# Effects of elevated ozone on chlorophyll *a* fluorescence in symptomatic and asymptomatic leaves of two tomato genotypes

E. DEGL'INNOCENTI, L. GUIDI\* and G.F. SOLDATINI

Dipartimento di Chimica e Biotecnologie Agrarie, University of Pisa, Via del Borghetto 80, I-56124 Pisa, Italy

# Abstract

Two different genotypes of Lycopersicon esculentum Mill. (cv. Cuor di Bue, O<sub>3</sub>-sensitive and line 93.1033/1,  $O_3$ -resistant) were treated with a single dose of ozone (150 mm<sup>3</sup> m<sup>-3</sup> for 3 h). The PS 2 activity was examined by measurements of chlorophyll a fluorescence on symptomatic and asymptomatic leaves. Symptoms were evident on the 4<sup>th</sup> leaves from the bottom, in both genotypes, while the 2<sup>nd</sup> leaves of the line 93.1033/1 were asymptomatic. In these leaves, the net photosynthetic rate  $(\tilde{P_N})$  did not change even if the  $F_v/F_m$  ratio significantly decreased. A strong reduction in P<sub>N2</sub> mostly due to the stomatal closure, was observed in Cuor di Bue. The non photochemical quenching coefficient  $(q_{NP})$  and the degree of PS 2 reaction centres closure  $(1-q_P)$  were higher, while the quantum efficiency of PS 2 photochemistry ( $\Phi_{PS2}$ ) and quantum efficiency of excitation energy capture ( $\Phi_{exc}$ ) were lower in O<sub>3</sub> treated leaves of both genotypes. The limitation of photosynthesis was shown also by a decrease in the parameter %P, which diminished compared to controls in both genotypes. The response of the two genotypes for the energy fraction dissipated as thermal energy in the PS 2 antennae (%D) was similar. The fraction of %P remained lower during the recovery in symptomatic leaves of the resistant line as compared to the controls, whereas %X, which represents the amount of light energy that is not utilized in photochemistry or dissipated in the PS 2 antennae, significantly rose in the asymptomatic leaves of this line and in both the leaves of Cuor di Bue. From data obtained we concluded that ozone affected the plants independently on the appearance of visible symptoms of injury because the leaves without visible symptoms of both the genotypes were negatively influenced.

Additional key words: Lycopersicon esculentum, net photosynthetic rate, quenching analysis.

## Introduction

Ozone is one of the most dangerous phytotoxic air pollutants (Matyssek *et al.* 1997). Its toxicity is principally attributed to the oxidizing potential which, in plants, determines the production of the activated species of oxygen (AOS) (Schraudner *et al.* 1998, Rao and Davis 1999).

Photosynthesis is particularly sensitive to ozone (Heath 1994, Schraudner *et al.* 1997). The reduction in the net photosynthetic rate can be determined by a direct

effect of  $O_3$  on the stomatal opening (Robinson *et al.* 1998), but also by an alteration of the electron transport rate and the biochemical activity of the Calvin cycle (Calatayud and Barreno 2001, Calatayud *et al.* 2002, Degl'Innocenti *et al.* 2002b).

The responses of plants to ozone differ among species and also among cultivars of the same species (Guidi *et al.* 2000). In a previous work (Guidi *et al.* 2005) we have reported the differing  $O_3$  sensitivity of two tomato

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Abbreviations:  $c_i$  - intercellular CO<sub>2</sub> concentration; E - transpiration rate; ETR - electron transport rate;  $F_0$  - minimum Chl fluorescence in dark adapted state;  $F_m$  - maximum Chl fluorescence in dark adapted state;  $F_m$ ' = maximum Chl fluorescence with all PS 2 reaction centres closed in light adapted state;  $F_s$  - Chl fluorescence in steady state conditions;  $F_v$  - variable Chl fluorescence in dark adapted state;  $F_v'$  - variable Chl fluorescence in light adapted state;  $F_v/F_m$  - photochemical PS 2 photochemistry in dark adapted state;  $g_s$  - stomatal conductance to water vapour; PFD - photon flux density;  $P_N$  - net photosynthetic rate; PS 2 - photosystem 2;  $q_{NP}$  - non-photochemical quenching;  $q_P$  - photochemical quenching;  $\Phi_{exc}$  - excitation capture efficiency of PS 2;  $\Phi_{PS2}$  - actual PS 2 efficiency; %D - fraction of light absorbed in PS 2 that is dissipated in the PS 2 antenna; %P - fraction of light absorbed that is used in photochemistry; %X - fraction of light absorbed that is not used or dissipated in the PS 2 antenna.

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<sup>\*</sup> Corresponding author; fax: (+39) 50 2216630; e-mail: guidilu@agr.unipi.it

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genotypes which are known also for their different susceptibility to pathogens: cv. Cuor di Bue, sensitive to *Verticillium* spp., *Fusarium* spp. and to the tobacco mosaic virus (TMV) and line 93.1033/1, resistant to the same pathogens. In that research we found that, in terms of visible symptoms of injury, the line 93.1033/1 showed resistance to ozone, while the pathogen-sensitive Cuor di Bue showed visible symptoms of a damage probably due

## Materials and methods

**Plant material:** Seeds of two genotypes of *Lycopersicon* esculentum Mill. (cv. Cuor di Bue and line 93.1033/1, sensitive and resistant to some pathogens, respectively) were sown in a sterilized soil. The seedlings emerged 20 -30 d later and were grown thereafter in a greenhouse (temperature of  $25 \pm 3$  °C, relative humidity  $75 \pm 5$  %, 12-h photoperiod at an irradiance of about 400 µmol m<sup>-2</sup> s<sup>-1</sup>). The tomato plants were grown to the 5<sup>th</sup> leaf stage and then placed in 2 growth chambers, one to permit ozone treatment, 24 h before the fumigation. In the experiment, we analyzed the 2<sup>nd</sup> (mature) and the 4<sup>th</sup> (young) leaves from the bottom. The measurements in symptomatic leaves were carried out on asymptomatic area.

**Ozone fumigation:** The O<sub>3</sub> treatments were performed in a controlled-environment chamber (*Cavallo*, Milan, Italy): more details of the O<sub>3</sub> exposure are reported in Guidi *et al* (2000). A single ozone fumigation was carried out for 3 h at an irradiance of 350 µmol m<sup>-2</sup> s<sup>-1</sup>, temperature of  $25 \pm 3.1$  °C and relative humidity  $81 \pm 7$  %. The plants were treated for 3 h with a single dose of O<sub>3</sub> ( $150 \pm 6.7$  and  $150 \pm 2.3$  mm<sup>3</sup> m<sup>-3</sup>, respectively for the first and second experiment) or maintained in charcoalfiltered air. After the end of the O<sub>3</sub> fumigation, ozonated plants were put in the same chamber with filtered air, where the control plants were placed.

**Gas exchange analysis:** Gas exchange measurements [net photosynthetic rate ( $P_N$ ), stomatal conductance ( $g_s$ ), transpiration rate (E) and intercellular CO<sub>2</sub> concentration ( $c_i$ )] were carried out at the end of the treatment on the second fully developed leaf from the bottom and using an open system (*CMS-400, Walz*, Effeltrich, Germany; for details see Guidi *et al.* 1997). Measurements were carried out at an irradiance of about 800 µmol m<sup>-2</sup> s<sup>-1</sup>, temperature 25 ± 2.3 °C, relative humidity 70 ± 5.2 %, CO<sub>2</sub> concentration 350 µmol mol<sup>-1</sup> and O<sub>2</sub> concentration 21 %.

**Chlorophyll fluorescence analysis:** Modulated chlorophyll *a* fluorescence measurements were made with a *PAM-2000* fluorometer (*Walz*, Effeltrich, Germany) on second and fourth leaves, counting from the bottom, at the end of the fumigation and 24 and 48 h after the end of O<sub>3</sub> treatment. Leaves were pre-darkened for 40 min in leaf-clips before measurement. Firstly a weak irradiance (< 0.1  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>) was used to obtain basic

to ozone. This different sensitivity to ozone was shown by seedlings which presented only two leaves.

The aim of this paper was the evaluation of the different responses of symptomatic and asymptomatic leaves of these two tomato genotypes to an acute ozone treatment in terms of changes in chlorophyll *a* fluorescence, including quenching analysis, and in gas exchange measurements.

fluorescence ( $F_0$ ). Successively, a 1-s saturating pulse of white light (15 000  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>) was given to determine maximum fluorescence (F<sub>m</sub>) when all PS 2 reaction centres are closed in the dark. These parameters were used for the calculation of the  $F_v/F_m$  ratio which means the maximum photochemical capacity of PS 2. The saturation pulse method was used for the analysis of quenching components according to Schreiber et al. (1986). Intermittent, short-term illumination by sufficiently strong radiation causes a transient, but complete removal of photochemical quenching, prompting a corresponding increase in variable fluorescence,  $F_v$  to  $F_v$ '; the residual quenching is assumed to have nonphotochemical nature. The photon flux density (PFD) of the actinic light was maintained at 300  $\mu$  umol m<sup>-2</sup> s<sup>-1</sup> and a sequence of saturating flashes of white light 15 000  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup> was given firstly every 20 s and then increases to 40 s and then to 80 s. The overall time of illumination during the quenching analysis was 20 min. After the saturating pulse, the maximum fluorescence yield reached the  $\bar{F_{m}}\sp{\prime}$  value and the actinic radiation allowed both steady-state photosynthesis and steady-state fluorescence yield  $(F_s)$ . Determination of quenching components q<sub>P</sub> and q<sub>NP</sub> was done as defined by Schreiber *et al.* (1986), *i.e.*  $q_P = (F_m' - F_s)/F_v'$  and  $q_{NP} = 1 - (F_v'/F_v)$ . Minimum fluorescence in the light-adapted state  $(F_0)$ was determined immediately after turning off the actinic source in the presence of a far-red (7  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>) background for 10-s to ensure maximum oxidation of PS 2 electron acceptors. The actual quantum yield of PS 2 photochemistry ( $\Phi_{PS2}$ ) and the quantum efficiency of open PS 2 reaction centres in light conditions ( $\Phi_{exc}$ ) were computed as  $(F_m' - F_s)/F_m'$  and  $F_v'/F_m'$ , respectively, as reported by Genty *et al.* (1989). Apparent rates of photosynthetic electron transport (ETR) were estimated as  $(\Phi_{PS2} \times 0.5) \times (PPFD \times 0.8)$  where factor 0.5 represents the excitation of both PS 2 and PS 1 and factor 0.8 represents the average value for leaf absorbance.

The fraction of radiation absorbed that are dissipated in the PS 2 antennae (%D) and utilized in PS 2 photochemistry (%P) were estimated as  $[1-(F_v'/F_m')] \times 100$  and  $[(F_v'/F_m') \times q_P] \times 100$ , respectively (Demmig-Adams and Adams 1996). It should be noted that although the parameter %P is proportional to  $\Phi_{PS2}$ , both are still commonly used in the literature. The fraction of radiation absorbed by PS 2 which is not utilized in photochemistry or dissipated in the PS 2 antennae (%X), was estimated from  $[(F_v'/F_m') \times (1-q_P)] \times 100$ , according to Demmig-Adams and Adams (1996).

**Statistical analysis:** Two experiments were done in two different periods of the year (May - June and July 2003). For each experiment, 2 growth chambers were used. The chambers were equivalent, but one was fed with filtered air and the other with ozone. Ten-fifteen potted plants were put in each chamber. After treatment, two plants, similar in their growth, were chosen from each chamber and the  $2^{nd}$  and the  $4^{th}$  leaves from every plant were utilized. The data obtained from each experiment were

#### Results

**Visual symptoms:** The day after the end of the fumigation, visible symptoms of injury due to the presence of  $O_3$  appeared on the young leaves on both genotypes. Symptoms were principally located on the third and fourth leaves, counting from the bottom, and were characterized by chlorotic and/or necrotic areas on the margins of the leaves.

**Gas exchange:** Measurements were carried out only in the asymptomatic leaves at the end of the  $O_3$  fumigation. In the sensitive Cuor di Bue a significant decrease in  $P_N$ , accompanied by a reduction in  $g_s$  was observed following the ozone treatment (Table 1). Also E and  $c_i$  decreased in  $O_3$ -treated plants. In the resistant line 93.1033/1, no changes in  $P_N$  and  $c_i$  were observed after the  $O_3$ fumigation, while a significant increase in  $g_s$  and E was recorded (Table 1).

**Chlorophyll** *a* **fluorescence:** Minimum fluorescence yield  $F_0$  in line 93.1033/1 was significantly greater in ozonated leaves than in controls (Table 2) while no effects were found in relation to the time and leaf age. There were no significant two-way or three-way interactive effects of different treatments on  $F_0$  in line

pooled, in order to obtain 4 replicates for both leaves  $(2^{nd} \text{ and } 4^{th})$ . Data were subjected to three way analysis of variance in which the age of leaves, ozone treatment and time represent the three factors of variability. Multiple comparison was conducted on significant (P<0.05) three-way interactions using least square means analysis. The significance of pairwise comparisons was determined by using a Bonferroni adjustment with  $\alpha = 0.05/14 = 0.0036$  for three-way interaction.

Data measured as a percentage were angularly transformed previously to the *ANOVA*. For gas exchange analysis, the one way *ANOVA* was applied with the  $O_3$  treatment as variability factor.

93.1033/1. Maximum fluorescence  $F_m$  and the ratio  $F_v/F_m$  significantly decreased because of the presence of O<sub>3</sub>, with the major effects detected on leaf No. 4 at the end of the experiment (Fig. 1).

Ozone induced alterations in the values of  $F_0$  and the ratio  $F_v/F_m$  in the 2<sup>nd</sup> and 4<sup>th</sup> leaves of Cuor di Bue, while no changes were evident in the values of  $F_m$  (Fig. 1, Table 2). The time factor affected maximum fluorescence in this tomato variety, but not the  $F_0$  and  $F_v/F_m$  ratio. Considering the significant leaf age-ozone interaction (P < 0.05), the comparison showed that the air pollutant increased the  $F_0$  in the symptomatic leaves compared to the 4<sup>th</sup> one (asymptomatic) but there was no effect of time. The three-way interaction was not significant for  $F_o$ ,  $F_m$  and  $F_v/F_m$  in Cuor di Bue (Table 2).

The quenching coefficients  $(1-q_P)$  and  $q_{NP}$  of both genotypes were significantly higher in the ozonated leaves than in controls, according to time and leaf age treatments (Fig. 2, Table 2). In line 93.1033/1 the reduction state of  $Q_A$ , *i.e.*  $(1-q_P)$ , significantly increased in both the leaves at the end of the fumigation and also during the recovery (Fig. 2). The behavior of  $q_{NP}$ coefficient was different, changing significantly in relation to the presence of  $O_3$  and the time of treatment.

Table 1. Gas exchange parameters determined in two genotypes of *Lycopersicon esculentum* leaves exposed to a single dose of  $O_3$  (150 mm<sup>3</sup> m<sup>-3</sup> for 3 h). Controls were represented by plants maintained in filtered air. Measurements were carried out at the end of the fumigation (O<sub>3</sub>). Analysis were carried out on the 2<sup>nd</sup> leaves counting from the bottom. Each data represents the mean of 4 replicates ± standard deviation. Analysis were made in saturating irradiance (about 800 µmol m<sup>-2</sup>s<sup>-1</sup>), temperature of 25 ± 2.3 °C, RH of 70 ± 5.2 % and 350 µmol(CO<sub>2</sub>) mol<sup>-1</sup>. P<sub>N</sub> - net photosynthetic rate; g<sub>s</sub> - stomatal conductance; E - transpiration rate; c<sub>i</sub> - intercellular CO<sub>2</sub> concentration. Data are subjected to the one way *ANOVA*; significant differences between ozonated and controls at the *P* < 0.05 (\*) and *P* < 0.01 (\*\*)

Genotype	Treatment	P <sub>N</sub> [μmol(CO <sub>2</sub> ) m <sup>-2</sup> s <sup>-1</sup> ]	g <sub>s</sub> [mmol(H <sub>2</sub> O) m <sup>-2</sup> s <sup>-1</sup> ]	E [mmol(H <sub>2</sub> O) m <sup>-2</sup> s <sup>-1</sup> ]	c <sub>i</sub> [µmol(CO <sub>2</sub> ) mol <sup>-1</sup> ]
93.1033/1	control	$5.19 \pm 1.31$	238 ± 47.1	$3.8 \pm 0.78$	$302 \pm 3.0$
Cuor di Bue	O <sub>3</sub> control O <sub>3</sub>	$6.22 \pm 0.70$ $5.20 \pm 0.81$ $3.80 \pm 0.95*$	$301 \pm 69.2^{*}$ $254 \pm 16.1$ $143 \pm 33.0^{**}$	$4.9 \pm 1.12^*$ $4.2 \pm 0.16$ $2.4 \pm 0.54^{**}$	$303 \pm 4.3$ $304 \pm 3.0$ $295 \pm 1.4**$

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Table 2. Analysis of variance for parameters of chlorophyll fluorescence kinetics in tomato leaves (93.1033/1 and Cuor di Bue). ns - not significant, \* - P < 0.05, \*\* - P < 0.01, \*\*\* - P < 0.001.

Genotype		d.f.	F <sub>0</sub>	F <sub>m</sub>	$F_v/F_m$	1-q <sub>P</sub>	q <sub>NP</sub>	$\Phi_{PS2}$	$\Phi_{ m exc}$	ETR
93.1033/1	ozone	1	57.11***	75.81***	149.72***	282.06***	115.37***	449.38***	198.13***	286.99***
	time	2	2.71ns	4.60*	13.10***	14.64***	11.90***	25.93***	17.99***	10.87***
	age	1	0.02ns	31.25***	37.66***	0.20ns	0.08ns	0.99ns	19.56***	6.08*
	age×time	2	1.08ns	12.65***	12.90***	6.50**	0.74ns	1.81ns	2.34ns	1.75ns
	age×ozone	1	1.08ns	53.31***	50.59***	3.32ns	1.80ns	6.76*	17.49***	13.48***
	time×ozone	2	2.71ns	4.60*	13.10***	14.63***	11.90***	25.92***	17.99***	10.87***
	time×age×ozone	2	0.15ns	12.65***	12.91***	6.50**	0.74ns	1.81ns	2.34ns	1.65ns
Cuor di Bue	ozone	1	70.85**	0.69ns	13.07***	155.32***	248.39***	1721.55***	441.44***	516.19***
	time	2	0.13ns	5.74**	1.95ns	30.80***	2.00ns	60.76***	14.04***	58.00***
	age	1	0.01ns	0.06ns	0.003ns	0.58ns	10.12**	0.04ns	8.72**	0.35ns
	age×time	2	1.68ns	0.05ns	0.31ns	0.83ns	0.19ns	0.25ns	0.58ns	0.18ns
	age×ozone	1	6.92*	2.95ns	0.45ns	0.49ns	12.17**	1.84ns	0.41ns	0.08ns
	time×ozone	2	2.52ns	5.63**	2.34ns	23.67***	0.53ns	60.76***	14.04***	58.01***
	time×age×ozone	2	0.53ns	0.06ns	0.31ns	0.43ns	0.65ns	0.25ns	0.58ns	0.18ns



Fig. 1. Minimum and maximum fluorescence yields ( $F_0$  and  $F_m$ ) and the ratio  $F_v/F_m$  in the 2<sup>nd</sup> and 4<sup>th</sup> leaves, counting from the bottom, of *Lycopersicon esculentum* 93.1033/1 and Cuor di Bue. Plants were exposed to a single dose of O<sub>3</sub> (150 mm<sup>3</sup> m<sup>-3</sup> for 3 h) (*grey bars*) or to charcoal-filtered air (*white bars*). The measurements were carried out at 0, 24 and 48 h after the end of the funigation. Each value represents the mean of four replicates. *Bars* indicate the standard deviation. Different letters over the bars indicate significant differences between means in pairwise comparison within a three-way interaction using a Bonferroni adjustment (P < 0.0036).

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Fig. 2. Reduction state  $(1-q_P)$  and the non photochemical quenching coefficient  $(q_{NP})$  in the 2<sup>nd</sup> and 4<sup>th</sup> leaves, counting from the bottom, of *Lycopersicon esculentum* 93.1033/1 and Cuor di Bue. Plants were exposed to a dose pulse of O<sub>3</sub> (150 mm<sup>3</sup> m<sup>3</sup> for 3 h) (*grey bars*) or to charcoal-filtered air (*white bars*). The measurements were carried out at 0, 24 and 48 hafter the end of the fumigation. Each value represents the mean of four replicates. *Bars* indicate the standard deviation. Different letters over the bars indicate significant differences (*P* < 0.0036).

The interaction between these two factors was also significant (Table 2). There were no other significant two-way or three-way interactive effects of treatments on the  $q_{NP}$  coefficient in line 93.1033/1.

In Cuor di Bue a significant effect on  $(1-q_P)$  was attributable to ozone and time and also the interaction between these two factors was significant (Table 2). This parameter did not change in relation to time and no effects were recorded for the other two-way or three-way interactive effects of treatments (Table 2). Nonphotochemical quenching  $q_{NP}$  was significantly higher in the 4<sup>th</sup> leaf of Cuor di Bue, while no differences were found in relation to time. Two-way interaction between ozone and leaf age was significant while time × ozone and the three-way interaction had no reflections in  $q_{NP}$ .

The actual PS 2 efficiency ( $\Phi_{PS2}$ ) significantly decreased following of ozone treatment in both the genotypes (Table 2, Fig. 3), with major effects recorded the days after the end of the treatment. However, there was no significant difference between the 2<sup>nd</sup> and 4<sup>th</sup> leaf in both the genotypes and the two-way interaction between leaf age and time was not significant. In line 93.1033/1 the interaction between leaf age and ozone and time × ozone was significant, with major effect attributable to ozone treatment in the 2<sup>nd</sup> leaf on the day after the end of the exposure to the pollutant (Table 2, Fig. 3). In Cuor di Bue the parameter  $\Phi_{PS2}$  significantly changed only in relation to time without effects due to the leaf age.

The values of  $\Phi_{exc.}$ , which reflect the excitation capture efficiency of open PS 2 centres in the light,

significantly decreased both in symptomatic and asymptomatic leaves of Cuor di Bue following the O<sub>3</sub> exposure compared to controls, as well as the line 93.1033/1 (Table 2, Fig. 3). The decrease in  $\Phi_{exc.}$  was strong during the days after the end of the fumigation (Fig. 3). There were significant two-way interactive effects of treatments (leaf age × ozone and ozone × time) on  $\Phi_{exc.}$  values in line 93.1033/1, while no significant three-way interactive effects were observed. In Cuor di Bue only the two-way interaction between time and ozone was significant (Table 2).

The electron transport rate significantly decreased also in both symptomatic and asymptomatic leaves of the line 93.1033/1 at the end of fumigation without recovering when plants were put in  $O_3$ -free air. Comparisons showed that ozone significantly decreased ETR in both the 2<sup>nd</sup> and the 4<sup>th</sup> leaves in line 93.1033/1, but there was no significant interactive three-way effect (Table 2). In Cuor di Bue the three factors of variability (ozone, leaf age and time) significantly reduced ETR values (Table 2, Fig. 3) and the only interactive effect was between leaf age and ozone (Table 2).

Following the ozone treatment, in the resistant line 93.1033/1 the amount of radiation absorbed by antennae and utilized in photochemistry (%P) significantly decreased during the time of the experiment (Table 3, Fig. 4), while no differences were found between the  $2^{nd}$  and  $4^{th}$  leaves. Only the two-way interactive effect of ozone × time was significant. A similar response was observed for %P in Cuor di Bue (Table 3).



Fig. 3. Actual photochemical efficiency ( $\Phi_{PS2}$ ), intrinsic PS 2 efficiency ( $\Phi_{exc}$ ) and electron transport rate (ETR) in the 2<sup>nd</sup> and 4<sup>th</sup> leaves of *Lycopersicon esculentum* 93.1033/1 and Cuor di Bue. Plants were exposed to a single dose of O<sub>3</sub> (150 mm<sup>3</sup> m<sup>-3</sup> for 3 h) (*grey bars*) or to charcoal-filtered air (*white bars*). The measurements were carried out at 0, 24 and 48 h after the end of the fumigation. Each value represents the mean of four replicates. *Bars* indicate the standard deviation.

The amount of dissipated radiation absorbed (%D), significantly increased in both the leaves of line 93.1033/1 following the ozone exposure and the increase was more marked the days after the end of fumigation. The highest values were recorded 24 h after the end of  $O_3$  exposure in the 4<sup>th</sup> leaf when %D reached values of about 70 % (Table 3, Fig. 4). No interactive three-way effects were detected in this line. In Cuor di Bue %D increased significantly in both leaves after the ozone fumigation with a stronger increase during the recovery in  $O_3$ -free air (Table 3, Fig. 4). The interactive effect of time × ozone was significant, whereas the other two- and three-way effects were not significant (Table 3).

## Discussion

This experiment was carried out on plants which had 5 leaves and the  $O_3$  fumigation induced visible symptoms of injury only in the young leaves of both genotypes (the

Finally, the amount of radiation absorbed, which is neither used for PS 2 photochemistry nor dissipated in the PS 2 antennae (%X), significantly increased in line 93.1033/1 at the end of the fumigation with ozone (Table 3). During the recovery time (24 and 48 h after the end of the fumigation) %X high without differences attributable to the leaf age in line 93.1033/1 (Table 3), while in Cuor di Bue, the %X significantly increased in both leaves in plants exposed to ozone, but no differences were found in relation to the leaf age (Table 3). The only interactive effects was between leaf age and ozone as %X increased in the 4<sup>th</sup> leaf (Fig. 4).

4<sup>th</sup> from the bottom). This indicates that the sensitivity of the plants to ozone fumigation was related to the age of the leaf, as the young leaves of both genotypes showed

Genotype		d.f.	%P	%D	%X
93.1033/1 Cuor di Bue	O <sub>3</sub> time age age×time age×O <sub>3</sub> time×O <sub>3</sub> time×age×O <sub>3</sub> ozone time age age×time age×C <sub>3</sub> time×O <sub>3</sub> time×O <sub>3</sub>	1 2 1 2 1 2 1 2 1 2 1 2 1 2 1 2 2	356.19* 1.24* 4.65ns 5.91ns 10.08ns 0.76*** 5.28ns 189.14*** 18.94*** 1.82ns 0.57ns 0.32ns 14.72*** 0.24ns	191.13*** 19.05*** 11.24** 0.71ns 8.76** 19.05*** 0.71ns 238.03*** 8.38** 4.99* 0.31ns 0.20ns 8.38** 0.31ns	19.77*** 13.89*** 3.93*** 0.76ns 0.46ns 13.89*** 0.76ns 20.08*** 9.43** 0.05ns 1.00ns 0.11ns 8.08** 0.04ns

symptoms. In order to evaluate if changes in PS 2 activity were related to the effects of ozone on photosynthesis, chlorophyll a fluorescence measurements were carried out in leaves with visible symptoms and leaves without them of both genotypes. As both genotypes were grown in the same conditions, the effects of ozone also in the asymptomatic leaves, could be present.

Results obtained from gas exchange analysis indicate that the asymptomatic (the 2<sup>nd</sup> counting from the bottom) leaves of Cuor di Bue showed an alteration in  $P_N$  mainly due to a reduced  $g_s$ . The behavior of the resistant genotype 93.1033/1 was different: no changes were recorded in the  $P_N$ , whereas the  $g_s$  significantly increased. The data obtained indicate that the greater O<sub>3</sub> tolerance of line 93.1033/1 is not attributable to lower absorption of the pollutant. These results agree with the findings of Degl'Innocenti *et al.* (2002a) in the tobacco-tolerant cultivar BelB and also with those of Pell and Pearson (1983) and Langebartels *et al.* (1991), who found that O<sub>3</sub> did not influence stomatal closure.

Acute  $O_3$  exposure caused a significant decrease in the dark-adapted variable-to-maximum fluorescence yield ratio ( $F_v/F_m$ ) only in the young and symptomatic leaves of



Fig. 4. Fraction of absorbed radiation utilized in PS 2 photochemistry (%P), fraction of absorbed radiation dissipated in the PS 2 antennae (%D) and absorbed radiation that is not utilized in photochemistry or dissipated in the PS 2 antennae (%X) in the 2<sup>nd</sup> and 4<sup>th</sup> leaves of *Lycopersicon esculentum* 93.1033/1 and Cuor di Bue. Plants were exposed to a single dose of O<sub>3</sub> (150 mm<sup>3</sup> m<sup>-3</sup> for 3 h) (*grey bars*) or to charcoal-filtered air (*white bars*). The measurements were carried out at 0, 24 and 48 h after the end of the fumigation. Each value represents the mean of four replicates. *Bars* indicate the standard deviation. Data are angularly transformed previous of the application of *ANOVA* test.

the resistant line 93.1033/1, while the effects were minor in the asymptomatic leaves which showed also an unaffected  $P_N$ . The decline in  $F_v/F_m$  observed in leaves of this genotype was related to the increase in  $F_0$  and a decrease in  $F_m$  values. This indicates that the photochemistry of PS 2 and its ability to reduce the primary acceptor  $Q_A$  were affected by ozone, as already reported (Calatayud and Barreno 2001).

In Cuor di Bue, only a mild increase in the values of  $F_0$  was observed, indicating that  $O_3$  induced modifications at the level of the antenna pigments or in the photochemical efficiency of the reaction centers of PS 2. These perturbations determined only slight changes in the ratio of  $F_v/F_m$ .

The increase of  $(1-q_P)$  in symptomatic and asymptomatic leaves of both the genotypes suggests that ozone caused a decrease in the fraction of open PS 2 reaction centers (Krause and Weiss 1991). This could be attributable to a slow re-oxidation of Q<sub>B</sub>, which may be due to the inhibition of Calvin cycle activity, as already reported about ozonated leaves of tobacco species (Degl'Innocenti et al. 2002a). However, we found a reduction in P<sub>N</sub> only in the asymptomatic leaves of Cuor di Bue at the end of the fumigation, which was mainly attributable to stomatal closure. These results indicate that the first limitation to the  $P_N$  response to  $O_3$  in Cuor di Bue is at stomatal level. Then the reduction in stomatal conductance determined a diminished availability of CO<sub>2</sub> at mesophyllic level. Also Calatayud et al. (2003) reported that in spinach there were no apparent effects on the ratio  $F_v/F_m$  even if the  $CO_2$  assimilation rate significantly decreased. Similar results have been obtained by other authors in other plant species (Gimeno et al. 1999).

The non photochemical quenching coefficient  $q_{NP}$  was higher in leaves showing visual symptoms, but increased also in leaves that remained free of visible injury. The increased  $q_{NP}$  in both the genotypes following ozone treatment corresponds to a higher activation of defense and reparative mechanisms of non-photochemical nature.

The efficiency of excitation energy capture by open PS 2 reaction centers ( $\Phi_{exc}$ ) significantly changed in both the leaves of the two genotypes. Although the decrease in excitation capture efficiency is prevalently accompanied with an increase in energy dissipation in the antenna (Demmig-Adams *et al.* 1995), the mechanisms whereby ozone induces the reduction of this parameter are unknown, as reported also by Carrasco Rodriguez and Del Valle-Tascon (2001).

Also the  $\Phi_{PS2}$ , closely correlated with the quantum yield of non-cyclic electron flow (Genty *et al.* 1989), was

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reduced in both tomato genotypes and both symptomatic and asymptomatic leaves by  $O_3$  exposure. It indicates an inhibition of electron chain and was confirmed also by the reduction of %P, *i.e.* the amount of radiation absorbed by PS 2 antennae and used for photochemistry. The reduction in  $\Phi_{PS2}$  is due both to the decline of  $q_P$  and the increase in  $q_{NP}$ .

The data obtained in this work seem to indicate that the different response of plants to ozone varies also in relation to the leaf age. In fact, these two tomato genotypes showed a different response when exposed to a single dose of ozone in terms of visible symptoms of injury, but also at a physiological level, as shown by the chlorophyll *a* fluorescence measurements.  $P_N$  was reduced only in Cuor di Bue while no change was observed in line 93.1033/1, in which mechanisms aimed at limiting the damage to the photosynthetic apparatus can be involved. On the other hand, a similar response was already observed in an O<sub>3</sub>-tolerant cultivar of clover (Degl'Innocenti et al. 2003). Many hypotheses have been proposed to explain the effects of ozone on plants (Pell et al. 1997, Schraudner et al. 1997). In this article we suggest that ozone-induced effects are different in two tomato genotypes. In the O<sub>3</sub>-resistant line 93.1033/1 the strong decrease in the optimal photochemical PS 2 efficiency  $(F_v/F_m)$  is linked to an increase in  $(1-q_P)$ , which determined also a large fraction of PS 2 reaction centres that are incapable of stable charge separation. This decreased ability of QA re-oxidation may be due to the large fraction of excitation energy which was dissipated mostly to heat (increased q<sub>NP</sub>) and no differences were found between leaves with or without symptoms. In Cuor di Bue, the F<sub>v</sub>/F<sub>m</sub> ratio did not change significantly due to the presence of O<sub>3</sub>, but the efficiency of PS 2 photochemistry in conditions of a steady state of photosynthesis was significantly affected. Even in Cuor di Bue, the leaf age did not significantly affect the response of the plants to the pollutant.

In conclusion, the data obtained in this work show that ozone affected the plants independently on the appearance of visible symptoms of injury because the leaves without symptoms of both the genotypes were negatively influenced. Also line 93.1033/1, which in a previous report was considered as resistant to  $O_3$ , showed symptoms on the 4<sup>th</sup> leaf and alteration in the photochemical efficiency of PS 2. Considering that in this genotype a stimulation of the phenylpropanoid metabolism was observed (Guidi *et al.* 2005), we can suggest that a defence  $O_3$ -induced mechanism of response was involved in line 93.1033/1.

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