PERSPECTIVES AND PARADIGMS



Evidence does not support the targeting of cryptic invaders at the subspecies level using classical biological control: the example of *Phragmites*

Erik Kiviat · Laura A. Meyerson · Thomas J. Mozdzer · Warwick J. Allen · Andrew H. Baldwin · Ganesh P. Bhattarai · Hans Brix · Joshua S. Caplan · Karin M. Kettenring · Carla Lambertini · Judith Weis · Dennis F. Whigham · James T. Cronin

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Abstract Classical biocontrol constitutes the importation of natural enemies from a native range to control a non-native pest. This is challenging when the target organism is phylogenetically close to a sympatric nontarget form. Recent papers have proposed and recommended that two European moths (*Archanara* spp.) be introduced to North America to control non-native *Phragmites australis australis*, claiming they would not adversely affect native *P. australis americanus*. We assert that these papers overlooked research

E. Kiviat (⊠) Hudsonia, P.O. Box 5000, Annandale, NY 12504, USA e-mail: kiviat@bard.edu

L. A. Meyerson Natural Resources Science, University of Rhode Island, Kingston, RI 02881, USA

T. J. Mozdzer Department of Biology, Bryn Mawr College, Bryn Mawr, PA 19010, USA

W. J. Allen The Bio-Protection Research Centre, Lincoln University, Lincoln 7647, New Zealand

A. H. Baldwin

Department of Environmental Science and Technology, University of Maryland, College Park, MD 20742, USA

G. P. Bhattarai

Department of Entomology, Kansas State University, Manhattan, KS 66506, USA

contradicting their conclusions and that the authors recommended release of the non-native moths despite results of their own studies indicating that attack on native *Phragmites* is possible after field release. Furthermore, their open-field, host-specificity tests were conducted in non-wetland fields in Switzerland using potted plants, reflecting considerably different conditions than those of North American wetlands. Also, native *Phragmites* in eastern North America has declined, increasing its potential vulnerability to any

H. Brix

Department of Bioscience, Aarhus University, Ole Worms Allé, 8000 Arhus C, Denmark

J. S. Caplan Department of Landscape Architecture and Horticulture, Temple University, Ambler, PA 19002, USA

K. M. Kettenring Department of Watershed Sciences and Ecology Center, Utah State University, Logan, UT 84322, USA

C. Lambertini Department of Agricultural and Food Sciences, University of Bologna, Bologna, Italy

J. Weis Department of Biological Sciences, Rutgers University, Newark, NJ 07102, USA

D. F. Whigham Smithsonian Environmental Research Center, Edgewater, MD 21037, USA new stressors. Because all inadvertently introduced, established, Phragmites-specialist, herbivorous insects have done more harm to native than nonnative Phragmites, native Phragmites may experience more intense herbivory than non-native Phragmites from the introduction of Archanara spp. due to demographic mechanisms (e.g., increase in density of the biocontrol agent and spillover onto alternate hosts) or because the herbivores may undergo genetic change. In addition to the risk to native Phragmites, significant biomass reduction of non-native Phragmites may decrease important ecosystem services, including soil accretion in wetlands affected by sea level rise. We strongly caution against the approval of Archanara spp. as biocontrol agents for non-native Phragmites in North America.

Keywords Ecosystem services · Herbivory · Host switching · Invasive species · Non-target impacts of biocontrol · *Phragmites australis*

Introduction

There has been controversy for many years about the risks of classical biological control to non-target species (Simberloff and Stiling 1996), although, during this period, biocontrol has progressed to a modern and circumspective science heralding numerous successes (Heimpel and Cock 2018). In the age of globalization, biological control practitioners are not only challenged by the dramatic increase in occurrence of non-native species (Meyerson and Mooney 2007; Stohlgren et al. 2011) but also by discoveries of "cryptic invaders" (non-native species, hybrids, or genotypes that cannot be easily distinguished from native species or genotypes; Gaskin and Schaal 2002; Saltonstall 2002; Ciotir et al. 2013). As we show in this paper, finding an effective biological control agent with the specificity to negatively affect only the targeted cryptic invader is a daunting task, especially given their genetic, morphological, or physiological similarity to native species or genotypes.

Department of Biological Sciences, Louisiana State University, Baton Rouge, LA 70803, USA The classical biological control program aimed at non-native *Phragmites* has been in development for 20 years (Blossey and McCauley 2000) and has been controversial for almost as long (Rooth and Windham 2000; Meyerson et al. 2009; Packer et al. 2017). The Old World or "European" form of *Phragmites australis* [called *P. australis* subsp. *australis* or "non-native *Phragmites*" to distinguish it from the native American *P. australis* subsp. *americanus* or "native *Phragmites*" (Saltonstall et al. 2004)] is an invasive plant in the United States and Canada and has been the target of widespread management efforts for more than 40 years (Hazelton et al. 2014).

Recently, Casagrande et al. (2018) advocated for biological control programs that target cryptic invaders at the subspecific level. To support their argument, they presented the case of Phragmites australis (Cav.) Trin. ex Steud. (hereafter Phragmites) in North America, in which a highly invasive, non-native lineage broadly overlaps and interbreeds with other native and non-native lineages (Lambertini et al. 2012; Meyerson et al. 2012; Saltonstall et al. 2014, 2016; Wu et al. 2015; Williams et al. 2019). Although there has never been a documented case of successful biological control of a weed at the subspecific level, Casagrande et al. (2018) suggested that it is possible because some biocontrol herbivores can exhibit such specificity. In particular, Blossey et al. (2018a), conducted greenhouse, laboratory, and open-field, host-specific trials on acceptance, larval growth, and oviposition by two European moth species on Phragmites. Many of their studies showed no differences between non-native and native Phragmites, but, in some, they observed reduced survival (by 40-50% in no-choice studies) and oviposition rates (on 6.5% of eggs laid in an openfield test with one-third of the plants being native Phragmites and the other two-thirds American or European non-native Phragmites) on native Phragmites. These findings were the basis for their conclusion that "P. australis americanus genotypes are within the physiological or fundamental host range of A. geminipuncta and A. neurica" and their decision to "recommend release of these two biocontrol agents in North America."

In this paper we review the available data and draw the opposite conclusion, namely that biological control of non-native *Phragmites*, and, more generally, any case of biological weed control at the subspecific level, remains a risky endeavor. We discuss the

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literature regarding the biological control of nonnative Phragmites in North America and the potential intended and unintended consequences if such a program were to be instituted. As we detail below, Casagrande et al. (2018) have downplayed the role that evolution can have on the adoption of new genotypes, subspecies, or species into an herbivore's diet. Moreover, they have not addressed the indirect species interactions (e.g., spillover effects and apparent competition) that operate at larger scales than can be tested in simple laboratory settings. Blossey et al. (2018a) have also acknowledged that, despite years of extensive host-specificity testing, they "cannot categorically exclude the possibility of attack on P. australis americanus after field release." Focusing on *Phragmites* specifically, there are additional issues at play, including the occurrence of other Phragmites lineages and hybrids in North America (see Saltonstall 2002; Lambertini et al. 2006, 2012; Lambertini 2016; Meyerson et al. 2012; Saltonstall et al. 2014; Packer et al. 2017 for a full review of Phragmites lineages and genotypes in North America). Conserving infraspecific (below the species level) genetic diversity is important for, among other reasons, its role in facilitating ecological processes such as trophic cascades (Des Roches et al. 2018). Further, non-native Phragmites has important societal benefits in some regions of the United States and native Phragmites has cultural significance-all of these are at risk if biological control agents targeting non-native Phragmites are introduced. Native Phragmites has declined in the eastern U.S. (Saltonstall and Stevenson 2007; Meyerson et al. 2010b), increasing its potential vulnerability to additional stressors. Finally, while it is not our intent to repeat previously published criticisms of *Phragmites* biocontrol (Bhattarai et al. 2016; Cronin et al. 2016) or the responses (Blossey and Casagrande 2016a, b), these publications were not cited by Casagrande et al. (2018) so we will reiterate some of the key elements here.

Evolution and ecology of novel interactions

Certainly, there have been relatively few documented cases of significant non-target attacks from biological control agents (van Klinken and Edwards 2002; Suckling and Sforza 2014; Wright and Bennett 2018). However, the number of documented cases is

likely to be an underestimation for several reasons: the monitoring of non-target species has been minimal historically (Simberloff and Stiling 1996), there may be a post-introduction lag phase before densitydependent host range expansion and coevolution, and we are aware of no prior biocontrol programs at the subspecific level. In other words, there are few data from which to estimate non-target attack rates. Furthermore, the arguments of Casagrande et al. (2018) (e.g., phylogenetic conservatism of host range) are almost all based on interspecific or higher taxonomic-level comparisons. This precludes consideration of the *infraspecific* variation that already complicates management of Phragmites and which only increases the likelihood that any introduced biological control agent would adopt the native Phragmites lineage into its host range. It is also important to note that the proposed biological control agents, Archanara geminipuncta (Haworth) and A. neurica (Hübner) (Lepidoptera: Noctuidae), are not specialists but are oligophagous on multiple Phragmites lineages and other wetland grasses, including some economically important species (Blossey et al. 2013, 2018a; Blossey 2014). Thus, a major concern remains about the Phragmites biological control program, namely that any introduced herbivores may form novel and damaging host associations with native Phragmites or other non-target species.

Casagrande et al. (2018) downplayed the ecological and evolutionary processes that underlie the incorporation of novel host species or genotypes into the host range in the wild. Such novel interactions can form in two main ways: ecological fitting or evolution. Ecological fitting describes the formation of novel species interactions as a result of pre-existing traits (e.g., phenology, palatability, defenses) but with no coevolutionary history. Such novel interactions have been observed across many taxa, trophic levels, and systems, with two of the better-known examples being California butterflies using non-native larval hosts (Graves and Shapiro 2003) and leaf chewers and miners feeding on introduced oaks (Pearse and Hipp 2009). Ecological fitting has also been reported for two major pests of forest trees, the emerald ash borer (Agrilus planipennis Fairmaire) (Cipollini and Peterson 2018) and the mountain pine beetle (Dendroctonus ponderosae Hopkins) (Erbilgin et al. 2014). Several biological control agents themselves are examples, such as Rhinocyllus conicus Frölich on native thistles

(Louda et al. 2003) and Chrysolina quadrigemina (Suffrian) on non-target, native and non-native Hypericum spp. (Andres 1985). Interestingly, novel interactions can often be predicted from phylogeny, because the host range of insect herbivores generally shows a strong phylogenetic signal (Pearse and Hipp 2009; Pearse et al. 2013; Gilbert et al. 2015; Parker et al. 2015). In other words, introduced herbivores are more likely to incorporate plants into their host ranges that are closely related to their original hosts. However, not all traits involved in ecological fitting will be phylogenetically conserved (e.g., ecophysiological traits; Desurmont et al. 2011; Whitfeld et al. 2012; Erbilgin et al. 2014; Cipollini and Peterson 2018). Thus, it is important to assess these traits in combination with phylogeny, which has led to innovative and successful approaches to predicting novel interactions (e.g., Pearse and Altermatt 2013). Moreover, theoretical and empirical research has demonstrated that the likelihood of host range expansion increases at high population densities of the natural enemy in question, and when primary and alternate hosts co-occur (Araujo et al. 2015; Castagneyrol et al. 2016). Finally, one research area yet to be addressed for Phragmites but previously identified as important (Blossey 2014) concerns the potential for Archanara moths to be incorporated into the diet of natural enemies (e.g., predators and parasitoids). Such an interaction could reduce their efficacy, have indirect effects on nontarget species (Pearson and Callaway 2005), or even promote feeding on native Phragmites. This may provide 'enemy free space' as it does for other internal stem feeders such as Lipara Meigen spp. (Diptera: Chloropidae) (Allen et al. 2015).

Evolution can also contribute to the formation of novel interactions and has been previously identified as the greatest risk to non-target species in weed biological control by some in the field (van Klinken and Edwards 2002). Indeed, rapid evolution has frequently occurred in species introduced to a new range (e.g., Maron et al. 2004; Prentis et al. 2008; Buswell et al. 2010; Turner et al. 2014; Stutz et al. 2018). This includes *Phragmites* (Guo et al. 2014, 2018; Cronin et al. 2015) as well as a number of biological control agents (Szűcs et al. 2012). However, the evolution of novel interactions is more challenging to demonstrate, with the best-known example being unrelated to biocontrol, namely that of soapberry bugs (*Jadera haematoloma* HerrichSchäffer) (Hemiptera: Rhopalidae), which evolved different length mouthparts to incorporate multiple introduced plant species into their host range (Carroll and Boyd 1992; Dingle et al. 2009; Cenzer 2016). In the case of the proposed biocontrol agents for Phragmites, A. geminipuncta and A. neurica, the native lineage of *Phragmites* is already known to be within the physiological and fundamental host range of both species (Blossey et al. 2018a). Because Archanara spp. already feed on several other plant species (e.g., in host range testing) as a result of ecological fitting (Blossey et al. 2013, 2018a; Blossey 2014), a major barrier has already been crossed. The only evolutionary step remaining for Archanara to shift to native Phragmites is that of changes in patterns of fundamental host use, which could be driven by high population density, decreasing frequency of the primary host (i.e., non-native Phragmites), and increasing relative frequency of potential alternate hosts (i.e., native Phragmites) (Murdoch 1969). Combining these demographic characteristics with the multiple phylogenetically and functionally similar non-target hosts that co-occur spatially and temporally with non-native Phragmites, and given that native Phragmites is within the fundamental host range of both Archanara species, it is entirely possible that the proposed biological control agents will evolve to be more effective against the non-target, native Phragmites. That said, we recognize that predicting postintroduction evolutionary trajectories remains a major challenge in ecology. Furthermore, potential also exists for non-native Phragmites to evolve resistance to the proposed biocontrol agents, especially given the novel admixture of genetic diversity present in North America (reviewed by Meyerson et al. 2012), high prevalence of sexual reproduction (McCormick et al. 2010), and the demonstrated rapid evolution in plant defenses of the non-native lineage (Cronin et al. 2015; Allen et al. 2017a, b; Bhattarai et al. 2017a). Such a phenomenon has received little consideration to date but has been observed in Canada with purple loosestrife (Lythrum salicaria L.), which has evolved resistance to its beetle biological control agent Neogalerucella calmariensis (L.) (Coleoptera: Chrysomelidae) (Stastny and Sargent 2017), as well as for the Argentine stem weevil (Listronotus bonariensis) (Kuschel) (Coleoptera: Curculionidae) and its (now ineffective) parasitoid Microctonus hyperodae

Loan (Hymenoptera: Braconidae) in New Zealand (Tomasetto et al. 2018).

Herbivory on *Phragmites australis* lineages in North America

The identity of Phragmites herbivores and the ecological and evolutionary consequences of herbivory are well-studied for all major lineages of the species in North America (e.g., Tewksbury et al. 2002; Lambert and Casagrande 2007; Lambert et al. 2007; Park and Blossey 2008; Saltonstall et al. 2014; Allen et al. 2015, 2017a, b; Cronin et al. 2015; Bhattarai et al. 2017a, b). A great majority of these herbivores include exotic specialist and semi-specialist species that were co-introduced to North America from their native ranges in Europe and Asia. All the introduced species that were specialists on the non-native Phragmites have apparently expanded their niches to include the native and Gulf Coast lineages of Phragmites in North America (Tewksbury et al. 2002; Blossey 2003; Lambert and Casagrande 2007; Lambert et al. 2007; Allen et al. 2015; Cronin et al. 2015). These cases strongly contradict the assertion of subspecies-level host-specificity of Phragmites herbivores. Although Blossey and Casagrande (2016a, b) and Casagrande et al. (2018) consider Lipara pullitarsis Doskocil and Chvala (Diptera: Chloropidae) as an example of subspecies-level host specificity, evidence of infestation on native plants has been documented by Allen et al. (2015). Another species accidentally introduced from Europe, Lasioptera hungarica Möhn (Diptera: Cecidomyiidae), purported to be a specialist of nonnative Phragmites, has been shown to feed on the hybrids of native and non-native lineages in the field (Saltonstall et al. 2014). For this and other species that exhibit a strong preference for the non-native lineage, the hybrid may serve as a bridge to native Phragmites (see Floate and Whitham 1993). Furthermore, plants of the native lineage are more palatable to a wide range of herbivores than plants of the non-native lineages (Allen et al. 2015; Cronin et al. 2015, 2016; Bhattarai et al. 2017a). These two key pieces of evidence, higher palatability of native Phragmites and an overwhelming record of host range expansion by the specialist herbivores of non-native Phragmites to include the native *Phragmites*, strongly suggest that the native lineage would be a highly susceptible naïve host to any new herbivores introduced to North America. Consequently, native *Phragmites* could experience the negative effects of spillover, associational susceptibility, and apparent competition if *Archanara* spp. were introduced as biocontrol agents. A recent study by Bhattarai et al. (2017b) supported this prediction, demonstrating strong apparent competition, mediated by three herbivore guilds, that disproportionately affected the native lineage. Interestingly, two of the herbivore guilds (a sucking insect [the aphid *Hyalopterus pruni*, (Geoffroy)] and gall-forming insects [*Lipara* spp.]) that mediated apparent competition involved introduced species (Bhattarai et al. 2017b). Such a possibility remains to be tested with *Archanara* spp.

Casagrande et al. (2018) stated that larvae of Archanara spp. could not survive through winter on native *Phragmites* because they oviposit beneath leaf sheaths, which typically abscise in autumn, whereas leaf sheaths of non-native Phragmites tend to persist through winter and thus offer greater protection. In fact, loose leaf sheaths are a typical but variable trait of native Phragmites, which is why using it as an identifying characteristic has given way to genetic analysis in cases where lineage needs to be determined unequivocally (Saltonstall 2003c; Tulbure et al. 2012; Guo et al. 2014). Some native populations retain leaf sheaths to a greater degree than others and Allen et al. (2017a, b) and Swearingen and Saltonstall (2012) have cautioned that morphological and phenological traits are subtle, sometimes subjective, and variable across the North American range of Phragmites. Moreover, Blossey et al. (2018a) found that leaf abscission in autumn less than halved larval attack rates of stems the next spring and did not eliminate the Archanara population entirely.

Research involving proposed biocontrol agents

Classical biological control programs often undergo a rigorous evaluation of intended and unintended consequences of proposed biocontrol agents in the introduced range. In the case of *Phragmites* and the proposed biocontrol agents *A. geminipuncta* and *A. neurica*, there is a paucity of scientific evidence (1) demonstrating the efficacy of those insects in control-ling non-native *Phragmites* in North America and (2) ensuring that there will be no negative consequences

to the members of natural ecosystems and agricultural areas. To the best of our knowledge, there has been no publication demonstrating the impact of either of those herbivores on growth, performance, and population dynamics of non-native Phragmites in North America. The only study presented in the context of biological control was a field survey and a common garden experiment evaluating Archanara damage on nonnative Phragmites in its native range in Switzerland (Häfliger et al. 2006). However, evolutionary changes of invasive plants in response to the novel environment (both biotic and abiotic) of the introduced range render those results less robust to predict their success in controlling invasive plants. In its non-native range, P. australis australis exhibits stronger defense responses against herbivores through leaf toughness, total phenolics, and nutrient concentrations (% C and N in leaf tissues) than in its native range (Cronin et al. 2015). Rigorous studies evaluating the impact of Archanara spp. on the performance, growth, and expansion of populations of non-native Phragmites are necessary before assessing whether they are worthy of consideration as biological control agents.

Furthermore, assessments of potential biocontrol introductions need to provide evidence that they do not have negative consequences for members of natural ecosystems and agricultural areas, and such evidence has not been presented for Archanara spp. in North America. Contrary to the expectation of specificity at the subspecies level, these species are not specialists, even at the genus level. In addition to both species consuming and ovipositing on native Phragmites, both species fed on several other species of wetland plants and economically important crops (Blossey et al. 2013, 2018a). Of 43 non-Phragmites species examined, A. geminipuncta was shown to feed on 9 species in no-choice tests including Arundinaria tecta (Walter) Muhl., Arundo donax L., Cortaderia selloana (Schult. & Schult.f.) Asch. & Graebn., Schoenoplectus americanus (Pers.) Volkart ex Schinz & R. Keller, Spartina alterniflora Loisel., S. cynosuroides L. (Roth), Zizania aquatica L., and the commercial crop species rice (Oryza sativa L.) and wheat (Triticum aestivum L.) (Blossey et al. 2013, 2018a: Tables 1 and 2, respectively). Similarly, A. neurica fed on 7 species, namely Eragrostis trichodes (Nutt.), Phalaris arundinacea L., Schoenoplectus acutus (Muhl. ex J.M.Bigelow) Á. Löve and D. Löve, S. alterniflora, Z. aquatica, Glyceria striata (Lam.) Hitchc., and sugarcane (Saccharum officinarum L.). Moreover, first instar larvae of both Archanara species survived on several of the test species (S. alterniflora, S. cynosuroides, Cortaderia selloana, Oryza sativa, and Schoenoplectus acutus) for the two week duration of the experiment (Blossey et al. 2018a; Table 3). Although the concerns over these results (e.g., Bhattarai et al. 2016; Cronin et al. 2016) have been dismissed as unimportant damage in no-choice experiments (Blossey and Casagrande 2016b), these results are critical in evaluating potential harmful ecological and economic consequences of the biocontrol agents through indirect effects (Simberloff 2012). Paynter et al. (2015) analyzed data from no-choice feeding and oviposition tests performed for arthropod agents in New Zealand and concluded that these tests were an efficient means of predicting if non-target species would experience serious damage. However, Paynter et al. presented an example of testing that failed to predict extensive damage to a native thistle by a weevil distributed to target Cirsium arvense (L.) Scop., and cautioned that the twelve cases they had analyzed may have had insufficient replication of tests to yield robust conclusions. Given the current lack of evidence, the introduction and release of Archanara spp., which are known to feed on multiple native plant species and economically important crops, should be postponed until further evidence is acquired. Moreover, several of the potential alternate plant hosts identified by Blossey et al. (2013)-S. alterniflora, Z. aquatica, and Schoenoplectus americanus-are foundation species in coastal and inland marsh restoration efforts. Such projects cost many millions in taxpayer dollars annually and restore critical ecosystem services to coastal residents, fisheries, and other wildlife (Barbier et al. 2011). A quantitative assessment of the potential effects that releasing any biological control agent could have should be undertaken to identify intended and unintended consequences.

After acknowledging the lack of host specificity, the solution proposed by Casagrande et al. (2018) and Blossey et al. (2018a) to infestation of native *Phragmites* by *Archanara* spp. was mowing and burning *Archanara*-infested stands. Given that mowing and burning are practices employed by land managers to contain or weaken non-native *Phragmites* (Hazelton et al. 2014 and citations within), exposure of native *Phragmites* to multiple stressors (biocontrol plus mowing or burning) could decimate native lineages. This approach is also impractical where the use of fire could threaten residential areas.

On a practical level, we agree with Blossey et al. (2018a) that open-field host-specificity tests are more realistic than those conducted in the lab, greenhouse, or outdoor cages. Nonetheless, the condition of test plants and the growing environment are critical to the realism of field trials (USDA 2016), yet a common problem in open-field trials is the suboptimal growth of non-target plants that are not typically cultivated (Schaffner et al. 2018). The attack on positive controls (i.e., *Phragmites australis*) found by Blossey et al. (2018a, b) demonstrated that conditions were suitable for the biocontrol agent but did not demonstrate that non-target plants experienced conditions similar to those of natural wetlands. The open-field tests used pot-grown plants, which are highly susceptible to nutrient or water stress (USDA 2016). Moreover, the soil type and fertilization regime were not described for the experiment (Blossey et al. 2018a, b), and it is not mentioned (and therefore unlikely) that native Phragmites pots were supplemented with reduced iron. In pot cultivation, it is important to alleviate iron limitation and its associated effects like chlorosis and growth reduction (Willson et al. 2017). Further, the pots were placed in a mown, non-wetland field, creating a drained hydrologic condition that would have further decreased the availability of reduced iron and could have thereby altered plant physiological processes. It is thus possible that the growth problems resulting in thin stems of P. a. berlandieri reported by Blossey et al. (2018a) were due to suboptimal soil, fertility, or moisture conditions rather than climate. The abundance of host plant species relative to the non-target plants can also influence test outcomes; in designs where host plant abundance is greater than that of non-target plants, the likelihood of non-target plants being attacked decreases (Schaffner et al. 2018). Native Phragmites comprised only one-third of the plants used in the open-field tests of Blossey et al. (2018a, b), with the other two-thirds being plants of European and North American non-native Phrag*mites*. Thus, the result that both moth species only laid 6.5% of eggs on native Phragmites should be considered in light of two considerations: (1) high host plant densities may have inhibited searching behavior of herbivores (Schaffner et al. 2018) and (2) egg-laying on native Phragmites would have been 33% even if egg-laying was identical across the three groups tested.

Based on our evaluation of the literature, our position is that several critical questions must be explicitly addressed before any biological control agent for non-native *Phragmites* could be safely approved for release in North America:

- 1. What are the expected geographic and ecological range limits for the proposed biocontrol agents in North America?
- 2. What are the impacts of the proposed biocontrol agents on the growth, performance, and population dynamics of non-native *Phragmites*?
- 3. What is the potential for negative non-target impacts by the proposed biocontrol agents, both behavioral (i.e., apparent competition) and evolutionary (i.e., genetic host switches)?
- 4. How likely are native and non-native *Phragmites* to evolve resistance to the proposed biocontrol agents?
- 5. What potential natural enemies do the proposed biological control agents have in North America, how do these vary geographically, and what indirect effects may be predicted from this knowledge?
- 6. How do the answers to the above questions vary among the major lineages, haplotypes, and hybrids present in North America?
- 7. If non-target impacts appear unavoidable, what is the relative risk of biological control vs. other management approaches in terms of the fate of the native *Phragmites* lineage?

Genetics

Both the native and non-native populations of *Phragmites* in North America exhibit high genetic and genomic diversity (Saltonstall 2002, 2003a; Saltonstall et al. 2010; Lambertini et al. 2006, 2012; Lambertini 2016; McCormick et al. 2010; Meyerson et al. 2010a, b, 2012, 2016a, b; Meyerson and Cronin 2013; Kulmatiski et al. 2011; Kettenring and Mock 2012; Colin and Eguiarte 2016). The native *Phragmites* is often referred to as a single lineage when contrasted with the non-native Old World lineage, but it includes 17 distinct haplotypes that have different distributions and range sizes in North America. These native haplotypes have different genetic distances among them and with the introduced lineage (Saltonstall 2003b; Lambertini 2016). This might imply large and still unexplored variation in the response to the proposed biocontrol agents. The western states are a center of diversity for native Phragmites, and native populations can be genetically very different even at the local scale (Kettenring and Mock 2012). This large genetic and geographic variation has not been considered in the laboratory host preference trials, which comprised up to three genotypes per lineage (Blossey et al. 2013). This subset is unlikely to represent the wide range of genetic, morphologic, and physiologic variation, as well as variation of environments, in North America. Moreover, other native, potentially endangered, *Phragmites* populations were recently discovered in México (Colin and Eguiarte 2016), where biocontrol could spread from the United States.

In addition, hybridization between native and nonnative lineages has been detected in multiple studies (Meyerson et al. 2010a; Paul et al. 2010; Saltonstall et al. 2014, 2016; Wu et al. 2015; Lambertini 2016). The occurrence and impacts of hybrids in populations should not be underestimated, as has been shown with hybrid complexes of a number of other plant species. Hybrids may be "sinks" for insects and divert the biocontrol from its target (Floate et al. 1993; Williams et al. 2014). On the other hand, hybrids could escape the biocontrol and replace the non-native populations. For example, Hallgren et al. (2003) found that phenolic compounds decreased with increasing introgression in Salix hybrids. In Tamarix spp., introgression increased with latitude and was highly correlated with resistance to insect attack (Williams et al. 2014). Hybrids may also have a different phenology from their parents and increase the fecundity of the herbivore by setting leaves earlier in the spring (Floate et al. 1993). In one case, grazing geese ignored Spartina hybrids and preferentially consumed native plants, contributing to the expansion of the hybrid invasive populations in a California estuary (Grosholz 2010). These findings indicate that hybridization and introgression play an important role in structuring herbivore communities. This aspect has not been considered in the studies of host specificity for the biocontrol of Phragmites.

Ecosystem services provided by Phragmites

The negative impacts of non-native *Phragmites* in North America are well-documented (Packer et al.

2017 and references within). However, *Phragmites* also provides many ecosystem services in North America (Hershner and Havens 2008; Kiviat 2013). These services prominently include nutrient sequestration in eutrophic environments (Meyerson et al. 2000; Mozdzer et al. 2010), phytoremediation of heavy metals and nutrients (Meyerson et al. 2000; Windham et al. 2003), protection of coastal soils from storms and enabling marshes to better keep up with sea level rise (Windham and Lathrop 1999; Rooth and Stevenson 2000; Rooth et al. 2003; Ravit et al. 2015; Knight et al. 2018), carbon sequestration (Caplan et al. 2015), providing faunal habitat (Kane 2001; Weis and Weis 2003; Kiviat 2013), enabling the continuation of cultural uses by Native Americans (Kiviat and Hamilton 2001; Long et al. 2003), and widespread use for sewage treatment and sludge dewatering (Begg et al. 2001; Rodríguez and Brisson 2015). Most of these services are due to non-native Phragmites, although the cultural uses by Native Americans are at least partly due to native Phragmites. An especially important example of soil stabilization exists on the Louisiana coast where a recently flourishing, nonnative scale insect is devastating large expanses of Phragmites and causing marsh collapse (Knight et al. 2018). Although Phragmites is invasive in coastal habitats, it may be one of the few species capable of keeping pace with accelerating sea level rise, though there is some evidence that it can also accelerate carbon loss (Bernal et al. 2016 but see Kirwan et al. 2016). If Archanara spp. were to remove substantial biomass from non-native Phragmites, a number of the aforementioned ecosystem services would be adversely affected. In particular, if high-density stands experienced widespread mortality, the belowground structure of wetlands may become destabilized and collapse, allowing the wetland to become permanently inundated.

Biocontrol generally reduces weed biomass substantially (Stiling and Cornelissen 2005). Classical biocontrol tends to be spatially nonselective, and, if the proposed biocontrol program is established, may greatly reduce the aboveground and belowground *Phragmites* biomass and structure that supports ecosystem services. The architecture of *Phragmites* stands after being affected by biocontrol is unpredictable and might not support the biodiversity and other desirable services now supported by non-native *Phragmites*. For example, reduced aboveground or belowground biomass might lessen the protection of tidal marsh sediments; shorter or more slender culms might provide less sturdy nest substrate for marsh wrens (Cistothorus palustris Wilson); and lack of seed production might affect the food supply of certain migrant or wintering songbirds (see Kiviat 2013 and references within). Native Phragmites, or other robust native graminoids such as Spartina alterniflora or Typha L., might replace some of the biologicallycontrolled non-native Phragmites. However, at least in some situations, non-native Phragmites has been shown to exceed native *Phragmites* in rhizosphere oxygenation, photosynthetic rate, photosynthetic canopy, specific leaf area, nitrogen content, length of growing season, sexual reproduction, shoot density and height, biomass, and relative growth rate (Mozdzer and Zieman 2010; Kettenring and Mock 2012; Tulbure et al. 2012; Mozdzer et al. 2013), but it is not known how these traits might drive differences in ecosystem services overall. Also, as explained above, native Phragmites may be adversely affected by biocontrol of non-native Phragmites. The proposed biocontrol program may substantially reduce biomass and cover of native as well as non-native Phragmites. We disagree strongly with Blossey et al. (2018a), who "consider the potential threat to P. australis americanus demography by A. geminipuncta and A. neurica to be far smaller than allowing continued expansion of invasive P. australis."

Conclusions

It is difficult to find host-specific biocontrol agents at the species level, and finding them at the subspecies level is especially challenging. Phragmites australis is not the only cryptic, subspecific invader; reed canary grass (Phalaris arundinacea L.) is another example (Nelson et al. 2014). We are concerned that Casagrande et al. (2018) and Blossey et al. (2018a) have downplayed our previously published analyses (Bhattarai et al. 2016; Cronin et al. 2016), namely that biocontrol insects targeting non-native Phragmites may cause substantial damage to native Phragmites, while not adequately addressing a well-established body of literature relevant to this question. Native Phragmites constitutes a unique and complex reservoir of genetic diversity and provides ecosystem services that could be lost with the introduction of a

risky biocontrol agent. Previous responses by Blossey and Casagrande (2016a, b), and the recent publications by Casagrande et al. (2018) and Blossey et al. (2018a), have not satisfactorily addressed concerns surrounding the potential for damage to native Phragmites. Inasmuch as substantial reduction of native as well as non-native Phragmites across the North American continent could result from the proposed classical biological control, all of the relevant ecological, social, and economic concerns must be considered explicitly. Because both beneficial and detrimental effects of *Phragmites* are site-specific, and because biocontrol may harm native lineages as well as nonnative Phragmites, management of Phragmites should be situational rather than general. We strongly disagree with the conclusions and recommendations of Casagrande et al. (2018) and Blossey et al. (2018a) and warn that approval of Archanara spp. as biocontrol agents for non-native Phragmites in North America could result in widespread ecological and socioeconomic impacts.

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Compliance with ethical standards

Conflict of interest The authors state they have no conflicts of interest.

References

- Allen WJ, Young RE, Bhattarai GP, Croy JR, Lambert AM, Meyerson LA, Cronin JT (2015) Multitrophic enemy escape of invasive *Phragmites australis* and its introduced herbivores in North America. Biol Invasions 17:3419–3432
- Allen GA, McCormick LJ, Jantzen JR, Marr KL, Brown BN (2017a) Distributional and morphological differences between native and introduced common reed (*Phragmites australis*, Poaceae) in western Canada. Wetlands 37:819–827
- Allen WJ, Meyerson LA, Cummings D, Anderson J, Bhattarai GP, Cronin JT (2017b) Biogeography of a plant invasion: drivers of latitudinal variation in local enemy release. Glob Ecol Biogeogr 26:435–446
- Andres LA (1985) Interaction of *Chrysolina quadrigemina* and *Hypericum* spp. in California. In: Delfosse ES (ed) Proceedings of the VI international symposium on biological control of weeds, August 1984, Agriculture Canada, Vancouver, pp 235–239
- Araujo SBL, Pires Braga M, Brooks DR, Agosta SJ, Hoberg EP, von Hartenthal FW, Boeger WA (2015) Understanding

host-switching by ecological fitting. PLoS ONE 10:e0139225

- Barbier EB, Hacker SD, Kennedy C, Koch EW, Stier AC, Silliman BR (2011) The value of estuarine and coastal ecosystem services. Ecol Monogr 81(2):169–193
- Begg JS, Lavigne RL, Veneman PLM (2001) Reed beds: constructed wetlands for municipal wastewater treatment plant sludge dewatering. Water Sci Technol 44:393–398
- Bernal B, Megongal JP, Mozdzer TJ (2016) An invasive wetland grass primes deep soil carbon pools. Glob Chang Biol 23(5):2104–2116
- Bhattarai GP, Allen WJ, Cronin JT, Kiviat E, Meyerson LA (2016) Response to Blossey and Casagrande—ecological and evolutionary processes make host specificity at the subspecies level exceedingly unlikely. Biol Invasions 18:2757–2758
- Bhattarai GP, Meyerson LA, Anderson J, Cummings D, Allen WJ, Cronin JT (2017a) Biogeography of a plant invasion: genetic variation and plasticity in latitudinal clines for traits related to herbivory. Ecol Monogr 87:57–75
- Bhattarai GP, Meyerson LA, Cronin JT (2017b) Geographic variation in apparent competition between native and invasive *Phragmites australis*. Ecology 98(2):349–358
- Blossey B (2003) A framework for evaluating potential ecological effects of implementing biological control of *Phragmites australis*. Estuaries 26:607–617
- Blossey B (2014) Identification, development, and release of insect biocontrol agents for the management of *Phragmites australis*. ERDC/EL CR-14-2. US Army Corps of Engineers, Washington
- Blossey B, Casagrande RA (2016a) Biological control of invasive *Phragmites* may safeguard native *Phragmites* and increase wetland conservation values. Biol Invasions 18(9):2753–2755
- Blossey B, Casagrande RA (2016b) Response to Bhattarai et al.: trait differences between native and introduced genotypes results in subspecies level specificity in select *Phragmites* herbivores. Biol Invasions 18:2759–2760
- Blossey B, McCauley J (2000) A plan for developing biological control of *Phragmites australis* in North America. Wetl J 12:23–28
- Blossey B, Casagrande RA, Tewksbury L, Hinz H, Häfliger P, Martin L, Cohen J (2013) Identifying, developing and releasing insect biocontrol agents for the management of *Phragmites australis*. ERDC/EL TN-13-3. U.S. Army Engineer Research and Development Center, Vicksburg
- Blossey B, Häfliger P, Tewksbury L, Dávalos A, Casagrande R (2018a) Host specificity and risk assessment of Archanara geminipuncta and Archanara neurica, two potential biocontrol agents for invasive Phragmites australis in North America. Biol Control 125:98–112
- Blossey B, Häfliger P, Tewksbury L, Dávalos A, Casagrande R (2018b) Complete host specificity test plant list and associated data to assess host specificity of Archanara geminipuncta and Archanara neurica, two potential biocontrol agents for invasive Phragmites australis in North America. Data in Brief 19:1755–1764. https://doi.org/10.1016/j.dib. 2018.06.068
- Buswell JM, Moles AT, Hartley S (2010) Is rapid evolution common in introduced plant species? J Ecol 99:214–224

- Caplan JS, Hager RN, Megonigal JP, Mozdzer TJ (2015) Global change accelerates carbon assimilation by a wetland ecosystem engineer. Environ Res Lett 10:115006. https:// doi.org/10.1088/1748-9326/10/11/115006
- Carroll S, Boyd C (1992) Host race radiation in the soapberry bug—natural history with the history. Evolution 46:1052–1069
- Casagrande RA, Häfliger P, Hinz HL, Tewksbury L, Blossey B (2018) Grasses as appropriate targets in weed biocontrol: is the common reed, *Phragmites australis*, an anomaly? Biocontrol 63:391–403. https://doi.org/10.1007/s10526-018-9871-y
- Castagneyrol B, Jactel H, Brockerhoff EG, Perrette N, Larter M, Delzon S, Piou D (2016) Host range expansion is density dependent. Oecologia 182:779–788
- Cenzer ML (2016) Adaptation to an invasive host is driving the loss of a native ecotype. Evolution 70:2296–2307
- Ciotir C, Kirk H, Row JR, Freeland JR (2013) Intercontinental dispersal of *Typha angustifolia* and *T. latifolia* between Europe and North America has implications for *Typha* invasions. Biol Invasions 15:1377–1390
- Cipollini D, Peterson DL (2018) The potential for host switching via ecological fitting in the emerald ash borer-host plant system. Oecologia 187:507–519
- Colin R, Eguiarte LE (2016) Phylogeographic analyses and genetic structure illustrate the complex evolutionary history of *Phragmites australis* in Mexico. Am J Bot 103(5):876–887
- Cronin JT, Bhattarai GP, Allen WJ, Meyerson LA (2015) Biogeography of a plant invasion: plant-herbivore interactions. Ecology 96:1115–1127
- Cronin JT, Kiviat E, Meyerson LA, Bhattarai GP, Allen WJ (2016) Biological control of invasive *Phragmites australis* will be detrimental to native *P. australis*. Biol Invasions 18:2749–2752
- Des Roches S, Post DM, Turley NE, Bailey JK, Hendry AP, Kinnison MT, Schweitzer JA, Palkovacs EP (2018) The ecological importance of intraspecific variation. Nat Ecol Evol 2:57–64
- Desurmont GA, Donoghue MJ, Clement WL, Agrawal AA (2011) Evolutionary history predicts plant defense against an invasive pest. Proc Natl Acad Sci 108:7070–7074
- Dingle H, Carroll SP, Famula TR (2009) Influence of genetic architecture on contemporary local evolution in the soapberry bug, *Jadera haematoloma*: artificial selection on beak length. J Evol Biol 22:2031–2040
- Erbilgin N, Ma C, Whitehouse C, Shan B, Najar A, Evenden M (2014) Chemical similarity between historical and novel host plants promotes range and host expansion of the mountain pine beetle in a naïve host ecosystem. New Phytol 201:940–950
- Floate KD, Whitham TG (1993) The hybrid bridge hypothesis host shifting via plant hybrid swarms. Am Nat 141:651–662
- Floate KD, Kaersley MJC, Whitham TG (1993) Elevated herbivory in plant hybrid zone: *Chrysomela confluens, Populus* and phenological sinks. Ecology 74:2056–2065
- Gaskin JF, Schaal BA (2002) Hybrid *Tamarix* widespread in U.S. invasion and undetected in native Asian range. Proc Natl Acad Sci 99:11256–11259

- Gilbert GS, Briggs HM, Magarey R (2015) The impact of plant enemies shows a phylogenetic signal. PLoS ONE. https:// doi.org/10.1371/journal.pone.0123758
- Graves SD, Shapiro AM (2003) Exotics as host plants of the California butterfly fauna. Biol Conserv 110:413–433
- Grosholz E (2010) Avoidance by grazers facilitates spread of an invasive hybrid plant. Ecol Lett 13:145–153
- Guo W-Y, Lambertini C, Nguyen LX, Li X-Z, Brix H (2014) Preadaptation and post-introduction evolution facilitate the invasion of *Phragmites australis* in North America. Ecol Evol 4:4567–4577
- Guo W-Y, Lambertini C, Pyšek P, Meyerson LA, Brix H (2018) Living in two worlds: evolutionary mechanisms act differently in the native and introduced ranges of an invasive plant. Ecol Evol 8:2440–2452
- Häfliger P, Schwarzländer M, Blossey B (2006) Impact of Archanara geminipuncta (Lepidoptera: Noctuidae) on aboveground biomass production of Phragmites australis. Biol Control 38:413–421
- Hallgren P, Ikonen A, Hjaelte J, Roininen H (2003) Inheritance patterns of phenolics in F1, F2 and back-cross hybrids of willows: implications for herbivore responses to hybrid plants. J Chem Ecol 29:1143–1158
- Hazelton EL, Mozdzer TJ, Burdick DM, Kettenring KM, Whigham DF (2014) *Phragmites australis* management in the United States: 40 years of methods and outcomes. AoB PLANTS 6:plu001. https://doi.org/10.1093/aob-pla/ plu001
- Heimpel GE, Cock MJW (2018) Shifting paradigms in the history of classical biological control. Biocontrol 63:27–37
- Hershner C, Havens KJ (2008) Managing invasive aquatic plants in a changing system: strategic consideration of ecosystem services. Conserv Biol 22(3):544–550
- Kane R (2001) *Phragmites* use by birds in New Jersey. Rec New Jersey Birds 26:122–124
- Kettenring KM, Mock KE (2012) Genetic diversity, reproductive mode, and dispersal differ between the cryptic invader, *Phragmites australis*, and its native conspecific. Biol Invasions 14:2489–2504
- Kirwan ML, Temmerman S, Skeehan EE, Guntenspergen GR, Fagherazzi S (2016) Overestimation of marsh vulnerability to sea level rise. Nat Clim Chang 6(3):253–260
- Kiviat E (2013) Ecosystem services of *Phragmites* in North America with emphasis on habitat functions. AoB PLANTS 5:plt008. https://doi.org/10.1093/aobpla/plt00
- Kiviat E, Hamilton E (2001) *Phragmites* use by Native North Americans. Aquat Bot 69(2–4):341–357
- Knight IA, Wilson BE, Gill M, Aveles L, Cronin JT, Nyman JA, Schneider SA, Diaz R (2018) Invasion of *Nipponaclerda biwakoensis* (Hemiptera: Aclerdidae) and associated *Phragmites australis* dieback in southern Louisiana. Biol Invasions 20:2739–2744
- Kulmatiski A, Beard KH, Meyerson LA, Gibson JR, Mock KE (2011) Nonnative *Phragmites australis* invasion into Utah wetlands. West N Am Nat 70(4):541–552
- Lambert AM, Casagrande RA (2007) Susceptibility of native and non-native common reed to the non-native mealy plum aphid (Homoptera: Aphididae) in North America. Environ Entomol 36:451–457

- Lambert AM, Winiarski K, Casagrande RA (2007) Distribution and impact of exotic gall flies (*Lipara* sp. [sic]) on native and exotic *Phragmites australis*. Aquat Bot 86:163–170
- Lambertini C (2016) Heteroplasmy due to chloroplast paternal leakage: another insight into *Phragmites* haplotypic diversity in North America. Biol Invasions 18:2443–2455
- Lambertini C, Gustafsson MHG, Frydenberg J, Lissner J, Speranza M, Brix H (2006) A phylogeographic study of the cosmopolitan genus *Phragmites* (Poaceae) based on AFLPs. Plant Syst Evol 258(3–4):161–182
- Lambertini C, Mendelssohn IA, Gustafsson MHG, Olesen B, Riis T, Sorrell BK, Brix H (2012) Tracing the origin of Gulf Coast *Phragmites* (Poaceae): a story of long-distance dispersal and hybridization. Am J Bot 99:538–551
- Long J, Tecle A, Burnette B (2003) Cultural foundations for ecological restoration on the White Mountain Apache Reservation. Conserv Ecol 8(1). Available via Ecology and Society http://www.consecol.org/vol8/iss1/art4
- Louda SM, Arnett AE, Rand TA, Russell FL (2003) Invasiveness of some biological control insects and adequacy of their ecological risk assessment and regulation. Conserv Biol 17(1):73–82
- Maron JL, Vilà M, Bommarco R, Elmendorf S, Beardsley P (2004) Rapid evolution of an invasive plant. Ecol Monogr 74:261–280
- McCormick MK, Kettenring KM, Baron HM, Whigham DF (2010) Spread of invasive *Phragmites australis* in estuaries with differing degrees of development: genetic patterns, Allee effects and interpretation. J Ecol 98:1369–1378
- Meyerson LA, Cronin JT (2013) Evidence for multiple introductions of *Phragmites australis* to North America: detection of a new non-native haplotype. Biol Invasions 15:2605–2608
- Meyerson LA, Mooney HA (2007) Invasive alien species in an era of globalization. Front Ecol Environ 5:199–208
- Meyerson LA, Saltonstall K, Windham L, Kiviat E, Findlay S (2000) A comparison of *Phragmites australis* in freshwater and brackish marsh environments in North America. Wetl Ecol Manag 8:89–103
- Meyerson LA, Saltonstall K, Chambers RM (2009) *Phragmites* australis in eastern North America: a historical and ecological perspective. In: Silliman BR, Grosholz E, Bertness MD (eds) Salt marshes under global siege. University of California Press, Oakland
- Meyerson LA, Lambert AM, Saltonstall K (2010a) A tale of three lineages: expansion of common reed (*Phragmites australis*) in the U.S. Southwest and Gulf Coast. Invasive Plant Sci Manag 3:515–520
- Meyerson LA, Viola D, Brown R (2010b) Hybridization of invasive *Phragmites australis* with a native subspecies in North America. Biol Invasions 12:103–111
- Meyerson LA, Lambertini C, McCormick M, Whigham DF (2012) Hybridization of common reed in North America? The answer is blowing in the wind. AoB PLANTS 2012:pls1022. https://doi.org/10.1093/aobpla/pls1022
- Meyerson LA, Cronin JT, Bhattarai GP, Brix H, Lambertini C, Lučanová M, Rinehart S, Suda J, Pyšek P (2016a) Do ploidy level and nuclear genome size and latitude of origin modify the expression of *Phragmites australis* traits and interactions with herbivores? Biol Invasions 18:2531–2549

- Meyerson LA, Cronin JT, Pyšek P (2016b) *Phragmites australis* as a model organism for studying plant invasions. Biol Invasions 18:2421–2431
- Mozdzer TJ, Zieman JC (2010) Ecophysiological differences between genetic lineages facilitate the invasion of nonnative *Phragmites australis* in North American Atlantic coast wetlands. J Ecol 98:451–458
- Mozdzer TJ, Zieman JC, McGlathery KJ (2010) Nitrogen uptake by native and invasive temperate coastal macrophytes: importance of dissolved organic nitrogen. Estuaries Coasts 33:784–797
- Mozdzer TJ, Brisson J, Hazelton EL (2013) Physiological ecology and functional traits of North American native and Eurasian introduced *Phragmites australis* lineages. AoB Plants 5:plt048. https://doi.org/10.1093/aobpla/plt048
- Murdoch WW (1969) Switching in general predators: experiments on predator specificity and stability of prey populations. Ecol Monogr 39:335–354
- Nelson MF, Anderson NO, Casler MD, Jakubowski AR (2014) Population genetic structure of N. American and European *Phalaris arundinacea* L. as inferred from inter-simple sequence repeat markers. Biol Invasions 16(2):353–363
- Packer J, Meyerson LA, Skálová H, Pyšek P, Kueffer C (2017) Biological flora of the British Isles: *Phragmites australis*. J Ecol 105:1123–1162
- Park MG, Blossey B (2008) Importance of plant traits and herbivory for invasiveness of *Phragmites australis* (Poaceae). Am J Bot 95:1557–1568
- Parker IM, Saunders M, Bontrager M, Weitz AP, Hendricks R, Magarey R, Suiter K, Gilbert GS (2015) Phylogenetic structure and host abundance drive disease pressure in communities. Nature 520:542–544
- Paul J, Vachon N, Garroway CJ, Freeland JR (2010) Molecular data provide strong evidence of natural hybridization between native and introduced lineages of *Phragmites australis* in North America. Biol Invasions 12:2967–2973
- Paynter Q, Fowler SV, Gourlay AH, Peterson PG, Smith LA, Winks CJ (2015) Relative performance on test and target plants in laboratory tests predicts the risk of non-target attack in the field for arthropod weed biocontrol agents. Biol Control 80:133–142
- Pearse IS, Altermatt F (2013) Predicting novel trophic interactions in a non-native world. Ecol Lett 16:1088–1094
- Pearse IS, Hipp AL (2009) Phylogenetic and trait similarity to a native species predict herbivory on non-native oaks. Proc Natl Acad Sci 106:18097–18102
- Pearse IS, Harris DJ, Karban R, Sih A (2013) Predicting novel herbivore-plant interactions. Oikos 122:1554–1564
- Pearson DE, Callaway RM (2005) Indirect nontarget effects of host-specific biological control agents: implications for biological control. Biol Control 35:288–298
- Prentis PJ, Wilson JRU, Dormontt EE, Richardson DM, Lowe AJ (2008) Adaptive evolution in invasive species. Trends Plant Sci 13:288–294
- Ravit B, Weis JS, Rounds D (2015) Is urban marsh sustainability compatible with the Clean Water Act? Environ Pract 17(1):46–56
- Rodríguez M, Brisson J (2015) Pollutant removal efficiency of native versus exotic common reed (*Phragmites australis*) in North American treatment wetlands. Ecol Eng 74:364–370

- Rooth JE, Stevenson JC (2000) Sediment deposition patterns in *Phragmites australis* communities: implications for coastal areas threatened by rising sea level. Wetl Ecol Manag 8:173–183
- Rooth JE, Windham L (2000) *Phragmites* on death row: is biocontrol really warranted? Wetl J 12:29–37
- Rooth JE, Stevenson JC, Cornwell JC (2003) Increased sediment accretion rates following invasion by *Phragmites australis*: the role of litter. Estuaries 26(2B):475–483
- Saltonstall K (2002) Cryptic invasion by a non-native genotype of the common reed, *Phragmites australis*, into North America. Proc Natl Acad Sci USA 99:2445–2449
- Saltonstall K (2003a) Microsatellite variation within and among North American lineages of *Phragmites australis*. Mol Ecol 12:1689–1702
- Saltonstall K (2003b) Genetic variation among North American populations of *Phragmites australis*: implications for management. Estuaries 26:444–451
- Saltonstall K (2003c) A rapid method for identifying the origin of North American *Phragmites* populations using RFLP analysis. Wetlands 23:1043–1047
- Saltonstall K, Stevenson JC (2007) The effect of nutrients on seedling growth of native and introduced *Phragmites australis*. Aquat Bot 86:331–336
- Saltonstall K, Peterson PM, Soreng RJ (2004) Recognition of *Phragmites australis* subsp. *americanus* (Poaceae: Arundinoideae) in North America: evidence from morphological and genetic analysis. SIDA Contrib Bot 21:683–692
- Saltonstall K, Lambert A, Meyerson LA (2010) Genetics and reproduction of common (*Phragmites australis*) and giant reed (*Arundo donax*). Invasive Plant Sci Manag 3:495–505
- Saltonstall K, Castillo HE, Blossey B (2014) Confirmed field hybridization of native and introduced *Phragmites australis* (Poaceae) in North America. Am J Bot 101:211–215
- Saltonstall K, Lambert AM, Rice N (2016) What happens in Vegas, better stay in Vegas: *Phragmites australis* hybrids in the Las Vegas Wash. Biol Invasions 18:2463–2474
- Schaffner U, Smith L, Cristofaro M (2018) A review of openfield host range testing to evaluate nontarget use by herbivorous biological control candidates. Biocontrol 63:405–416
- Simberloff D (2012) Risks of biological control for conservation purposes. Biocontrol 57:263–276
- Simberloff D, Stiling P (1996) How risky is biological control? Ecology 77:1965–1974
- Stastny M, Sargent RD (2017) Evidence for rapid evolutionary change in an invasive plant in response to biological control. J Evol Biol 30:1042–1052
- Stiling P, Cornelissen T (2005) What makes a successful biocontrol agent? A meta-analysis of biological control agent performance. Biol Control 34:236–246
- Stohlgren TJ, Pyšek P, Kartesz J, Nishino M, Pauchard A, Winter M, Pino J, Richardson DM, Wilson JRU, Murray BR, Phillips ML, Ming-yang L, Celesti-Grapow L, Font X (2011) Widespread plant species: natives versus aliens in our changing world. Biol Invasions 13:1931–1944
- Stutz S, Mráz P, Hinz HL, Müller-Schärer H, Schaffner U (2018) Biological invasion of oxeye daisy (*Leucanthemum* vulgare) in North America: pre-adaptation, post-introduction evolution, or both? PLoS ONE 13:e0190705

- Suckling DM, Sforza RFH (2014) What magnitude are observed non-target impacts from weed biocontrol? PLoS ONE 9:e84847
- Swearingen J, Saltonstall K (2012) *Phragmites* field guide: distinguishing native and exotic forms of common reed (*Phragmites australis*) in the United States. Technical Note, Natural Resources Conservation Service, US Department of Agriculture
- Szűcs M, Schaffner U, Price WJ, Schwarzländer M (2012) Postintroduction evolution in the biological control agent *Longitarsus jacobaeae* (Coleoptera: Chrysomelidae). Evol Appl 5:858–868
- Tewksbury L, Casagrande R, Blossey B, Häfliger P, Schwarzländer M (2002) Potential for biological control of *Phragmites australis* in North America. Biol Control 23(2):191–212
- Tomasetto F, Cianciullo S, Reale M, Attorre F, Olaniyan O, Goldson SL (2018) Breakdown in classical biological control of Argentine stem weevil: a matter of time. Biocontrol 63:521–531
- Tulbure MG, Ghioca-Robrecht DM, Johnston CA, Whigham DF (2012) Inventory and ventilation efficiency of nonnative and native *Phragmites australis* (common reed) in tidal wetlands of the Chesapeake Bay. Estuaries Coasts 35:1353–1359
- Turner KG, Hufbauer RA, Rieseberg LH (2014) Rapid evolution of an invasive weed. New Phytol 202:309–321
- USDA (U.S. Department of Agriculture) (2016) Technical advisory group for biological control agents of weeds manual. Interim edition. USDA, Washington, DC. https:// www.aphis.usda.gov/import_export/plants/manuals/ domestic/downloads/tag-bcaw_manual.pdf. Accessed 1 Apr 2018
- van Klinken RD, Edwards OR (2002) Is host specificity of weed biocontrol agents likely to evolve rapidly following establishment? Ecol Lett 5:590–595
- Weis JS, Weis P (2003) Is the invasion of the common reed, *Phragmites australis*, into tidal marshes of the eastern US an ecological disaster? Mar Pollut Bull 46(7):816–820

- Whitfeld TJS, Novotny V, Miller SE, Hrcek J, Klimes P, Weiblen GD (2012) Predicting tropical insect herbivore abundance from host plant traits and phylogeny. Ecology 93:S211–S222
- Williams WI, Friedman JM, Gaskin JF, Norton AP (2014) Hybridization of an invasive shrub affects tolerance and resistance to defoliation by a biological control agent. Evol Appl 1:11. https://doi.org/10.1111/eva.12134
- Williams J, Lambert AM, Long R, Saltonstall K (2019) Does hybrid *Phragmites australis* differ from native and introduced lineages in reproductive, genetic, and morphological traits? Am J Bot 106:29–41
- Willson KG, Perantoni AN, Berry ZC, Eicholtz MI, Tamukong YB, Yarwood SA, Baldwin AH (2017) Influences of reduced iron and magnesium on growth and photosynthetic performance of *Phragmites australis* subsp. *americanus* (North American common reed). Aquat Bot 137:30–38
- Windham L, Lathrop RG (1999) Effects of *Phragmites australis* (common reed) invasion on aboveground biomass and soil properties in brackish tidal marsh of the Mullica River, New Jersey. Estuaries 22:927–935
- Windham L, Weis JS, Weis P (2003) Uptake and distribution of metals in two dominant salt marsh macrophytes, *Spartina alterniflora* (cordgrass) and *Phragmites australis* (common reed). Estuar Coast Shelf Sci 56:63–72
- Wright MG, Bennett GM (2018) Evolution of biological control agents following introduction to new environments. Biocontrol 63:105–116
- Wu CA, Murray LA, Heffernan KE (2015) Evidence for natural hybridization between native and introduced lineages of *Phragmites australis* in the Chesapeake Bay watershed. Am J Bot 102:805–812

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