

Id4 protein is highly expressed in triple-negative breast carcinomas: possible implications for BRCA1 downregulation

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Received: 12 April 2012 / Accepted: 16 April 2012 / Published online: 27 April 2012
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Abstract *BRCA1* germline mutation carriers usually develop ER, PR and HER2 negative breast carcinoma. Somatic *BRCA1* mutations are rare in sporadic breast cancers, but other mechanisms could impair *BRCA1* functions in these tumors, particularly in triple-negative breast carcinomas (TNBCs). Id4, a helix-loop-helix DNA binding factor, blocks *BRCA1* gene transcription in vitro and could downregulate *BRCA1* in vivo. We compared Id4 immunoreactivity in 101 TNBCs versus 113 non-TNBCs, and correlated the results with tumor morphology and immunoreactivity for CK5/6, CK14, EGFR, and androgen receptor (AR). Id4 was present in 76 out of 101 (75 %) TNBCs: 40 (40 %) TNBCs displayed Id4 positivity in >50 % of neoplastic cells, 23 (23 %) in 5–50 %, and 13

(13 %) in <5 %. In contrast, only 6 (5 %) of 113 non-TNBCs showed focal Id4 positivity, limited to fewer than 5 % of the tumor ($p < 0.0001$). Id4 expression significantly associated with high histologic grade ($p = 0.0002$) and mitotic rate ($p = 0.006$). Id4 decorated all 12 TNBCs with large central acellular zone of necrosis in our series, with positive staining in 10–90 % of the cells. Id4 signal strongly correlated with cytokeratin CK14 reactivity ($p < 0.0001$), but not with CK5/6 and EGFR. All apocrine carcinomas in our series were positive for AR and most for EGFR, but they were negative for CK5/6, CK14, and Id4, with only two exceptions. Our results document substantial expression of Id4 in most TNBCs, which could result in functional downregulation of *BRCA1* pathways in these tumors.

Part of this work was presented at United States and Canadian Academy of Pathology (USCAP) 99th Annual Meeting in Washington, DC, March 2010.

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Keywords Cytokeratin 5/6 · Cytokeratin 14 · Basal-like carcinoma · Epidermal growth factor receptor · Androgen receptor

Introduction

Triple-negative breast carcinomas (TNBCs), defined by lack of expression for estrogen receptor (ER), progesterone receptor (PR) and human epidermal growth factor receptor 2 (HER2), represent about 10–15 % of all breast cancers [1]. They are frequent in *BRCA1* germline mutation carriers [2], and common in women under 40 years of age and in African-American women [3]. TNBCs show aggressive clinical behavior characterized by early recurrence, frequent brain and lung involvement [3], and poor survival [4]. These tumors demonstrate a high rate of pathologic complete response post-neoadjuvant treatment [5, 6], but patients with residual disease have poor survival [6] and

early relapse is common [7]. At present, TNBCs constitute the most challenging group of breast cancers due to the lack of effective targeted therapies. Morphologically, TNBCs are a very heterogeneous group, which consists predominantly of high grade invasive ductal carcinoma of no special type (NOS), but includes special subtypes such as medullary, metaplastic, and apocrine carcinomas. Gene expression studies have classified breast cancers into specific molecular subtypes, including basal-like breast carcinomas which show significant morphologic and immunophenotypic overlap with TNBCs [8]. The use of immunohistochemical stains for ER, HER2, basal cytokeratins CK 5/6 or CK 14 and EGFR in different combinations has been proposed as a surrogate method for the identification of basal-like breast carcinomas [9, 10].

Breast cancers arising in *BRCA1* germline mutation carriers are usually triple-negative and display basal-like gene expression profiles [11]. Similarities between breast cancers occurring in *BRCA1* germline mutation carriers and sporadic TNBCs suggest that *BRCA1* inactivation also plays a role in the pathogenesis of some sporadic tumors [12–15]. Silencing of the *BRCA1* gene by promoter methylation has been reported in some non-familial medullary carcinoma and metaplastic carcinomas [12, 16]. Furthermore, Turner et al. [12] showed a two-fold reduction of *BRCA1* mRNA levels in basal-like breast cancers compared to matched non-basal-like carcinomas.

Id4 is a negative regulator of the basic helix-loop-helix group of transcription factors [17], and downregulates the *BRCA1* promoter in vitro [18]. Recent studies have also demonstrated a regulatory interplay of Id4 with microRNAs (including miR 335 and miR 9) and p53 [19–22]. We have previously reported an inverse correlation between Id4 mRNA and ER immunoreactivity in normal breast epithelium and in ER-positive breast carcinomas [23]. Turner et al. [12] have demonstrated a 9.1-fold increase in Id4 mRNA levels in basal-like carcinoma compared to control tumors by PCR analysis. These data suggest that Id4 could play an important role in the functional down-regulation of *BRCA1* in TNBCs. In this study, we used immunohistochemistry to assess the expression of Id4 in TNBCs and correlated Id4 reactivity with tumor morphology, basal phenotype, and patient outcome.

Materials and methods

Patient selection and tissue microarrays (TMAs)

The study was approved by the Institutional Review Board. TNBCs were defined as having ER and PR nuclear staining in <1 % of the tumor cells, and either negative (0 or 1+) HER2 staining or equivocal (2+) HER2 staining and no

HER2 gene amplification by FISH [24, 25]. A search of our pathology database identified patients with TNBC who had surgical excision of the primary carcinoma at our center between 2002 and 2004. Our study cohort consists of 101 patients with tissue blocks of the primary TNBC available for immunohistochemical analysis. Information about patient age, race and clinical follow-up was obtained from the electronic medical records. One of the study pathologists (YHW) reviewed all available slides for the patients in the study and selected representative tumor slides, which were reviewed together with the other study pathologist (EB). The morphologic characteristics of the TNBCs (tumor type, size, grade, growth pattern, presence or absence of associated in situ carcinoma, calcifications, necrosis, lymphocytic infiltrate, lymphovascular invasion, and lymph node metastasis) were noted. The lymphocytic infiltrate associated with invasive carcinoma was scored according to Kreike et al. [26] as absent = no notable lymphocytic infiltrate; minimal = scattered lymphocytes (<10 lymphocytes/400× power field); moderate = lymphocytes easily identified, but no large aggregates; extensive = large aggregates of lymphocytes in >50 % of the tumor. As reference cases, we used 113 non-TNBCs, which included 11 ER–/HER2+ carcinomas (HER2 3+ by immunohistochemistry or HER2 gene amplified by FISH) and 102 consecutive non-TNBCs (84 ER+/HER2–; 17 ER+/HER2+; 1 ER–/HER2+). The 102 tumors, available as triplicate 0.6 mm cores in TMAs, were all greater than 1 cm in size. The 1 cm cutoff had been arbitrarily chosen before preparation of the TMAs to ensure availability of residual carcinoma for possible future clinical use.

Immunohistochemistry

We performed immunoperoxidase stains for Id4, CK5/6, CK14, EGFR, AR, ER, PR, and HER2, with appropriate positive and negative controls for each staining. Normal breast epithelium expressing Id4 was used as positive control for Id4 staining [23]. Sources and dilutions of the primary antibodies are summarized in Table 1. ER, PR, and HER2 stains were repeated in all TNBCs and re-assessed according to the current American Society of Clinical Oncology/College of American Pathologist guideline recommendations [24, 25]. Immunoperoxidase studies for Id4 were performed on whole tissue sections (1–2 representative tumor blocks from each case) of all TNBCs and of 11 ER–/HER2+ breast carcinomas, and on TMAs of the remaining 102 non-TNBCs. The results of Id4 immunoreactivity on TMAs were validated by immunoperoxidase stain on whole tissue sections of 23/102 (22.5 %) reference tumors, including all non-TNBCs showing any Id4 positivity in the TMAs.

Table 1 Antibody resources and dilutions for immunohistochemical stains

Antibody	Clone	Manufacturer	Dilution
ER	1D5	Dako	1:100
PR	PGR636	Dako	1:100
HER2	P185 ^{Her2}	Dako	Pre-diluted
CK5/6	D5-16B4	AbCam	1:50
CK14	LL002	AbCam	1:100
EGFR	31G7	Zymed	1:20
AR	AR441	Dako	1:300
Id4	82-12	Biocheck	1:50

We recorded the percentage of Id4-positive tumor cells in each case and defined Id4 positivity as nuclear staining in at least 5 % of the tumor cells. EGFR was graded according to the scoring guidelines for HER2, and EGFR overexpression was defined as 2+ and 3+ reactivity [27, 28]. Carcinomas showing any cytoplasmic immunoreactivity for CK5/6 and CK14 were classified as positive for these markers.

Statistical analysis

Clinicopathological characteristics of the TNBC and non-TNBC cohorts were compared using the *t* test or χ^2 or if warranted, the Fisher's exact test or Wilcoxon rank sum test. Similar tests were used to compare the clinicopathological characteristics in the TNBC cohort, by Id4 positivity, where Id4 positivity was defined at a 5 % cutoff. Kaplan–Meier methods were used to estimate distant metastasis-free survival and overall survival. Distant metastasis-free survival was defined as (a) the time from diagnosis to distant metastasis, or (b) the time to death or (c) time to last follow-up, according to patient status. Overall survival was defined as the time from initial diagnosis to death or to last follow-up. The log-rank test was used to assess survival differences between groups with low and high Id4 levels again using 5 % of tumor cells with positive staining as the cutoff; 5 % cutoff was chosen based on the results of exploratory analysis using different threshold values (data not shown). For all analyses, a *p* value <0.05 was considered as statistically significant. All statistical analysis was performed in SAS 9.2 and R 2.11.1.

Results

Study cohort

All patients in the study were women, with mean age of 54 years (range 29–95). Patients with TNBC and non-TNBC were similar with regard to age, tumor size, and

length of follow-up (Table 2). Lymph node metastases were less common in patients with TNBCs than in patients with non-TNBCs, but visceral metastases were more frequent in TNBCs. Table 2 summarizes the clinicopathologic features of TNBCs and non-TNBCs. Information about patient ethnicity was available for all patients with TNBC: 86 (85 %) had been classified as Caucasian, including 21 (21 %) Jewish patients; 11 (11 %) were African-American and 4 (4 %) were Asian.

Morphology of TNBCs and non-TNBCs

Most TNBCs were invasive ductal carcinomas NOS (86/101; 85 %). Twelve of 101 (12 %) carcinomas had a large central acellular zone of necrosis occupying more than 30 % of the tumor mass (LCAZ) [29, 30]. Eleven tumors (11 %) were apocrine carcinomas by morphological assessment, and showed abundant eosinophilic cytoplasm and prominent nucleoli. [31] The remaining TNBCs consisted of two metaplastic carcinomas (2 %), and two carcinomas with medullary features (2 %).

Ninety-one of 113 (81 %) non-TNBCs were invasive ductal carcinomas NOS; the rest included 9 (8 %) invasive lobular carcinomas, 6 (5 %) invasive mammary carcinomas with mixed ductal and lobular features, 4 (4 %) invasive micropapillary carcinomas, 2 (2 %) mixed mucinous carcinomas, and 1 (1 %) invasive papillary carcinoma.

Id4 expression in TNBCs versus non-TNBCs

Id4 immunoreactivity was detected in 76 out of 101 (75 %) TNBCs, but the percentage of Id4-positive cells varied widely. Sixty-three (62.4 %) of the 101 TNBCs showed Id4 positivity in 5 % or more of the neoplastic cells, including 40 (39.6 %) cases with nuclear staining in at least 50 % of the tumor (Fig. 1) and 23 (22.8 %) in 5–50 %.

Table 2 Clinicopathologic features of TNBCs and non-TNBCs

	TNBCs (<i>N</i> = 101)	Non-TNBCs (<i>N</i> = 113)
Mean age (range, years)	55 (29–95)	54 (30–87)
Patients \leq 40 years of age (age range, years)	<i>N</i> = 19 (29–39)	<i>N</i> = 19 (30–38)
Mean tumor size (range, cm)	2.4 (0.3–28)	2.4 (1.1–11)
Carcinomas \leq 1 cm (size range, cm)	<i>N</i> = 20 (0.3–1)	None
Median follow-up (range, month)	73 (4–136)	79 (3–134)
Patients with lymph node metastases	<i>N</i> = 34 (34 %)	<i>N</i> = 66 (58 %)
Patients with visceral metastases	<i>N</i> = 22 (22 %)	<i>N</i> = 13 (12 %)

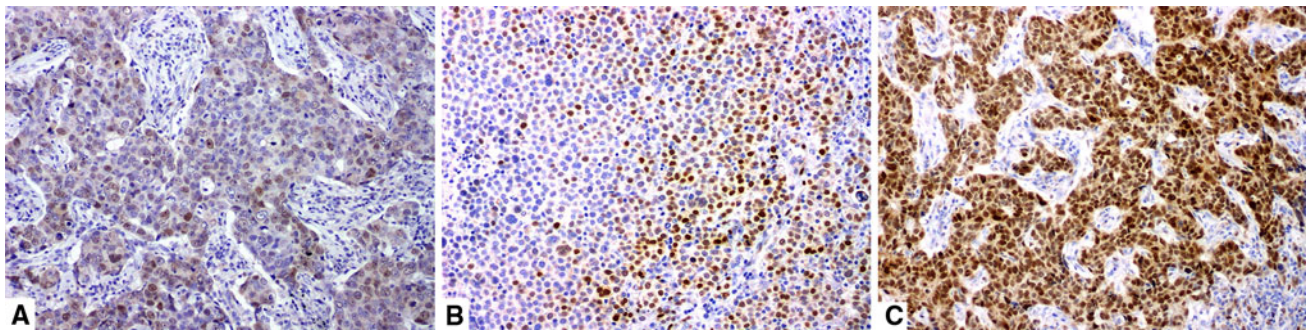


Fig. 1 Range of Id4 immunoreactivity in TNBCs. **a** Focal nuclear Id4 expression in <5 % of the tumor cells. **b** Nuclear Id4 is present in over 5 % but <50 % of the tumor cells. **c** Strong and diffuse nuclear Id4 in more than 50 % of the tumor cells. ($\times 200$)

Thirteen (12.7 %) TNBCs displayed focal weak nuclear reactivity in fewer than 5 % of the tumor cells. Twenty-five TNBCs (24.8 %) had no nuclear stain for Id4.

Only 6 out of 113 (5 %) non-TNBCs (4 ER+/HER2– and 2 ER–/HER2+ tumors) showed some immunoreactivity for Id4. The staining was focal and limited to fewer than 5 % of the tumor cells in all six cases when confirmed on the whole tissue sections.

The difference in Id4 expression between TNBCs and non-TNBCs was statistically significant ($p < 0.0001$) independent of the percentage of Id4-positive tumor cells used as cutoff.

Id4 expression and tumor morphology in TNBCs

Id4 positivity in breast carcinomas is significantly associated with high histologic grade, however, all but 4 TNBCs had modified Bloom–Richardson grade 3. Id4 is also significantly associated with high mitotic rate (Table 3). All 12 TNBCs with LCAZ showed strong positivity for Id4 (Fig. 2), ranging from 10 to 90 % of the tumor cells, with 6 cases showing strong nuclear staining in 50 % or more of the tumor (Table 4).

We observed a significant inverse correlation between Id4 positivity and apocrine morphology. Out of 11 apocrine TNBCs, only two cases showed very focal Id4 staining, which was limited to only 1 and 5 % of the tumor. The 9 remaining apocrine carcinomas were completely negative for Id4 (Table 5). All 11 apocrine TNBCs displayed strong and diffuse immunoreactivity for AR.

Patient age, tumor size, lymphovascular invasion, nodal and visceral metastases did not correlate with Id4 positivity in TNBCs.

Id4 expression and basal-like markers in TNBCs

Of the 101 TNBCs, 47 (47 %) were positive for CK14, 50 (50 %) for CK5/6, and 68 (67 %) for EGFR. Seventy-eight TNBCs (77 %) were positive for CK5/6 and/or EGFR, and 63 (62 %) for CK5/6 and/or CK14. Positive reactivity for

Id4 significantly associated with positivity for CK14 (Table 6), but showed no correlation with CK5/6 and EGFR, either alone or in combination. We observed a significant relationship between Id4 staining and positivity for CK5/6 and/or CK14, but this finding is probably secondary to the strong positive correlation between Id4 and CK14. A case demonstrating positive reactivity for Id4, CK14, CK5/6 and EGFR is illustrated in Fig. 3. All 12 TNBCs with LCAZ were positive for Id4 and CK14, and showed variable positivity for CK5/6 and EGFR (Table 4).

Table 3 Clinicopathological features of TNBCs in relation to Id4 expression

	Id4 \geq 5 % (N = 63)	Id4 < 5 % (N = 38)	p
Mean age, year	54	57	NS
<i>Race</i>			
Caucasian	54	32	NS
African-American	6	5	NS
Asian	3	1	NS
<i>Jewish ethnicity</i>			
Yes	17	4	0.048
No	46	34	
Mean tumor size, cm	2.7	2.0	NS
MBR grade 2	4	14	NS
MBR grade 3	59	24	0.0002
Mitoses/10 hpf	26	16	0.0060
LCAZ	12	0	0.0031
Lymphocytic infiltrate, moderate to extensive	44	20	NS
Pushing border	42	19	NS
Apocrine carcinoma	1	10	0.0002
LVI	18	10	NS
Lymph node metastases	19	18	NS
Visceral metastases	16	6	NS

NS not significant, MBR modified Bloom–Richardson, hpf high power fields, LCAZ large central acellular zone of necrosis, LVI lymphovascular invasion

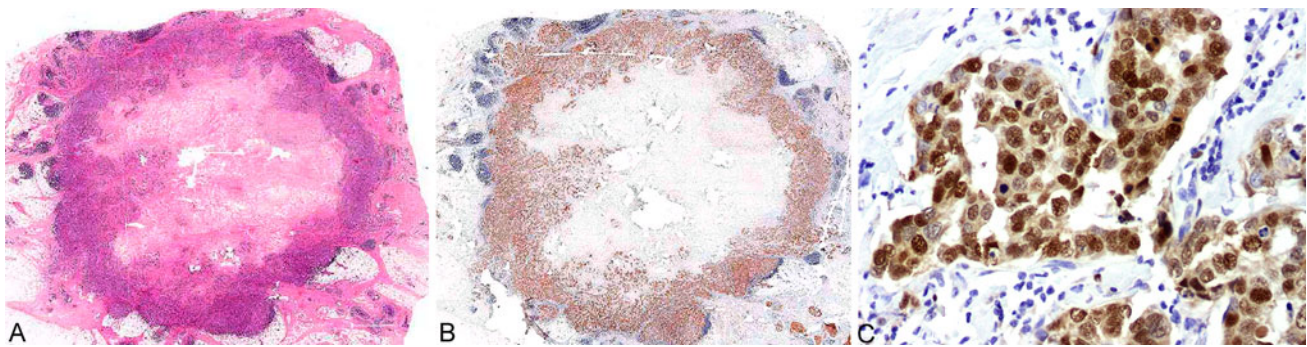


Fig. 2 Id4 expression in a triple-negative breast carcinoma with large central acellular zone of necrosis (LCAZ). **a** Morphology of a TNBC with LCAZ (hematoxylin and eosin stain) ($\times 10$). **b, c** Strong and diffuse nuclear staining of Id4 in the same tumor (**b** $\times 10$, **c** $\times 400$)

Table 4 Id4 and basal markers in 12 cases with LCAZ

Case#	Age (years)	Size (cm)	MBR grade	Id4 %	CK14	CK5/6	EGFR
1	53	2	2	10	Pos	Pos	Pos
2	39	1.2	2	20	Pos	Pos	Pos
3	42	2.0	3	20	Pos	Neg	Pos
4	44	0.7	3	30	Pos	Neg	Neg
5	61	2.5	3	30	Pos	Pos	Pos
6	36	1.8	3	30	Pos	Pos	Neg
7	50	1.7	3	50	Pos	Pos	Neg
8	49	2.5	3	70	Pos	Neg	Pos
9	50	8.0	3	80	Pos	Pos	Pos
10	56	1.8	3	80	Pos	Pos	Pos
11	62	1.1	3	90	Pos	Neg	Neg
12	60	2.5	3	90	Pos	Pos	Pos
Summary	Mean age 50	Mean size 2.3	Grade 2 (2/12) Grade 3 (10/12)	Positive 12/12 Mean % positivity 50	Positive 12/12	Positive 8/12 negative 4/12	Positive 8/12 negative 4/12

MBR grade modified Bloom–Richardson grade, Pos positive, Neg negative

Table 5 Id4 and basal markers in 11 apocrine carcinomas

Case#	Age (years)	Size (cm)	MBR Grade	Id4 %	CK14	CK5/6	EGFR
1	54	2.5	2	0	Neg	Pos	Pos
2	82	2.2	2	0	Neg	Neg	Pos
3	64	0.6	1	0	Neg	Neg	Neg
4	57	2.5	2	0	Neg	Neg	Pos
5	73	0.7	2	0	Neg	Neg	Pos
6	75	2.5	2	0	Neg	Neg	Pos
7	76	0.7	2	0	Neg	Neg	Pos
8	58	0.9	2	0	Neg	Pos	Pos
9	78	1.4	3	0	Neg	Neg	Pos
10	78	1.4	2	1 (focal)	Neg	Pos	Pos
11	55	1.4	2	5	Neg	Neg	Pos
Summary	Mean age 68	Mean size 1.5	Grade 1 (1/11) Grade 2 (9/11) Grade 3 (1/11)	Id4 negative (9/11) Focally positive (2/11) (1 and 5 %)	Negative 11/11	Positive 3/11 Negative 8/11	Positive 10/11 Negative 1/11

MBR grade modified Bloom–Richardson grade, Pos positive, Neg negative

Table 6 Id4 and basal markers

	Id4+	Id4−	<i>p</i>
CK14			
Positive	43	4	<0.0001
Negative	33	21	
CK5/6			
Positive	44	6	0.0993
Negative	32	19	
EGFR			
Positive	51	17	0.7774
Negative	25	7	
CK5/6 and/or EGFR			
Positive	61	17	0.7822
Negative	15	7	
CK5/6 and/or CK14			
Positive	55	8	0.0041
Negative	21	17	

All 11 apocrine TNBCs were negative for CK14 and 8/11 (73 %) were also negative for CK5/6. EGFR was positive in 10 out of 11 apocrine carcinomas, with 7 showing uniform and strong membranous immunoreactivity in >30 % of the tumor cells.

Id4 expression and patient ethnicity

The TNBCs of 17/21 (80 %) patients of Jewish ethnicity showed Id4 positivity in over 5 % of the tumor cells versus TNBCs in 46/80 (58 %) patients from other ethnic groups ($p = 0.048$). Aside from Jewish patients, the TNBC in patients of non-Jewish ethnicity, including African-American, did not show significant differences with regard to Id4 positivity.

Survival analysis

Follow-up data were available for 88/101 (87 %) patients with TNBC. The median follow-up time was 72.9 months (range 3.7–136.4 months). We found no statistically significant correlation between Id4 positivity and patient outcome. Overall survival and disease-free survival of patients with Id4 positive TNBC were not significantly different from those of patients with Id4 negative TNBC, independent of the percentage (5, 20 and 50 % or greater) of Id4 positivity used as cutoff for analysis.

Follow-up data were available for all 113 patients with non-TNBC. The median follow-up time was 79 months (range 3–134 months). All 6 patients with non-TNBC showing focal Id4 positivity were alive with no evidence of disease at last follow-up (median 84 months, range 77–103 months). Due to the low number of Id4-positive

non-TNBCs and lack of events in this group of patients, we did not perform statistical analysis to compare the outcome of Id4-positive versus Id4-negative non-TNBCs.

Discussion

TNBCs are a morphologically heterogeneous group of tumors. They consist mainly of high grade invasive ductal carcinoma of NOS, but include special subtypes such as medullary, metaplastic, and apocrine carcinomas. Morphologic features common to most TNBCs include nodular growth with a pushing border of invasion, poorly differentiated histology, high mitotic rate, prominent lymphocytic infiltrate, and extensive areas of necrosis such as geographic necrosis or LCAZ [29, 32].

Basal-like breast carcinomas are classified by gene expression profiling [33]. They overlap significantly with TNBCs, but the two groups of tumors are not identical [8]. Investigators have proposed the use of immunohistochemical markers as a surrogate method for the identification of basal-like breast carcinomas. Nielsen [34] and Livasy [32] identified basal-like breast cancer as ER and HER2 negative tumors that express CK5/6 and/or EGFR, whereas Rakha et al. [10] used positivity for a basal cytokeratin (CK5/6, CK17, CK14). At present, however, no immunohistochemical panel identifies all basal-like breast carcinomas with 100 % sensitivity and specificity. Homologous DNA repair is critically altered in most familial breast carcinomas associated with *BRCA1* germline mutation, which constitute the prototype of basal-like carcinomas. Although somatic mutation of the *BRCA1* gene is not commonly encountered in sporadic breast carcinomas, some of the morphologic features of *BRCA1*-deficient tumors [35] occur in TNBCs not associated with *BRCA1* germline mutation. Functional inactivation of the *BRCA1* gene could play an important role in the pathogenesis of these tumors.

Id4 is a member of the Id (inhibitor of DNA binding) family of proteins. Id proteins inhibit the functions of basic helix-loop-helix transcription factors by blocking the ability of these factors to bind DNA [17], and exert an important role in mammalian embryogenesis [36], angiogenesis [22, 37–39], and the maintenance of cancer stem cells [40]. Increased Id4 mRNA levels are found in small cell lung cancer [41], and positive Id4 nuclear immunoreactivity is present in glioblastoma [42] and malignant rhabdoid tumors [43]. Conversely, few studies have documented Id4 epigenetic inactivation by promoter hypermethylation in a wide range of human malignancies including mammary [44, 45], gastric [46], colorectal [47] and prostate adenocarcinoma [48], leukemia, [49] and lymphoma [50], suggesting that Id4 is a tumor-suppressor.

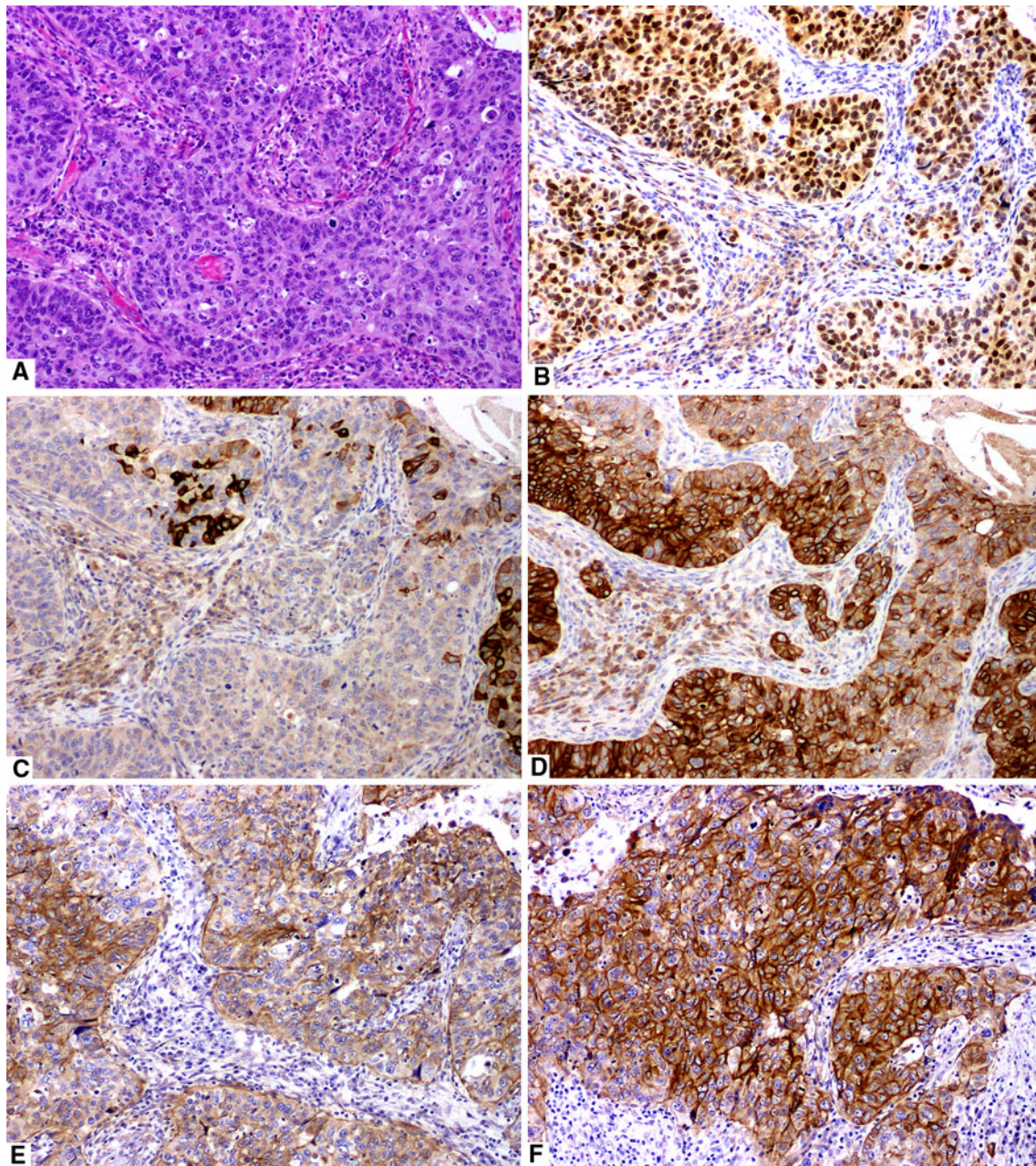


Fig. 3 Expression of Id4 and of the basal markers CK5/6, CK14 and EGFR in a triple-negative breast carcinoma. **a** Tumor morphology (hematoxylin and eosin stain). **b–e** Id4, CK5/6, CK14, and EGFR

immunoreactivity in the same tumor. Note that Id4 (**b**) CK5/6 (**c**) and CK14 (**d**) colocalizes in the same area, whereas EGFR positivity is present in a different area in **f** ($\times 200$)

Id4 downregulates *BRCA1* gene expression in vitro [18]. In turn, it appears that *BRCA1* can downregulate the expression of Id4, as part of a regulatory loop balancing the expression of both genes [51].

microRNAs participate in the modulation of *BRCA1* signal by Id4. microRNA-335 simultaneously upregulates the known *BRCA1* activators ER α , IGF1R, SP1, and downregulates Id4, a *BRCA1* repressor [19]. In one study, overexpression of microRNA-335 was associated with a significant increase of *BRCA1* mRNA level and with

marked reduction in Id4 mRNA, supporting a functional predominance of Id4 in *BRCA1* gene regulation [19]. In the same study, microRNA-335 levels positively correlated with ER and *BRCA1* expression in a group of 30 sporadic breast cancers [19]. Positive feedback regulation of microRNA-335 expression by estrogens was also documented in breast cancer cell line [19].

We have previously reported an inverse correlation between Id4 mRNA and ER positivity in the normal breast epithelium [23], consistent with a role of Id4 in the

physiologic modulation of the mammary gland. We have also documented an inverse relationship between ER immunoreactivity and Id4 mRNA signal in a series of ER-positive breast carcinomas, and in ER-positive ductal carcinoma in situ and atypical ductal hyperplasia, non-obligate precursors of ER-positive breast cancer [23]. In this setting, Id4 appears to counterbalance the activities of BRCA1 and ER, opposing ER-driven tumorigenesis. Vice versa, Id4 expression could promote tumorigenesis via downregulation of BRCA1 in ER-negative carcinomas.

To evaluate this hypothesis, we assessed the expression of Id4 in TNBCs. Our data demonstrate that Id4 is highly expressed in TNBCs, in marked contrast to ER-positive cancers. We note, however, that Id4 levels in TNBCs vary widely with about 40 % of tumors showing positivity in more than half of the neoplastic cells and about one-fourth displaying positivity in 5–50 %. Our findings are in agreement with those of Turner et al. [12], who found that Id4 mRNA levels are 9.1-fold higher in basal-like breast cancer than in matched non-basal-like control tumors. Turner et al. identified basal-like carcinomas as positive for CK5/6, but our results show stronger correlation between Id4 and CK14 than between Id4 and CK5/6. We also observed that TNBCs with LCAZ, a subset of TNBCs with distinctive morphology and characterized by very poor prognosis [30, 52], show strong positivity for both Id4 and CK14.

When subject to unsupervised gene array analysis, apocrine carcinomas consistently cluster together as a distinct subtype [26, 53]. In our study, most (8/11; 73 %) apocrine carcinomas were negative for the basal cytokeratin CK5/6, consistent with prior data [26]. All apocrine carcinomas in our study were also CK14 negative, but showed positive EGFR staining, consistent with a recent report by Vranic et al. [54]. We found that Id4 expression is extremely rare in apocrine TNBCs. Apocrine carcinomas are unique among TNBCs because they express AR, which documents activity of a hormonal pathway. Based on our prior observation that in breast carcinomas ER and Id4 are inversely correlated [23], we speculate that a similar inverse relationship also exists between Id4 and AR, but our hypothesis needs further testing.

No information regarding *BRCA1* mutation status was available for the patients in our study. Even though in our series Id4 positivity was statistically higher in patients with TNBC and Jewish ethnicity, the latter group represents only a fifth of all our cases, whereas about 75 % of all TNBC were Id4 positive. *BRCA1* (and *BRCA2*) mutations are identified in less than 10 % of all patients with breast cancer [55–57], and *BRCA* germline mutations carriers account for only about 11–12 % of young patients with TNBC [58, 59]. Based on these data, it is reasonable to assume that most of the patients in our series were not *BRCA1* (or *BRCA2*) germline mutation carriers. Although

it would be interesting to know the distribution of Id4 in TNBCs occurring in *BRCA1* germline mutation carriers, the validity of our data documenting overexpression of Id4 in TNBCs remains unaltered.

Id4 positive TNBCs did not have worse clinical outcome than Id4 negative tumors. These data are consistent with the findings in two recent series, which have reported that the overall prognosis of TNBCs does not significantly differ in patients with or without *BRCA*-germline mutation [60, 61]. A recent study has reported high Id4 expression in the cancer stem cells of a mouse mammary cancer cell line. In the same study, knockdown of Id4 expression suppressed the stem cell properties [40]. The expression of stem cell markers is reported to be high in basal-like breast carcinoma compared to other breast cancer subtypes [62, 63]. Our finding of high Id4 expression in basal-like breast carcinoma links these observations. However, assuming that Id4 positivity in TNBC reflects a stem cell enriched population, the stem cell phenotype does not appear to predict clinical outcome, as we found no significant difference in overall survival and disease-free survival between Id4 positive and Id4 negative TNBCs.

In summary, our results document high expression of intranuclear Id4 protein in TNBCs, in contrast to most non-triple-negative tumors. These findings suggest that Id4 overexpression plays a role in the downregulation of *BRCA1* in sporadic TNBCs of patients without *BRCA1* germline mutation, and provide new insight into the biology of these tumors.

Disclosure The authors declare that neither pharmaceutical nor industry support was provided for this work. No funding for this project was received from any of the following organizations: National Institutes of Health (NIH); Wellcome Trust; Howard Hughes Medical Institute (HHMI); or other(s).

Conflict of interest The authors declare no conflict of interest.

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