

# *In Vitro* Study of Neuroprotective Properties of GK-2, a New Original Nerve Growth Factor Mimetic

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New nerve growth factor (NGF) mimetic GK-2, a substituted dimeric dipeptide, in a concentration of up to  $10^{-9}$  M produced a protective effect on the culture of immortalized mouse hippocampal neurons (line HT-22) after addition of  $H_2O_2$  and glutamate. GK-2 in a concentration of  $10^{-8}$  M protected rat PC-12 pheochromocytoma cells from the neurotoxin MPTP. The neuroprotective effect of this peptide on the model of oxidative stress was also observed in the primary culture of embryonic rat hippocampal neurons.

**Key Words:** neuroprotection; GK-2; oxidative stress; glutamate toxicity; MPTP

Over the past decades, numerous studies have shown that neurotrophins NGF and BDNF are involved in the pathogenesis of neurodegenerative diseases [5,13]. The neuroprotective effect of NGF and BDNF was confirmed by pharmacological researches. The use of NGF and BDNF in pharmacotherapy is limited because of difficulties in crossing the blood-brain barrier, influence of endogenous peptidases, and other problems associated with systemic treatment with these drugs [2].

Recent studies at the V. V. Zakusov Institute of Pharmacology were directed towards the construction of a small molecule, which possesses the properties of NGF. A low-molecular-weight mimetic of the 4th loop of NGF (GK-2) was synthesized [1]. GK-2 is a substituted dimeric dipeptide, bis-(N-succinyl-glutamyl-lysine) hexamethylenediamine.

This work was designed to study the *in vitro* neuroprotective properties of GK-2 on the models of oxidative stress (OS), glutamate toxicity, and MPTP-induced neuronal damage.

## MATERIALS AND METHODS

Experiments were performed on the culture of immortalized mouse hippocampal neurons (line HT-22), primary culture of rat hippocampal neurons (HN), and

dopamine-positive PC-12 cells of rat pheochromocytoma (tumor of the adrenal cortex).

Primary culture of HN was obtained from 18-day-old embryos of MR (Maudsley reactive) rats. Dissection of the brain and isolation of midbrain neurons were performed in Hank's medium (BioloT) [6]. The isolated HN were put in centrifuge tubes with DMEM/F-12 medium (ICN), resuspended, and centrifuged twice at 1500 rpm for 10 min. The cells were repeatedly resuspended, counted, and inoculated in 48-well culture plates (density 350,000-400,000 cells per well) with the same medium. Experiments were conducted on the 6th day after cell inoculation. HT-22 cells were inoculated (3500 cells per well) in 96-well plates with DMEM medium (ICN) containing 2 mM glutamine (ICN) and 5% FBS (ICN). Incubation was performed until the formation of monolayer. The cells were cultured at 37°C and 5%  $CO_2$ .

OS was induced by  $H_2O_2$  in a final concentration of 1.5 mM [7]. HT-22 cells were incubated with  $H_2O_2$  at 37°C and 5%  $CO_2$  for 30 min. The culture medium with  $H_2O_2$  was replaced for normal medium. Cell viability was estimated after 4 h.

For modeling MPTP-induced damage in culture of PC12 cells, neurotoxin MPTP (Sigma) in a final concentration of 1 mM was added to the cell medium containing 1% FBS (24 h after passage of cells) [12]. The peptide was put to the cell medium simultaneously or 24 h before addition of MPTP.

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Glutamate toxicity was induced by adding glutamate (final concentration 5 mM) to the culture medium. After 24-h incubation at 37°C and 5% CO<sub>2</sub>, the medium was replaced with normal medium. Cell viability was estimated after 24 h [9,14]. The peptide was added 24 h before damage or immediately after washing out glutamic acid.

Cell viability was evaluated in the test with bromide 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium (MTT, Sigma) [7]. Optical density was measured on a Multiscan EX spectrophotometer (Thermo).

For evaluation of the neuroprotective effect of GK-2, the peptide was added 24 h before or immediately after the exposure to adverse factors. NGF (Sigma) in a final concentration of 100 ng/ml was used as the positive control.

The significance of differences between the treatment and control groups was estimated by Student's *t* test.

## RESULTS

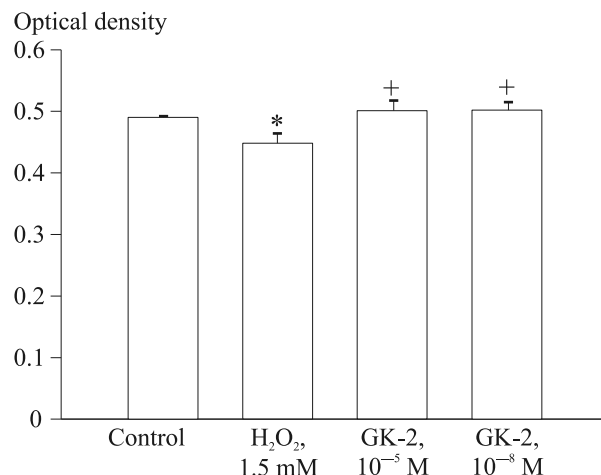
Addition of GK-2 after treatment with H<sub>2</sub>O<sub>2</sub> prevented death of immortalized mouse HT-22 hippocampal neurons (Table 1). NGF has a similar effect, which is probably related to modulation of antioxidant enzyme activity in cells [15].

Addition of GK-2 (final concentration up to 10<sup>-9</sup> M) 24 h before treatment with H<sub>2</sub>O<sub>2</sub> produced a neuroprotective effect on HT-22 cells (Table 1). The neuroprotective effect is also observed in experiments with addition of NGF 24 h before OS, which results from an increase in the synthesis of neuroprotective heat-

**TABLE 1.** Effect of GK-2 on Viability of Mouse HT-22 HN after OS (*M±m*, *n*=12)

Group	Optical density	
	24 h before OS	immediately after addition of H <sub>2</sub> O <sub>2</sub>
Control	0.36±0.07	0.47±0.07
H <sub>2</sub> O <sub>2</sub> , 1.5 mM	0.24±0.02*	0.35±0.05*
NGF, 100 ng/ml	0.34±0.07 <sup>+</sup>	0.460±0.047 <sup>+</sup>
GK-2, 10 <sup>-5</sup> M	0.32±0.07 <sup>+</sup>	0.44±0.05 <sup>+</sup>
GK-2, 10 <sup>-6</sup> M	—	0.37±0.06
GK-2, 10 <sup>-7</sup> M	—	0.39±0.07
GK-2, 10 <sup>-8</sup> M	0.35±0.07 <sup>+</sup>	0.47±0.07 <sup>+</sup>
GK-2, 10 <sup>-9</sup> M	—	0.41±0.06 <sup>+</sup>
GK-2, 10 <sup>-10</sup> M	—	0.38±0.06

**Note.** \**p*≤0.05 compared to the control; <sup>+</sup>*p*≤0.05 compared to H<sub>2</sub>O<sub>2</sub>.



**Fig. 1.** Effect of GK-2 on viability of HN from 18-day-old rat embryos (*n*=12) after OS. The peptide is added 24 h before treatment with H<sub>2</sub>O<sub>2</sub>. \**p*≤0.05 compared to the control; <sup>+</sup>*p*≤0.05 compared to H<sub>2</sub>O<sub>2</sub>.

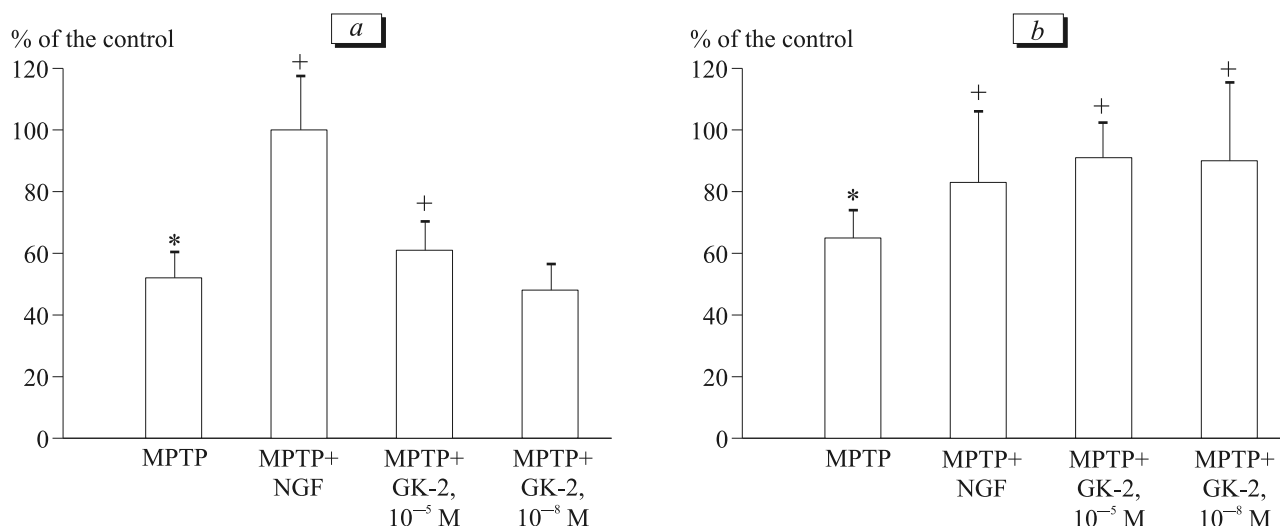
shock proteins [4,10].

Experiments on the model of OS showed that GK-2 in the specified concentrations produced a neuroprotective effect not only on immortalized neurons, but also on the primary culture of embryonic HN (Fig. 1). The protective effect of GK-2 was also observed upon exposure to a variety of adverse factors.

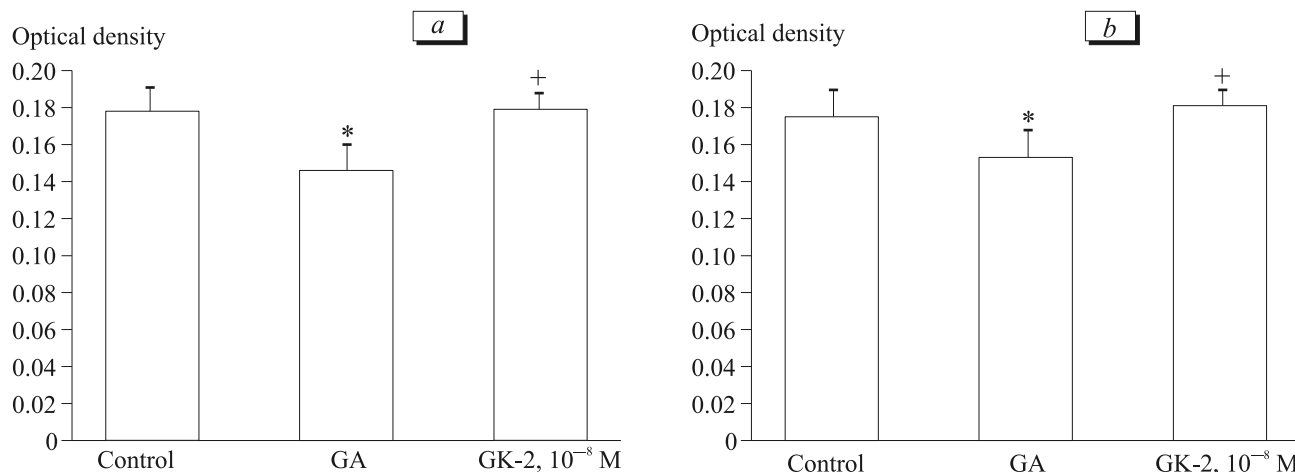
The neurotoxin MPTP is used for modeling of the cellular stage of Parkinson's disease. This agent causes a selective damage to dopaminergic neurons due to mitochondrial dysfunction and impairment of energy metabolism in the neuron [11]. In these experiments, GK-2 was added simultaneously or 24 h before treatment with 1 mM MPTP [12]. GK-2 exhibited protective properties under these conditions (Fig. 2).

The neuroprotective effect of KG-2 was also revealed under conditions of glutamate toxicity. The peptide was added 24 h before or immediately after washing out glutamic acid. GK-2 in the test concentration had a strong protective effect on cells under both conditions of treatment with glutamic acid (Fig. 3).

We conclude that GK-2 in low concentrations produces a neuroprotective effect on the models of OS, glutamate toxicity, and MPTP-induced cell damage. Activity of GK-2 is similar to that of NGF. GK-2 is effective in a concentration of 10<sup>-9</sup> M. It should be emphasized that the most potent agonist of non-peptide TrkA receptors, Gambogic amide [8], only in concentrations of up to 0.5×10<sup>-6</sup> M protects the culture of T17 and SN56 cells from apoptotic death. *In vitro* study showed that L1L4 (strong peptide agonist of NGF) only in a concentration of 5×10<sup>-6</sup> M exhibits NGF-like properties [3]. L1L4 is a 14-membered bicyclic peptide with a molecular weight >2000 Da. The dimeric dipeptide GK-2 has a molecular weight of 830 Da. A smaller number and variability of peptide



**Fig. 2.** Effect of GK-2 on PC12 cell viability during MPTP-induced damage. (a) Addition of GK-2 simultaneously with MPTP; (b) addition of GK-2 24 h before treatment with MPTP. \* $p \leq 0.05$  compared to the control (100%); † $p \leq 0.05$  compared to MPTP.



**Fig. 3.** Effect of GK-2 on the viability of mouse HT-22 HN under conditions of glutamate toxicity. (a) Addition of the peptide 24 h before treatment with glutamic acid (GA); (b) addition of the peptide immediately after washing out GA. \* $p \leq 0.05$  compared to the control; † $p \leq 0.05$  compared to glutamic acid.

bonds in GK-2 determine higher resistance of this agent to peptidases. The ability of GK-2 to cross the blood-brain barrier is associated with its relatively low molecular weight.

The properties of GK-2 should be evaluated in *in vitro* and *in vivo* experiments.

## REFERENCES

- S. B. Seredenin and T. A. Gudasheva, Invention Application No. 2009105176, *Byull. Izobr.*, No. 24 (2010).
- C. A. Altar, M. P. Vawter, and S. D. Ginsberg, *Neuropsychopharmacology*, **34**, No. 1, 18-54 (2009).
- A. M. Colangelo, M. R. Bianco, L. Vitagliano, *et al.*, *J. Neurosci.*, **28**, No. 11, 2698-2709 (2008).
- N. D'Ambrosi, B. Murra, F. Vacca, and C. Volonte, *Prog. Brain Res.*, **4**, 93-100 (2004).
- D. Dawbarn and S. J. Allen, *Neuropathol. Appl. Neurobiol.*, **29**, No. 3, 211-230 (2003).
- T. L. Fletcher, P. De Camilli, and G. Banker, *J. Neurosci.*, **14**, No. 11, Pt. 1, 6695-6706 (1994).
- G. R. Jackson, K. Werrbach-Perez, E. L. Ezell, *et al.*, *Brain Res.*, **592**, Nos. 1-2, 239-248 (1992).
- S. W. Jang, M. Okada, I. Sayeed, *et al.*, *Proc. Natl. Acad. Sci. USA*, **104**, No. 41, 16,329-16,334 (2007).
- J. F. Kerr, G. C. Gobé, C. M. Winterford, and B. V. Harmon, *Methods Cell Biol.*, **46**, 1-27 (1995).
- H. Liu, R. Nowak, W. Chao, and K. D. Bloch, *J. Neurochem.*, **86**, No. 6, 1553-1563 (2003).
- N. Schmidt and B. Ferger, *J. Neural. Transm.*, **108**, No. 11, 1263-1282 (2001).
- K. Shimoke and H. Chiba, *J. Neurosci. Res.*, **63**, No. 5, 402-409 (2001).
- G. J. Siegel and N. B. Chauhan, *Brain Res. Brain Res. Rev.*, **33**, Nos. 2-3, 199-227 (2000).
- S. Tan, M. Wood, and P. Maher, *J. Neurochem.*, **71**, No. 1, 95-105 (1998).
- Z. Zhou, H. Chen, K. Zhang, *et al.*, *J. Basic Clin. Physiol. Pharmacol.*, **14**, No. 3, 217-224 (2003).