

Spatial and temporal variability in methane emissions from tree stems of *Fraxinus mandshurica* in a cool-temperate floodplain forest

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Abstract We performed field measurements on the spatial and temporal variability in CH₄ emissions from stem surfaces of mature *Fraxinus mandshurica* Rupr. trees in a floodplain forest of northern Japan. Stem CH₄ fluxes were measured by a static closed-chamber method at ca. 15 cm above ground on ten selected trees to test among-individual variability, and the diurnal and seasonal changes in three representative trees. Daytime stem CH₄ emission rates varied between 81 and 1,305 μg CH₄ m⁻² h⁻¹ among individual trees, and showed a spatial gradient apparently corresponding to the difference in water table depth at the experimental site. Stem CH₄ fluxes were quite stable throughout a 24 h period for foliated trees in August and were similar for defoliated trees in November. Large differences were observed in the magnitude of seasonal changes in stem CH₄ flux among individual trees; one sampled tree showed no clear seasonal

changes in stem CH₄ flux, while another tree exhibited drastic seasonal changes ranging larger than one order of magnitude. Results demonstrated the high variability in stem CH₄ emissions in space and time, and suggested the importance of soil temperature, water table depth and porewater CH₄ concentration as possible environmental factors controlling stem CH₄ emissions from temperate forested wetlands.

Keywords Diurnal and seasonal change · Forested wetland · *Fraxinus* · Methane emissions · Tree stem

Introduction

Methane (CH₄) has a strong greenhouse effect, and its global warming potential is 25 times greater than carbon dioxide for a 100 year time horizon (WMO 2013a). Global abundance of CH₄ as a mole fraction was 1,819 ± 1 ppb in 2012, which is 260 % of its preindustrial level of 700 ppb; an annual increase averaging 3.7 ppb year⁻¹ has been observed during the last 10 years (WMO 2013b).

Anaerobic soils in natural wetlands and rice paddies are the greatest producers of atmospheric CH₄; bottom-up global estimates of CH₄ emissions from those two sources for the decade of 2000–2009 were 177–284 Tg year⁻¹ and 33–40 Tg year⁻¹, respectively; these constitute substantial parts of the total global emission of 542–852 Tg year⁻¹ (Ciais et al.

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2013). In these ecosystems, methanogens produce CH_4 in anoxic soils. The CH_4 is subsequently released into the atmosphere by three different pathways: molecular diffusion at soil/water–atmosphere interfaces; ebullition of gas bubbles; and plant-mediated transport (Carmichael et al. 2014; Joabsson et al. 1999; Schütz et al. 1991). Among these three pathways, plant-mediated gas transport through the internal air space of plant bodies makes the greatest contribution to total CH_4 flux from soil to the atmosphere, causing more than 90 % of the total flux in various ecosystems (Schütz et al. 1989; Shannon et al. 1996; Whiting and Chanton 1992).

Studies have documented plant-mediated CH_4 transport in rice plants (Cicerone and Shetter 1981; Holzappel-Pschorn et al. 1986; Nouchi et al. 1990; Schütz et al. 1989; Seiler et al. 1984), aquatic plants (Chanton et al. 1992; Dacey and Klug 1979; Sebacher et al. 1985), and wetland herbaceous plants (King et al. 1998; Shannon et al. 1996; Whiting and Chanton 1992). These plants develop aerenchyma tissue, i.e., internal lacunae formed by cell separation or cell breakdown, in their roots and culms through which atmospheric oxygen can be transported downward to the anoxic rhizosphere (Armstrong 1979). Plant-mediated CH_4 emissions, thus, are the upward transportation of soil-borne CH_4 through the aerenchyma tissue of plants that have adapted well to anoxic soil environments (Schütz et al. 1991).

As a response to soil inundation, several flood-tolerant woody tree species, such as *Fraxinus* or *Alnus* spp. in temperate forests and some species in tropical regions, can develop various adaptive changes in the anatomy and morphology of roots and stems including aerenchyma formation that allows their roots to survive anoxic belowground environments (De Simone et al. 2002; Grosse and Schröder 1984; Hook 1984; Yamamoto et al. 1995a; Yamamoto et al. 1995b). Therefore, woody species could also most likely mediate the transport of CH_4 from the soil to the atmosphere (Carmichael et al. 2014; Schütz et al. 1991). In fact, some experimental studies in laboratories or mesocosms have demonstrated that CH_4 is transported through bodies of tree seedlings or saplings from the submerged soil layer or methane-enriched root zone to the atmosphere (Garnet et al. 2005; Machacova et al. 2013; Pangala et al. 2014; Rice et al. 2010; Rusch and Rennenberg 1998; Vann and Megonigal 2003). Furthermore, in recent field studies,

emissions of CH_4 , which were presumably produced anaerobically in the submerged soil layer, from the stem surfaces of mature trees have been measured in temperate wetland forests (Gauci et al. 2010; Terazawa et al. 2007) and a tropical forested peatland (Pangala et al. 2013), as have CH_4 emissions from pneumatophores of mangroves (Kreuzwieser et al. 2003; Purvaja et al. 2004) and cypress knees (Pulliam 1992). The field study by Pangala et al. (2013) noted CH_4 emissions from tree stems accounted for at least 62 % of the total CH_4 emissions from the tropical forested peatland in Southeast Asia (Pangala et al. 2013). To date, however, details related to the nature of CH_4 emissions from tree stems in natural wetlands remain unclear. To our knowledge, a very few field studies have addressed the variability of CH_4 emission rates, factors controlling those rates and the underlying mechanisms of CH_4 emissions from tree stems (Pangala et al. 2013), although some experimental studies have attempted to tackle these issues (Pangala et al. 2014).

In this paper, we present the results of a field study on the spatial and temporal variability of CH_4 emissions from stem surfaces of mature *Fraxinus* trees in a temperate floodplain forest. The specific aims of this paper are to show: (1) the variations of stem CH_4 emission rates among individual trees in a site with a spatial gradient of belowground water regime; (2) diurnal changes in stem CH_4 emission rates in both foliate and defoliate periods of deciduous *Fraxinus* trees; (3) seasonal changes in stem CH_4 emission rates and possible related changes in the belowground environment, such as soil temperature, water table depth and porewater CH_4 concentration.

Materials and methods

Study site

This study was conducted in a mature stand of *F. mandshurica* Rupr., a deciduous tall tree species, in central Hokkaido, northern Japan. The research site was located on a floodplain along a small mountain stream (43°22'N, 141°36'E; 60 m a.s.l.). The canopy was dominated by *F. mandshurica* trees planted in 1931, and the subcanopy was composed of *Salix udensis* Trautv. et C. A. Mey. Mean height, mean diameter at breast height (DBH), tree density, and

relative dominance in the basal area of *F. mandshurica* were 28.1 ± 1.7 m (mean \pm SD), 30.7 ± 4.0 cm, 183 trees ha^{-1} , and 74 %, respectively. From June through August, the forest floor was mostly covered by large perennial herbs such as *Filipendula camtschatica* (Pall.) Maxim., *Fallopia sachalinensis* (F. Schmidt) Ronse Decr., and *Phragmites australis* (Cav.) Trin. ex Steud. The maximum height of forest floor vegetation reached ca. 4 m above ground in August.

The annual mean air temperature at the study site is 6.0 °C, with the highest monthly mean temperature in August (19.5 °C) and the lowest in January (-6.8 °C) (Japan Meteorological Agency 2002). Total annual precipitation is 1,460 mm, of which approximately 470 mm falls in the form of snow from December to March (Japan Meteorological Agency 2002).

The soil at the study site was composed of alluvial deposits with depths exceeding 1.5 m and classified as a Gleysol (IUSS Working Group WRB 2006). The mostly loam and clay-loam soil contained no gravel and had a pH of 5.4 at the surface.

Variation in stem CH_4 flux among individual trees

A 20×60 m experimental plot was set up in the stand (Fig. 1). This plot was established by extending the former experimental quadrat from our previous work (Terazawa et al. 2007). Ten dominant *F. mandshurica* trees within the plot were selected for the

measurements of CH_4 flux from the stem surfaces (referred as “stem CH_4 flux”, hereafter). Sampled trees ranged from 26 to 31 m (mean 28.0 ± 1.8 m) in height and 26–39 cm (mean 30.7 ± 4.4 cm) in DBH. Each sampled tree had a single trunk without branches and apparent scars from injury or stem disease up to at least 3 m above ground level.

Stem CH_4 flux was measured using a static closed-chamber method in the daytime (between 10:00 and 16:00) on July 20–22, 2011. A custom-made stainless steel chamber (70 mm wide \times 180 mm long \times 80 mm high; Alfa Kikai Co. Ltd., Ibaraki, Japan) with an acrylic lid was used for the flux measurements. A chamber was attached to a stem surface at ca. 15 cm above ground using a urethane frame placed between the chamber and bark. Gaps between the chamber, frame, and tree bark were sealed with pulp clay (Kutsuwa Co., Ltd., Osaka, Japan). The chamber body had a fringe (10 mm in width) on the edge of its open end, and an acrylic lid was fixed firmly on the fringe with silicone rubber packing and spring clips when gas flux measurement was started. Air inside the chamber was sampled at 0, 10, and 20 min after chamber closure through a silicone rubber septum attached on the lid using a 50 ml gas-tight plastic syringe. Samples of gases (40 ml each) were injected into pre-evacuated 30 ml glass vials with butyl rubber stoppers. A Tedlar® bag containing 120 ml air was attached to each chamber to allow the inside pressure to equilibrate with atmospheric pressure during gas sampling.

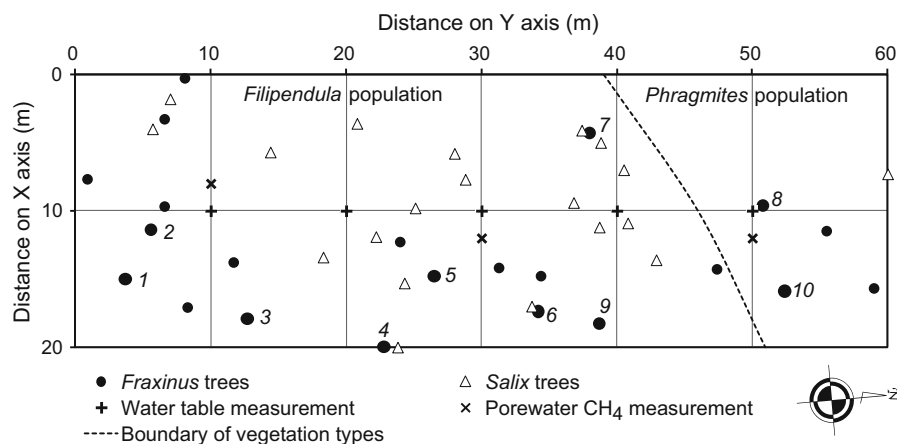


Fig. 1 Locations of trees and measuring points for below-ground environment in the experimental plot for the study of stem CH_4 flux in a floodplain forest of northern Japan. The plot is 20×60 m. A dashed line indicates the boundary between

two vegetation types of the forest floor, i.e., *Filipendula* and *Phragmites* populations. Larger closed circles with Arabic numbers indicate the trees for the CH_4 flux measurement at the stem surface

Chambers were shaded by aluminum foil during measurement to prevent overheating from direct sunlight.

Diurnal change in stem CH₄ flux

Diurnal changes in stem CH₄ flux from the stem surfaces were measured for the three representative individuals of the ten dominant *F. mandshurica* trees that were also used for the measurement of among-tree variability in stem CH₄ flux described above. Among these three individuals, two trees (Trees 2 and 5) were located within the site where a *Filipendula* population dominated the forest floor, while another individual (Tree 8) was located within a *Phragmites* population (Fig. 1). The three individuals (Trees 2, 5, and 8) were located within several meters (≤ 6 m) from each corresponding point that was used for measurement of the belowground environment, i.e., 10, 30 and 50 m points (Fig. 1). Heights of the sampled trees (Trees 2, 5, and 8) were 28, 26 and 30 m, and DBHs were 29, 26 and 34 cm, respectively.

Stem CH₄ fluxes of the three sampled trees were concurrently measured seven times at 4-h intervals during each 24 h on August 9–10 and November 1–2, 2011. The August and November measurements represented the foliate and defoliate periods of *Fraxinus* trees, respectively. Procedures for CH₄ flux measurement were the same as described above.

Seasonal change in stem CH₄ flux

Seasonal changes in the stem CH₄ emission rate were measured for the same three representative trees of *F. mandshurica* (Trees 2, 5, and 8) on which the diurnal changes in stem CH₄ flux were measured. The flux measurements were conducted from May 25 to October 30 in 2012, and again from August 2 to November 1 in 2013 with an almost 1-month interval between measurements. Measurements of stem CH₄ flux in May, October and November were conducted in the leafless season of canopy trees, and the sampled trees had no intact leaves on their crowns on these measurement days. Procedures for CH₄ flux measurement were the same as described above.

Stem CH₄ fluxes were measured regularly at the fixed position on a stem in each sampled tree throughout the study. To test the effect of position of

CH₄ flux measurement in a circumferential direction on a stem surface, CH₄ fluxes were measured at two different positions on the same height (ca. 15 cm above ground) for each sampled tree on August 30, 2012. The two positions (Position A and B) were separated at least a quarter of a circle on a stem; the Position A was the regular position for stem CH₄ measurements in this study.

Environmental variables

Water table depths and soil temperatures at 112.5 cm below the soil surface were monitored at five points with 10 m-intervals along the center line of the experimental plot (Fig. 1) using water height data loggers (TruTrack WT-HR 1500; Intech Instruments Ltd., Christchurch, New Zealand), which were set in perforated PVC pipes, in 2011 and 2012. In 2013, the number of measuring points for water table depth and soil temperature was reduced to three, with measurements taken at 10, 30 and 50 m points within the plot (Fig. 1). Soil temperatures were also monitored at 25 and 50 cm below the soil surface by thermistor sensors and data loggers (UIZ3633, Uizin, Tokyo, Japan) at the center of the experimental plot. Water table depth and soil temperature at 112.5 cm were recorded at 6 h intervals, and those for soil temperature at 25 and 50 cm were 1 h intervals.

On each day of the stem CH₄ flux measurements, air temperature and relative humidity at 1 m above ground were recorded by a temperature/relative humidity logger (HOBO U23-001; Onset Computer Corporation, Bourne, MA., USA) with 15 min intervals at the center of the experimental plot. For the measurements of diurnal change in stem CH₄ flux, in particular, vapor pressure deficit (VPD) was calculated from the temperature and relative humidity data, and the photosynthetic photon flux density (PPFD) was monitored at the height of 5 m above ground at an open site close to the experimental plot by a quantum sensor (PAR-001; Prede Co., Ltd., Tokyo, Japan) and a portable recorder (Model 3057; Yokogawa Electric Corporation, Tokyo, Japan).

Daily precipitation and daily mean temperature data from May to November in 2012 and 2013 were obtained from the Tsukigata Regional Meteorological Station, which was located 6 km southeast of the study site.

Porewater CH₄ concentration

On the day or following day of the stem CH₄ flux measurement in 2012 and 2013, porewater CH₄ concentration in the soil was measured by the following procedures based on in situ sampling method for dissolved components in sediment porewater (Hesslein 1976). A ceramic porous cup (55 mm long, 18 mm in external diameter, 1.5 mm thick; Sekiya Rika Co., Ltd., Tokyo, Japan) attached to a short acrylic pipe (60 mm long, 22 mm in external diameter, 3 mm thick) was filled with 17 ml distilled-deionized water. The open end of the pipe was plugged by a silicone stopper with a careful attention to purge any bubbles inside the pipe as much as possible through an injection needle inserted in the stopper. After the plugging was completed, the injection needle was removed from the stopper. This water-filled porous cup was attached at the end of a 150-cm-long PVC pipe, buried at a given depth in the soil and allowed to equilibrate with porewater for at least 13 days. The porous cup was collected from the soil on the day the porewater CH₄ concentration was measured. Immediately after the porous cup collection, 10 ml water was sampled from the cup by a 50 ml gas-tight plastic syringe without any exposure of the sampled water to ambient air. Then, 40 ml of N₂ gas was added to the syringe, and the syringe was shaken by hand for 3 min to equilibrate the gas concentration between the gaseous and liquid phases in the syringe. A 15 ml sample of gas was taken from the headspace and injected into a pre-evacuated 10 ml glass vial through a butyl rubber stopper.

Porewater CH₄ concentrations were measured at 80 and 120 cm depths in 2012 and at 120 cm depth in 2013 at three points in the experimental plot (Fig. 1). These points for measurement of the porewater CH₄ concentration were set at 2 m apart from each corresponding point for water table measurement (10, 30 and 50 m point) to avoid possible interference from the perforated pipes for water depth measurement.

Soil profile and carbon content

Soil cores were taken by a 1.5 m long soil auger within a distance of 2 m from each measurement point for the belowground environment (10, 30, and 50 m point; Fig. 1). On each soil core, the soil profile was carefully observed with special attention to noting the depths of

the gley horizon with or without oxidized iron mottles. Soil samples taken from each 10 cm depth of the soil cores were used for carbon content determination. Carbon content of the air-dried soil samples was determined by an NC analyzer (Sumigraph NC-220F, Sumika Chemical Analysis Service, Ltd., Tokyo, Japan).

Laboratory gas analysis and calculation of stem CH₄ flux and porewater CH₄ concentration

For the determination of stem CH₄ flux and porewater CH₄ concentration, the CH₄ concentration of each gas sample was determined using a gas chromatograph (GC-14BPTF; Shimadzu Co., Kyoto, Japan) equipped with a flame ionization detector and a Unibeads C column (2 m × 1.5 mm diameter; GL Science, Tokyo, Japan).

Stem CH₄ flux was calculated using a linear regression for the time course of gas concentration in the chamber. Of the regression coefficients for linear regressions, 93 % of the individual measurements were higher than 0.999, and the minimum regression coefficient value was 0.993 except for the lowest value of 0.890 which was deleted from further analysis. The porewater CH₄ concentration was calculated using the Henry's Law from the headspace CH₄ concentration inside the syringe and the ambient temperature at the time when the syringe was shaken.

Statistical analysis

A generalized linear model (GLM) was used to examine the effects of environmental variables on the variation in stem CH₄ flux for the three representative trees (Trees 2, 5, and 8). The explanatory variables included to the models were soil temperature at 50 cm below soil surface, water table depth, porewater CH₄ concentration, tree individuals, and interaction terms between tree individuals and each environmental variable. Soil temperature at 25 and 112.5 cm below soil surface were not included to the model because of the multicollinearity with soil temperature at 50 cm below soil surface. Selection of explanatory variables was conducted based on Akaike's information criterion (AIC). The response variable was assumed to follow a gamma distribution and a reciprocal function was adopted as a link function in the GLM analysis. The statistical software

R version 3.1.1 (R Core Team 2014) was used for the analysis.

Results

Variation in stem CH₄ flux among individual trees

Substantial stem CH₄ emissions were detected for all ten trees measured. Daytime stem CH₄ emission rates at the height of ca. 15 cm above ground varied between 81 and 1,305 $\mu\text{g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$ among the trees (Fig. 2a).

Stem CH₄ emission rates seemed to have a spatial gradient within the experimental plot, increasing with the distance on the Y-axis of the plot (Fig. 2a). The difference in stem CH₄ flux corresponded well to both the difference in belowground water table depth and forest floor vegetation within the experimental plot (Fig. 2a, b). Eight individual trees in the *Filipendula* population, where water table depths were around 100 cm below the soil surface, showed relatively low stem CH₄ emission rates ranging between 81 and 512 $\mu\text{g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$. In contrast, two individual trees growing in a *Phragmites* population, where the water table laid just a few centimeters below soil

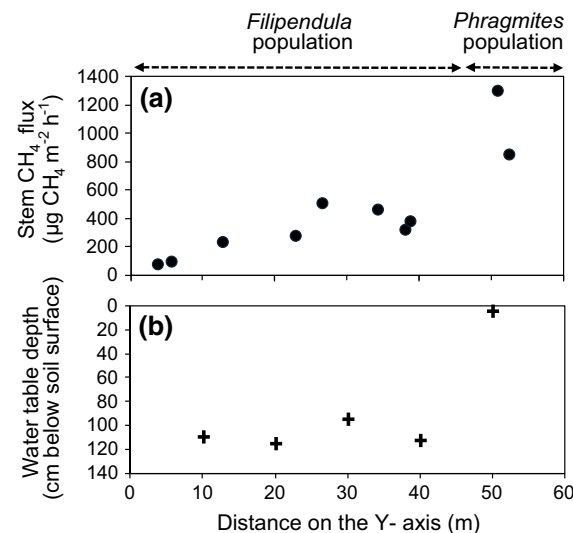


Fig. 2 Stem CH₄ flux of ten canopy trees of *F. mandshurica* measured in late July, 2011 (a), and water table depth measured at 10 m-interval along the center line of the experimental plot on the same day of flux measurement (b), in a floodplain forest of northern Japan

surface, emitted CH₄ at a rate exceeding 800 $\mu\text{g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$. No clear correlation was found between stem CH₄ emission rates and DBH of the trees.

Diurnal change in stem CH₄ flux

In both sets of measurements in August and November, stem CH₄ emissions were observed throughout a 24 h period (Fig. 3a, e). Diurnal variations of stem CH₄ emission rates were relatively small for all of the measurement series. For Trees 2, 5 and 8, average stem CH₄ emission rates were 119, 215 and 189 $\mu\text{g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$ on August 9–10, and 84, 154, 494 $\mu\text{g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$ on November 1–2, respectively. The coefficient of variance of stem CH₄ emission rates of each measurement series fell between 0.02 and 0.08. No distinct pattern of variation was found in the diurnal change of stem CH₄ emission rates, except for Tree 8 in the November measurement which showed apparent slight decrease in stem CH₄ flux at night (from 18:00 to 6:00).

Air temperature varied gently between 20.8 and 27.1 °C during the stem CH₄ measurements in August, corresponding to the diurnal change in PPFD (Fig. 3b, c). In November, air temperature changed more drastically during the stem CH₄ measurements, ranging between -0.9 °C and 15.3 °C with a consistent decline throughout the night (Fig. 3g). Changes in VPD were similar to those of PPFD in both August and November (Fig. 3c, g). Soil temperatures were stable during the stem CH₄ measurements even at the upper most measuring depth, i.e., 25 cm (Fig. 3d, h).

Water table depths on the day of stem CH₄ flux measurements on August 9–10 were 115, 89 and 11 cm below soil surface, at the 10, 30 and 50 m points, respectively. Water table depths on November 1–2 were 69, 46, and 20 cm, at the 10, 30, and 50 m points, respectively.

Seasonal change in stem CH₄ flux

Large differences were observed in the range of seasonal variation of stem CH₄ emission rates among the three sampled trees. Stem CH₄ emission rates of Tree 2 were rather stable throughout the entire period of measurement, ranging between 59 and 124 $\mu\text{g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$ (Fig. 4a, e). In contrast, Tree 8 showed a drastic change in stem CH₄ emission rates during the measuring period. In 2012, the stem CH₄

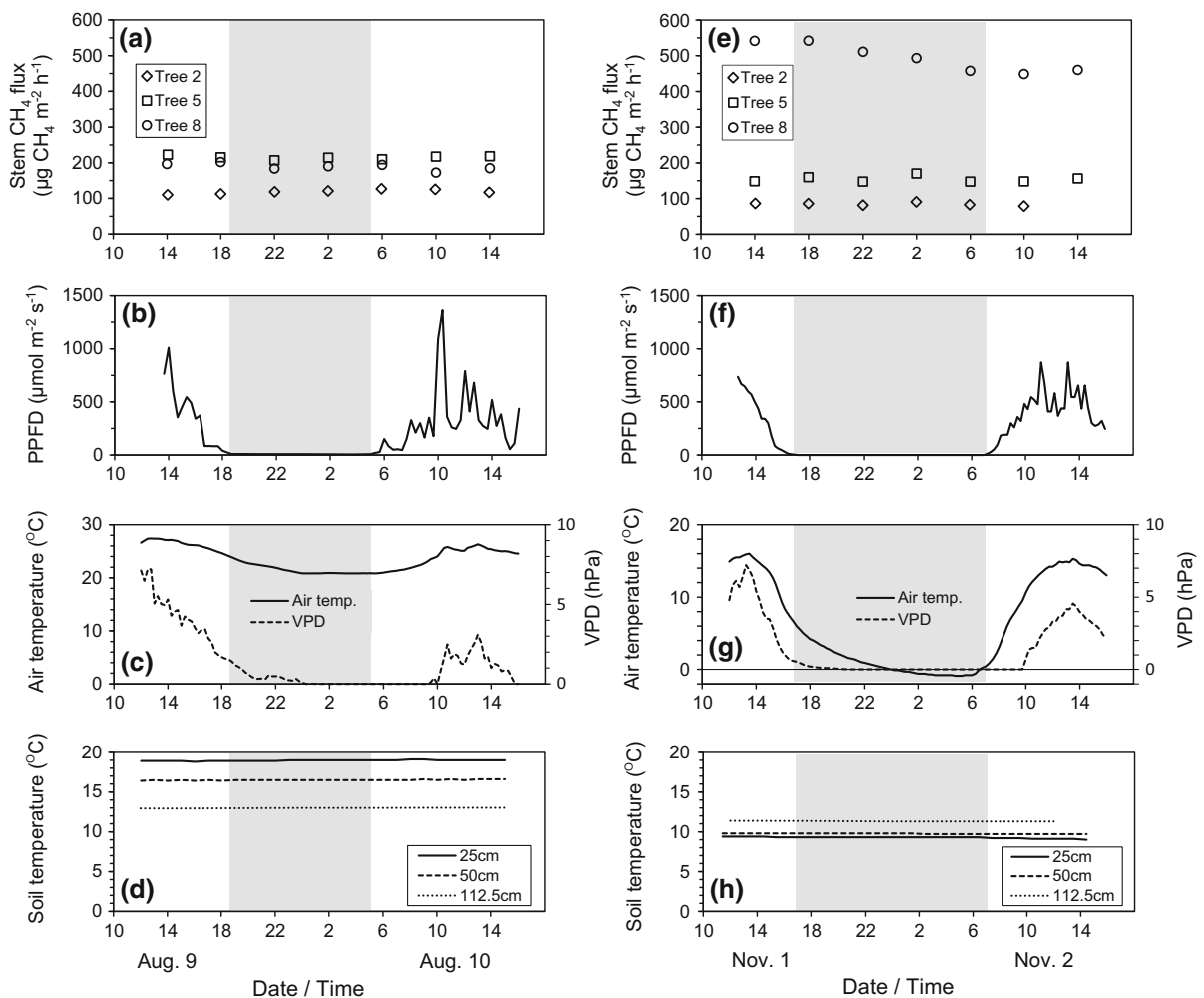


Fig. 3 Diurnal changes in stem CH₄ flux of *F. mandshurica* trees (a, e), PPFD (photosynthetic photon flux density) (b, f), air temperature and VPD (vapor pressure deficit) (c, g), and soil temperature (d, h) in a floodplain forest of northern Japan. Stem CH₄ fluxes were measured on three canopy trees (Trees 2, 5, and

8; Fig. 1) at 4-h intervals for 24 h on August 9–10 (a), and November 1–2 (e), 2011. Measurement intervals of soil temperatures were 1 h for 25 and 50 cm, and 6 h for 112.5 cm depths. Night is shown by the gray belt in each graph

emission rate of Tree 8 peaked in late August at 1,514 $\mu\text{g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$ (Fig. 4a); similarly in 2013, it peaked in late September at 1,492 $\mu\text{g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$ (Fig. 4e). The magnitude of seasonal change in stem CH₄ emission rates of Tree 5 was intermediate between the other two individuals (Trees 2 and 8), ranging between 139 and 331 $\mu\text{g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$ (Fig. 4a, e). A late-summer to early-autumn increase in the stem CH₄ emission rate observed in Tree 5 was less drastic than that observed in Tree 8.

On August 30, 2012, differences in stem CH₄ emission rates measured at two positions at the same

height in Trees 2, 5, and 8 were 106, 32 and 162 $\mu\text{g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$, accounting for 60, 9 and 10 % of the mean stem CH₄ emission rate of each respective tree on that day (Fig. 5).

The ranges of fluctuation in the water table depth differed among the three measuring points (Fig. 4b, f); the fluctuation range during the period between May 25 and November 1 in each year was smallest at the 10 m point (38–70 cm below soil surface in 2012, and 55–76 cm in 2013). It was the largest at the 50 m point (18–72 cm below soil surface in 2012, and 16–85 cm in 2013). Intermediate measurements were observed at

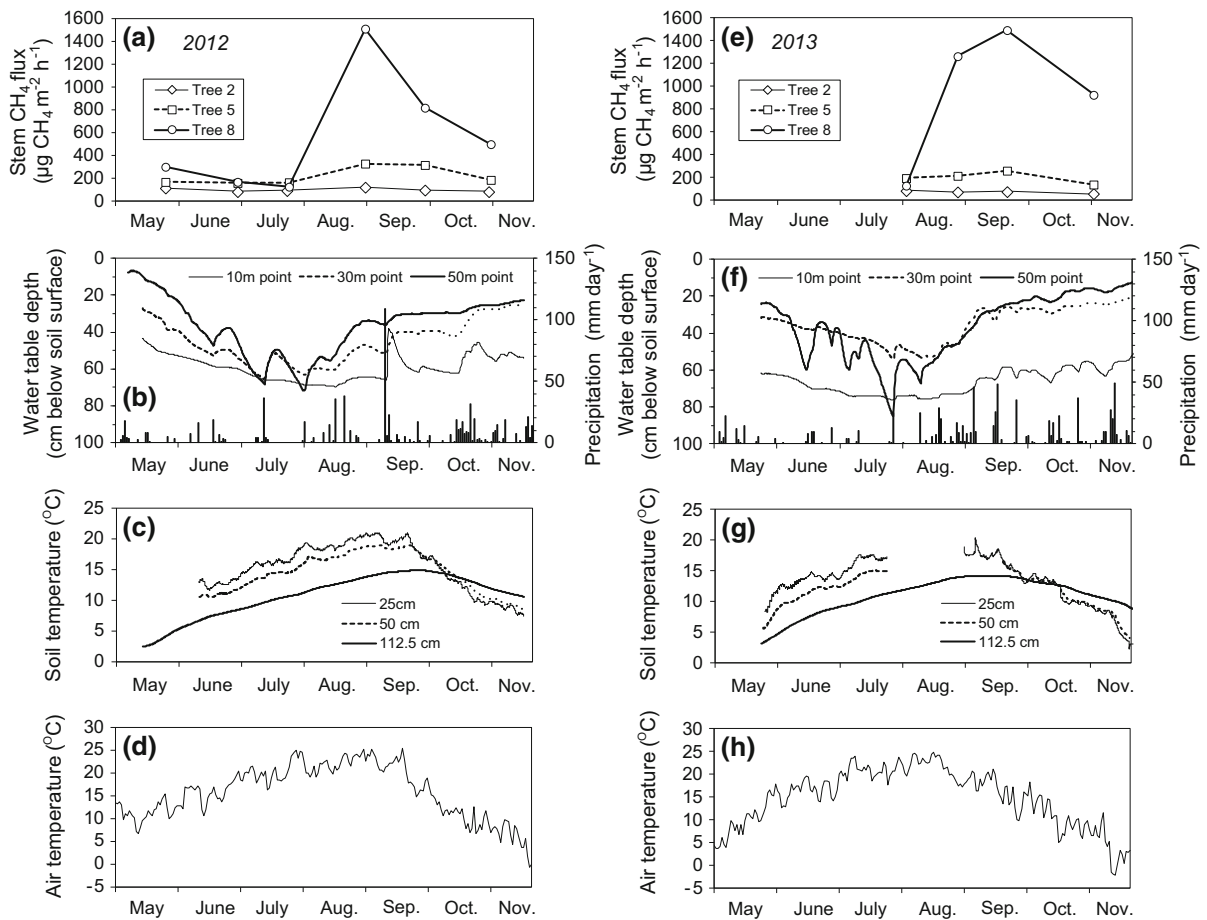


Fig. 4 Seasonal changes in stem CH₄ flux of *F. mandshurica* trees in a floodplain forest of northern Japan in 2012 (a) and 2013 (e). Water table depth and precipitation (b, f), soil temperature (c, g), and air temperature (d, h) during the corresponding period are also shown. Stem CH₄ fluxes were measured at three canopy trees (Trees 2, 5, and 8). Each

measuring point of water table depth (10, 30 and 50 m points) corresponded to each tree for stem CH₄ flux measurement, Trees 2, 5, and 8, respectively. Precipitation and air temperature data were obtained from the Tsukigata Regional Meteorological Station located 6 km southeast of the study site

the 30 m point (28–65 cm below soil surface in 2012, and 24–54 cm in 2013). Mean depths of the water table during the study period at the 10, 30 and 50 m point were 60, 48 and 40 cm below the soil surface in 2012, respectively, and 68, 37 and 39 cm below the soil surface in 2013, respectively.

Soil temperatures at 112.5 cm in depth peaked in late-September in 2012 and in early-September in 2013 (Fig. 4c, g).

Porewater CH₄ concentration

Large differences were observed in porewater CH₄ concentration among the measuring points at both 80 and 120 cm below the soil surface (Fig. 6).

Porewater CH₄ concentrations were consistently highest at the 50 m point, exceeding 600 μmol l⁻¹ throughout the study period at both depths. In contrast, at the 10 m point, porewater CH₄ concentrations were 10⁻⁴ to 10⁻³-fold lower than those of the 50 m point during the initial stage of the experiment period. Porewater CH₄ concentrations at the 30 m point were intermediate between 50 and 10 m points at both depths.

Porewater CH₄ concentrations increased consistently from late May to late September in 2012 at both depths of the 10 and 30 m points (Fig. 6a, b). A consistent increase in porewater CH₄ concentration was also observed at the 120 cm deep of the 10 m point in 2013 (Fig. 6c).

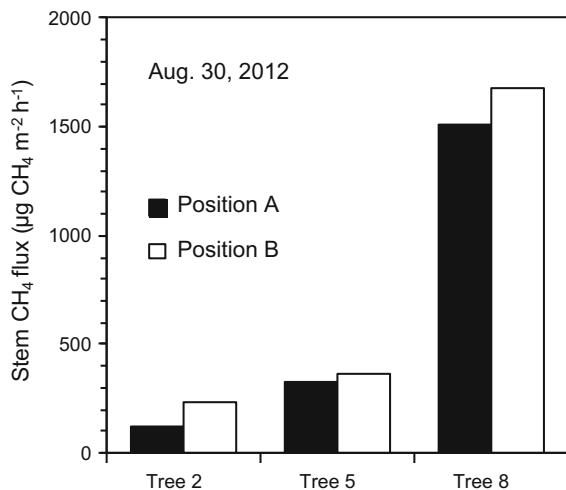


Fig. 5 Stem CH₄ flux of *F. mandshurica* trees measured at two different circumferential positions on a stem on August 30, 2012. Position A and B were separated at least a quarter of a circle of a stem at the same height (ca. 15 cm above ground)

Effects of environmental variables on stem CH₄ flux

As a result of the GLM analysis, soil temperature at 50 cm deep, water table depth, porewater CH₄ concentration at 120 cm below the soil surface, tree individuals, and an interaction between porewater CH₄ and tree individuals were selected as important factors in the best model for stem CH₄ flux (Table 1). Since a reciprocal function was adopted as a link function in the GLM analysis, a negative value of the estimated parameter represents a positive effect of the variable on the stem CH₄ flux, and vice versa. Soil temperature and water table depth tended to have a positive and a negative effect on stem CH₄ flux, respectively (Table 1, Fig. 7). Porewater CH₄ concentration tended to have positive effects on stem CH₄ flux (Table 1, Fig. 7). However, the effect of porewater CH₄ concentration on stem CH₄ flux differed between tree individuals, because an interaction term (porewater CH₄ × tree) was in the model (Table 1). Stem CH₄ flux also differed between tree individuals (Table 1).

Soil profile and carbon content

The gley horizon and the layer with mottles of oxidized iron were observed in each soil core taken

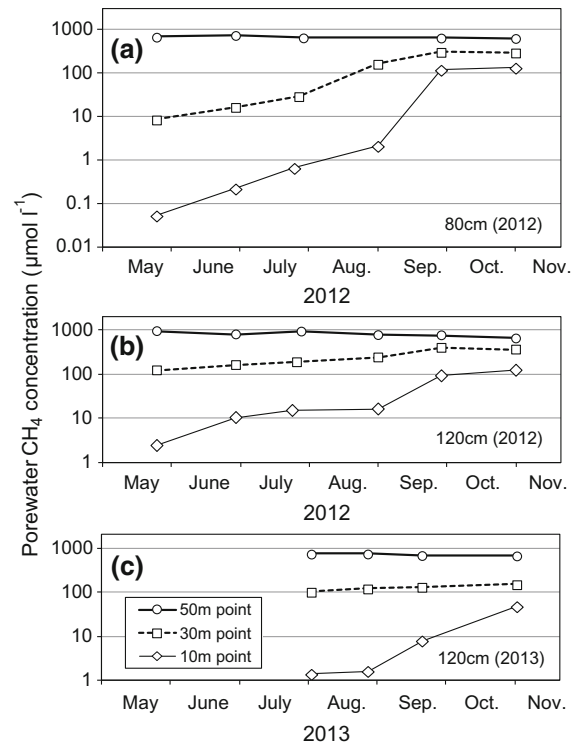


Fig. 6 Seasonal changes in porewater CH₄ concentrations at 80 cm (a) and 120 cm (b) below the soil surface in 2012, and 120 cm below the soil surface in 2013 (c) in a floodplain forest of northern Japan. Each measuring point for porewater CH₄ concentration (10, 30, and 50 m points) corresponded to each tree used for stem CH₄ flux measurement, Trees 2, 5, and 8, respectively

from the 10, 30 and 50 m points. Depths of these reduced layers in the soil cores differed among sampling points. A gley horizon (5Y 4/1–7.5GY 4/1 in Munsell color notation) without mottles was observed from a depth deeper than 100 cm at the 10 m point, while they were observed from shallower depths at the 30 and 50 m points, appearing at 75 and 70 cm below the soil surface, respectively. The pseudogley horizon (2.5Y 4/2–2.5Y 4/3) containing mottles of oxidized iron was observed below 60 cm deep at the 10 m point, while it appeared at the much shallower depth of 20 cm below the soil surface at both the 30 and 50 m points.

Total carbon content of the surface soil was 3.8–4.3 %, and decreased exponentially with soil depth. Carbon content was higher at the 50 m point when compared with the 10 and 30 m points to a depth of 60 cm.

Table 1 Estimated parameters of the selected model for stem CH₄ flux analysed by the GLM with gamma distribution and reciprocal link function

Variable	Estimate ^a	SE
Intercept	0.0131	0.001805
Soil temperature (50 cm)	-0.00022	0.000057
Water table depth	0.000020	0.000013
Porewater CH ₄ (120 cm)	-0.0000018	0.000013
Tree		
Tree 5	-0.0055	0.001938
Tree 8	-0.0190	0.002876
Porewater CH ₄ × Tree		
Porewater CH ₄ × Tree 5	-0.0000002	0.000014
Porewater CH ₄ × Tree 8	0.0000156	0.000014

AIC: 484.51

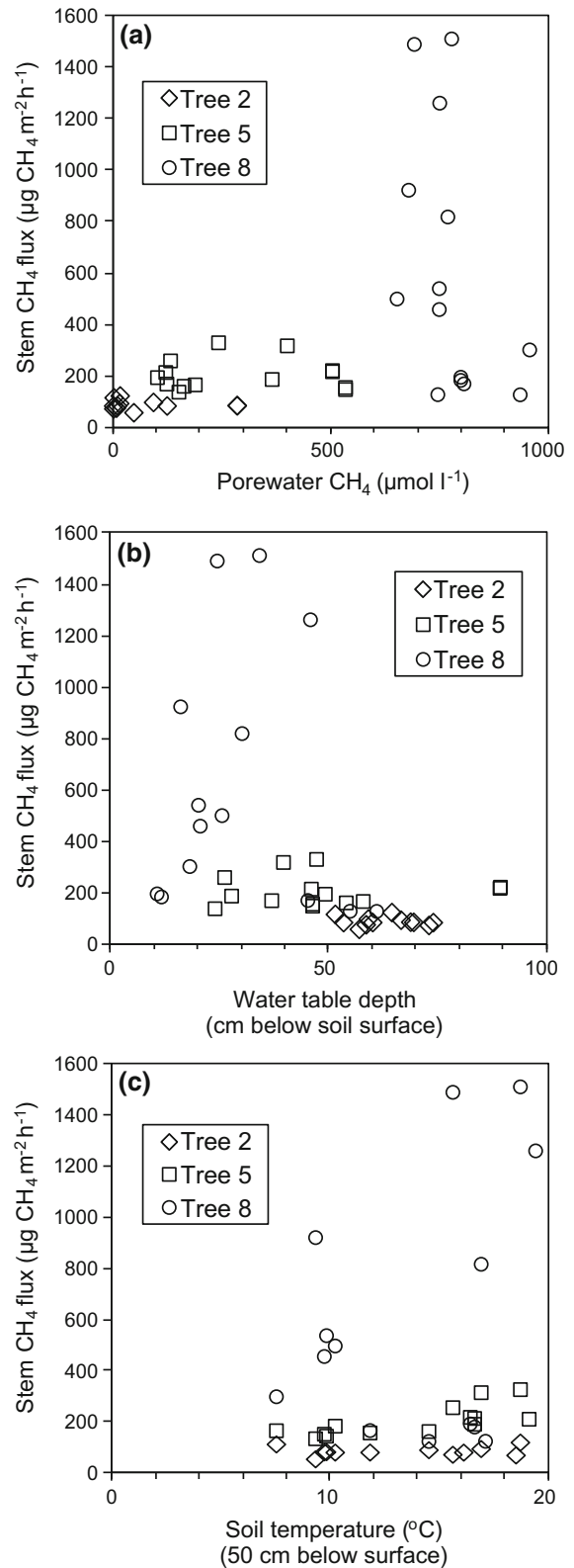
Deviance explained: 0.817

^a As a reciprocal function was adopted as a link function, a negative value of the coefficient represents a positive effect of the variable on the stem CH₄ flux, and vice versa

Discussion

In the present study, large variations were observed in stem CH₄ emission rates among individual trees within a site; the CH₄ emission rates differed greater than one order of magnitude between individual trees in July, 2011 (Fig. 2a) and in late-summer to early-autumn in 2012 and 2013 (Fig. 4a, e). This result simply suggests that we should pay attention to the possibility of inter-individual or spatial variability in stem CH₄ emissions, when we attempt to precisely evaluate the impact of stem CH₄ emissions on the CH₄ budget at an ecosystem or global scale by scaling up the CH₄ flux measured on individual trees in a small area. When compared with these large variations among individual trees, differences in CH₄ emission rates measured at two different horizontal positions on an individual tree stem were relatively small (Fig. 5); this allowed us to discuss the inter-individual and spatial variability in stem CH₄ flux in a site in more detail.

The variation in stem CH₄ emission rates among ten sampled trees in the present study showed a spatial gradient, which apparently corresponded to the difference in water table depth (Fig. 2). This result implies that one of the possible rate-controlling environmental factors of stem CH₄ emissions may



◀ **Fig. 7** Relationships between stem CH₄ flux of *F. mandshurica* trees and environmental variables in a floodplain forest of northern Japan: porewater CH₄ concentration at 120 cm below the soil surface (a), water table depth (b), and soil temperature at 50 cm below the soil surface (c). Data collected during the daytime measurements of diurnal changes (August and November, 2011) and seasonal changes (May, 2012–November, 2013) in stem CH₄ flux were plotted

be the depth of the water table, which alters the magnitude of anaerobic/aerobic conditions in the soil, resulting in an alteration in CH₄ production/oxidation rates and CH₄ concentration in the root zone (e.g., Christensen 2010). Water table depth was also selected as an important explanatory variable in the GLM analysis which was performed to examine the effects of environmental variables on the stem CH₄ flux for the three representative trees (Trees 2, 5, and 8; Table 1; Fig. 7). Pangala et al. (2014) recently reported an experimental study showing water table depth significantly influenced CH₄ emissions from tree stems of pot-grown saplings of *Alnus glutinosa*.

In the present study, stem CH₄ fluxes were quite stable throughout a 24-h period, and no distinct patterns of change in stem CH₄ flux were found in both August and November measurements on foliated and defoliated trees, respectively, except for one case (Tree 8 in November, 2011), (Fig. 3a, e). Many of the previous studies on CH₄ emissions from natural wetlands with herbaceous plants or rice paddies reported considerable diurnal changes in CH₄ emission rates (e.g., Holzapfel-Pschorn and Seiler 1986; Shannon et al. 1996; van der Nat et al. 1998; Whiting and Chanton 1996). These changes were mainly associated with soil or peat temperature, which is one of the factors responsible for the belowground CH₄ production rate (Holzapfel-Pschorn and Seiler 1986; Shannon et al. 1996). Changes in CH₄ emission rates have also been associated with light regime for some plant species in which convective CH₄ transport occurs during the day (van der Nat et al. 1998; Whiting and Chanton 1996). Why, then, were stem CH₄ emission rates so stable during a day in the present study? One possible reason may be the diel stability of soil temperature in the deeper layer where CH₄ was produced under water-saturated conditions. Water table depths on the days of the August and November measurements in the present study were 11 and 20 cm at highest, and 115 and 69 cm at lowest, respectively,

and the soil temperatures at 25 cm and deeper were quite stable during each day of stem CH₄ flux measurement (Fig. 3d, h). The results of the GLM analysis for the stem CH₄ flux also suggests soil temperature is an important factor responsible for controlling stem CH₄ emission rates (Table 1; Fig. 7). No marked diel variation of stem CH₄ emissions was similarly reported from the experimental study using *Alnus glutinosa* saplings under flooded soil conditions (Pangala et al. 2014).

In the present study, almost identical rates of CH₄ emissions were observed at both night and day (Fig. 3a, e). In addition, comparable or even higher CH₄ emission rates were measured in November when all sampled trees had no intact leaves on their crowns as compared with August measurements for foliated trees (Fig. 3a, e). These results suggest that the transpirational stream in tree stems may have no or only a very small contribution to the CH₄ emissions from tree stems. CH₄ probably could not be transported in a dissolved form in the upward transpirational stream. Instead, CH₄ was most likely transported in a gaseous form through internal air spaces in the tree body, such as aerenchyma tissues or intercellular spaces associated with a flooded soil condition (De Simone et al. 2002; Yamamoto et al. 1995a, b). As for anatomical evidence for a possible gas conduit in a mature tree body of *F. mandshurica* at the present experimental site, development of the aerenchyma tissues was observed in the cortex of fine roots, which were sampled from 10–60 cm deep in the soil at the north edge of the site (Yamamoto et al. unpublished data). Similarly, in their experimental study of CH₄ and N₂O emissions from seedling stems of *Fagus sylvatica* and *Alnus glutinosa*, Machacova et al. (2013) assumed that CH₄ was transported through the aerenchyma system rather than via the xylem sap because of the low solubility of CH₄ in water. Pangala et al. (2013) also reported a significant positive relationship between stem CH₄ flux and wood specific density, which is an indicator of wood properties including porosity and anatomical composition, for seven tree species coexisting in a tropical forested peatland in Southeast Asia. Their studies suggested the possibility of the existence of a gaseous transport of CH₄ through tree bodies in these species.

Large differences were found in the magnitude and pattern of seasonal changes in stem CH₄ flux among three sampled trees (Fig. 4a, e). Tree 2, which showed

the lowest stem CH₄ emission rate with no clear seasonal changes, was located near the 10 m point where the water table depth remained below 50 cm deep during most of the study period (Fig. 4b, f). In contrast, Tree 8, which showed drastic seasonal changes in the stem CH₄ emission rate, was located at the 50 m point where water table depth and its range of variation were much higher than those of the 10 m point (Fig. 4b, f). The contrasting water regimes between the 10 and 50 m points were clearly reflected to their soil profiles, i.e., depth of the gley horizon and thickness of the layer with mottles of oxidized iron. The water table depth and its seasonal fluctuation have been frequently reported as predominant factors relating to the plant-mediated CH₄ emission rates at various wetland ecosystems (Shannon and White 1994; Treat et al. 2007; Turetsky et al. 2008; Waddington et al. 1996). Therefore, the difference in magnitude and patterns of seasonal changes in stem CH₄ emissions among the sampled trees in the present study may be, in part, the result of the different water regimes within the experimental plot. The GLM analysis for the stem CH₄ flux also supported this hypothesis (Table 1; Fig. 7).

Comparable large seasonal changes in stem CH₄ emission rates to those in the present study were reported in *Alnus glutinosa* trees in a wetland in the United Kingdom (Gauci et al. 2010). In that study, one of the sampled trees showed an increase of the stem CH₄ emission rate of over 40 times during almost 1 month from the early May to early June (Gauci et al. 2010). Relatively small seasonal changes in the average stem CH₄ flux measured in 2005 at the same stand of the present study (Terazawa et al. 2007) may be attributed to the location of the measurement. The measurements in 2005 were conducted on five *Fraxinus* trees including Trees 2 and 5 in the southern half of the present experimental plot where water table was relatively low and fluctuated less.

Large differences in the magnitude and pattern of seasonal changes in porewater CH₄ concentrations were also found (Fig. 6). The porewater in the 50 m point, in the immediate vicinity of Tree 8, was apparently supersaturated with CH₄ throughout the study period, given the solubility of CH₄ in distilled water (ca. 1,850 μmol l⁻¹ at 12 °C; Yamamoto et al. 1976). The consistent increases of porewater CH₄ concentrations at the 10 and 30 m points during the study period may have been caused by the

accumulation of CH₄ produced in the saturated soil layer over time (Schütz et al. 1989). Seasonal increases in the porewater CH₄ concentration were reported from other temperate wetlands (Shannon et al. 1996; Sun et al. 2012), although the magnitudes of increase were much smaller than observed in the present study.

Results of the GLM analysis related to the stem CH₄ flux showed that porewater CH₄ concentration had a positive effect on the stem CH₄ emission rates with an interaction with tree individuals (Table 1; Fig. 7). The results are in agreement with other recent studies related to CH₄ emissions from tree stems in which positive relationships were detected between the stem CH₄ flux and the porewater CH₄ concentration in mesocosms (Machacova et al. 2013; Pangala et al. 2014) as well as in a natural tropical wetland (Pangala et al. 2013). The interaction between porewater CH₄ concentration and tree individuals in the present study implies that structural or/and physiological properties of a tree might have an effect on stem CH₄ flux along with porewater CH₄ concentration. However, no evidence was obtained in the present study on the parameters that were responsible for the difference in tree properties related to stem CH₄ flux.

Results of the present field study suggest that the CH₄ emitted from stem surfaces of wetland trees may be produced in the submerged soil layer, and it may be transported in a gaseous form through internal air spaces in tree bodies, as has also been observed for various wetland herbaceous plants and rice (e.g., Cicerone and Shetter 1981; Nouchi et al. 1990; Shannon et al. 1996; Whiting and Chanton 1992). Forested wetlands represent up to 60 % of the total wetland area worldwide (Matthews and Fung 1987). According to recent studies of upscaled CH₄ emissions from wetland trees, the estimated annual amount of tree-mediated CH₄ flux on a global scale ranged from 2 Tg yr⁻¹ (Carmichael et al. 2014) to 60 ± 20 Tg yr⁻¹ (Rice et al. 2010), which could account for 0.2 and 10 % of the global CH₄ source, respectively. To gain a better understanding and more accurate estimation of CH₄ emission rates from woody plants in forested wetlands, further field and experimental studies will be needed. Such studies should particularly focus on (i) biogeochemical processes and factors involved in controlling stem CH₄ emissions, (ii) vertical variations in CH₄ flux on a tree stem up to the canopy level, which mostly showed a decreasing

tendency as far as it was measured below the breast height level (Pangala et al. 2013; Terazawa et al. 2007), for more accurate evaluation of the contribution of CH₄ emissions from entire trees to the total CH₄ flux of a forested wetland, and (iii) the possibility and magnitude of CH₄ production in living trees and its release to the atmosphere (Covey et al. 2012; Zeikus and Ward 1974).

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