

Natural enemies associated with *Tuta absoluta* and functional biodiversity in vegetable crops

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Abstract Tuta absoluta Meyrick (Lepidoptera: Gelechiidae) is considered one of the main pests threatening tomato production worldwide. In the Mediterranean, the use of predatory mirids is widespread, and although several larval parasitoid species have been recorded in the area, their contribution to the biological control of the pest is often neglected. With the general objective of improving the biological control of T. absoluta, our field study aimed to determine the relative abundance of natural enemies associated with T. absoluta in tomato fields and to assess whether insectary plants placed in the vicinity of vegetable crops would help to improve functional biodiversity in the farms. The study was conducted during two years in six commercial tomato fields in Northeast Spain that were managed using an integrated pest management program based on predatory mirid conservation. Our results indicated that Necremnus tutae Ribes & Bernardo (Hymenoptera: Eulophidae) was by far the main natural enemy responsible for larval parasitism of T. absoluta. The flowering plants

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Y. Aparicio Lausanne, Switzerland used in the present study were attractive to several natural enemies that are of interest not only for tomato but also for other vegetable crops coexisting at the farm level, including parasitoid wasps, hoverflies, and other important beneficials, such as *Orius* spp., ladybeetles and predatory thrips.

Keywords Conservation biological control · Tomato · *Lobularia maritima* · *Sinapis alba* · *Achillea millefolium* · *Fagopyrum esculentum*

Introduction

Tuta absoluta Meyrick (Lepidoptera: Gelechiidae) is considered one of the main tomato pests globally. After invading Spain in 2006, it quickly spread to several countries beyond its area of origin (Desneux et al. 2010, 2011; Biondi et al. 2018; Han et al. 2019), threating tomato production. In the Mediterranean, the use of predatory mirid bugs following either conservation or inoculation strategies is one of the most widely used methods for biological control of the pest (Urbaneja et al. 2012). These predators are polyphagous and feed not only on *T. absoluta* but also on other vegetable pests (Arnó et al. 2018a). In the Mediterranean, several predators have been described as feeding efficiently on *T. absoluta* eggs but as poor predators of *T. absoluta* larvae, such as *Macrolophus* *pygmaeus* Rambur, *Nesidiocoris tenuis* (Reuter), *Dicyphus errans* (Wolff), and *Dicyphus bolivari* Lindberg (Hemiptera: Miridae) (Arnó et al. 2009; Urbaneja et al. 2009; Ingegno et al. 2019).

Regarding parasitoids, several species have been reported worldwide (Zappalà et al. 2013; Biondi et al. 2018; Mansour et al. 2018; Salas Gervassio et al. 2019). In the Spanish Mediterranean area, Gabarra et al. (2014) described a number of species attacking T. absoluta larvae belonging to the five families also recorded in other parts of the Mediterranean (Biondi et al. 2018; Mansour et al. 2018). These authors also reported the occurrence of egg parasitoids of the genus Trichogramma. Among the larval parasitoids, the genus Necremnus has been found the most abundant and widespread. This genus was reviewed by Gebiola et al. (2015), who found that several of the individuals that had been classified as Necremnus artynes (Walker) (Hymenoptera: Eulophidae) or N. nr. artynes actually corresponded to a new species they called Necremnus tutae Ribes & Bernardo.

The biology of *N. tutae* (referred to as *N. artynes*) has been extensively studied (Calvo et al. 2013, 2016; Chailleux et al. 2014; Bodino et al. 2016). Fitness of this parasitoids benefits from feeding on sugar-reach diet such as honey and honeydew (Arnó et al. 2018b; de Campos et al. 2020). This species has been considered for inoculative release as a control strategy against *T. absoluta* by several companies that mass rear natural enemies. Although currently it is not commercially available, it occurs spontaneously and abundantly in many tomato fields (Abbes et al. 2014; Crisol-Martínez and van der Blom 2019). Therefore, its promotion could be a good strategy to improve the biological control of *T. absoluta*.

The use of floral strips around crops is a recommended practice to promote natural enemies close to the fields in conservation biological control (Blaauw and Isaacs 2012). In particular, the use of strips of *Calendula officinalis* L. (Asteraceae) close to tomato fields to boost mirid predator populations and to be their refuge in non-crop periods is a practice that is quickly becoming common in the horticultural farms in Northeast Spain (Arnó et al. 2018a; Ardanuy et al. 2021; Agustí et al. 2020). However, this plant does not seem particularly attractive to parasitoids (Arnó et al. 2012). Including other plants that better contribute to conserving and increasing parasitoids of *T. absoluta* and other pests might enhance biological control in tomato and other vegetables. In this regard, Arnó et al. (2018b) and Balzan and Wäckers (2013) demonstrated in the laboratory that some plants, such as Lobularia maritima L. (Brassicaceae), could be useful in improving the survival and reproductive potential of N. tutae and other T. absoluta larval parasitoids. Arnó et al. (2018b) also reported benefits regarding the fitness of the larval parasitoids of T. absoluta when fed on Sinapis alba L. (Brassicaceae) and Fagopyrum esculentum Moench (Polygonaceae). In a horticultural production system with coexisting crops, the functionality of insectary plants must go beyond the promotion of a single group of parasitoid and extend to a wider range of natural enemies. Wäckers and van Rijn (2005) summarized the importance of flowers as food providers for several groups of natural enemies, including those listed in terms of interest in conservation biological control in vegetables (Riudavets et al. 2020). Considering this background and with the general objective of improving the biological control of T. absoluta, our field study aimed to determine the relative abundance of natural enemies associated with T. absoluta in tomato fields and to assess whether the use of insectary flowering plants close to the crops would help to improve functional biodiversity in vegetable farms.

Materials and methods

Study area and experimental set-up

The study was conducted in six tomato open-air fields (three in 2015 and three in 2016) of 0.25 ± 0.05 ha located in the Maresme county (Barcelona, Spain) from the second half of July to the last week of September. The fields were managed with an integrated pest management (IPM) program based on the conservation of predatory bugs and the use of selective insecticides when necessary (Arnó et al. 2018a). As in Aparicio et al. (2021), four patches (1 m^2) with four plant containers (40 cm \emptyset) were arranged along one of the sides of each field at approximately 0.5-1.8 m apart from the first row of tomatoes. These patches were separated by an approximate distance of 2 m. In each container, four to eight plants of the following species were planted: L. maritima, S. alba, Achillea millefolium L. (Asteraceae), and F. esculentum. Hence, in each patch, all four plant species were

present. These species were chosen according to the previous results of our research group (Arnó et al. 2012, 2018b). All plants were grown from seed in seed-starting pots with potting soil in a greenhouse, transplanted to the containers, and taken to the field at week 28 (mid-July) in 2015 and 2016 when they were already in bloom. Containers with the insectary plants were kept in the sites until the end of the samplings in week 39 (end of September). Fertigation was provided throughout the experiment simultaneously to the tomato crop. The phenological stage of the different plant species was recorded biweekly (Table 1).

Natural enemies of Tuta absoluta

To estimate the abundance of natural enemies associated with T. absoluta, samplings were carried out using yellow sticky traps and sentinel plants. Yellow sticky traps (24.5 cm tall and 20 cm wide; ATRA-PAXON, Projar S.A, Spain) were biweekly displayed for one week in the field from week 29 to week 39. Eight traps were placed in each field at a distance of approximately 3.5 m inside the crop: four in front of the four patches with the insectary plants (near) and four apart at a distance of 45-240 m (far) depending on the field size. The number of predatory mirid bugs and hymenopteran parasitoids were counted in all the yellow sticky traps in the laboratory. The number of M. pygmaeus and N. tenuis, the two most abundant mirid predators colonizing tomato fields in the area (Arnó et al. 2018a), was recorded separately. The abundance of the different families of T. absoluta parasitoids have been estimated in traps of two fields in weeks 29, 33, and 37 of both years. Hymenoptera were identified at family level when possible using the taxonomic keys of Grissell and Schauff (1990) and Hanson and Gauld (2006). Only families with species previously identified as *T. absoluta* parasitoids in Spain and in the Mediterranean basin were taken into account. Thus, target families were Eulophidae, Braconidae, Chalcididae, Ichneumonidae, Pteromalidae, and Trichogrammatidae (Gabarra et al 2014; Biondi et al. 2018; Mansour et al. 2018).

Sentinel plants were used to assess the parasitism of the larval stages of T. absoluta during the weeks when no yellow sticky traps were in the field to ensure that the two sampling systems would not interfere with each other. They were potted tomato plants of approximately 50 cm high, which were isolated in aerated cages ($60 \times 75 \times 100$ cm) with adults of T. absoluta for seven days at approximately 25° C. After removing the adults, the plants were kept in cages for another week in order to obtain plants with second to third instar T. absoluta larvae. Then, plants were enclosed in a sleeve (50 \times 30 cm) made of fine net $(7 \times 8 \text{ threads } \text{cm}^{-2})$, which was chosen to prevent the exit of T. absoluta adults and the colonization by predatory mirid bugs. Preliminary tests done prior to the field experiment confirmed that this net allowed the entrance of *N. tutae*, which is the most abundant larval parasitoid in the Mediterranean area (Gabarra et al. 2014; Biondi et al. 2018). To avoid plant desiccation, potted plants were placed into bigger pots filled with water. The outside of the pots were sprayed with insect-trapping glue to prevent ants and other non-winged predators from climbing the plant. Similar to the yellow sticky traps, eight plants were placed in each field at a distance of approximately 2 m inside the crop, four near to the insectary plants and four far from them. The plants were taken to the fields at 15 day intervals and deployed for one week in the fields starting in week 30 and ending

Table 1 Number of containers with insectary plants on bloom for each plant species during the sampling periods of 2015 and 2016
out of 12 containers per each insectary plants polling all fields together

Insectary plants	Week of the year 2015					Week of the year 2016						
	29	31	33	35	37	39	29	31	33	35	37	39
A. millefolium	12	12	12	12	11	12	12	12	12	11	11	10
F. esculentum	12	12	12	12	12	12	12	12	12	12	12	12
L. maritima	12	12	12	12	9	12	12	12	12	10	10	10
S. alba	12	8	6	5*	3*	3	12	12	12	0	0	0

*The total number of containers were 11 instead of 12

in week 38 (beginning of October) for both years. In 2016, plants were not available in week 34 due to a failure in the climatic chamber. Therefore, data from this week was missing. Once in the laboratory, galleries were inspected under a stereomicroscope, and the number of live, dead but apparently not parasitized (hereafter "dead"), and ectoparasitized T. absoluta larvae was recorded. All larvae were kept in the climatic chambers for at least four weeks and parasitoids that reached the adult stage were collected and stored in alcohol. A sample of emerged parasitoids individuals were identified to genus level following morphological characterization, and out of them, a subsample was identified to species level following the taxonomic keys of Askew (1968) and Gebiola et al. (2015).

Presence of natural enemies in flower patches

The attractiveness of the insectary plants to natural enemies of T. absoluta and to other insects relevant to the biological control of vegetable pests was evaluated on the same dates that the yellow sticky traps were deployed in the field using two different methods. For hoverflies, which can be easily distinguished during their flight, visual observations of the number of adults that effectively contacted each insectary plant (landing) was recorded for 3 min. For insects that could not be seen during their flight and that were usually within the plant foliage or within the flowers, the beating tray method (hereafter "beating") was used. A bunch of each plant species was separately hand-beaten three consecutive times on a white plastic trav $(24 \times 35 \text{ cm})$. Insects that fell onto the tray were visually classified in situ as hymenopteran parasitoids, predatory mirid bugs (Hemiptera: Miridae), Orius spp. (Hemiptera: Anthocoridae), ladybeetles (Coleoptera: Coccinellidae), lacewings (Neuroptera: Chrysopidae), and Aelothrips spp. (Thysanoptera: Aeolothripidae). In the same samples, the number of aphids, thrips other than Aelothrips spp. (hereafter "thrips"), and Eurydema spp. (Hemiptera: Pentatomidae) was also recorded. Other insect groups were discarded. Both methods were conducted biweekly on the same day, between 9h00 and 12h00, avoiding cloudy and rainy days.

Statistical analysis

Data were analyzed for both years, using a generalized linear mixed-effects model (GLMM) with a negative binomial response distribution using the R function "glmer.nb" and a log link function. The number of insects per beating sample and per yellow sticky trap, and the number of hoverflies landings per insectary plant species were the response variables. The two different distances (treatment factor), nearness to the insectary plants and farness from them, was the main fixed effect. The fields, patches, weeks and the interaction treatment × weeks were the random factors. Statistical inferences for fixed effects were based on Wald-type tests assuming a standard normal distribution. We tested the significance (and contribution) of entire grouping factors of the model using the "Anova" function from package "car" that calculates Wald χ^2 tests (Fox and Weisberg 2019), and pairwise comparisons (post-hoc tests) were conducted with the Tukey's method for multiple comparisons. The statistical analysis was performed using the software R (R Core Team 2020), using the packages "lme4" (Bates et al. 2015), MASS (Venables and Ripley 2002), "emmeans" (Russell Lenth 2020) and "ggplot2" (Wickham 2016). For all statistical tests a nominal significance level of 5% (P < 0.05) was used for the study.

Results

Natural enemies of Tuta absoluta

During the two years of sampling, 3,091 hymenopteran parasitoids were caught in the yellow sticky traps, with number per trap significantly higher near to than far away from the insectary plants patches (12.34 \pm 0.66; 9.84 \pm 0.60, z = 2.89, P = 0.0039). Traps used to estimate the abundance of the different families of T. absoluta parasitoids yielded a total of 940 individuals that were identified to family level. Eighty-nine of them belonged to the target families Eulophidae, Braconidae, Ichneumonidae, Pteromalidae, and Trichogrammatidae, whereas no Chalcididae specimen was identified in the traps. Significant differences in abundance were found among the $(\chi^2 = 69.05,$ families mentioned df = 4, P < 0.0001), with Eulophidae being the most abundant (0.69 \pm 0.12 individuals per trap) compared to the other target families with similar abundances (between 0.01 \pm 0.01 and 0.16 \pm 0.06 individuals per trap). However, the number of individuals of these target families recorded in the yellow sticky traps was not significantly different between those placed near to and far from the insectary plant patches for none of the families and for the total (Table 2).

A total of 817 predatory mirid bugs were caught in the yellow sticky traps for the entire sampling period. When comparing the abundances of both mirid bugs in the yellow sticky traps placed close and far to the insectary plant patches, no significant differences were observed for any of the two species (0.58 \pm 0.15 vs. 0.40 \pm 0.07, z = 0.869, P = 0.3847 for *M. pygmaeus*, and 2.97 \pm 0.56 vs. 1.89 \pm 0.33, z = 0.486, P = 0.6271 for *N. tenuis*).

From the sentinel plants placed within the tomato crop, 3,557 T. absoluta larvae were recovered (671 ectoparasitized, 977 alive and 1,909 dead). Regarding the number of ectoparasitized larvae per plant in the sentinel plants placed near to and far from the flower patches, no significant differences were found (2.73 \pm 0.23 and 3.07 \pm 0.29, respectively; z = 1.105, P = 0.2691). A total of 462 hymenoptera adults were reared from the sentinel plants. Four were identified as belonging to the family Braconidae, subfamily Microgastrinae. Of the remaining 458, a randomly selected sample of 159 individuals were identified to genus level, and 155 were morphologically compatible with Necremnus spp. Out of the Necremnus spp., 42 individuals were identified to the species level, with all but one being identified as N. tutae and the other as N. artynes.

Presence of natural enemies in flower patches

The total number of hymenopteran parasitoids and predatory mirid bugs recorded by beating each insectary plant is shown in Fig. 1. Results revealed that hymenopteran parasitoids were significantly more abundant on *A. millefolium* than on *S. alba*, whereas *F. esculentum* and *L. maritima* had intermediate values ($\chi^2 = 19.61$, df = 3, P = 0.0002). Regarding predatory mirid bugs, recorded populations were very low (< 0.10 individuals per beating sample), and no significant differences were found among the different plant species tested ($\chi^2 = 0.16$, df = 3, P = 0.9845).

Samplings conducted on the different plants also provided information regarding the potential of the insectary plants as a food resource for other biological control agents that, although not abundant in tomato crops and not T. absoluta control agents, may be relevant in other vegetable crops. In this vein, except hymenopteran parasitoids, the most abundant natural enemies recorded with the beating method were Orius spp. (30%) and ladybeetles (18%). In addition, some predatory thrips (5%) were collected. Altogether, 947 individuals were accounted for. No differences in numbers of natural enemies among the different insectary plant species were found (Table 3). On the contrary, the number of hoverfly landings recorded during the visual sampling was significantly higher on F. esculentum than in L. maritima and A. millefollium. Flowers of S. alba registered significantly less landings than any of the other three insectary plants.

Regarding phytophagous insects, no significant differences were found for thrips numbers among the four insectary plants tested. Significantly higher

Families	Treatment	z	Р	
	Far	Near		
Eulophidae	0.62 ± 0.15	0.75 ± 0.19	0.454	0.650
Braconidae	0.23 ± 0.11	0.08 ± 0.04	1.395	0.163
Pteromalidae	0.04 ± 0.03	0.04 ± 0.03	0.000	1.000
Ichneumonidae	0.04 ± 0.03	0.02 ± 0.02	0.566	0.571
Trichogrammatidae	0.00 ± 0.00	0.02 ± 0.02	0.002	0.998
Total of target families	0.94 ± 0.19	0.92 ± 0.22	0.225	0.822

Table 2 Number of hymenopteran individuals (mean \pm SE) of the different target families per yellow sticky trap placed near to and far from the insectary plant patches and results of the generalized linear mixed-effects model based on the Wald tests

The P value > 0.05 indicates no significant differences between values in a row. N = 48 traps in each treatment

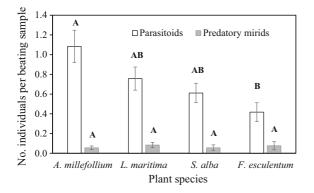


Fig. 1 Number of parasitoids and predatory mirids (mean \pm SE) per beating sample found in the different plants included in the study. Different capital letters above the bars indicate significant differences between the different plants for the two types of natural enemies separately (P < 0.05). The number of samples for each plant species is 144

number of aphids were recorded on *A. millefollium* and *F. esculentum* than on *L. maritima* and *S. alba*. By contrast, significantly more *Eurydema* spp. individuals were found on *L. maritima* and *S. alba* than on *A. millefollium* and *F. esculentum* (Table 4).

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Discussion

The main groups of natural enemies of T. absoluta described in the literature (Biondi et al. 2018), namely predatory mirid bugs and larval parasitoids, were present in the sampled tomato fields. The presence of mirids was foreseeable, since the IPM program used in the sampled fields was based in the conservation of generalist predatory mirids, mainly M. pygmaeus and N. tenuis that contributed to the control of several tomato pests (Arnó et al. 2018a). Soon after the invasion of this pest in Spain, Arnó et al. (2009) and Urbaneja et al. (2009) had already determined the great impact of these predators on T. absoluta eggs. More than three times as many N. tenuis than M. pygmaeus were recorded. The relative abundance of N. tenuis over M. pygmaeus has increased in recent years in the tomato crops in the area of study (RG, JA, JR unpublished data; Matas 2014), a phenomenon probably associated with climate change since N. *tenuis* is a clearly thermophilic species (Sánchez et al. 2009).

Table 3 Number of natural enemies (mean \pm SE) per beating sample and hoverfly landings recorded during the visual observations of the four insectary plants species included in the study, and results of the Wald χ^2 tests

Natural enemies	A. millefollium	F. esculentum	L. maritima	S. alba	χ^2	df	Р
Beating sampling							
Orius spp.	0.54 ± 0.13	0.44 ± 0.08	0.57 ± 0.10	0.37 ± 0.09	3.087	3	0.3784
Ladybeetles	0.32 ± 0.09	0.26 ± 0.06	0.19 ± 0.04	0.40 ± 0.14	1.613	3	0.6564
Predatory thrips	0.06 ± 0.02	0.09 ± 0.03	0.10 ± 0.03	0.04 ± 0.02	6.321	3	0.0969
Visual sampling							
Hoverflies	$1.06\pm0.25\mathrm{b}$	$5.58\pm0.85a$	$1.45\pm0.37\mathrm{b}$	$0.31\pm0.09\mathrm{c}$	73.172	3	< 0.0001

Within a row, different letters indicate significant differences between the different plants. If no letters are indicated in a row, the plant comparisons were not significant (P > 0.05). Number of samples for each plant is 144

Table 4 Number of phytophagous insects (mean \pm SE) per beating sample of the four insectary plants species included in the study, and results of the Wald χ^2 tests

Phytophagous	A. millefollium	L. maritima	S. alba	F. esculentum	χ^2	df	Р
Thrips	0.70 ± 0.17	0.65 ± 0.15	0.33 ± 0.10	0.73 ± 0.16	0.7528	3	0.8607
Aphids	$1.26\pm0.56a$	$0.08\pm0.04\mathrm{b}$	$0.09\pm0.05\mathrm{b}$	$1.49\pm0.48a$	11.478	3	0.0094
Eurydema spp.	$0.29\pm0.08b$	$2.67\pm0.33a$	$1.60\pm0.25a$	$0.49\pm0.10a$	11.636	3	0.0087

Within a row, different letters indicate significant differences between the different plants. If no letters are indicated in a row, the plant comparisons were not significant (P > 0.05). Number of samples for each plant is 144

The number of hymenopteran parasitoids was by far the highest among the potential natural enemies caught in the sticky traps. However, the percentage of individuals belonging to the target families that include known *T. absoluta* parasitoids (i.e., Eulophidae, Braconidae, Ichneumonidae, Pteromalidae, and Trichogrammatidae) represented less than 9% of total hymenopterans. Eulophidae were the most abundant followed by Braconidae, which is in accordance with Zappalà et al. (2013) and Biondi et al. (2018). These authors indicated that these two families were the best represented among parasitoids naturally attacking *T. absoluta* in the Mediterranean in terms of both the number of species and their geographical distribution.

Most of the parasitoids recruited with the sentinel plants were Necremnus spp., and, according to the sample of individuals identified to the species level, 98% were N. tutae. The higher prevalence of N. tutae in the sentinel plants was consistent with the higher prevalence of Eulophidae in the yellow sticky traps over other families with potential parasitoids of T. absoluta larvae. The net sleeve around the sentinel plants might have biased the results in regards to the parasitoid guild. Preliminary studies to verify the permeability of the net were only conducted with N. tutae. Therefore, it cannot be ruled out that parasitization by other bigger hymenopterans was prevented. Additionally, plants were taken to the field when second to third instar larvae were present, which are the preferred instars for N. tutae (Calvo et al. 2013). Therefore, the bias, if any, does not seem to have substantially modified the parasitoid species composition obtained with the sentinel plants. In fact, our results are consistent with previous studies that suggest that N. tutae is widely distributed in the Mediterranean (Ferracini et al. 2012; Zappalà et al. 2013; Gabarra et al. 2014; Gebiola et al. 2015; Biondi et al. 2018). This species was considered as a potential biological control agent to be released for pest control (Calvo et al. 2013, 2016; Chailleux et al. 2014; Bodino et al. 2016). Although it is not currently commercialized for inoculation, several authors have acknowledged its contribution to the conservation biological control of T. absoluta. In the present study, the parasitism rate in the sentinel plants was close to 20%. In the same area, a survey in tomato crops conducted in 2018 yielded a parasitism rate of approximately 14% (unpublished results), and parasitism rates were up to 73% in tomato greenhouses in the south of Spain (Crisol-Martínez and van der Blom 2019). Similarly, Abbes et al. (2014) reported an average parasitism rate of 26% in Tunisian open field crops using sentinel plants and between 11 and 15% when directly sampling tomato leaves from the crop.

The number of captures on yellow sticky traps placed close to the insectary plants were approximately the same than those placed far from the blooming plants, both in the case of parasitoids and predators. Likewise, the number of ectoparasitized T. absoluta larvae, and therefore the presence of T. absoluta parasitoids, were similar in sentinel plants located near to and far away from insectary plants placed in one side of the field. Both results indicated that in our experiment set-up, the distribution of natural enemies within the fields was uniform. This is probably because the size of the commercial fields in the area hampered the ability to differentiate between the effects of nearness to versus farness from the insectary plants due to the operational range of some parasitoids as has been suggested by Heimpel (2019).

Attractiveness of natural enemies on flowers is a first step to achieve biocontrol-related benefits from habitat manipulation. Among the insectary plants tested, A. millefolium was the most attractive for parasitoids, followed by F. esculentum and L. maritima, whereas S. alba registered the lowest populations. Attraction of parasitoids to A. millefolium has been previously reported in field experiments (Dib et al. 2012; El-Nabawy et al. 2015; Arnó et al. 2012; Gibson et al. 2019; Lundin et al. 2019). This statement disagrees with findings by Wäckers (2004), who in laboratory experiments demonstrated the repellency of A. millefolium flowers for two species of Braconidae and one Ichneumonidae. This repellent effect prevented these species from feeding nectar. In fact, Arnó et al (2018b) demonstrated that access to A. millefolium flowers did not provide any benefit to the survival and reproduction of three T. absoluta parasitoids, i.e., N. tutae, Stenomesius nr. japonicus Ashmed (Hymenoptera: Eulophidae), and Bracon nr. nigricans Szépligeti (Hymenoptera: Braconidae). Further studies will be necessary to clarify the divergent results from field and laboratory experiments regarding the role of A. millefolium as an insectary plant.

Sinapis alba, a spring oilseed crop typically harvested in early summer (Sáez-Bastante et al. 2016), registered the lower parasitoid abundance, which might be linked to the low flower presence

during the sampling period (July–September). However, the presence of this plant when in bloom might be of interest. Laboratory experiments have proven its positive effects on the egg load of *S*. nr. *japonicus* (Arnó et al. 2018b) and the longevity and fecundity of *Cotesia glomerata* (L.) (Hymenoptera: Braconidae) and *Diadegma semiclausum* Hellén (Hymenoptera: Ichneumonidae) (Winkler et al. 2009). Furthermore, field surveys have corroborated the value of this flowering plant to enhance parasitism of beetles (Manojlovic et al. 2000) and cereal aphids by *Aphidius* spp., especially when hosts and other food resources are limited (Damien et al. 2017).

During our samplings, *F. esculentum* and *L. maritima* had an intermediate position related to the attractiveness for parasitoids. These are two of the most commonly used insectary plants to enhance natural enemy populations, due to the good results on parasitoid fitness obtained when tested in laboratory experiments (Fiedler et al. 2008; Badenes-Pérez 2019). Arnó et al. (2018b) reported these two species to be the most suitable to enhance survival and reproduction of *T. absoluta* parasitoids. Similarly, Balzan and Wäckers (2013) reported survival increases for *N. artynes*.

None of the tested insectary plants was more attractive than others for the predatory mirid bugs considered in the survey, namely, M. pygameus and N. tenuis, and number of individuals was low (< 0.10individuals per sample) although the insectary plants were close to tomato fields were mirid populations were well established. By contrast, in one of the farms in 2015, C. officinalis yielded an average of 4.3 ± 0.7 adults of *M. pygmaeus* using the same beating method (unpublished results). This suggests that plants tested in our field experiments were much less attractive than the tomato crop for these two species of predatory mirids although scientific reports have suggested that their fitness benefits when feeding on pollen, nectar, or other sugary foods (Vandekerkhove and De Clercq 2010; Portillo et al. 2012; Put et al. 2012; Urbaneja-Bernat et al. 2015). Therefore, none of the four insectary plants examined in this study can be considered as a candidate to replace C. officinalis, but they could complement the floral margins of this plant to promote the presence of a more varied guild of natural enemies, in particular parasitoids.

Regarding other predators that are interesting biological control agents in vegetables other than

tomato and that were found in the insectary plants during the samplings, only hoverflies have shown preference for some species over others: namely, F. esculentum, which was the most preferred, and S. alba, which was the less preferred. Fagopyrum esculentum, and, to a lesser extent, A. millefolium, were ranked as suitable flowers to feed zoophagous hoverflies due to their nectar accessibility, which influences fitness and attractiveness (van Rijn and Wäckers 2016). On the other hand, L. maritima has been previously described as very attractive for adult hoverflies (reviewed by Badenes-Pérez 2019). The low number of visits to the yellow S. alba flowers was probably related to the poor blooming during the sampling period, which might have biased the preference among different insectary plant species as discussed by Ambrosino et al. (2006). Carreck and Williams (1997) reported S. alba being attractive to syrphid adults when present in flower mixtures. On the other hand, yellow flowers elicited landing more than other colors (Rodríguez-Gasol et al. 2019).

Insectary plants behaved differently as hosts of aphids and Eurydema spp., but not of thrips. Aphids were mostly abundant on A. millefollium and F. esculentum, and the pentatomid on the two Brasicaceae species. The presence of herbivores in insectary plants placed in the vicinity of crops may have an ambivalent role in biological control. On the one hand, they may act as refuges for pest to further colonize the crop, and on the other, as alternative prey or hosts to boost natural enemy populations. Furthermore, they may also act as trap plants that attract and retain target insects and reduce damage in the main crop. This might be the case for S. alba and L. maritima and Eurydema spp. in our study. Badenes-Pérez et al. (2017) found another Brasicaceae, Barbarea vulgaris, useful to reduce damage produced by Eurydema ornata L. (Hemiptera: Pentatomidae) in cauliflower. Therefore, their negative or positive role for the crops nearby deserve further investigation.

To summarize, results obtained in the present study indicate that 17 years after the invasion of *T. absoluta* in Northeast Spain, the eulophid *N. tutae* is the main species responsible for larval parasitism of *T. absoluta*, a phenomenon which has been recorded in other Mediterranean production areas (Gebiola et al. 2015). According to our results, the role of parasitoids belonging to other target families in the control of the pest was minimal: they were scarcely present in the fields and were not found parasitizing *T. absoluta* larvae, apart from a few Braconidae. Similarly, the low number of Trichogrammatidae captured in the yellow sticky traps is in accordance with Gabarra et al. (2014), which found egg rates of parasitism to be very low. Therefore, in the area of study, *T. absoluta* biological control is the result of egg predation by *M. pygmaeus* and *N. tenuis* and parasitation of larvae by *N. tutae*.

Moreover, the flowering plants used in the present study were attractive to several natural enemies that are of interest not only for tomatoes but also for other vegetable crops. Although A. millefollium was the most attractive for parasitoid wasps and F. esculentum for hoverflies, all of them were visited by these two groups of beneficials and other important predators (i.e., Orius spp., ladybeetles, and predatory thrips) in vegetable production systems characterized by the coexistence of several crops. Therefore, inclusion of these insectary plants in the C. officinalis margins that are already used in the area will surely contribute to increasing functional biodiversity in the farms and benefiting biological control of several pests in different crops. Considering the number of hymenopteran parasitoids in the insectary plants as a proxy for attractiveness to T. absoluta parasitoids, our results together with those of Arnó et al. (2018a), which reported benefits of these plant species in the fitness of T. absoluta larval parasitoids, suggest that the presence of these insectary plants might benefit the control of this pest. Therefore, it would be necessary to define methods of incorporating these plants into the horticultural agro-ecosystem in order to improve the biological control of pests.

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Authors contributions JA, RG, and JR conceived and designed the research. All authors conducted experiments and analyzed data. JA and JR wrote the manuscript. All authors read and approved the manuscript.

Availability of data and material The datasets generated during and/or analyzed during the current study are available from the corresponding author upon reasonable request.

Declarations

Conflict of interest The authors declare that they have no conflicts of interest.

Ethical approval This article does not contain any study with human participants or vertebrate animals.

References

- Abbes K, Biondi A, Zappalà L, Chermiti B (2014) Fortuitous parasitoids of the invasive tomato leafminer *Tuta absoluta* in Tunisia. Phytoparasitica 42:85–92
- Agustí N, Castañé C, Fraile I, Alomar O (2020) Development of a PCR-based method to monitor arthropod dispersal in agroecosystems: *Macrolophus pygmaeus* (Hemiptera: Miridae) from banker plants to tomato crops. Insect Sci 27:1125–1134
- Ambrosino MD, Luna JM, Jepson PC, Wratten SD (2006) Relative frequencies of visits to selected insectary plants by predatory hoverflies (Diptera: Syrphidae), other beneficial insects, and herbivores. Environ Entomol 35:394–400
- Aparicio Y, Riudavets J, Gabarra R, Agustí N, Rodríguez-Gasol N, Alins G, Blasco-Moreno A, Arnó J (2021) Can insectary plants enhance the presence of natural enemies of the Green Peach Aphid (Hemiptera: Aphididae) in Mediterranean Peach Orchards? J Econ Entomol 114:784–793
- Ardanuy A, Figueras M, Matas M, Madeira F, Arnó J, Alomar O, Albajes R, Gabarra R (2021) Banker plants and landscape composition influence colonisation precocity of tomato greenhouses by mirid predators. J Pest Sci. https:// doi.org/10.1007/s10340-021-01387-y
- Arnó J, Castañé C, Alomar O, Riudavets J, Agustí N, Gabarra R, Albajes R (2018a) Forty years of biological control in Mediterranean tomato greenhouses: the story of success. Isr J Entomol 48:209–226
- Arnó J, Gabarra R, Alomar O (2012) Hymenoptera abundance on candidate plants for conservation biological control. IOBC/WPRS Bull 75:13–16
- Arnó J, Oveja MF, Gabarra R (2018b) Selection of flowering plants to enhance the biological control of *Tuta absoluta* using parasitoids. Biol Control 122:41–50
- Arnó J, Sorribas R, Prat M, Matas M, Pozo C, Rodríguez D, Garreta A, Gómez A, Gabarra R (2009) *Tuta absoluta*, a new pest in IPM tomatoes in the northeast of Spain. IOBC/ WPRS Bull 9:203–208
- Askew RR (1968) Handbooks for the identification of British insects. Vol. VIII. Hymenoptera 2. Chalcidoidea section (b). Royal Entomological Society of London. London

- Badenes-Pérez FR (2019) Trap crops and insectary plants in the order Brassicales. Ann Entomol Soc Am 112:318–329
- Badenes-Pérez FR, Márquez BP, Petitpierre E (2017) Can flowering *Barbarea* spp. (Brassicaceae) be used simultaneously as a trap crop and in conservation biological control? J Pest Sci 90:623–633
- Balzan MV, Wäckers FL (2013) Flowers to selectively enhance the fitness of a host-feeding parasitoid: adult feeding by *Tuta absoluta* and its parasitoid *Necremnus artynes*. Biol Control 67:21–31
- Bates D, Maechler M, Bolker B, Walker S (2015) Fitting linear mixed-effects models using lme4. J Stat Softw 67:1–48
- Biondi A, Guedes RNC, Wan FH, Desneux N (2018) Ecology, worldwide spread, and management of the invasive South American tomato pinworm, *Tuta absoluta*: past, present, and future. Ann Rev Entomol 63:239–258
- Blaauw BR, Isaacs R (2012) Larger wildflower plantings increase natural enemy density, diversity, and biological control of sentinel prey, without increasing herbivore density. Ecol Entomol 37:386–394
- Bodino N, Ferracini C, Tavella L (2016) Is host selection influenced by natal and adult experience in the parasitoid *Necremnus tutae* (Hymenoptera: Eulophidae)? Anim Behav 112:221–228
- Calvo FJ, Soriano JD, Bolckmans K, Belda JE (2013) Host instar suitability and life-history parameters under different temperature regimes of *Necremnus artynes* on *Tuta absoluta* Biocontrol. Sci Technol 23:803–815
- Calvo FJ, Soriano JD, Stansly PA, Belda JE (2016) Can the parasitoid *Necremnus tutae* (Hymenoptera: Eulophidae) improve existing biological control of the tomato leafminer *Tuta aboluta* (Lepidoptera: Gelechiidae)? Bull Entomol Res 106:502–511
- Carreck NL, Williams IH (1997) Observations on two commercial flower mixtures as food sources for beneficial insects in the UK. J Agric Sci Camb 128:397–403
- Chailleux A, Desneux N, Arnó J, Gabarra R (2014) Biology of two key Palearctic larval ectoparasitoids when parasitizing the invasive pest *Tuta absoluta*. J Pest Sci 87:441–448
- Crisol-Martínez E, van der Blom J (2019) *Necremnus tutae* (Hymenoptera, Eulophidae) is widespread and efficiently controls *Tuta absoluta* in tomato greenhouses in SE Spain. IOBC/WPRS Bull 147:22–29
- Damien M, Le Lann C, Desneux N, Alford L, Al Hassana D, Georges R, van Baaren J (2017) Flowering cover crops in winter increase pest control but not trophic link diversity. Agric Ecosyst Environ 247:418–425
- de Campos MR, Monticelli LS, Béarez P, Amiens-Desneux E, Wang Y, Lavoir AV, Zappalà L, Biondi A, Desneux N (2020) Impact of a shared sugar food source on biological control of *Tuta absoluta* by the parasitoid *Necremnus tutae*. J Pest Sci 93:207–218
- Desneux N, Wajnberg E, Wyckhuys KAG, Burgio G, Arpaia S, Narváez-Vasquez C, González-Cabrera J, Catalán Ruescas D, Tabone E, Frandon J, Pizzol J, Poncet C, Cabello T, Urbaneja A (2010) Biological invasion of European tomato crops by *Tuta absoluta*: Ecology, geographic expansion and prospects for biological control. J Pest Sci 83:197–215
- Desneux N, Luna MG, Guillemaud T, Urbaneja A (2011) The invasive South American tomato pinworm, *Tuta absoluta*,

continues to spread in Afro-Eurasia and beyond: the new threat to tomato world production. J Pest Sci 84:403–408 Dib H. Libourel G. Warlon E (2012) Entomological and func-

- Dib H, Libourel G, Warlop F (2012) Entomological and functional role of floral strips in an organic apple orchard: Hymenopteran parasitoids as a case study. J Insect Conserv 16:315–318
- El-Nabawy EM, Tsuda K, Sakamaki Y (2015) Attractiveness of spiders and insect predators and parasitoids to flowering plants. Egypt J Biol Pest Cont 25:245–250
- Ferracini C, Ingegno BL, Navone P, Ferrari E, Mosti M, Tavella L, Alma A (2012) Adaptation of indigenous larval parasitoids to *Tuta absoluta* (Lepidoptera: Gelechiidae) in Italy. J Econ Entomol 105:1311–1319
- Fiedler AK, Landis DA, Wratten SD (2008) Maximizing ecosystem services from conservation biological control: the role of habitat management. Biol Control 45:254–271
- Fox J, Weisberg S (2019) An R companion to applied regression, 3rd edn. Sage, Thousand Oaks. https://socialsciences. mcmaster.ca/jfox/Books/Companion
- Gabarra R, Arnó J, Lara L, Verdú MJ, Ribes A, Beitia F, Urbaneja A, Téllez MM, Mollá O, Riudavets J (2014) Native parasitoids associated with *Tuta absoluta* in the tomato production areas of the Spanish Mediterranean Coast. BioControl 59:45–54
- Gebiola M, Bernardo U, Ribes A, Gibson GAP (2015) An integrative study of *Necremnus* Thomson (Hymenoptera: Eulophidae) associated with invasive pests in Europe and North America: Taxonomic and ecological implications. Zool J Linn Soc-Lond 173:352–423
- Gibson DR, Rowe K, Isaacs R, Landis DA (2019) Screening drought-tolerant native plants for attractiveness to arthropod natural enemies in the U.S. Great Lakes Region. Environ Entomol 48:1469–1480
- Grissell EE, Schauff ME (1990) A handbook of the families of Nearctic Chalcidoidea (Hymenoptera). Entomol. Soc. Wash, Washington, DC
- Han P, Bayram Y, Shaltiel-Harpaz L, Sohrabi F, Saji A, Tair Esenali U, Jalilov A, Ali A, Shashank PR, Ismoilov K, Lu ZZ, Wang S, Zhang GF, Wan FH, Biondi A, Desneux N (2019) *Tuta absoluta* continues to disperse in Asia: Damage, ongoing management and future challenges. J Pest Sci 92:1317–1327
- Hanson PE, Gauld ID (2006) Hymenoptera de la región Neotropical. Memoirs of the American Entomological Institute 77. The American Entomological Institute. Gainesville
- Heimpel GE (2019) Linking parasitoid nectar feeding and dispersal in conservation biological control. Biol Control 132:36–41
- Ingegno BL, Messelink GJ, Bodino N, Iliadou A, Driss L, Woelke JB, Leman A, Tavella L (2019) Functional response of the mirid predators *Dicyphus bolivari* and *Dicyphus errans* and their efficacy as biological control agents of *Tuta absoluta* on tomato. J Pest Sci 92:1457–1466
- Lundin O, Ward KL, Williams NM (2019) Identifying native plants for coordinated habitat management of arthropod pollinators, herbivores and natural enemies. J Appl Ecol 56:665–676
- Manojlovic B, Zabel A, Kostic M, Stankovic S (2000) Effect of nutrition of parasites with nectar of melliferous plants on

parasitism of the elm bark beetles (Col., Scolytidae). J Appl Entomol 124:155–161

- Mansour R, Brévault T, Chailleux A, Cherif A, Grissa-Lebdi K, Haddi K, Mohamed SA, Nofemela RS, Oke A, Sylla S, Tonnang HE (2018) Occurrence, biology, natural enemies and management of *Tuta absoluta* in Africa. Entomol Gen 38:83–112
- Matas M (2014) Control biológico en el tomate en el Maresme. Conservación de enemigos naturales y gestión de infraestructuras ecológicas. Phytoma-España 262:56
- Portillo N, Alomar O, Wäckers FL (2012) Nectarivory by the plant-tissue feeding predator *Macrolophus pygmaeus* Rambur (Heteroptera: Miridae): nutritional redundancy or nutritional benefit? J Insect Physiol 58:397–401
- Put K, Bollens T, Wäckers FL, Pekas A (2012) Type and spatial distribution of food supplements impact population development and dispersal of the omnivore predator *Macrolophus pygmaeus* (Rambur) (Hemiptera: Miridae). Biol Control 63:172–180
- R Core Team (2020) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna. https://www.r-project.org/
- Riudavets J, Moerman E, Vila E (2020) Implementation of integrated pest and disease management in greenhouses: from research to the consumer. In: Gullino ML, Albajes R, Nicot P (eds) Integrated pest and disease management in greenhouse crops. Plant pathology in the 21st Century, vol 9. Springer, Cham, pp 457–485
- Rodríguez-Gasol N, Avilla J, Aparicio Y, Arnó J, Gabarra R, Riudavets J, Alegre S, Lordán J, Alins G (2019) The contribution of surrounding margins in the promotion of natural enemies in Mediterranean apple orchards. Insects 10:148
- Russell Lenth (2020) emmeans: estimated marginal means, aka least-squares means. R package version 1.5.1. https://cran. r-project.org/web/packages/emmeans/index.html.
- Sáez-Bastante J, Fernández-García P, Saavedra M, López-Bellido L, Dorado MP, Pinzi S (2016) Evaluation of *Sinapis alba* as feedstock for biodiesel production in Mediterranean climate. Fuel 184:656–664
- Salas Gervassio NG, Aquino D, Vallina C, Biondi A, Luna MG (2019) A re-examination of *Tuta absoluta* parasitoids in South America for optimized biological control. J Pest Sci 92:1343–1357
- Sanchez JA, Lacasa A, Arnó J, Castañé C, Alomar O (2009) Life history parameters for *Nesidiocoris tenuis* (Reuter) (Het., Miridae) under different temperature regimes. J Appl Entomol 133:125–132
- Urbaneja A, Montón H, Mollá O (2009) Suitability of the tomato borer *Tuta absoluta* as prey for *Macrolophus pygmaeus* and *Nesidiocoris tenuis*. J Appl Entomol 133:292–296
- Urbaneja-Bernat P, Mollá O, Alonso M, Bolkcmans K, Urbaneja A, Tena A (2015) Sugars as complementary alternative food for the establishment of *Nesidiocoris tenuis* in greenhouse tomato. J Appl Entomol 139:161–167
- Urbaneja A, González-Cabrera J, Arnó J, Gabarra R (2012) Prospects for the biological control of *Tuta absoluta* in tomatoes of the Mediterranean basin. Pest Manag Sci 68:1215–1222
- van Rijn PCJ, Wäckers FL (2016) Nectar accessibility determines fitness, flower choice and abundance of hoverflies that provide natural pest control. J Appl Ecol 53:925–933

- Vandekerkhove B, De Clercq P (2010) Pollen as an alternative or supplementary food for the mirid predator *Macrolophus pygmaeus*. Biol Control 53:238–242
- Venables WN, Ripley BD (2002) Modern applied statistics with S, 4th edn. Springer, New York
- Wäckers FL (2004) Assessing the suitability of flowering herbs as parasitoid food sources: flower attractiveness and nectar accessibility. Biol Control 29:307–314
- Wäckers FL, van Rijn PCJ (2005) Food for protection: an introduction. In: Wäckers FL, van Rijn PCJ, Bruin J (eds) Plant-provided food for carnivorous insects: a protective mutualism and its applications. Cambridge University Press, Cambridge, pp 1–14
- Wickham H (2016) ggplot2: elegant graphics for data analysis. Springer, New York
- Winkler K, Wäckers FL, Kaufman LV, Larraz V, van Lenteren JC (2009) Nectar exploitation by herbivores and their parasitoids is a function of flower species and relative humidity. Biol Control 50:299–306
- Zappalà L, Biondi A, Alma A, Al-Jboory IJ, Arnó J, Bayram A, Chailleux A, El-Arnaouty A, Gerling D, Guenaoui Y, Shaltiel-Harpaz L, Siscaro G, Stavrinides M, Tavella L, Vercher-Aznar R, Urbaneja A, Desneux N (2013) Natural enemies of the South American moth, *Tuta absoluta*, in Europe, North Africa and Middle East, and their potential use in pest control strategies. J Pest Sci 86:635–647

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