doi.org/10.1097/ ju.000000000001570
- Wang, Z., Wang, T., Hong, D., Dong, B., Wang, Y., Huang, H., Zhang, W., Lian, B., Ji, B., Shi, H., et al. (2022). Single-cell transcriptional regulation and genetic evolution of neuroendocrine prostate cancer. *iScience*, 25, 104576. https://doi.org/10.1016/j. isci.2022.104576
- 22. Lv, S., Wu, Z., Luo, M., Zhang, Y., Zhang, J., Pascal, L. E., Wang, Z., & Wei, Q. (2022). Integrated analysis reveals FOXA1 and Ku70/Ku80 as targets of ivermectin in prostate cancer. *Cell Death & Disease*, 13, 754. https://doi.org/10.1038/ s41419-022-05182-0
- Tang, D. G. (2022). Understanding and targeting prostate cancer cell heterogeneity and plasticity. *Seminars in Cancer Biology*, 82, 68–93. https://doi.org/10.1016/j.semcancer.2021.11.001
- Cyrta, J., Prandi, D., Arora, A., Hovelson, D. H., Sboner, A., Rodriguez, A., Fedrizzi, T., Beltran, H., Robinson, D. R., Gopalan, A., et al. (2022). Comparative genomics of primary prostate cancer and paired metastases: Insights from 12 molecular case studies. *The Journal of pathology*, 257, 274–284. https://doi.org/ 10.1002/path.5887
- Sun, G., Ma, S., Zheng, Z., Wang, X., Chen, S., Chang, T., Liang, Z., Jiang, Y., Xu, S., & Liu, R. (2022). Multi-omics analysis of expression and prognostic value of NSUN members in prostate cancer. *Frontiers in Oncology*, *12*, 965571. https://doi.org/10. 3389/fonc.2022.965571
- 26. Jia, D., Zhou, Z., Kwon, O. J., Zhang, L., Wei, X., Zhang, Y., Yi, M., Roudier, M. P., Regier, M. C., Dumpit, R., et al. (2022). Stromal FOXF2 suppresses prostate cancer progression and metastasis by enhancing antitumor immunity. *Nature Communications*, *13*, 6828. https://doi.org/10.1038/s41467-022-34665-z
- Chang, M., He, Y., Liu, C., Lin, R., Huang, X., Liang, D., Zhang, J., & Lu, Y. (2022). Downregulation of SEPTIN5 inhibits prostate cancer progression by increasing CD8(+) T cell infiltration. *International Journal of Biological Sciences*, 18, 6035–6051. https://doi.org/10.7150/ijbs.76573
- Ding, L., Wang, R., Zheng, Q., Shen, D., Wang, H., Lu, Z., Luo, W., Xie, H., Ren, L., Jiang, M., et al. (2022). circPDE5A regulates prostate cancer metastasis via controlling WTAP-dependent N6-methyladenisine methylation of EIF3C mRNA. *Journal of Experimental & Clinical Cancer Research : CR, 41*, 187. https:// doi.org/10.1186/s13046-022-02391-5
- 29. Li, Z., Li, B., Yu, H., Wang, P., Wang, W., Hou, P., Li, M., Chu, S., Zheng, J., Mao, L., et al. (2022). DNMT1-mediated epigenetic silencing of TRAF6 promotes prostate

cancer tumorigenesis and metastasis by enhancing EZH2 stability. *Oncogene*, *41*, 3991–4002. https://doi.org/10.1038/ s41388-022-02404-9

- Vasan, N., Baselga, J., & Hyman, D. M. (2019). A view on drug resistance in cancer. *Nature*, 575, 299–309. https://doi.org/10. 1038/s41586-019-1730-1
- Goldie, J. H., & Coldman, A. J. (1984). The genetic origin of drug resistance in neoplasms: Implications for systemic therapy. *Cancer Research*, 44, 3643–3653.
- 32. Fisher, B., Slack, N. H., & Bross, I. D. (1969). Cancer of the breast: Size of neoplasm and prognosis. *Cancer*, 24, 1071–1080. https://doi.org/10.1002/1097-0142(196911)24:5%3c1071::aidcncr2820240533%3e3.0.co;2-h
- Skipper, H. E., Schabel, F. M., Jr., & Wilcox, W. S. (1964). Experimental evaluation of potential anticancer agents. XIII. On the criteria and kinetics associated with "curability" of experimental leukemia. *Cancer Chemotherapy Reports*, 35, 1–111.
- Goldie, J. H., & Coldman, A. J. (1979). A mathematic model for relating the drug sensitivity of tumors to their spontaneous mutation rate. *Cancer Treatment Reports*, 63, 1727–1733.
- Luria, S. E., & Delbrück, M. (1943). Mutations of bacteria from virus sensitivity to virus resistance. *Genetics*, 28, 491–511. https://doi.org/10.1093/genetics/28.6.491
- Tannock, I. F. (2015). Cancer: Resistance through repopulation. Nature, 517, 152–153. https://doi.org/10.1038/nature14075
- 37 Nowell, P. C. (1976). The clonal evolution of tumor cell populations. *Science (New York, N.Y.), 194*, 23–28. https://doi.org/10. 1126/science.959840
- Alexandrov, L. B., Nik-Zainal, S., Wedge, D. C., Aparicio, S. A., Behjati, S., Biankin, A. V., Bignell, G. R., Bolli, N., Borg, A., Børresen-Dale, A. L., et al. (2013). Signatures of mutational processes in human cancer. *Nature*, 500, 415–421. https://doi.org/10.1038/nature12477
- Lengauer, C., Kinzler, K. W., & Vogelstein, B. (1998). Genetic instabilities in human cancers. *Nature*, 396, 643–649. https://doi. org/10.1038/25292
- Stephens, P. J., Greenman, C. D., Fu, B., Yang, F., Bignell, G. R., Mudie, L. J., Pleasance, E. D., Lau, K. W., Beare, D., Stebbings, L. A., et al. (2011). Massive genomic rearrangement acquired in a single catastrophic event during cancer development. *Cell*, 144, 27–40. https://doi.org/10.1016/j.cell.2010.11.055
- Sansregret, L., Vanhaesebroeck, B., & Swanton, C. (2018). Determinants and clinical implications of chromosomal instability in cancer. *Nature Reviews. Clinical Oncology*, *15*, 139–150. https://doi.org/10.1038/nrclinonc.2017.198
- 42 Jain, R. K. (2005). Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science (New York, N.Y.)*, 307, 58–62. https://doi.org/10.1126/science.1104819
- Minchinton, A. I., & Tannock, I. F. (2006). Drug penetration in solid tumours. *Nature Reviews. Cancer*, 6, 583–592. https://doi. org/10.1038/nrc1893
- Makker, V., Rasco, D., Vogelzang, N. J., Brose, M. S., Cohn, A. L., Mier, J., Di Simone, C., Hyman, D. M., Stepan, D. E., Dutcus, C. E., et al. (2019). Lenvatinib plus pembrolizumab in patients with advanced endometrial cancer: An interim analysis of a multicentre, open-label, single-arm, phase 2 trial. *The Lancet. Oncology*, 20, 711–718. https://doi.org/10.1016/s1470-2045(19) 30020-8
- 45. Rini, B. I., Plimack, E. R., Stus, V., Gafanov, R., Hawkins, R., Nosov, D., Pouliot, F., Alekseev, B., Soulières, D., Melichar, B., et al. (2019). Pembrolizumab plus axitinib versus sunitinib for advanced renal-cell carcinoma. *The New England Journal* of Medicine, 380, 1116–1127. https://doi.org/10.1056/NEJMo a1816714
- Liu, H. J., & Xu, P. (2022). Strategies to overcome/penetrate the BBB for systemic nanoparticle delivery to the brain/brain tumor.

Advanced Drug Delivery Reviews, 191, 114619. https://doi.org/ 10.1016/j.addr.2022.114619

- Holohan, C., Van Schaeybroeck, S., Longley, D. B., & Johnston, P. G. (2013). Cancer drug resistance: An evolving paradigm. *Nature Reviews Cancer*, 13, 714–726. https://doi.org/10.1038/ nrc3599
- Mirzaei, S., Gholami, M. H., Hashemi, F., Zabolian, A., Farahani, M. V., Hushmandi, K., Zarrabi, A., Goldman, A., Ashrafizadeh, M., & Orive, G. (2022). Advances in understanding the role of P-gp in doxorubicin resistance: Molecular pathways, therapeutic strategies, and prospects. *Drug Discovery Today*, 27, 436–455. https://doi.org/10.1016/j.drudis.2021.09.020
- Gottesman, M. M., Fojo, T., & Bates, S. E. (2002). Multidrug resistance in cancer: Role of ATP–dependent transporters. *Nature Reviews Cancer*, 2, 48–58.
- Ambudkar, S. V., Dey, S., Hrycyna, C. A., Ramachandra, M., Pastan, I., & Gottesman, M. M. (1999). Biochemical, cellular, and pharmacological aspects of the multidrug transporter. *Annual Review of Pharmacology and Toxicology*, 39, 361–398.
- Choi, C.-H. (2005). ABC transporters as multidrug resistance mechanisms and the development of chemosensitizers for their reversal. *Cancer Cell International*, 5, 1–13.
- Thomas, H., & Coley, H. M. (2003). Overcoming multidrug resistance in cancer: An update on the clinical strategy of inhibiting p-glycoprotein. *Cancer Control*, 10, 159–165.
- 53. Pusztai, L., Wagner, P., Ibrahim, N., Rivera, E., Theriault, R., Booser, D., Symmans, F. W., Wong, F., Blumenschein, G., & Fleming, D. R. (2005). Phase II study of tariquidar, a selective P-glycoprotein inhibitor, in patients with chemotherapy-resistant, advanced breast carcinoma. *Cancer*, 104, 682–691.
- Meijer, C., Mulder, N. H., Timmer-Bosscha, H., Sluiter, W. J., Meersma, G. J., & de Vries, E. G. (1992). Relationship of cellular glutathione to the cytotoxicity and resistance of seven platinum compounds. *Cancer Research*, *52*, 6885–6889.
- Schwartz, P. M., Moir, R. D., Hyde, C. M., Turek, P. J., & Handschumacher, R. E. (1985). Role of uridine phosphorylase in the anabolism of 5-fluorouracil. *Biochemical Pharmacology*, 34, 3585–3589.
- Houghton, J. A., & Houghton, P. J. (1983). Elucidation of pathways of 5-fluorouracil metabolism in xenografts of human colorectal adenocarcinoma. *European Journal of Cancer and Clinical Oncology*, 19, 807–815.
- Malet-Martino, M., & Martino, R. (2002). Clinical studies of three oral prodrugs of 5-fluorouracil (capecitabine, UFT, S-1): a review. *The Oncologist*, 7, 288–323.
- Kosuri, K., Wu, X., Wang, L., Villalona-Calero, M., & Otterson, G. (2010). An epigenetic mechanism for capecitabine resistance in mesothelioma. *Biochemical and Biophysical Research Communications*, 391, 1465–1470.
- Bélanger, A.-S., Tojcic, J., Harvey, M., & Guillemette, C. (2010). Regulation of UGT1A1 and HNF1 transcription factor gene expression by DNA methylation in colon cancer cells. *BMC Molecular Biology*, 11, 1–11.
- 60. Toffoli, G., Cecchin, E., Gasparini, G., D'Andrea, M., Azzarello, G., Basso, U., Mini, E., Pessa, S., De Mattia, E., & Lo Re, G. (2010). Genotype-driven phase I study of irinotecan administered in combination with fluorouracil/leucovorin in patients with metastatic colorectal cancer. *Journal of Clinical Oncology*, 28, 866–871.
- Ward, R. A., Fawell, S., Floc'h, N., Flemington, V., McKerrecher, D., & Smith, P. D. (2020). Challenges and opportunities in cancer drug resistance. *Chemical Reviews*, *121*, 3297–3351.
- Joyce, H., McCann, A., Clynes, M., & Larkin, A. (2015). Influence of multidrug resistance and drug transport proteins on chemotherapy drug metabolism. *Expert Opinion on Drug Metabolism & Toxicology, 11*, 795–809.

- Wood, K. C. (2015). Mapping the pathways of resistance to targeted therapies. *Cancer Research*, 75, 4247–4251.
- Kandoth, C., McLellan, M. D., Vandin, F., Ye, K., Niu, B., Lu, C., Xie, M., Zhang, Q., McMichael, J. F., & Wyczalkowski, M. A. (2013). Mutational landscape and significance across 12 major cancer types. *Nature*, 502, 333–339.
- 65. Luo, J., Emanuele, M. J., Li, D., Creighton, C. J., Schlabach, M. R., Westbrook, T. F., Wong, K.-K., & Elledge, S. J. (2009). A genome-wide RNAi screen identifies multiple synthetic lethal interactions with the Ras oncogene. *Cell*, *137*, 835–848.
- Zhang, T., Brazhnik, P., & Tyson, J. J. (2009). Computational analysis of dynamical responses to the intrinsic pathway of programmed cell death. *Biophysical Journal*, 97, 415–434.
- Pommier, Y., Sordet, O., Antony, S., Hayward, R. L., & Kohn, K. W. (2004). Apoptosis defects and chemotherapy resistance: Molecular interaction maps and networks. *Oncogene*, 23, 2934–2949.
- Zhivotovsky, B., & Orrenius, S. (2003). Defects in the apoptotic machinery of cancer cells: Role in drug resistance. In: Proceedings of the Seminars in cancer biology, pp. 125–134.
- Indran, I. R., Tufo, G., Pervaiz, S., & Brenner, C. (2011). Recent advances in apoptosis, mitochondria and drug resistance in cancer cells. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1807, 735–745.
- Liang, X., Zhang, H., Wang, Z., Zhang, X., Dai, Z., Zhang, J., Luo, P., Zhang, L., Hu, J., Liu, Z., et al. (2022). JMJD8 Is an M2 macrophage biomarker, and it associates with DNA damage repair to facilitate stemness maintenance, chemoresistance, and immunosuppression in pan-cancer. *Frontiers in immunology*, *13*, 875786. https://doi.org/10.3389/fimmu.2022.875786
- Zhou, W., Xu, Y., Zhang, J., Zhang, P., Yao, Z., Yan, Z., Wang, H., Chu, J., Yao, S., Zhao, S., et al. (2022). MiRNA-363-3p/ DUSP10/JNK axis mediates chemoresistance by enhancing DNA damage repair in diffuse large B-cell lymphoma. *Leukemia*, *36*, 1861–1869. https://doi.org/10.1038/s41375-022-01565-6
- Su, Y., Wu, C., Chang, Y., Li, L., Chen, Y., Jia, X., Wang, X., Lv, Y., Yu, B., & Yuan, J. (2022). USP17L2-SIRT7 axis regulates DNA damage repair and chemoresistance in breast cancer cells. *Breast Cancer Research and Treatment, 196*, 31–44. https://doi. org/10.1007/s10549-022-06711-3
- Tang, C., Qiu, S., Mou, W., Xu, J., & Wang, P. (2022). Excessive activation of HOXB13/PIMREG axis promotes hepatocellular carcinoma progression and drug resistance. *Biochemical and Biophysical Research Communications*, 623, 81–88. https://doi. org/10.1016/j.bbrc.2022.07.066
- Nicholson, H. A., Sawers, L., Clarke, R. G., Hiom, K. J., Ferguson, M. J., & Smith, G. (2022). Fibroblast growth factor signalling influences homologous recombination-mediated DNA damage repair to promote drug resistance in ovarian cancer. *British Journal of Cancer*, *127*, 1340–1351. https://doi.org/10.1038/ s41416-022-01899-z
- Zhang, C., Chen, L., Sun, L., Jin, H., Ren, K., Liu, S., Qian, Y., Li, S., Li, F., Zhu, C., et al. (2023). BMAL1 collaborates with CLOCK to directly promote DNA double-strand break repair and tumor chemoresistance. *Oncogene*, 42, 967–979. https://doi.org/ 10.1038/s41388-023-02603-y
- Jin, J., Ma, M., Shi, S., Wang, J., Xiao, P., Yu, H. F., Zhang, C., Guo, Q., Yu, Z., Lou, Z., et al. (2022). Copper enhances genotoxic drug resistance via ATOX1 activated DNA damage repair. *Cancer Letters*, 536, 215651. https://doi.org/10.1016/j.canlet. 2022.215651
- 77. Yang, L., Shi, P., Zhao, G., Xu, J., Peng, W., Zhang, J., Zhang, G., Wang, X., Dong, Z., Chen, F., et al. (2020). Targeting cancer stem cell pathways for cancer therapy. *Signal Transduction and Targeted Therapy*, 5, 8. https://doi.org/10.1038/s41392-020-0110-5

- Chen, W., Dong, J., Haiech, J., Kilhoffer, M. C., & Zeniou, M. (2016). Cancer stem cell quiescence and plasticity as major challenges in cancer therapy. *Stem Cells International*, 2016, 1740936. https://doi.org/10.1155/2016/1740936
- 79. Lapidot, T., Sirard, C., Vormoor, J., Murdoch, B., Hoang, T., Caceres-Cortes, J., Minden, M., Paterson, B., Caligiuri, M. A., & Dick, J. E. (1994). A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature*, *367*, 645–648. https://doi.org/10.1038/367645a0
- Bonnet, D., & Dick, J. E. (1997). Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nature Medicine*, *3*, 730–737. https://doi.org/ 10.1038/nm0797-730
- Shimokawa, M., Ohta, Y., Nishikori, S., Matano, M., Takano, A., Fujii, M., Date, S., Sugimoto, S., Kanai, T., & Sato, T. (2017). Visualization and targeting of LGR5(+) human colon cancer stem cells. *Nature*, 545, 187–192. https://doi.org/10.1038/natur e22081
- Shibata, M., Hoque, M. O. (2019). Targeting cancer stem cells: A strategy for effective eradication of cancer. *Cancers*, 11. https:// doi.org/10.3390/cancers11050732
- Visvader, J. E., & Lindeman, G. J. (2012). Cancer stem cells: Current status and evolving complexities. *Cell Stem Cell*, 10, 717–728. https://doi.org/10.1016/j.stem.2012.05.007
- Ajani, J. A., Song, S., Hochster, H. S., & Steinberg, I. B. (2015). Cancer stem cells: The promise and the potential. *Seminars in Oncology*, 42(Suppl 1), S3-17. https://doi.org/10.1053/j.semin oncol.2015.01.001
- Chen, P. M., Wong, C. N., Wong, C. N., Chu, P. Y. (2023). Actinlike protein 6A expression correlates with cancer stem cell-like features and poor prognosis in ovarian cancer. *International Journal of Molecular Sciences*, 24. https://doi.org/10.3390/ijms2 4032016
- Sun, K., Shen, H., He, S., & Liu, Y. (2022). MASM inhibits cancer stem cell-like characteristics of EpCAM(+) cells via AKT/ GSK3β/β-catenin signaling. *American Journal of Translational Research*, 14, 8380–8389.
- Chen, Y., Yang, Z., He, X., Zhu, W., Wang, Y., Li, J., Han, Z., Wen, J., Liu, W., Yang, Y., et al. (2023). Proanthocyanidins inhibited colorectal cancer stem cell characteristics through Wnt/β-catenin signaling. *Environmental Toxicology*. https://doi. org/10.1002/tox.23924
- Gonzalez-Callejo, P., Guo, Z., Ziglari, T., Claudio, N. M., Nguyen, K. H., Oshimori, N., Seras-Franzoso, J., & Pucci, F. (2023). Cancer stem cell-derived extracellular vesicles preferentially target MHC-II-macrophages and PD1+ T cells in the tumor microenvironment. *PLoS ONE*, *18*, e0279400. https://doi.org/10. 1371/journal.pone.0279400
- Liao, W., Zhang, L., Chen, X., Xiang, J., Zheng, Q., Chen, N., Zhao, M., Zhang, G., Xiao, X., Zhou, G., et al. (2023). Targeting cancer stem cells and signalling pathways through phytochemicals: A promising approach against colorectal cancer. *Phytomedicine : International Journal of Phytotherapy and Phytopharmacology*, *108*, 154524. https://doi.org/10.1016/j.phymed.2022. 154524
- Ashrafizadeh, M., Zhang, W., Zou, R., Sethi, G., Klionsky, D. J., & Zhang, X. (2023). A bioinformatics analysis, pre-clinical and clinical conception of autophagy in pancreatic cancer: Complexity and simplicity in crosstalk. *Pharmacological Research*, 194, 106822. https://doi.org/10.1016/j.phrs.2023.106822
- 91. Qin, Y., Ashrafizadeh, M., Mongiardini, V., Grimaldi, B., Crea, F., Rietdorf, K., Győrffy, B., Klionsky, D. J., Ren, J., Zhang, W., et al. (2023). Autophagy and cancer drug resistance in dialogue: Pre-clinical and clinical evidence. *Cancer Letters*, 570, 216307. https://doi.org/10.1016/j.canlet.2023.216307

- 92. Zhang, S., Liu, X., Abdulmomen Ali Mohammed, S., Li, H., Cai, W., Guan, W., Liu, D., Wei, Y., Rong, D., Fang, Y., et al. (2022). Adaptor SH3BGRL drives autophagy-mediated chemoresistance through promoting PIK3C3 translation and ATG12 stability in breast cancers. *Autophagy*, *18*, 1822–1840. https://doi.org/10. 1080/15548627.2021.2002108
- 93. Luo, M., Deng, X., Chen, Z., & Hu, Y. (2023). Circular RNA circPOFUT1 enhances malignant phenotypes and autophagyassociated chemoresistance via sequestrating miR-488-3p to activate the PLAG1-ATG12 axis in gastric cancer. *Cell Death & Disease*, 14, 10. https://doi.org/10.1038/s41419-022-05506-0
- 94. Zhang, S. R., Li, J., Chen, J. X., Chen, G., Chen, J. Y., Fu, H. W., & Zhou, B. (2022). SMC4 enhances the chemoresistance of hepatoma cells by promoting autophagy. *Annals of Translational Medicine*, 10, 1308. https://doi.org/10.21037/atm-22-3623
- 95. Sun, Y., Shen, W., Hu, S., Lyu, Q., Wang, Q., Wei, T., Zhu, W., & Zhang, J. (2023). METTL3 promotes chemoresistance in small cell lung cancer by inducing mitophagy. *Journal of Experimental & Clinical Cancer Research : CR*, 42, 65. https://doi.org/10. 1186/s13046-023-02638-9
- 96. Shen, Z., Zhou, L., Zhang, C., & Xu, J. (2020). Reduction of circular RNA Foxo3 promotes prostate cancer progression and chemoresistance to docetaxel. *Cancer Letters*, 468, 88–101. https://doi.org/10.1016/j.canlet.2019.10.006
- 97. Zhou, Q., Chen, X., He, H., Peng, S., Zhang, Y., Zhang, J., Cheng, L., Liu, S., Huang, M., Xie, R., et al. (2021). WD repeat domain 5 promotes chemoresistance and programmed deathligand 1 expression in prostate cancer. *Theranostics*, 11, 4809– 4824. https://doi.org/10.7150/thno.55814
- Bai, L., Li, X., Yang, Y., Zhao, R., White, E. Z., Danaher, A., Bowen, N. J., Hinton, C. V., Cook, N., Li, D., et al. (2023). Bromocriptine monotherapy overcomes prostate cancer chemoresistance in preclinical models. *Translational Oncology*, 34, 101707. https://doi.org/10.1016/j.tranon.2023.101707
- 99. Xie, C., Wang, Z., Ba, Y., Aguilar, J., Kyan, A., Zhong, L., & Hao, J. (2023). BMP signaling inhibition overcomes chemoresistance of prostate cancer. *American Journal of Cancer Research*, *13*, 4073–4086.
- 100. Ma, B., Shao, H., Jiang, X., Wang, Z., Wu, C. C., Whaley, D., & Wells, A. (2021). Akt isoforms differentially provide for chemoresistance in prostate cancer. *Cancer Biology & Medicine*, 19, 635–650. https://doi.org/10.20892/j.issn.2095-3941.2020.0747
- Orellana-Serradell, O., Herrera, D., Castellón, E. A., & Contreras, H. R. (2019). The transcription factor ZEB1 promotes chemoresistance in prostate cancer cell lines. *Asian Journal of Andrology*, 21, 460–467. https://doi.org/10.4103/aja.aja_1_19
- 102. Wang, W., Wang, L., Mizokami, A., Shi, J., Zou, C., Dai, J., Keller, E. T., Lu, Y., & Zhang, J. (2017). Down-regulation of E-cadherin enhances prostate cancer chemoresistance via Notch signaling. *Chinese Journal of Cancer*, 36, 35. https://doi.org/10. 1186/s40880-017-0203-x
- 103. Li, X., Gera, L., Zhang, S., Chen, Y., Lou, L., Wilson, L. M., Xie, Z. R., Sautto, G., Liu, D., Danaher, A., et al. (2021). Pharmacological inhibition of noncanonical EED-EZH2 signaling overcomes chemoresistance in prostate cancer. *Theranostics*, 11, 6873–6890. https://doi.org/10.7150/thno.49235
- 104. Luo, S., Shao, L., Chen, Z., Hu, D., Jiang, L., & Tang, W. (2020). NPRL2 promotes docetaxel chemoresistance in castration resistant prostate cancer cells by regulating autophagy through the mTOR pathway. *Experimental Cell Research*, 390, 111981. https://doi.org/10.1016/j.yexcr.2020.111981
- 105. Liu, C., Li, Z., Bi, L., Li, K., Zhou, B., Xu, C., Huang, J., & Xu, K. (2014). NOTCH1 signaling promotes chemoresistance via regulating ABCC1 expression in prostate cancer stem cells. *Molecular and Cellular Biochemistry*, 393, 265–270. https://doi. org/10.1007/s11010-014-2069-4

- 106. Biernacka, K. M., Uzoh, C. C., Zeng, L., Persad, R. A., Bahl, A., Gillatt, D., Perks, C. M., & Holly, J. M. (2013). Hyperglycaemiainduced chemoresistance of prostate cancer cells due to IGFBP2. *Endocrine-Related Cancer*, 20, 741–751. https://doi.org/10.1530/ erc-13-0077
- 107. Kato, T., Fujita, Y., Nakane, K., Kojima, T., Nozawa, Y., Deguchi, T., & Ito, M. (2012). ETS1 promotes chemoresistance and invasion of paclitaxel-resistant, hormone-refractory PC3 prostate cancer cells by up-regulating MDR1 and MMP9 expression. *Biochemical and Biophysical Research Communications*, 417, 966–971. https://doi.org/10.1016/j.bbrc.2011.12.047
- Stafford, M. Y. C., McKenna, D. J. (2023). MiR-182 is upregulated in prostate cancer and contributes to tumor progression by targeting MITF. *International Journal of Molecular Sciences*, 24. https://doi.org/10.3390/ijms24031824
- 109. Bozgeyik, E., Arslan, A., Temiz, E., Batar, B., Koyuncu, I., & Tozkir, H. (2022). miR-320a promotes p53-dependent apoptosis of prostate cancer cells by negatively regulating TP73-AS1 invitro. *Biochemical and Biophysical Research Communications*, 619, 130–136. https://doi.org/10.1016/j.bbrc.2022.06.034
- 110. Wang, C., Li, W., Hu, Q., Feng, N., Liu, C., Shi, N., Chen, S., Chen, M., Guan, H., You, Z., et al. (2022). Transgenic construction and functional miRNA analysis identify the role of miR-7 in prostate cancer suppression. *Oncogene*, 41, 4645–4657. https:// doi.org/10.1038/s41388-022-02461-0
- 111. Wang, Y., Lieberman, R., Pan, J., Zhang, Q., Du, M., Zhang, P., Nevalainen, M., Kohli, M., Shenoy, N. K., Meng, H., et al. (2016). miR-375 induces docetaxel resistance in prostate cancer by targeting SEC23A and YAP1. *Molecular Cancer*, 15, 70. https://doi.org/10.1186/s12943-016-0556-9
- 112. Fujita, Y., Kojima, K., Hamada, N., Ohhashi, R., Akao, Y., Nozawa, Y., Deguchi, T., & Ito, M. (2008). Effects of miR-34a on cell growth and chemoresistance in prostate cancer PC3 cells. *Biochemical and Biophysical Research Communications*, 377, 114–119. https://doi.org/10.1016/j.bbrc.2008.09.086
- 113. Gujrati, H., Ha, S., Waseem, M., Wang, B. D. (2022). Downregulation of miR-99b-5p and upregulation of nuclear mTOR cooperatively promotes the tumor aggressiveness and drug resistance in African American prostate cancer. *International Journal* of Molecular Sciences, 23. https://doi.org/10.3390/ijms23179643
- 114. Paskeh, M. D. A., Entezari, M., Mirzaei, S., Zabolian, A., Saleki, H., Naghdi, M. J., Sabet, S., Khoshbakht, M. A., Hashemi, M., Hushmandi, K., et al. (2022). Emerging role of exosomes in cancer progression and tumor microenvironment remodeling. *Journal of Hematology & Oncology*, 15, 83. https://doi.org/10. 1186/s13045-022-01305-4
- 115. Cao, Z., Xu, L., & Zhao, S. (2019). Exosome-derived miR-27a produced by PSC-27 cells contributes to prostate cancer chemoresistance through p53. *Biochemical and Biophysical Research Communications*, 515, 345–351. https://doi.org/10.1016/j.bbrc. 2019.05.120
- 116. Shi, G. H., Ye, D. W., Yao, X. D., Zhang, S. L., Dai, B., Zhang, H. L., Shen, Y. J., Zhu, Y., Zhu, Y. P., Xiao, W. J., et al. (2010). Involvement of microRNA-21 in mediating chemo-resistance to docetaxel in androgen-independent prostate cancer PC3 cells. *Acta Pharmacologica Sinica*, 31, 867–873. https://doi.org/10. 1038/aps.2010.48
- 117. Zhu, J., Qin, P., Cao, C., Dai, G., Xu, L., & Yang, D. (2021). Use of miR-145 and testicular nuclear receptor 4 inhibition to reduce chemoresistance to docetaxel in prostate cancer. *Oncology Reports*, 45, 963–974. https://doi.org/10.3892/or.2021.7925
- Zhou, P., Ma, L., Zhou, J., Jiang, M., Rao, E., Zhao, Y., & Guo, F. (2016). miR-17-92 plays an oncogenic role and conveys chemoresistance to cisplatin in human prostate cancer cells. *International Journal of Oncology*, 48, 1737–1748. https://doi.org/10. 3892/ijo.2016.3392

- Zhong, J., Huang, R., Su, Z., Zhang, M., Xu, M., Gong, J., Chen, N., Zeng, H., Chen, X., & Zhou, Q. (2017). Downregulation of miR-199a-5p promotes prostate adeno-carcinoma progression through loss of its inhibition of HIF-1α. *Oncotarget*, *8*, 83523– 83538. https://doi.org/10.18632/oncotarget.18315
- 120. Tseng, J. C., Huang, S. H., Lin, C. Y., Wang, B. J., Huang, S. F., Shen, Y. Y., & Chuu, C. P. (2020). ROR2 suppresses metastasis of prostate cancer via regulation of miR-199a-5p-PIAS3-AKT2 signaling axis. *Cell Death & Disease*, 11, 376. https://doi.org/ 10.1038/s41419-020-2587-9
- 121. Chen, L., Cao, H., & Feng, Y. (2018). MiR-199a suppresses prostate cancer paclitaxel resistance by targeting YES1. World Journal of Urology, 36, 357–365. https://doi.org/10.1007/ s00345-017-2143-0
- 122. Xu, X., Wang, Y., Deng, H., Liu, C., Wu, J., & Lai, M. (2018). HMGA2 enhances 5-fluorouracil chemoresistance in colorectal cancer via the Dvl2/Wnt pathway. *Oncotarget*, 9, 9963–9974. https://doi.org/10.18632/oncotarget.24133
- 123. Wu, C., Miao, C., Tang, Q., Zhou, X., Xi, P., Chang, P., Hua, L., & Ni, H. (2020). MiR-129-5p promotes docetaxel resistance in prostate cancer by down-regulating CAMK2N1 expression. *Journal of Cellular and Molecular Medicine*, 24, 2098–2108. https://doi.org/10.1111/jcmm.14050
- 124. Yang, Z., Chen, J. S., Wen, J. K., Gao, H. T., Zheng, B., Qu, C. B., Liu, K. L., Zhang, M. L., Gu, J. F., Li, J. D., et al. (2017). Silencing of miR-193a-5p increases the chemosensitivity of prostate cancer cells to docetaxel. *Journal of Experimental & Clinical Cancer Research : CR*, 36, 178. https://doi.org/10.1186/s13046-017-0649-3
- 125. Mirzaei, S., Paskeh, M. D. A., Okina, E., Gholami, M. H., Hushmandi, K., Hashemi, M., Kalu, A., Zarrabi, A., Nabavi, N., Rabiee, N., et al. (2022). Molecular landscape of LncR-NAs in prostate cancer: A focus on pathways and therapeutic targets for intervention. *Journal of Experimental & Clinical Cancer Research : CR*, 41, 214. https://doi.org/10.1186/ s13046-022-02406-1
- 126. Liu, B., Li, X., Wang, D., Yu, Y., Lu, D., Chen, L., Lv, F., Li, Y., Cheng, L., Song, Y., et al. (2022). CEMIP promotes extracellular matrix-detached prostate cancer cell survival by inhibiting ferroptosis. *Cancer Science*, 113, 2056–2070. https://doi.org/10. 1111/cas.15356
- 127. Cheng, L., He, Q., Liu, B., Chen, L., Lv, F., Li, X., Li, Y., Liu, C., Song, Y., & Xing, Y. (2023). SGK2 promotes prostate cancer metastasis by inhibiting ferroptosis via upregulating GPX4. *Cell Death & Disease*, 14, 74. https://doi.org/10.1038/ s41419-023-05614-5
- 128. Jiang, X., Guo, S., Xu, M., Ma, B., Liu, R., Xu, Y., & Zhang, Y. (2022). TFAP2C-mediated lncRNA PCAT1 inhibits ferroptosis in docetaxel-resistant prostate cancer through c-Myc/miR-25-3p/ SLC7A11 signaling. *Frontiers in Oncology*, *12*, 862015. https:// doi.org/10.3389/fonc.2022.862015
- 129. Jiang, H., Xiong, W., Chen, L., Lv, Z., Yang, C., & Li, Y. (2019). Knockdown of the long noncoding RNA HOTTIP inhibits cell proliferation and enhances cell sensitivity to cisplatin by suppressing the Wnt/β-catenin pathway in prostate cancer. *Journal* of Cellular Biochemistry, 120, 8965–8974. https://doi.org/10. 1002/jcb.27851
- 130. Cao, C., Sun, G., & Liu, C. (2020). Long non-coding RNA SNHG6 regulates the sensitivity of prostate cancer cells to paclitaxel by sponging miR-186. *Cancer Cell International*, 20, 381. https://doi.org/10.1186/s12935-020-01462-x
- 131. Xing, Z., Li, S., Xing, J., Yu, G., Wang, G., & Liu, Z. (2022). Silencing of LINC01963 enhances the chemosensitivity of prostate cancer cells to docetaxel by targeting the miR-216b-5p/ TrkB axis. *Laboratory Investigation; A Journal of Technical*

Methods and Pathology, 102, 602-612. https://doi.org/10.1038/ s41374-022-00736-4

- 132. Gao, W., Lin, S., Cheng, C., Zhu, A., Hu, Y., Shi, Z., Zhang, X., & Hong, Z. (2019). Long non-coding RNA CASC2 regulates Sprouty2 via functioning as a competing endogenous RNA for miR-183 to modulate the sensitivity of prostate cancer cells to docetaxel. *Archives of Biochemistry and Biophysics*, 665, 69–78. https://doi.org/10.1016/j.abb.2018.01.013
- 133. Li, X., Han, X., Wei, P., Yang, J., & Sun, J. (2020). Knockdown of lncRNA CCAT1 enhances sensitivity of paclitaxel in prostate cancer via regulating miR-24-3p and FSCN1. *Cancer Biology & Therapy*, 21, 452–462. https://doi.org/10.1080/15384047.2020. 1727700
- 134. Wang, C., Ding, T., Yang, D., Zhang, P., Hu, X., Qin, W., & Zheng, J. (2021). The lncRNA OGFRP1/miR-149-5p/IL-6 axis regulates prostate cancer chemoresistance. *Pathology, Research* and Practice, 224, 153535. https://doi.org/10.1016/j.prp.2021. 153535
- 135. Shi, T., Li, R., Duan, P., Guan, Y., Zhang, D., Ding, Z., & Ruan, X. (2022). TRPM2-AS promotes paclitaxel resistance in prostate cancer by regulating FOXK1 via sponging miR-497-5p. *Drug Development Research*, 83, 967–978. https://doi.org/10.1002/ddr. 21924
- 136. Wang, Y. Y., & Chen, C. (2022). lncRNA-DANCR promotes Taxol resistance of prostate cancer cells through modulating the miR-33b-5p-LDHA axis. *Disease Markers*, 2022, 9516774. https://doi.org/10.1155/2022/9516774
- 137. Gu, P., Chen, X., Xie, R., Han, J., Xie, W., Wang, B., Dong, W., Chen, C., Yang, M., Jiang, J., et al. (2017). IncRNA HOXD-AS1 regulates proliferation and chemo-resistance of castrationresistant prostate cancer via recruiting WDR5. *Molecular therapy : The Journal of the American Society of Gene Therapy*, 25, 1959–1973. https://doi.org/10.1016/j.ymthe.2017.04.016
- Ding, X., Sun, J., & Zhang, X. (2022). Circ_0076305 facilitates prostate cancer development via sponging miR-411-5p and regulating PGK1. *Andrologia*, 54, e14406. https://doi.org/10.1111/ and.14406
- Zhang, G., Liu, Y., Yang, J., Wang, H., & Xing, Z. (2022). Inhibition of circ_0081234 reduces prostate cancer tumor growth and metastasis via the miR-1/MAP 3 K1 axis. *The Journal of Gene Medicine*, 24, e3376. https://doi.org/10.1002/jgm.3376
- 140. Cai, F., Li, J., Zhang, J., & Huang, S. (2022). Knockdown of Circ_CCNB2 sensitizes prostate cancer to radiation through repressing autophagy by the miR-30b-5p/KIF18A axis. *Cancer Biotherapy & Radiopharmaceuticals*, 37, 480–493. https://doi. org/10.1089/cbr.2019.3538
- 141. Yu, T., Du, H., & Sun, C. (2023). Circ-ABCC4 contributes to prostate cancer progression and radioresistance by mediating miR-1253/SOX4 cascade. *Anti-Cancer Drugs*, 34, 155–165. https://doi.org/10.1097/cad.00000000001361
- 142. Zhang, Y., Liu, F., Feng, Y., Xu, X., Wang, Y., Zhu, S., Dong, J., Zhao, S., Xu, B., & Feng, N. (2022). CircRNA circ_0006156 inhibits the metastasis of prostate cancer by blocking the ubiquitination of S100A9. *Cancer Gene Therapy*, 29, 1731–1741. https://doi.org/10.1038/s41417-022-00492-z
- 143. Chen, L., Song, Y., Hou, T., Li, X., Cheng, L., Li, Y., & Xing, Y. (2022). Circ_0004087 interaction with SND1 promotes docetaxel resistance in prostate cancer by boosting the mitosis error correction mechanism. *Journal of Experimental & Clinical Cancer Research : CR*, 41, 194. https://doi.org/10.1186/ s13046-022-02404-3
- 144. Gao, Y., Liu, J., Huan, J., & Che, F. (2020). Downregulation of circular RNA hsa_circ_0000735 boosts prostate cancer sensitivity to docetaxel via sponging miR-7. *Cancer Cell International*, 20, 334. https://doi.org/10.1186/s12935-020-01421-6

- 145. Gu, H., & Duan, Z. (2022). Silencing of circDPP4 suppresses cell progression of human prostate cancer and enhances docetaxel cytotoxicity through regulating the miR-564/ZIC2 axis. *The Journal Of Gene Medicine*, 24, e3403. https://doi.org/10. 1002/jgm.3403
- 146. Zheng, P., Gao, H., Xie, X., & Lu, P. (2022). Plasma exosomal hsa_circ_0015286 as a potential diagnostic and prognostic biomarker for gastric cancer. *Pathology Oncology Research : POR*, 28, 1610446. https://doi.org/10.3389/pore.2022.1610446
- 147. Zheng, Y., Li, P., Ma, J., Yang, C., Dai, S., & Zhao, C. (2022). Cancer-derived exosomal circ_0038138 enhances glycolysis, growth, and metastasis of gastric adenocarcinoma via the miR-198/EZH2 axis. *Translational Oncology*, 25, 101479. https://doi. org/10.1016/j.tranon.2022.101479
- 148. Tan, X., Song, X., Fan, B., Li, M., Zhang, A., & Pei, L. (2022). Exosomal circRNA Scm-like with four malignant brain tumor domains 2 (circ-SFMBT2) enhances the docetaxel resistance of prostate cancer via the microRNA-136-5p/tribbles homolog 1 pathway. *Anti-Cancer Drugs*, *33*, 871–882. https://doi.org/10. 1097/cad.00000000001365
- 149. Chen, H., Li, H., & Chen, Q. (2016). INPP4B reverses docetaxel resistance and epithelial-to-mesenchymal transition via the PI3K/ Akt signaling pathway in prostate cancer. *Biochemical and Biophysical Research Communications*, 477, 467–472. https://doi. org/10.1016/j.bbrc.2016.06.073
- 150. Ni, J., Cozzi, P., Hao, J., Beretov, J., Chang, L., Duan, W., Shigdar, S., Delprado, W., Graham, P., Bucci, J., et al. (2013). Epithelial cell adhesion molecule (EpCAM) is associated with prostate cancer metastasis and chemo/radioresistance via the PI3K/Akt/ mTOR signaling pathway. *The International Journal of Biochemistry & Cell Biology*, 45, 2736–2748. https://doi.org/10.1016/j. biocel.2013.09.008
- 151. Sharma, P. K., Singh, R., Novakovic, K. R., Eaton, J. W., Grizzle, W. E., & Singh, S. (2010). CCR9 mediates PI3K/AKT-dependent antiapoptotic signals in prostate cancer cells and inhibition of CCR9-CCL25 interaction enhances the cytotoxic effects of etoposide. *International Journal of Cancer*, *127*, 2020–2030. https://doi.org/10.1002/ijc.25219
- 152. Lee, J. T., Jr., Steelman, L. S., & McCubrey, J. A. (2004). Phosphatidylinositol 3'-kinase activation leads to multidrug resistance protein-1 expression and subsequent chemoresistance in advanced prostate cancer cells. *Cancer research*, 64, 8397–8404. https://doi.org/10.1158/0008-5472.Can-04-1612
- 153. Hour, T. C., Chung, S. D., Kang, W. Y., Lin, Y. C., Chuang, S. J., Huang, A. M., Wu, W. J., Huang, S. P., Huang, C. Y., & Pu, Y. S. (2015). EGFR mediates docetaxel resistance in human castrationresistant prostate cancer through the Akt-dependent expression of ABCB1 (MDR1). Archives of Toxicology, 89, 591–605. https:// doi.org/10.1007/s00204-014-1275-x
- 154. Ni, J., Cozzi, P. J., Hao, J. L., Beretov, J., Chang, L., Duan, W., Shigdar, S., Delprado, W. J., Graham, P. H., Bucci, J., et al. (2014). CD44 variant 6 is associated with prostate cancer metastasis and chemo-/radioresistance. *The Prostate*, 74, 602–617. https://doi.org/10.1002/pros.22775
- 155. Zhou, W., Su, Y., Zhang, Y., Han, B., Liu, H., & Wang, X. (2020). Endothelial cells promote docetaxel resistance of prostate cancer cells by inducing ERG expression and activating Akt/ mTOR signaling pathway. *Frontiers in Oncology*, 10, 584505. https://doi.org/10.3389/fonc.2020.584505
- 156. Song, C., Zhang, J., Liu, X., Li, M., Wang, D., Kang, Z., Yu, J., Chen, J., Pan, H., Wang, H., et al. (2022). PTEN loss promotes Warburg effect and prostate cancer cell growth by inducing FBP1 degradation. *Frontiers in Oncology*, *12*, 911466. https://doi.org/ 10.3389/fonc.2022.911466
- 157. Ferraldeschi, R., Nava Rodrigues, D., Riisnaes, R., Miranda, S., Figueiredo, I., Rescigno, P., Ravi, P., Pezaro, C., Omlin,

A., Lorente, D., et al. (2015). PTEN protein loss and clinical outcome from castration-resistant prostate cancer treated with abiraterone acetate. *European urology*, *67*, 795–802. https://doi.org/10.1016/j.eururo.2014.10.027

- 158. Hong, Z., Wu, G., Xiang, Z. D., Xu, C. D., Huang, S. S., Li, C., Shi, L., & Wu, D. L. (2019). KDM5C is transcriptionally regulated by BRD4 and promotes castration-resistance prostate cancer cell proliferation by repressing PTEN. *Biomedicine & Pharmacotherapy = Biomedecine & Pharmacotherapie*, 114, 108793. https://doi.org/10.1016/j.biopha.2019.108793
- Yuan, X. J., & Whang, Y. E. (2002). PTEN sensitizes prostate cancer cells to death receptor-mediated and drug-induced apoptosis through a FADD-dependent pathway. *Oncogene*, 21, 319–327. https://doi.org/10.1038/sj.onc.1205054
- 160. Huang, H., Cheville, J. C., Pan, Y., Roche, P. C., Schmidt, L. J., & Tindall, D. J. (2001). PTEN induces chemosensitivity in PTEN-mutated prostate cancer cells by suppression of Bcl-2 expression. *The Journal of Biological Chemistry*, 276, 38830–38836. https://doi.org/10.1074/jbc.M103632200
- 161. Zhao, W., Ning, L., Wang, L., Ouyang, T., Qi, L., Yang, R., & Wu, Y. (2021). miR-21 inhibition reverses doxorubicinresistance and inhibits PC3 human prostate cancer cells proliferation. *Andrologia*, 53, e14016. https://doi.org/10.1111/and. 14016
- 162. Sekino, Y., Han, X., Kawaguchi, T., Babasaki, T., Goto, K., Inoue, S., Hayashi, T., Teishima, J., Shiota, M., Yasui, W. et al. (2019). TUBB3 reverses resistance to docetaxel and cabazitaxel in prostate cancer. *International Journal of Molecular Sciences*, 20. https://doi.org/10.3390/ijms20163936
- 163. Sekino, Y., Han, X., Babasaki, T., Miyamoto, S., Kobatake, K., Kitano, H., Ikeda, K., Goto, K., Inoue, S., Hayashi, T., et al. (2021). TUBB3 is associated with PTEN, neuroendocrine differentiation, and castration resistance in prostate cancer. *Urologic Oncology*, 39(368), e361-368.e369. https://doi.org/10.1016/j. urolonc.2021.03.001
- 164. Thalappil, M. A., Butturini, E., Carcereri de Prati, A., Bettin, I., Antonini, L., Sapienza, F. U., Garzoli, S., Ragno, R., Mariotto, S. (2022). Pinus mugo essential oil impairs STAT3 activation through oxidative stress and induces apoptosis in prostate cancer cells. *Molecules (Basel, Switzerland)*, 27. https://doi.org/10. 3390/molecules27154834
- 165 Zhang, K., Yin, W., Ma, L., Liu, Z., & Li, Q. (2023). HSPB8 facilitates prostate cancer progression via activating the JAK/ STAT3 signaling pathway. *Biochemistry and Cell Biology = Biochimie et Biologie Cellulaire*, 101, 1–11. https://doi.org/10.1139/ bcb-2022-0205
- 166. Li, X., He, S., Liang, W., Zhang, W., Chen, X., Li, Q., Yang, X., Liu, Y., Zhu, D., Li, L., et al. (2023). Marsdenia tenacissima injection induces the apoptosis of prostate cancer by regulating the AKT/GSK3β/STAT3 signaling axis. *Chinese Journal of Natural Medicines*, 21, 113–126. https://doi.org/10.1016/s1875-5364(23)60389-9
- 167. Ji, Y., Liu, B., Chen, L., Li, A., Shen, K., Su, R., Zhang, W., Zhu, Y., Wang, Q., & Xue, W. (2023). Repurposing ketotifen as a therapeutic strategy for neuroendocrine prostate cancer by targeting the IL-6/STAT3 pathway. *Cellular Oncology (Dordrecht)*. https://doi.org/10.1007/s13402-023-00822-9
- 168. Zhong, W., Wu, K., Long, Z., Zhou, X., Zhong, C., Wang, S., Lai, H., Guo, Y., Lv, D., Lu, J., et al. (2022). Gut dysbiosis promotes prostate cancer progression and docetaxel resistance via activating NF-κB-IL6-STAT3 axis. *Microbiome*, 10, 94. https://doi.org/ 10.1186/s40168-022-01289-w
- 169. Cheteh, E. H., Sarne, V., Ceder, S., Bianchi, J., Augsten, M., Rundqvist, H., Egevad, L., Östman, A., & Wiman, K. G. (2020). Interleukin-6 derived from cancer-associated fibroblasts attenuates the p53 response to doxorubicin in prostate

cancer cells. *Cell Death Discovery*, *6*, 42. https://doi.org/10. 1038/s41420-020-0272-5

- 170. Liu, C., Zhu, Y., Lou, W., Cui, Y., Evans, C. P., & Gao, A. C. (2014). Inhibition of constitutively active Stat3 reverses enzalutamide resistance in LNCaP derivative prostate cancer cells. *The Prostate*, 74, 201–209. https://doi.org/10.1002/pros.22741
- 171. Fu, Z., Zhao, P. Y., Yang, X. P., Li, H., Hu, S. D., Xu, Y. X., & Du, X. H. (2023). Cannabidiol regulates apoptosis and autophagy in inflammation and cancer: A review. *Frontiers in Pharmacology*, 14, 1094020. https://doi.org/10.3389/fphar. 2023.1094020
- 172. Thomas, S., & Shah, G. (2005). Calcitonin induces apoptosis resistance in prostate cancer cell lines against cytotoxic drugs via the Akt/survivin pathway. *Cancer Biology & Therapy*, 4, 1226–1233. https://doi.org/10.4161/cbt.4.11.2093
- 173. Wu, G., Wang, J., Chen, G., & Zhao, X. (2017). microRNA-204 modulates chemosensitivity and apoptosis of prostate cancer cells by targeting zinc-finger E-box-binding homeobox 1 (ZEB1). *American Journal of Translational Research*, 9, 3599–3610.
- 174. Chen, L., Cai, J., Huang, Y., Tan, X., Guo, Q., Lin, X., Zhu, C., Zeng, X., Liu, H., & Wu, X. (2020). Identification of cofilin-1 as a novel mediator for the metastatic potentials and chemoresistance of the prostate cancer cells. *European Journal of Pharmacology*, 880, 173100. https://doi.org/10.1016/j.ejphar.2020. 173100
- 175. Sun, M. Y., Xu, B., Wu, Q. X., Chen, W. L., Cai, S., Zhang, H., & Tang, Q. F. (2021). Cisplatin-resistant gastric cancer cells promote the chemoresistance of cisplatin-sensitive cells via the exosomal RPS3-mediated PI3K-Akt-Cofilin-1 signaling axis. *Frontiers in Cell and Developmental Biology*, 9, 618899. https:// doi.org/10.3389/fcell.2021.618899
- 176. Hsu, W. C., Ramesh, S., Shibu, M. A., Chen, M. C., Wang, T. F., Day, C. H., Chen, R. J., Padma, V. V., Li, C. C., Tseng, Y. C., et al. (2021). Platycodin D reverses histone deacetylase inhibitor resistance in hepatocellular carcinoma cells by repressing ERK1/2-mediated cofilin-1 phosphorylation. *Phytomedicine : International Journal of Phytotherapy and Phytopharmacology*, 82, 153442. https://doi.org/10.1016/j.phymed.2020.153442
- 177. Xiao, G., Wang, X., & Yu, Y. (2017). CXCR4/Let-7a axis regulates metastasis and chemoresistance of pancreatic cancer cells through targeting HMGA2. *Cellular Physiology and Biochemistry : International Journal of Experimental Cellular Physiology, Biochemistry, and Pharmacology, 43*, 840–851. https://doi.org/10.1159/000481610
- 178. Dong, J., Wang, R., Ren, G., Li, X., Wang, J., Sun, Y., Liang, J., Nie, Y., Wu, K., Feng, B., et al. (2017). HMGA2-FOXL2 axis regulates metastases and epithelial-to-mesenchymal transition of chemoresistant gastric cancer. *Clinical Cancer Research : An Official Journal of the American Association for Cancer Research*, 23, 3461–3473. https://doi.org/10.1158/1078-0432. Ccr-16-2180
- 179. Cai, J., Shen, G., Liu, S., & Meng, Q. (2016). Downregulation of HMGA2 inhibits cellular proliferation and invasion, improves cellular apoptosis in prostate cancer. *Tumour Biology : The Journal of the International Society for Oncodevelopmental Biology and Medicine*, 37, 699–707. https://doi.org/10.1007/ s13277-015-3853-9
- 180. Liu, Y., Xu, X., Lin, P., He, Y., Zhang, Y., Cao, B., Zhang, Z., Sethi, G., Liu, J., Zhou, X., et al. (2019). Inhibition of the deubiquitinase USP9x induces pre-B cell homeobox 1 (PBX1) degradation and thereby stimulates prostate cancer cell apoptosis. *The Journal of Biological Chemistry*, 294, 4572–4582. https://doi. org/10.1074/jbc.RA118.006057
- 181. Ye, Q. F., Zhang, Y. C., Peng, X. Q., Long, Z., Ming, Y. Z., & He, L. Y. (2012). siRNA-mediated silencing of Notch-1 enhances docetaxel induced mitotic arrest and apoptosis in prostate cancer

cells. Asian Pacific Journal of Cancer Prevention : APJCP, 13, 2485–2489. https://doi.org/10.7314/apjcp.2012.13.6.2485

- 182. Mancini, M. C. S., Morelli, A. P., Severino, M. B., Pavan, I. C. B., Zambalde, É. P., Góis, M. M., Silva, L., Quintero-Ruiz, N., Romeiro, C. F., Dos Santos, D. F. G., et al. (2022). Knockout of NRF2 triggers prostate cancer cells death through ROS modulation and sensitizes to cisplatin. *Journal of Cellular Biochemistry*, 123, 2079–2092. https://doi.org/10.1002/jcb.30333
- 183. Zhang, Y., Wang, Y., Yuan, J., Qin, W., Liu, F., Wang, F., Zhang, G., & Yang, X. (2012). Toll-like receptor 4 ligation confers chemoresistance to docetaxel on PC-3 human prostate cancer cells. *Cell Biology and Toxicology*, 28, 269–277. https://doi.org/ 10.1007/s10565-012-9221-2
- Qian, S., Zhang, S., Wu, Y., Ding, Y., & Li, X. (2020). Protein disulfide isomerase 4 drives docetaxel resistance in prostate cancer. *Chemotherapy*, 65, 125–133. https://doi.org/10.1159/00051 1505
- 185. Liu, J., Chen, Z., Guo, J., Wang, L., Liu, X. (2019). Ambra1 induces autophagy and desensitizes human prostate cancer cells to cisplatin. *Bioscience Reports*, 39. https://doi.org/10.1042/ bsr20170770
- 186. Tang, Q., Fang, J., Lai, W., Hu, Y., Liu, C., Hu, X., Song, C., Cheng, T., Liu, R., & Huang, X. (2022). Hippo pathway monomerizes STAT3 to regulate prostate cancer growth. *Cancer Science*, 113, 2753–2762. https://doi.org/10.1111/cas.15463
- 187. Sheng, W., Ding, J., Liu, L., Wang, N., Lu, B., You, X., He, Q., & Zhou, Q. (2022). Curcumol inhibits the development of prostate cancer by miR-125a/STAT3 axis. *Evidence-based Complementary and Alternative Medicine : ECAM*, 2022, 9317402. https:// doi.org/10.1155/2022/9317402
- 188. Yu, C., Fan, Y., Zhang, Y., Liu, L., & Guo, G. (2022). LINC00893 inhibits the progression of prostate cancer through miR-3173-5p/SOCS3/JAK2/STAT3 pathway. *Cancer Cell International*, 22, 228. https://doi.org/10.1186/s12935-022-02637-4
- 189. Hu, F., Zhao, Y., Yu, Y., Fang, J. M., Cui, R., Liu, Z. Q., Guo, X. L., & Xu, Q. (2018). Docetaxel-mediated autophagy promotes chemoresistance in castration-resistant prostate cancer cells by inhibiting STAT3. *Cancer Letters*, 416, 24–30. https://doi.org/ 10.1016/j.canlet.2017.12.013
- 190. Chang, P. C., Wang, T. Y., Chang, Y. T., Chu, C. Y., Lee, C. L., Hsu, H. W., Zhou, T. A., Wu, Z., Kim, R. H., Desai, S. J., et al. (2014). Autophagy pathway is required for IL-6 induced neuroendocrine differentiation and chemoresistance of prostate cancer LNCaP cells. *PLoS ONE*, *9*, e88556. https://doi.org/10. 1371/journal.pone.0088556
- 191. Liao, H., Xiao, Y., Hu, Y., Xiao, Y., Yin, Z., Liu, L., Kang, X., & Chen, Y. (2016). Methylation-induced silencing of miR-34a enhances chemoresistance by directly upregulating ATG4Binduced autophagy through AMPK/mTOR pathway in prostate cancer. *Oncology Reports*, 35, 64–72. https://doi.org/10.3892/or. 2015.4331
- 192. Zeng, J., Liu, W., Fan, Y. Z., He, D. L., & Li, L. (2018). PrLZ increases prostate cancer docetaxel resistance by inhibiting LKB1/AMPK-mediated autophagy. *Theranostics*, 8, 109–123. https://doi.org/10.7150/thno.20356
- 193. Xie, J., Chen, X., Wang, W., Guan, Z., Hou, J., & Lin, J. (2022). Long non-coding RNA PCDRInc1 confers docetaxel resistance in prostate cancer by promoting autophagy. *Journal of Cancer*, 13, 2138–2149. https://doi.org/10.7150/jca.65329
- 194. Lamprou, I., Tsolou, A., Kakouratos, C., Mitrakas, A. G., Xanthopoulou, E. T., Kassela, K., Karakasiliotis, I., Zois, C. E., Giatromanolaki, A., & Koukourakis, M. I. (2021). Suppressed PLIN3 frequently occurs in prostate cancer, promoting docetaxel resistance via intensified autophagy, an event reversed by chloroquine. *Medical Oncology (Northwood, London, England), 38*, 116. https://doi.org/10.1007/s12032-021-01566-y

- 195. Li, M., Chen, X., Wang, X., Wei, X., Wang, D., Liu, X., Xu, L., Batu, W., Li, Y., Guo, B., et al. (2021). RSL3 enhances the antitumor effect of cisplatin on prostate cancer cells via causing glycolysis dysfunction. *Biochemical Pharmacology*, 192, 114741. https://doi.org/10.1016/j.bcp.2021.114741
- 196. Martínez-Martínez, D., Soto, A., Gil-Araujo, B., Gallego, B., Chiloeches, A., & Lasa, M. (2019). Resveratrol promotes apoptosis through the induction of dual specificity phosphatase 1 and sensitizes prostate cancer cells to cisplatin. Food and Chemical Toxicology : An International Journal Published for the British Industrial Biological Research Association, 124, 273–279. https://doi.org/10.1016/j.fct.2018.12.014
- 197. Zhu, Q., Li, H., Liu, Y., & Jiang, L. (2017). Knockdown of CFTR enhances sensitivity of prostate cancer cells to cisplatin via inhibition of autophagy. *Neoplasma*, 64, 709–717. https:// doi.org/10.4149/neo_2017_508
- 198. Kim, K. Y., Yun, U. J., Yeom, S. H., Kim, S. C., Lee, H. J., Ahn, S.C., Park, K. I., Kim, Y. W. (2021). Inhibition of autophagy promotes hemistepsin A-induced apoptosis via reactive oxygen species-mediated AMPK-dependent signaling in human prostate cancer cells. *Biomolecules*, 11. https://doi.org/10.3390/biom11121806
- 199. Lin, J. Z., Wang, W. W., Hu, T. T., Zhu, G. Y., Li, L. N., Zhang, C. Y., Xu, Z., Yu, H. B., Wu, H. F., & Zhu, J. G. (2020). FOXM1 contributes to docetaxel resistance in castrationresistant prostate cancer by inducing AMPK/mTOR-mediated autophagy. *Cancer Letters*, 469, 481–489. https://doi.org/10. 1016/j.canlet.2019.11.014
- 200. Chen, Z., Jiang, Q., Zhu, P., Chen, Y., Xie, X., Du, Z., Jiang, L., & Tang, W. (2019). NPRL2 enhances autophagy and the resistance to Everolimus in castration-resistant prostate cancer. *The Prostate*, 79, 44–53. https://doi.org/10.1002/pros.23709
- 201. Chen, J., Xu, D., Wang, T., Yang, Z., Yang, Y., He, K., Zhao, L. (2022). HMGB1 promotes the development of castration-resistant prostate cancer by regulating androgen receptor activation. *Oncology Reports*, 48. https://doi.org/10.3892/or. 2022.8412
- 202. Lv, D. J., Song, X. L., Huang, B., Yu, Y. Z., Shu, F. P., Wang, C., Chen, H., Zhang, H. B., & Zhao, S. C. (2019). HMGB1 promotes prostate cancer development and metastasis by interacting with Brahma-related gene 1 and activating the Akt signaling pathway. *Theranostics*, 9, 5166–5182. https://doi.org/10.7150/thno.33972
- 203. Jung, A. R., Kim, G. E., Kim, M. Y., Ha, U. S., Hong, S. H., Lee, J. Y., Kim, S. W., & Park, Y. H. (2021). HMGB1 promotes tumor progression and invasion through HMGB1/TNFR1/NF-κB axis in castration-resistant prostate cancer. *American Journal of Cancer Research*, 11, 2215–2227.
- 204. Zhang, Y. X., Yuan, Y. Q., Zhang, X. Q., Huang, D. L., Wei, Y. Y., & Yang, J. G. (2017). HMGB1-mediated autophagy confers resistance to gemcitabine in hormone-independent prostate cancer cells. *Oncology Letters*, 14, 6285–6290. https://doi.org/10. 3892/ol.2017.6965
- 205. Yu, Y., Yang, F. H., Zhang, W. T., Guo, Y. D., Ye, L., & Yao, X. D. (2021). Mesenchymal stem cells desensitize castrationresistant prostate cancer to docetaxel chemotherapy via inducing TGF-β1-mediated cell autophagy. *Cell & Bioscience*, 11, 7. https://doi.org/10.1186/s13578-020-00494-0
- 206. Wang, Q., He, W. Y., Zeng, Y. Z., Hossain, A., & Gou, X. (2018). Inhibiting autophagy overcomes docetaxel resistance in castration-resistant prostate cancer cells. *International Urology and Nephrology*, 50, 675–686. https://doi.org/10.1007/ s11255-018-1801-5
- 207. Nguyen, H. G., Yang, J. C., Kung, H. J., Shi, X. B., Tilki, D., Lara, P. N., Jr., DeVere White, R. W., Gao, A. C., & Evans, C. P. (2014). Targeting autophagy overcomes Enzalutamide resistance in castration-resistant prostate cancer cells and improves

therapeutic response in a xenograft model. *Oncogene*, *33*, 4521–4530. https://doi.org/10.1038/onc.2014.25

- 208. Peng, K., Sun, A., Zhu, J., Gao, J., Li, Y., Shao, G., Yang, W., & Lin, Q. (2021). Restoration of the ATG5-dependent autophagy sensitizes DU145 prostate cancer cells to chemotherapeutic drugs. *Oncology Letters*, 22, 638. https://doi.org/10.3892/ol. 2021.12899
- 209. Mortezavi, A., Salemi, S., Kranzbühler, B., Gross, O., Sulser, T., Simon, H. U., & Eberli, D. (2019). Inhibition of autophagy significantly increases the antitumor effect of Abiraterone in prostate cancer. *World Journal of Urology*, 37, 351–358. https:// doi.org/10.1007/s00345-018-2385-5
- Losada-García, A., Salido-Guadarrama, I., Cortes-Ramirez, S. A., Cruz-Burgos, M., Morales-Pacheco, M., Vazquez-Santillan, K., Rodriguez-Martinez, G., González-Ramírez, I., Gonzalez-Covarrubias, V., Perez-Plascencia, C., et al. (2023). SFRP1 induces a stem cell phenotype in prostate cancer cells. *Frontiers* in Cell and Developmental Biology, 11, 1096923. https://doi.org/ 10.3389/fcell.2023.1096923
- 211. Song, X. L., Huang, B., Zhou, B. W., Wang, C., Liao, Z. W., Yu, Y., & Zhao, S. C. (2018). miR-1301–3p promotes prostate cancer stem cell expansion by targeting SFRP1 and GSK3β. *Biomedicine & Pharmacotherapy* = *Biomedecine & Pharmacotherapie*, 99, 369–374. https://doi.org/10.1016/j.biopha.2018.01.086
- 212. Pérez, G., López-Moncada, F., Indo, S., Torres, M. J., Castellón, E. A., Contreras, H. R. (2021). Knockdown of ZEB1 reverses cancer stem cell properties in prostate cancer cells. *Oncology Reports*, 45. https://doi.org/10.3892/or.2021.8009
- 213. Cheng, J. W., Duan, L. X., Yu, Y., Wang, P., Feng, J. L., Feng, G. Z., & Liu, Y. (2021). Bone marrow mesenchymal stem cells promote prostate cancer cell stemness via cell-cell contact to activate the Jagged1/Notch1 pathway. *Cell & Bioscience*, 11, 87. https://doi.org/10.1186/s13578-021-00599-0
- 214. Zhang, Y., He, L., Sadagopan, A., Ma, T., Dotti, G., Wang, Y., Zheng, H., Gao, X., Wang, D., DeLeo, A. B., et al. (2021). Targeting radiation-resistant prostate cancer stem cells by B7–H3 CAR T cells. *Molecular Cancer Therapeutics*, 20, 577–588. https://doi.org/10.1158/1535-7163.mct-20-0446
- 215. Castellón, E. A., Indo, S., Contreras, H. R. (2022). Cancer stemness/epithelial-mesenchymal transition axis influences metastasis and castration resistance in prostate cancer: Potential therapeutic target. *International Journal of Molecular Sciences*, 23. https://doi.org/10.3390/ijms232314917
- Li, P., Yang, R., & Gao, W. Q. (2014). Contributions of epithelial-mesenchymal transition and cancer stem cells to the development of castration resistance of prostate cancer. *Molecular Cancer, 13*, 55. https://doi.org/10.1186/1476-4598-13-55
- 217. Yan, J., & Tang, D. (2014). Prostate cancer stem-like cells proliferate slowly and resist etoposide-induced cytotoxicity via enhancing DNA damage response. *Experimental Cell Research*, 328, 132–142. https://doi.org/10.1016/j.yexcr.2014.08.016
- 218. Jeter, C. R., Liu, B., Liu, X., Chen, X., Liu, C., Calhoun-Davis, T., Repass, J., Zaehres, H., Shen, J. J., & Tang, D. G. (2011). NANOG promotes cancer stem cell characteristics and prostate cancer resistance to androgen deprivation. *Oncogene*, 30, 3833– 3845. https://doi.org/10.1038/onc.2011.114
- Ramesh, V., Brabletz, T., & Ceppi, P. (2020). Targeting EMT in cancer with repurposed metabolic inhibitors. *Trends in Cancer*, *6*, 942–950. https://doi.org/10.1016/j.trecan.2020.06.005
- 220. Guo, Z., Ashrafizadeh, M., Zhang, W., Zou, R., Sethi, G., & Zhang, X. (2023). Molecular profile of metastasis, cell plasticity and EMT in pancreatic cancer: A pre-clinical connection to aggressiveness and drug resistance. *Cancer Metastasis Reviews*. https://doi.org/10.1007/s10555-023-10125-y
- He, P., Dai, Q., & Wu, X. (2023). New insight in urological cancer therapy: From epithelial-mesenchymal transition (EMT)

to application of nano-biomaterials. *Environmental Research*, 229, 115672. https://doi.org/10.1016/j.envres.2023.115672

- 222. Zhang, Z. H., Liu, M. D., Yao, K., Xu, S., Yu, D. X., Xie, D. D., & Xu, D. X. (2023). Vitamin D deficiency aggravates growth and metastasis of prostate cancer through promoting EMT in two β-catenin-related mechanisms. *The Journal of Nutritional Biochemistry*, 111, 109177. https://doi.org/10. 1016/j.jnutbio.2022.109177
- 223. Qiao, P., & Tian, Z. (2022). Atractylenolide I inhibits EMT and enhances the antitumor effect of cabozantinib in prostate cancer via targeting Hsp27. *Frontiers in Oncology*, *12*, 1084884. https://doi.org/10.3389/fonc.2022.1084884
- 224. Pang, X., Zhang, J., He, X., Gu, Y., Qian, B. Z., Xie, R., Yu, W., Zhang, X., Li, T., Shi, X., et al. (2021). SPP1 promotes enzalutamide resistance and epithelial-mesenchymal-transition activation in castration-resistant prostate cancer via PI3K/AKT and ERK1/2 pathways. Oxidative Medicine and Cellular Longevity, 2021, 5806602. https://doi.org/10.1155/2021/5806602
- 225. Chen, B., Zhang, Y., Li, C., Xu, P., Gao, Y., & Xu, Y. (2021). CNTN-1 promotes docetaxel resistance and epithelial-tomesenchymal transition via the PI3K/Akt signaling pathway in prostate cancer. *Archives of Medical Science : AMS, 17*, 152–165. https://doi.org/10.5114/aoms.2020.92939
- 226. Rajput, M., Singh, R., Singh, N., & Singh, R. P. (2021). EGFRmediated Rad51 expression potentiates intrinsic resistance in prostate cancer via EMT and DNA repair pathways. *Life Sciences*, 286, 120031. https://doi.org/10.1016/j.lfs.2021.120031
- 227. Choi, J. D., Kim, T. J., Jeong, B. C., Jeon, H. G., Jeon, S. S., Kang, M. Y., Yeom, S. Y., & Seo, S. I. (2021). ISL1 promotes enzalutamide resistance in castration-resistant prostate cancer (CRPC) through epithelial to mesenchymal transition (EMT). *Scientific Reports*, *11*, 21984. https://doi.org/10.1038/s41598-021-01003-0
- 228. Zhang, B., Li, Y., Wu, Q., Xie, L., Barwick, B., Fu, C., Li, X., Wu, D., Xia, S., Chen, J., et al. (2021). Acetylation of KLF5 maintains EMT and tumorigenicity to cause chemoresistant bone metastasis in prostate cancer. *Nature Communications*, *12*, 1714. https://doi.org/10.1038/s41467-021-21976-w
- 229. Chen, D., Chou, F. J., Chen, Y., Huang, C. P., Tian, H., Wang, Y., Niu, Y., You, B., Yeh, S., Xing, N., et al. (2022). Targeting the radiation-induced ARv7-mediated circNHS/miR-512-5p/XRCC5 signaling with Quercetin increases prostate cancer radiosensitivity. *Journal of Experimental & Clinical Cancer Research : CR*, 41, 235. https://doi.org/10.1186/ s13046-022-02287-4
- Zhang, X., Huang, J., Yu, C., Xiang, L., Li, L., Shi, D., & Lin, F. (2020). Quercetin enhanced paclitaxel therapeutic effects towards PC-3 prostate cancer through ER stress induction and ROS production. *OncoTargets and Therapy*, *13*, 513–523. https://doi.org/ 10.2147/ott.S228453
- 231. Shu, Y., Xie, B., Liang, Z., & Chen, J. (2018). Quercetin reverses the doxorubicin resistance of prostate cancer cells by downregulating the expression of c-met. *Oncology Letters*, 15, 2252–2258. https://doi.org/10.3892/ol.2017.7561
- 232. Lu, X., Yang, F., Chen, D., Zhao, Q., Chen, D., Ping, H., & Xing, N. (2020). Quercetin reverses docetaxel resistance in prostate cancer via androgen receptor and PI3K/Akt signaling pathways. *International Journal of Biological Sciences*, 16, 1121–1134. https://doi.org/10.7150/ijbs.41686
- 233. Serttas, R., Koroglu, C., & Erdogan, S. (2021). Eupatilin inhibits the proliferation and migration of prostate cancer cells through modulation of PTEN and NF-κB signaling. *Anti-cancer Agents in Medicinal Chemistry*, 21, 372–382. https://doi.org/10.2174/ 1871520620666200811113549
- Wang, P., Henning, S. M., Heber, D., & Vadgama, J. V. (2015). Sensitization to docetaxel in prostate cancer cells by green tea

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and quercetin. *The Journal of Nutritional Biochemistry*, 26, 408–415. https://doi.org/10.1016/j.jnutbio.2014.11.017

- 235. Bao, X., Zhu, J., Ren, C., Zhao, A., Zhang, M., Zhu, Z., Lu, X., Zhang, Y., Li, X., Sima, X., et al. (2021). β-elemonic acid inhibits growth and triggers apoptosis in human castration-resistant prostate cancer cells through the suppression of JAK2/STAT3/ MCL-1 and NF-κB signal pathways. *Chemico-Biological Interactions*, 342, 109477. https://doi.org/10.1016/j.cbi.2021.109477
- 236. Liu, Y. Q., Wang, S. K., Xu, Q. Q., Yuan, H. Q., Guo, Y. X., Wang, Q., Kong, F., Lin, Z. M., Sun, D. Q., Wang, R. M., et al. (2019). Acetyl-11-keto-β-boswellic acid suppresses docetaxelresistant prostate cancer cells in vitro and in vivo by blocking Akt and Stat3 signaling, thus suppressing chemoresistant stem cell-like properties. *Acta Pharmacologica Sinica*, 40, 689–698. https://doi.org/10.1038/s41401-018-0157-9
- 237. Cao, H., Wang, D., Gao, R., Feng, Y., & Chen, L. (2022). Qi Ling decreases paclitaxel resistance in the human prostate cancer by reversing tumor-associated macrophages function. *Aging*, 14, 1812–1821. https://doi.org/10.18632/aging.203904
- 238. Thaper, D., Vahid, S., Kaur, R., Kumar, S., Nouruzi, S., Bishop, J. L., Johansson, M., & Zoubeidi, A. (2018). Galiellalactone inhibits the STAT3/AR signaling axis and suppresses Enzalutamide-resistant Prostate Cancer. *Scientific Reports*, *8*, 17307. https://doi.org/10.1038/s41598-018-35612-z
- 239. Liu, Q., Tong, D., Liu, G., Xu, J., Do, K., Geary, K., Zhang, D., Zhang, J., Zhang, Y., Li, Y., et al. (2017). Metformin reverses prostate cancer resistance to enzalutamide by targeting TGF-β1/ STAT3 axis-regulated EMT. *Cell Death & Disease*, *8*, e3007. https://doi.org/10.1038/cddis.2017.417
- 240. Ge, J., Wang, P., Ma, H., & Zhang, J. (2022). Solamargine inhibits prostate cancer cell growth and enhances the therapeutic efficacy of docetaxel via Akt signaling. *Journal of Oncology*, 2022, 9055954. https://doi.org/10.1155/2022/9055954
- 241. Erdogan, S., Turkekul, K., Serttas, R., & Erdogan, Z. (2017). The natural flavonoid apigenin sensitizes human CD44(+) prostate cancer stem cells to cisplatin therapy. *Biomedicine & Pharmacotherapy = Biomedecine & Pharmacotherapie*, 88, 210–217. https://doi.org/10.1016/j.biopha.2017.01.056
- 242. Thamilselvan, V., Menon, M., & Thamilselvan, S. (2011). Anticancer efficacy of deguelin in human prostate cancer cells targeting glycogen synthase kinase-3 β/β-catenin pathway. *International Journal of Cancer*, *129*, 2916–2927. https://doi.org/10. 1002/ijc.25949
- 243. Hellsten, R., Stiehm, A., Palominos, M., Persson, M., & Bjartell, A. (2022). The STAT3 inhibitor GPB730 enhances the sensitivity to enzalutamide in prostate cancer cells. *Translational Oncology*, 24, 101495. https://doi.org/10.1016/j.tranon.2022.101495
- Liu, C., Lou, W., Armstrong, C., Zhu, Y., Evans, C. P., & Gao, A. C. (2015). Niclosamide suppresses cell migration and invasion in enzalutamide resistant prostate cancer cells via Stat3-AR axis inhibition. *The Prostate*, 75, 1341–1353. https://doi.org/10.1002/ pros.23015
- 245. Chen, Y., Gera, L., Zhang, S., Li, X., Yang, Y., Mamouni, K., Wu, A. Y., Liu, H., Kucuk, O., & Wu, D. (2019). Small molecule BKM1972 inhibits human prostate cancer growth and overcomes docetaxel resistance in intraosseous models. *Cancer Letters*, 446, 62–72. https://doi.org/10.1016/j.canlet.2019.01.010
- 246. Imrali, A., Mao, X., Yeste-Velasco, M., Shamash, J., & Lu, Y. (2016). Rapamycin inhibits prostate cancer cell growth through cyclin D1 and enhances the cytotoxic efficacy of cisplatin. *American Journal of Cancer Research*, 6, 1772–1784.
- 247. Alshaker, H., Wang, Q., Kawano, Y., Arafat, T., Böhler, T., Winkler, M., Cooper, C., & Pchejetski, D. (2016). Everolimus (RAD001) sensitizes prostate cancer cells to docetaxel by downregulation of HIF-1α and sphingosine kinase 1. *Oncotarget*, 7, 80943–80956. https://doi.org/10.18632/oncotarget.13115

- 248. Luo, Y., Li, M., Zuo, X., Basourakos, S. P., Zhang, J., Zhao, J., Han, Y., Lin, Y., Wang, Y., Jiang, Y., et al. (2018). β-catenin nuclear translocation induced by HIF-1α overexpression leads to the radioresistance of prostate cancer. *International Journal* of Oncology, 52, 1827–1840. https://doi.org/10.3892/ijo.2018. 4368
- 249. Zhang, P., Singh, A., Yegnasubramanian, S., Esopi, D., Kombairaju, P., Bodas, M., Wu, H., Bova, S. G., & Biswal, S. (2010). Loss of Kelch-like ECH-associated protein 1 function in prostate cancer cells causes chemoresistance and radioresistance and promotes tumor growth. *Molecular Cancer Therapeutics*, 9, 336–346. https://doi.org/10.1158/1535-7163. Mct-09-0589
- 250. Huang, Z. C., Huang, J., Huang, C. K., Hou, Y., & Zhu, B. (2023). Euchromatic histone lysine methyltransferase 2 facilitates radioresistance in prostate cancer by repressing endoplasmic reticulum protein 29 transcription. *The Kaohsiung Journal* of Medical Sciences. https://doi.org/10.1002/kjm2.12661
- 251. El Bezawy, R., Tinelli, S., Tortoreto, M., Doldi, V., Zuco, V., Folini, M., Stucchi, C., Rancati, T., Valdagni, R., Gandellini, P., et al. (2019). miR-205 enhances radiation sensitivity of prostate cancer cells by impairing DNA damage repair through PKCe and ZEB1 inhibition. *Journal of Experimental & Clinical Cancer Research : CR, 38*, 51. https://doi.org/10.1186/ s13046-019-1060-z
- 252. Marampon, F., Gravina, G., Ju, X., Vetuschi, A., Sferra, R., Casimiro, M., Pompili, S., Festuccia, C., Colapietro, A., Gaudio, E., et al. (2016). Cyclin D1 silencing suppresses tumorigenicity, impairs DNA double strand break repair and thus radiosensitizes androgen-independent prostate cancer cells to DNA damage. *Oncotarget*, 7, 5383–5400. https://doi.org/10.18632/oncotarget. 6579
- 253. Wu, K., Wu, M., Yang, H., Diao, R., & Zeng, H. (2023). Hypoxia promotes conversion to a stem cell phenotype in prostate cancer cells by activating HIF-1α/Notch1 signaling pathway. *Clinical* & Translational Oncology : Official Publication of the Federation of Spanish Oncology Societies and of the National Cancer Institute of Mexico. https://doi.org/10.1007/s12094-023-03093-w
- 254. Owari, T., Tanaka, N., Nakai, Y., Miyake, M., Anai, S., Kishi, S., Mori, S., Fujiwara-Tani, R., Hojo, Y., Mori, T., et al. (2022). 5-Aminolevulinic acid overcomes hypoxia-induced radiation resistance by enhancing mitochondrial reactive oxygen species production in prostate cancer cells. *British Journal of Cancer*, 127, 350–363. https://doi.org/10.1038/s41416-022-01789-4
- 255. Gu, H., Liu, M., Ding, C., Wang, X., Wang, R., Wu, X., & Fan, R. (2016). Hypoxia-responsive miR-124 and miR-144 reduce hypoxia-induced autophagy and enhance radiosensitivity of prostate cancer cells via suppressing PIM1. *Cancer Medicine*, 5, 1174–1182. https://doi.org/10.1002/cam4.664
- 256. Shang, Z. F., Wei, Q., Yu, L., Huang, F., Xiao, B. B., Wang, H., Song, M., Wang, L., Zhou, J., Wang, J., et al. (2016). Suppression of PC-1/PrLZ sensitizes prostate cancer cells to ionizing radiation by attenuating DNA damage repair and inducing autophagic cell death. *Oncotarget*, 7, 62340–62351. https://doi. org/10.18632/oncotarget.11470
- 257. Schwarz, F. M., Schniewind, I., Besso, M. J., Lange, S., Linge, A., Patil, S. G., Löck, S., Klusa, D., Dietrich, A., Voss-Böhme, A., et al. (2022). Plasticity within aldehyde dehydrogenase-positive cells determines prostate cancer radiosensitivity. *Molecular Cancer Research : MCR*, 20, 794–809. https://doi.org/10.1158/ 1541-7786.Mcr-21-0806
- 258. Xu, Z., Zhang, Y., Ding, J., Hu, W., Tan, C., Wang, M., Tang, J., & Xu, Y. (2018). miR-17-3p Downregulates mitochondrial antioxidant enzymes and enhances the radiosensitivity of prostate cancer cells. *Molecular Therapy. Nucleic Acids*, 13, 64–77. https://doi.org/10.1016/j.omtn.2018.08.009

- Fan, Y., Fan, H., Quan, Z., & Wu, X. (2021). Ionizing radiation combined with PARP1 inhibitor reduces radioresistance in prostate cancer with RB1/TP53 loss. *Cancer Investigation*, 39, 423–434. https://doi.org/10.1080/07357907.2021.1899200
- Pawar, J. S., Al-Amin, M. Y., & Hu, C. D. (2023). JNJ-64619178 radiosensitizes and suppresses fractionated ionizing radiationinduced neuroendocrine differentiation (NED) in prostate cancer. *Frontiers in Oncology*, 13, 1126482. https://doi.org/10.3389/ fonc.2023.1126482
- 261. He, Z., Yuan, J., Shen, F., Zeng, F., Qi, P., Wang, Z., & Zhai, Z. (2020). Atorvastatin enhances effects of radiotherapy on prostate cancer cells and xenograft tumor mice through triggering interaction between Bcl-2 and MSH2. *Medical Science Monitor : International Medical Journal of Experimental and Clinical Research*, 26, e923560. https://doi.org/10.12659/msm.923560
- Rajput, M., Mishra, D., Kumar, K., & Singh, R. P. (2022). Silibinin radiosensitizes EGF receptor-knockdown prostate cancer cells by attenuating DNA repair pathways. *Journal of Cancer Prevention*, 27, 170–181. https://doi.org/10.15430/jcp.2022.27.3. 170
- 263. Li, J., Wang, Z., Li, H., Cao, J., Nan, N., Zhai, X., Liu, Y., & Chong, T. (2022). Resveratrol inhibits TRAF6/PTCH/SMO signal and regulates prostate cancer progression. *Cytotechnology*, 74, 549–558. https://doi.org/10.1007/s10616-022-00544-0
- 264. Zhang, L., Lin, Z., Chen, Y., Gao, D., Wang, P., Lin, Y., Wang, Y., Wang, F., Han, Y., & Yuan, H. (2022). Co-delivery of Docetaxel and Resveratrol by liposomes synergistically boosts antitumor efficiency against prostate cancer. *European Journal of Pharmaceutical Sciences : Official Journal of the European Federation for Pharmaceutical Sciences*, 174, 106199. https:// doi.org/10.1016/j.ejps.2022.106199
- 265. Han, D. S., Lee, H. J., & Lee, E. O. (2022). Resveratrol suppresses serum-induced vasculogenic mimicry through impairing the EphA2/twist-VE-cadherin/AKT pathway in human prostate cancer PC-3 cells. *Scientific Reports*, 12, 20125. https://doi.org/ 10.1038/s41598-022-24414-z
- 266. Fang, Y., DeMarco, V. G., & Nicholl, M. B. (2012). Resveratrol enhances radiation sensitivity in prostate cancer by inhibiting cell proliferation and promoting cell senescence and apoptosis. *Cancer Science*, 103, 1090–1098. https://doi.org/10.1111/j.1349-7006.2012.02272.x
- 267. Zhang, T., Zhang, L., Zhang, T., Fan, J., Wu, K., Guan, Z., Wang, X., Li, L., Hsieh, J. T., He, D., et al. (2014). Metformin sensitizes prostate cancer cells to radiation through EGFR/p-DNA-PKCS in vitro and in vivo. *Radiation Research*, 181, 641–649. https:// doi.org/10.1667/rr13561.1
- 268. Potiron, V. A., Abderrahmani, R., Giang, E., Chiavassa, S., Di Tomaso, E., Maira, S. M., Paris, F., & Supiot, S. (2013). Radiosensitization of prostate cancer cells by the dual PI3K/mTOR inhibitor BEZ235 under normoxic and hypoxic conditions. *Radiotherapy and Oncology : Journal of the European Society* for Therapeutic Radiology and Oncology, 106, 138–146. https:// doi.org/10.1016/j.radonc.2012.11.014
- 269. Zhang, Y., Xu, Z., Ding, J., Tan, C., Hu, W., Li, Y., Huang, W., & Xu, Y. (2018). HZ08 suppresses RelB-activated MnSOD expression and enhances radiosensitivity of prostate cancer cells. *Journal of Experimental & Clinical Cancer Research : CR*, 37, 174. https://doi.org/10.1186/s13046-018-0849-5
- 270. Chen, Y. A., Tzeng, D. T. W., Huang, Y. P., Lin, C. J., Lo, U. G., Wu, C. L., Lin, H., Hsieh, J. T., Tang, C. H., Lai, C. H. (2018). Antrocin sensitizes prostate cancer cells to radiotherapy through inhibiting PI3K/AKT and MAPK signaling pathways. *Cancers*, *11*. https://doi.org/10.3390/cancers11010034
- 271. Ma, X., Wang, Z., Ren, H., Bao, X., Zhang, Y., Wang, B., & Ruan, D. (2020). Long non-coding RNA GAS5 suppresses tumor progression and enhances the radiosensitivity of prostate cancer

through the miR-320a/RAB21 axis. Cancer Management and Research, 12, 8833–8845. https://doi.org/10.2147/cmar.S244123

- 272. Zhu, L., Zhu, B., Yang, L., Zhao, X., Jiang, H., & Ma, F. (2014). RelB regulates Bcl-xl expression and the irradiation-induced apoptosis of murine prostate cancer cells. *Biomedical Reports*, 2, 354–358. https://doi.org/10.3892/br.2014.250
- 273. Fan, L., Xu, S., Zhang, F., Cui, X., Fazli, L., Gleave, M., Clark, D. J., Yang, A., Hussain, A., Rassool, F., et al. (2020). Histone demethylase JMJD1A promotes expression of DNA repair factors and radio-resistance of prostate cancer cells. *Cell Death & Disease, 11*, 214. https://doi.org/10.1038/s41419-020-2405-4
- 274. Chang, L., Graham, P. H., Hao, J., Ni, J., Bucci, J., Cozzi, P. J., Kearsley, J. H., & Li, Y. (2014). PI3K/Akt/mTOR pathway inhibitors enhance radiosensitivity in radioresistant prostate cancer cells through inducing apoptosis, reducing autophagy, suppressing NHEJ and HR repair pathways. *Cell Death & Disease*, 5, e1437. https://doi.org/10.1038/cddis.2014.415
- 275. Chen, Y. Y., Luo, L. P., & Deng, K. C. (2023). Circular RNA LPAR3 targets JPT1 via microRNA-513b-5p to facilitate glycolytic activation but repress prostate cancer radiosensitivity. *Acta Biochimica Polonica*, 70, 153–162. https://doi.org/10.18388/abp. 2020_6379
- 276. Hoey, C., Ray, J., Jeon, J., Huang, X., Taeb, S., Ylanko, J., Andrews, D. W., Boutros, P. C., & Liu, S. K. (2018). miRNA-106a and prostate cancer radioresistance: A novel role for LITAF in ATM regulation. *Molecular Oncology*, *12*, 1324–1341. https:// doi.org/10.1002/1878-0261.12328
- 277. Chen, X., Chen, F., Ren, Y., Weng, G., Keng, P. C., Chen, Y., & Lee, S. O. (2019). Glucocorticoid receptor upregulation increases radioresistance and triggers androgen independence of prostate cancer. *The Prostate*, *79*, 1386–1398. https://doi.org/10.1002/ pros.23861
- 278. Wang, W., Liu, M., Guan, Y., & Wu, Q. (2016). Hypoxia-responsive Mir-301a and Mir-301b promote radioresistance of prostate cancer cells via downregulating NDRG2. *Medical Science Monitor : International Medical Journal of Experimental and Clinical Research*, 22, 2126–2132. https://doi.org/10.12659/msm.896832
- Kajanne, R., Miettinen, P., Tenhunen, M., & Leppä, S. (2009). Transcription factor AP-1 promotes growth and radioresistance in prostate cancer cells. *International Journal of Oncology*, 35, 1175–1182. https://doi.org/10.3892/ijo_00000434
- 280. Du, S., Zhang, P., Ren, W., Yang, F., & Du, C. (2020). Circ-ZNF609 accelerates the radioresistance of prostate cancer cells by promoting the glycolytic metabolism through miR-501-3p/ HK2 axis. *Cancer Management and Research*, *12*, 7487–7499. https://doi.org/10.2147/cmar.S257441
- 281. Wang, T., Huang, J., Vue, M., Alavian, M. R., Goel, H. L., Altieri, D. C., Languino, L. R., & FitzGerald, T. J. (2019). α(v)β(3) Integrin mediates radioresistance of prostate cancer cells through regulation of survivin. *Molecular Cancer Research : MCR*, *17*, 398–408. https://doi.org/10.1158/1541-7786.Mcr-18-0544
- 282. Broustas, C. G., Duval, A. J., Chaudhary, K. R., Friedman, R. A., Virk, R. K., & Lieberman, H. B. (2020). Targeting MEK5 impairs nonhomologous end-joining repair and sensitizes prostate cancer to DNA damaging agents. *Oncogene*, *39*, 2467–2477. https://doi.org/10.1038/s41388-020-1163-1
- 283. Yao, B., Liu, B., Shi, L., Li, X., Ren, C., Cai, M., Wang, W., Li, J., Sun, Y., Wu, Y., et al. (2017). PAFR selectively mediates radioresistance and irradiation-induced autophagy suppression in prostate cancer cells. *Oncotarget*, 8, 13846–13854. https://doi. org/10.18632/oncotarget.14647
- 284. Ciccarelli, C., Di Rocco, A., Gravina, G. L., Mauro, A., Festuccia, C., Del Fattore, A., Berardinelli, P., De Felice, F., Musio, D., Bouché, M., et al. (2018). Disruption of MEK/ERK/c-Myc signaling radiosensitizes prostate cancer cells in vitro and in vivo.

Journal of Cancer Research and Clinical Oncology, 144, 1685–1699. https://doi.org/10.1007/s00432-018-2696-3

- 285. Chen, X., Chen, F., Ren, Y., Weng, G., Xu, L., Xue, X., Keng, P. C., Lee, S. O., & Chen, Y. (2019). IL-6 signaling contributes to radioresistance of prostate cancer through key DNA repair-associated molecules ATM, ATR, and BRCA 1/2. *Journal of Cancer Research and Clinical Oncology*, 145, 1471–1484. https://doi. org/10.1007/s00432-019-02917-z
- 286. Ruan, H., Bao, L., Tao, Z., & Chen, K. (2021). Flightless I homolog reverses enzalutamide resistance through PD-L1-mediated immune evasion in prostate cancer. *Cancer Immunol*ogy Research, 9, 838–852. https://doi.org/10.1158/2326-6066. Cir-20-0729
- 287. Kolijn, K., Verhoef, E. I., Smid, M., Böttcher, R., Jenster, G. W., Debets, R., & van Leenders, G. (2018). Epithelial-mesenchymal transition in human prostate cancer demonstrates enhanced immune evasion marked by IDO1 expression. *Cancer Research*, 78, 4671–4679. https://doi.org/10.1158/0008-5472.Can-17-3752
- 288. Chen, Q. H., Li, B., Liu, D. G., Zhang, B., Yang, X., & Tu, Y. L. (2020). LncRNA KCNQ1OT1 sponges miR-15a to promote immune evasion and malignant progression of prostate cancer via up-regulating PD-L1. *Cancer Cell International*, 20, 394. https://doi.org/10.1186/s12935-020-01481-8
- Kan, L., Huang, Y., Liu, Z. (2023). JUN and ATF3 are deficient in prostate cancer patients and their delivery in vivo via lipid nanoparticles has therapeutic efficacy by enhancing immune surveillance. *Pharmacological Research*, 106753. https://doi.org/10. 1016/j.phrs.2023.106753
- 290. Drehmer, D., Mesquita Luiz, J. P., Hernandez, C. A. S., Alves-Filho, J. C., Hussell, T., Townsend, P. A., & Moncada, S. (2022). Nitric oxide favours tumour-promoting inflammation through mitochondria-dependent and -independent actions on macrophages. *Redox Biology*, 54, 102350. https://doi.org/10.1016/j. redox.2022.102350
- 291. Won, H., Moreira, D., Gao, C., Duttagupta, P., Zhao, X., Manuel, E., Diamond, D., Yuan, Y. C., Liu, Z., Jones, J., et al. (2017). TLR9 expression and secretion of LIF by prostate cancer cells stimulates accumulation and activity of polymorphonuclear MDSCs. *Journal of Leukocyte Biology*, *102*, 423–436. https:// doi.org/10.1189/jlb.3MA1016-451RR
- 292. Groeger, S., Wu, F., Wagenlehner, F., Dansranjav, T., Ruf, S., Denter, F., & Meyle, J. (2022). PD-L1 up-regulation in prostate cancer cells by Porphyromonas gingivalis. *Frontiers in Cellular* and Infection Microbiology, 12, 935806. https://doi.org/10.3389/ fcimb.2022.935806
- 293. Li, N., Liu, Q., Han, Y., Pei, S., Cheng, B., Xu, J., Miao, X., Pan, Q., Wang, H., Guo, J., et al. (2022). ARID1A loss induces polymorphonuclear myeloid-derived suppressor cell chemotaxis and promotes prostate cancer progression. *Nature Communications*, 13, 7281. https://doi.org/10.1038/s41467-022-34871-9
- 294. Imamura, R., Kitagawa, S., Kubo, T., Irie, A., Kariu, T., Yoneda, M., Kamba, T., & Imamura, T. (2021). Prostate cancer C5a receptor expression and augmentation of cancer cell proliferation, invasion, and PD-L1 expression by C5a. *The Prostate*, *81*, 147–156. https://doi.org/10.1002/pros.24090
- 295. Zhang, Y., Zhu, S., Du, Y., Xu, F., Sun, W., Xu, Z., Wang, X., Qian, P., Zhang, Q., Feng, J., et al. (2022). RelB upregulates PD-L1 and exacerbates prostate cancer immune evasion. *Journal of Experimental & Clinical Cancer Research : CR*, 41, 66. https://doi.org/10.1186/s13046-022-02243-2
- 296. Wise, D. R., Schneider, J.A., Armenia, J., Febles, V. A., McLaughlin, B., Brennan, R., Thoren, K. L., Abida, W., Sfanos, K. S., De Marzo, A. M., et al. (2020). Dickkopf-1 can lead to immune evasion in metastatic castration-resistant prostate cancer. *JCO Precision Oncology*, 4. https://doi.org/10.1200/po.20.00097

- 297. Su, W., Han, H. H., Wang, Y., Zhang, B., Zhou, B., Cheng, Y., Rumandla, A., Gurrapu, S., Chakraborty, G., Su, J., et al. (2019). The polycomb repressor complex 1 drives double-negative prostate cancer metastasis by coordinating stemness and immune suppression. *Cancer Cell*, *36*, 139-155.e110. https://doi.org/10. 1016/j.ccell.2019.06.009
- 298. Bancaro, N., Calì, B., Troiani, M., Elia, A. R., Arzola, R. A., Attanasio, G., Lai, P., Crespo, M., Gurel, B., Pereira, R., et al. (2023). Apolipoprotein E induces pathogenic senescent-like myeloid cells in prostate cancer. *Cancer Cell*, *41*, 602-619.e611. https://doi.org/10.1016/j.ccell.2023.02.004
- 299. Moon, S. J., Jeong, B. C., Kim, H. J., Lim, J. E., Kim, H. J., Kwon, G. Y., Jackman, J. A., & Kim, J. H. (2021). Bruceantin targets HSP90 to overcome resistance to hormone therapy in castration-resistant prostate cancer. *Theranostics*, 11, 958–973. https://doi.org/10.7150/thno.51478
- 300. Castilla, C., Congregado, B., Conde, J. M., Medina, R., Torrubia, F. J., Japón, M. A., & Sáez, C. (2010). Immunohistochemical expression of Hsp60 correlates with tumor progression and hormone resistance in prostate cancer. *Urology*, 76(1017), e1011-1016. https://doi.org/10.1016/j.urology.2010.05.045
- 301. Karantanos, T., Karanika, S., Wang, J., Yang, G., Dobashi, M., Park, S., Ren, C., Li, L., Basourakos, S. P., Hoang, A., et al. (2016). Caveolin-1 regulates hormone resistance through lipid synthesis, creating novel therapeutic opportunities for castrationresistant prostate cancer. *Oncotarget*, 7, 46321–46334. https:// doi.org/10.18632/oncotarget.10113
- 302. Zhang, Y., Linn, D., Liu, Z., Melamed, J., Tavora, F., Young, C. Y., Burger, A. M., & Hamburger, A. W. (2008). EBP1, an ErbB3-binding protein, is decreased in prostate cancer and implicated in hormone resistance. *Molecular Cancer Therapeutics*, 7, 3176–3186. https://doi.org/10.1158/1535-7163.Mct-08-0526
- 303. Castilla, C., Congregado, B., Chinchón, D., Torrubia, F. J., Japón, M. A., & Sáez, C. (2006). Bcl-xL is overexpressed in hormoneresistant prostate cancer and promotes survival of LNCaP cells via interaction with proapoptotic Bak. *Endocrinology*, 147, 4960–4967. https://doi.org/10.1210/en.2006-0502
- 304. Wilkinson, S., Ye, H., Karzai, F., Harmon, S. A., Terrigino, N. T., VanderWeele, D. J., Bright, J. R., Atway, R., Trostel, S. Y., Carrabba, N. V., et al. (2021). Nascent prostate cancer heterogeneity drives evolution and resistance to intense hormonal therapy. *European Urology*, 80, 746–757. https://doi.org/10.1016/j. eururo.2021.03.009
- 305. Cheng, Q., Butler, W., Zhou, Y., Zhang, H., Tang, L., Perkinson, K., Chen, X., Jiang, X. S., McCall, S. J., Inman, B. A., et al. (2022). Pre-existing castration-resistant prostate cancer-like cells in primary prostate cancer promote resistance to hormonal therapy. *European Urology*, *81*, 446–455. https://doi.org/10.1016/j. eururo.2021.12.039
- 306. Edlind, M. P., & Hsieh, A. C. (2014). PI3K-AKT-mTOR signaling in prostate cancer progression and androgen deprivation therapy resistance. *Asian Journal of Andrology*, 16, 378–386. https://doi.org/10.4103/1008-682x.122876
- 307. Li, Q., Wang, M., Hu, Y., Zhao, E., Li, J., Ren, L., Wang, M., Xu, Y., Liang, Q., Zhang, D., et al. (2021). MYBL2 disrupts the Hippo-YAP pathway and confers castration resistance and metastatic potential in prostate cancer. *Theranostics*, 11, 5794–5812. https://doi.org/10.7150/thno.56604
- 308. Mateo, J., Seed, G., Bertan, C., Rescigno, P., Dolling, D., Figueiredo, I., Miranda, S., Nava Rodrigues, D., Gurel, B., Clarke, M., et al. (2020). Genomics of lethal prostate cancer at diagnosis and castration resistance. *The Journal of Clinical Investigation*, 130, 1743–1751. https://doi.org/10.1172/jci132031
- 309. Cheng, C., Wang, J., Xu, P., Zhang, K., Xin, Z., Zhao, H., Ji, Z., Zhang, M., Wang, D., He, Y., et al. (2022). Gremlin1 is a therapeutically targetable FGFR1 ligand that regulates

lineage plasticity and castration resistance in prostate cancer. *Nature Cancer*, *3*, 565–580. https://doi.org/10.1038/ s43018-022-00380-3

- Tcatchoff, L., Nespoulous, C., Pernollet, J. C., & Briand, L. (2006). A single lysyl residue defines the binding specificity of a human odorant-binding protein for aldehydes. *FEBS Letters*, 580, 2102–2108. https://doi.org/10.1016/j.febslet.2006.03.017
- 311. Jeong, J. H., Zhong, S., Li, F., Huang, C., Chen, X., Liu, Q., Peng, S., Park, H., Lee, Y.M., Dhillon, J., et al. (2023). Tumorderived OBP2A promotes prostate cancer castration resistance. *The Journal of Experimental Medicine*, 220. https://doi.org/10. 1084/jem.20211546
- 312. Sharp, A., Coleman, I., Yuan, W., Sprenger, C., Dolling, D., Rodrigues, D. N., Russo, J. W., Figueiredo, I., Bertan, C., Seed, G., et al. (2019). Androgen receptor splice variant-7 expression emerges with castration resistance in prostate cancer. *The Journal of Clinical Investigation*, *129*, 192–208. https://doi.org/10. 1172/jci122819
- 313. Liu, Y., Yu, C., Shao, Z., Xia, X., Hu, T., Kong, W., He, X., Sun, W., Deng, Y., Liao, Y., et al. (2021). Selective degradation of AR-V7 to overcome castration resistance of prostate cancer. *Cell Death & Disease*, *12*, 857. https://doi.org/10.1038/ s41419-021-04162-0
- 314. Yuan, S., He, S. H., Li, L. Y., Xi, S., Weng, H., Zhang, J. H., Wang, D. Q., Guo, M. M., Zhang, H., Wang, S. Y., et al. (2023). A potassium-chloride co-transporter promotes tumor progression and castration resistance of prostate cancer through m(6)A reader YTHDC1. *Cell Death & Disease*, 14, 7. https://doi.org/10.1038/ s41419-022-05544-8
- 315. Zhang, N., Huang, D., Ruan, X., Ng, A. T., Tsu, J. H., Jiang, G., Huang, J., Zhan, Y., & Na, R. (2023). CRISPR screening reveals gleason score and castration resistance related oncodriver ring finger protein 19 A (RNF19A) in prostate cancer. *Drug Resistance Updates : Reviews and Commentaries in Antimicrobial and Anticancer Chemotherapy*, 67, 100912. https://doi.org/10.1016/j. drup.2022.100912
- 316. Sun, Y., Cronin, M. F., Mendonça, M. C. P., Guo, J., & O'Driscoll, C. M. (2023). Sialic acid-targeted cyclodextrin-based nanoparticles deliver CSF-1R siRNA and reprogram tumourassociated macrophages for immunotherapy of prostate cancer. *European Journal of Pharmaceutical Sciences : Official Journal* of the European Federation for Pharmaceutical Sciences, 185, 106427. https://doi.org/10.1016/j.ejps.2023.106427
- 317. Tanaudommongkon, I., Tanaudommongkon, A., Prathipati, P., Nguyen, J. T., Keller, E. T., Dong, X. (2020). Curcumin nanoparticles and their cytotoxicity in docetaxel-resistant castrationresistant prostate cancer cells. *Biomedicines*, 8. https://doi.org/ 10.3390/biomedicines8080253
- 318. Hoang, B., Ernsting, M. J., Tang, W. S., Bteich, J., Undzys, E., Kiyota, T., & Li, S. D. (2017). Cabazitaxel-conjugated nanoparticles for docetaxel-resistant and bone metastatic prostate cancer. *Cancer Letters*, 410, 169–179. https://doi.org/10.1016/j.canlet. 2017.09.029
- 319. Peng, S., Zhang, X., Huang, H., Cheng, B., Xiong, Z., Du, T., Wu, J., & Huang, H. (2022). Glutathione-sensitive nanoparticles enhance the combined therapeutic effect of checkpoint kinase 1 inhibitor and cisplatin in prostate cancer. *APL Bioengineering*, 6, 046106. https://doi.org/10.1063/5.0126095
- 320. Yan, J., Wang, Y., Zhang, X., Liu, S., Tian, C., & Wang, H. (2016). Targeted nanomedicine for prostate cancer therapy: Docetaxel and curcumin co-encapsulated lipid-polymer hybrid nanoparticles for the enhanced anti-tumor activity in vitro and in vivo. *Drug Delivery*, 23, 1757–1762. https://doi.org/10.3109/ 10717544.2015.1069423
- 321. Gao, Z., Huang, J., Xie, Z., Xin, P., Huang, H., Du, T., Wu, J., & Huang, H. (2022). Delivery of enzalutamide via nanoparticles for

effectively inhibiting prostate cancer progression. *Biomaterials Science*, 10, 5187–5196. https://doi.org/10.1039/d2bm00697a

- 322. Xu, H., Sheng, G., Lu, L., Wang, C., Zhang, Y., Feng, L., Meng, L., Min, P., Zhang, L., Wang, Y., et al. (2021). GRPr-mediated photothermal and thermodynamic dual-therapy for prostate cancer with synergistic anti-apoptosis mechanism. *Nanoscale*, 13, 4249–4261. https://doi.org/10.1039/d0nr07196j
- 323. Wei, C. G., Zhang, R., Wei, L. Y., Pan, P., Zu, H., Liu, Y. Z., Wang, Y., & Shen, J. K. (2022). Calcium phosphate-based nanomedicine mediated CRISPR/Cas9 delivery for prostate cancer therapy. *Frontiers in Bioengineering and Biotechnology*, 10, 1078342. https://doi.org/10.3389/fbioe.2022.1078342
- 324. Zhang, X., He, Z., Xiang, L., Li, L., Zhang, H., Lin, F., & Cao, H. (2019). Codelivery of GRP78 siRNA and docetaxel via RGD-PEG-DSPE/DOPA/CaP nanoparticles for the treatment of castration-resistant prostate cancer. *Drug Design, Development and Therapy*, 13, 1357–1372. https://doi.org/10.2147/dddt.S198400
- 325. Liang, S., Han, L., Mu, W., Jiang, D., Hou, T., Yin, X., Pang, X., Yang, R., Liu, Y., & Zhang, N. (2018). Carboplatin-loaded SMNDs to reduce GSH-mediated platinum resistance for prostate cancer therapy. *Journal of Materials Chemistry. B*, 6, 7004– 7014. https://doi.org/10.1039/c8tb01721b
- 326. Guo, Q., Dong, Y., Zhang, Y., Fu, H., Chen, C., Wang, L., Yang, X., Shen, M., Yu, J., Chen, M., et al. (2021). Sequential release of pooled siRNAs and paclitaxel by aptamer-functionalized shellcore nanoparticles to overcome paclitaxel resistance of prostate cancer. ACS Applied Materials & Interfaces, 13, 13990–14003. https://doi.org/10.1021/acsami.1c00852
- 327. Nagesh, P. K. B., Chowdhury, P., Hatami, E., Boya, V. K. N., Kashyap, V. K., Khan, S., Hafeez, B. B., Chauhan, S. C., Jaggi, M., Yallapu, M. M. (2018). miRNA-205 nanoformulation sensitizes prostate cancer cells to chemotherapy. *Cancers*, *10*. https:// doi.org/10.3390/cancers10090289
- 328. Singh, S. K., Lillard, J. W., Jr., & Singh, R. (2018). Reversal of drug resistance by planetary ball milled (PBM) nanoparticle

loaded with resveratrol and docetaxel in prostate cancer. *Cancer Letters*, 427, 49–62. https://doi.org/10.1016/j.canlet.2018.04.017

- 329. Hara, D., Tao, W., Schmidt, R. M., Yang, Y. P., Daunert, S., Dogan, N., Ford, J. C., Pollack, A., Shi, J. (2022). Boosted radiation bystander effect of PSMA-targeted gold nanoparticles in prostate cancer radiosensitization. *Nanomaterials (Basel, Switzerland)*, 12. https://doi.org/10.3390/nano12244440
- 330. Zhang, X., Liu, N., Shao, Z., Qiu, H., Yao, H., Ji, J., Wang, J., Lu, W., Chen, R. C., & Zhang, L. (2017). Folate-targeted nanoparticle delivery of androgen receptor shRNA enhances the sensitivity of hormone-independent prostate cancer to radiotherapy. *Nanomedicine : Nanotechnology, Biology, and Medicine, 13*, 1309–1321. https://doi.org/10.1016/j.nano.2017.01.015
- 331. Mirjolet, C., Boudon, J., Loiseau, A., Chevrier, S., Boidot, R., Oudot, A., Collin, B., Martin, E., Joy, P. A., Millot, N., et al. (2017). Docetaxel-titanate nanotubes enhance radiosensitivity in an androgen-independent prostate cancer model. *International Journal of Nanomedicine*, *12*, 6357–6364. https://doi.org/10. 2147/ijn.S139167
- 332. Mortensen, M. M., Høyer, S., Lynnerup, A.-S., Ørntoft, T. F., Sørensen, K. D., Borre, M., & Dyrskjøt, L. (2015). Expression profiling of prostate cancer tissue delineates genes associated with recurrence after prostatectomy. *Scientific Reports*, 5, 16018.

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