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Tumor vasculogenic mimicry predicts poor prognosis in cancer patients: a meta-analysis

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Abstract

Background Vasculogenic mimicry (VM) is the formation of vascular channels by tumor cells or tumor cellderived, trans-differentiated cells in highly aggressive, solid tumors. However, the disease features and prognostic value of VM for overall survival of cancer patients remain controversial.

Method To systematically investigate the roles of VM in cancer progression and its prognostic values, we performed a meta-analysis based on 36 studies (33 eligible articles) including 3609 patients. The pooled hazard ratios (HRs) with 95 % confidence intervals (95 % CIs) were used to assess the relationship between VM and overall survival in cancer patients.

Results Vasculogenic mimicry was significantly associated with cancer differentiation, lymph node metastasis,

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distant metastasis, and TNM stage. The prognostic value of VM was significant in overall survival (HR 2.16; 95 % CI 1.98–2.38; $P \lt 0.001$). Analyses stratified by confounders, such as cancer type, ethnicity, VM detection methods, sample size, and Newcastle–Ottawa quality score, found similar significant results.

Conclusions The presence of VM predicts poorer survival outcomes in cancer patients.

Keywords Vasculogenic mimicry - Solid tumor - Angiogenesis - Survival - Meta-analysis

Introduction

The tumor vasculature system supplies blood for tumor growth and hematogenous dissemination, which have long been regarded as hallmarks of tumorigenesis [[1,](#page-7-0) [2\]](#page-7-0). The well-known theory of vascularization in tumors is a complex process that involves angiogenesis (sprouting new vessels from existing vessels), vasculogenesis (recruiting circulating endothelial progenitor cells), co-option (hijacking of the existing vasculature) [\[3–6](#page-7-0)], and tumor cellderived vasculature development known as vasculogenic mimicry (VM) $[6-9]$.

Tumor VM refers to the plasticity of highly aggressive cancer cells that imitate endothelial cells and form vessellike structure with the characteristic of positive PAS and negative endothelial cell markers, including CD31 or CD34 [\[8](#page-7-0)]. VM has been identified in many malignancies, such as melanoma [[7,](#page-7-0) [10](#page-7-0)–[13\]](#page-8-0), gastric cancer [\[14–17\]](#page-8-0), sarcoma [[10,](#page-7-0) [18](#page-8-0)], ovarian tumor [\[19–21](#page-8-0)], hepatocellular carcinoma [[22,](#page-8-0) [23](#page-8-0)], breast cancer [[24,](#page-8-0) [25](#page-8-0)], head and neck cancer [[26,](#page-8-0) [27](#page-8-0)], colorectal cancer [[28,](#page-8-0) [29\]](#page-8-0), glioma [[30–33\]](#page-8-0), lung cancer [[34,](#page-8-0) [35](#page-8-0)], testicular germ cell malignant tumors [[36\]](#page-8-0), gallbladder cancer [\[37](#page-8-0)], prostate cancer [\[38](#page-8-0)], esophageal cancer [\[39](#page-8-0)], renal carcinoma [[40\]](#page-8-0), and osteosarcoma [[41\]](#page-8-0).

Vasculogenic mimicry may be important in the biological behavior of various tumors because it can establish an adequate vascular supply to enhance tumor growth and metastasis [[8\]](#page-7-0). Growing evidence supports the notion of VM as a prognostic factor for poor clinical outcomes in various types of cancer, such as melanoma $[7, 10, 42]$ $[7, 10, 42]$ $[7, 10, 42]$ $[7, 10, 42]$ $[7, 10, 42]$ $[7, 10, 42]$, sarcoma $[10]$ $[10]$, ovarian tumor [[20,](#page-8-0) [21](#page-8-0)], hepatocellular carcinoma [[22,](#page-8-0) [23](#page-8-0)], and breast cancer [\[24](#page-8-0), [25\]](#page-8-0). Nonetheless, some of these studies have not confirmed the association between VM and cancer survival [\[10](#page-7-0), [12,](#page-8-0) [18](#page-8-0), [19,](#page-8-0) [38,](#page-8-0) [43](#page-8-0)]. As a result, the prognostic value of VM in tumors is still inconclusive.

Therefore, we conducted a meta-analysis to determine the prognostic value of VM on overall survival in cancer patients.

Methods

Search strategy

This meta-analysis was conducted according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement [\[44–46](#page-8-0)]. Relevant articles were identified from PubMed, EMBASE, and the Web of Science databases with the following keywords: "cancer,"

''tumor,'' ''neoplasm,'' ''vasculogenic mimicry,'' ''vascular mimicry," "VM," "tumor cell-lined vessels," "tumor derived endothelial cells," "prognosis," "survival," and ''outcome.'' The last search was performed up to March 10, 2015, without restriction on language. All relevant publications in reference lists were also searched manually to find the additional eligible articles.

Selection criteria

To be included, studies had to have met several criteria: (1) the diagnosis of cancer must have been confirmed histologically or pathologically; (2) the association between VM and overall survival (OS) or disease features, such as tumor differentiation, TNM stages, lymph node metastasis, and distant metastasis, had to be evaluated; and (3) data had to be sufficient to estimate hazard ratios (HRs) for overall survival and risk ratios (RRs) for disease features. In case of duplicated articles $[10, 47]$ $[10, 47]$ $[10, 47]$, only the most complete ones were included [\[10](#page-7-0)]. Narrative reviews, abstracts, letters, comments, editorials, and case reports were excluded.

Data extraction

Two investigators (JP Yang and YD Liao) separately extracted the data on authors, year of publication, races

Fig. 1 Flow chart of study selection in the meta-analysis of the effects of vasculogenic mimicry on tumor features and survival in cancer patients

Table 1 Summary of 36 studies (from 33 articles) included in a meta-analysis of the association of vasculogenic mimicry with disease characteristics and overall survival in cancer patients

References	Population	Tumor type	VM assay methods	Cases	No. of cases		NOS score	Quality ^a
					$VM+$	$VM-$		
1. Maniotis [7]	Non-Asian	Melanoma	PAS	234	106	128	$\overline{2}$	Low
2. Shirakawa [24]	Asian	Breast cancer	PAS	331	26	305	3	Low
3. Sun $[10]$	Asian	Sarcomas	CD31/PAS	37	10	27	3	Low
4. Sun $[10]$	Asian	Sarcomas	CD31/PAS	69	13	56	5	High
5. Sun [10]	Asian	Sarcomas	CD31/PAS	81	11	70	$\overline{4}$	Low
6. Sun $[10]$	Asian	Melanoma	CD31/PAS	190	10	180	3	Low
7. Sun [22]	Asian	HCC	PAS	99	12	87	$\overline{4}$	Low
8. Sun [14]	Asian	Gastric cancer	CD31/PAS	84	21	63	3	Low
9. Liu [26]	Asian	HNC	Pan-cytokeratin/CD34	112	41	71	$\overline{2}$	Low
10. Hillen [43]	Non-Asian	Melanoma	PAS	58	22	36	2	Low
11. Gao [19]	Asian	Ovarian tumor	CD31/PAS	84	36	48	2	Low
12. Zhang	Asian	Melanoma	PAS	124	54	70	3	Low
$[11]$								
13. Baeten [28]	Non-Asian	Colorectal cancer	PAS	117	23	94	\overline{c}	Low
14. Wang [18]	Asian	Sarcomas	CD34/PAS	45	25	20	$\overline{2}$	Low
15. Wang [36]	Asian	TGCMT	CD34/PAS	40	22	18	$\overline{4}$	Low
16. Li $[15]$	Asian	Gastric cancer	CD31/PAS	173	40	133	6	High
17. Wang [27]	Asian	HNC	CD31/PAS	203	44	159	$\overline{7}$	High
18. Liu [23]	Asian	HCC	CD34/PAS	151	31	120	$\overline{4}$	Low
19. Liu [30]	Asian	Glioma	CD34/PAS	101	13	88	5	High
20. Sun [37]	Asian	Gallbladder cancer	CD31/PAS	71	18	53	6	High
21. Liu [38]	Asian	Prostate cancer	CD31/PAS	96	24	72	6	High
22. Liu [29]	Asian	Colorectal cancer	CD34/PAS	203	39	164	$\overline{4}$	Low
23. Van Beurden [12]	Non-Asian	Melanoma	PAS	123	42	81	5	High
24. Wu [34]	Asian	Lung cancer	CD34/PAS	305	109	196	6	High
25. Yu [20]	Asian	Ovarian tumor	CD31/PAS	87	50	37	5	High
26. Chai [39]	Asian	Esophageal cancer	CD34/PAS	188	78	110	τ	High
27. Wang [31]	Asian	Glioma	CD34/PAS	86	23	63	6	High
28. Zhang [42]	Asian	HNC	CD34/PAS	42	18	24	\overline{c}	Low
29. Zhang [40]	Asian	Renal carcinoma	CD34/PAS	110	30	80	τ	High
30. Liu [25]	Asian	Breast cancer	CD31/PAS	90	26	64	6	High
31. Ren [41]	Asian	Osteosarcoma	CD34/PAS	66	15	51	6	High
32. Qi [57]	Asian	Gastric cancer	CD31/PAS	60	19	41	6	High
33. Song [16]	Asian	Gastric cancer	CD31/PAS	60	19	41	6	High
34. Sun [21]	Asian	Ovarian tumor	CD34/PAS	53	19	34	6	High
35. Li [35]	Asian	Lung cancer	CD34/PAS	51	17	34	3	Low
36. Zhou [17]	Asian	Gastric cancer	CD34/PAS	261	70	191	τ	High

VM+ patients with positive vasculogenic mimicry, VM- patients without vasculogenic mimicry, NOS score score of Newcastle-Ottawa scale, PAS periodic acid–Schiff staining, HCC hepatocellular carcinoma, HNC head and neck cancer, TGCMT testicular germ cell malignant tumors ^a Low-quality studies had NOS scores of zero to 4; high-quality studies had scores of 5–9

(Asian or non-Asian), tumor types, VM assay methods, disease information (including cancer differentiation, TNM stages, lymph node metastasis, and distant metastasis), sample sizes, and number of VM-positive or VM-negative patients. HRs with 95 % CIs were extracted from multivariable analyses. When HRs and 95 % CIs could not been directly extracted from the original studies, they were calculated using the method of Tierney et al. [\[45](#page-8-0)].

Two investigators assessed the quality of included studies using the Newcastle–Ottawa scale [\[48\]](#page-8-0). This scale consists of eight items assessing three aspects of a study: patient selection, comparability of study groups, and ascertainment of outcome. Each item could be awarded 1 point except for the item on comparability, which allowed 2 points. Studies with scores of five to nine were considered high quality; those with scores of zero to four were considered low quality.

Tumor differentiation grade was categorized as well (grade 1) or moderately/poorly differentiated (grades 2 and 3). TNM stage was dichotomized as I and II or III and IV. Lymph node metastasis and distant metastasis were both scored as positive or negative, respectively. Discrepancies were resolved by consensus.

Statistical methods

Hazard ratios (HRs) and 95 % CIs were used to evaluate the impact of VM on overall survival [[49,](#page-8-0) [50\]](#page-8-0). In addition, risk ratios were used to summarize the association between VM and disease features: TNM stage, differentiation, lymphatic metastasis, and distant metastasis [[49,](#page-8-0) [50\]](#page-8-0). The heterogeneity of pooled HRs and RRs was estimated using the Chi-square-based Cochrane's Q test ($P < 0.10$ was considered significant) and the I^2 index [\[51\]](#page-9-0). When heterogeneity was not so severe (l^2 values <50 %), a fixed-effect model was used. Otherwise, the random-effect model was applied to pooled data. A Galbraith plot was constructed to identify the main sources of heterogeneity [\[52\]](#page-9-0).

Sensitivity analysis that assesses the stability of results was conducted by sequentially removing each individual study and recalculating the results [\[53](#page-9-0)]. Cumulative meta-analysis was conducted by publication times [\[54](#page-9-0)]. Publication bias was evaluated by the Begg's funnel plot and the Egger's test [\[55,](#page-9-0) [56\]](#page-9-0).

All data were analyzed with the STATA software (version 12.0; STATA Corporation, College Station, TX). Unless otherwise stated, all P values were two-tailed and alpha was 0.05.

Results

Study characteristics

Of the 1056 articles identified in the literature search, 36 studies (described in 33 articles) representing 3609 patients were included in the meta-analysis (Fig. [1\)](#page-1-0) [[7,](#page-7-0) [10](#page-7-0), [12,](#page-8-0) [14](#page-8-0)– [31](#page-8-0), [34–43,](#page-8-0) [57\]](#page-9-0). The most common cancers studied were melanoma and gastric cancers (Table [1\)](#page-2-0). Vascular mimicry was detected in 8–56 % of patients. All but four studies were of Asian populations. Methods for detecting VM were PAS ($n = 7$), CD31/PAS ($n = 14$), CD34/PAS ($n = 14$), and pan-cytokeratin/CD34 $(n = 1)$ staining. Of the 36 studies, 18 (50 %) were high-quality studies (Table [1](#page-2-0)) [[12,](#page-8-0) [15–17](#page-8-0), [20](#page-8-0), [25,](#page-8-0) [27,](#page-8-0) [30,](#page-8-0) [31](#page-8-0), [34](#page-8-0), [37–41,](#page-8-0) [57\]](#page-9-0). Hazards ratios with 95 % CIs were extracted directly from 13 studies [[14,](#page-8-0) [16](#page-8-0), [17](#page-8-0), [20](#page-8-0), [23](#page-8-0), [25](#page-8-0), [34](#page-8-0), [36](#page-8-0), [37](#page-8-0), [40](#page-8-0), [41](#page-8-0), [57](#page-9-0)] and calculated for the remaining 23.

Disease features associated with vascular mimicry

Several studies evaluated associations between VM and cancer differentiation status (14 studies [[15,](#page-8-0) [17](#page-8-0), [19](#page-8-0), [21,](#page-8-0) [23,](#page-8-0) [25](#page-8-0), [27](#page-8-0), [29](#page-8-0), [30](#page-8-0), [34](#page-8-0), [37,](#page-8-0) [39,](#page-8-0) [40,](#page-8-0) [42\]](#page-8-0); 2032 patients), lymph metastasis (10 studies [[15,](#page-8-0) [17,](#page-8-0) [24,](#page-8-0) [25,](#page-8-0) [27](#page-8-0), [34](#page-8-0), [35](#page-8-0), [37–39](#page-8-0)]; 1735 patients), distant metastasis (13 studies [[14,](#page-8-0) [15](#page-8-0), [18,](#page-8-0) [19](#page-8-0), [21,](#page-8-0) [22,](#page-8-0) [24,](#page-8-0) [27,](#page-8-0) [36–38](#page-8-0), [40](#page-8-0), [42](#page-8-0)]; 1616 patients), and TNM stage (13 studies [\[15](#page-8-0), [17,](#page-8-0) [19,](#page-8-0) [21](#page-8-0), [23](#page-8-0), [25,](#page-8-0) [27,](#page-8-0) [29](#page-8-0), [34](#page-8-0), [35,](#page-8-0) [37,](#page-8-0) [39](#page-8-0), [40\]](#page-8-0); 1915 patients).

We found statistically significant associations between VM and cancer differentiation (well vs. moderate/poor: RR: 1.08; 95 % CI 1.04–1.12; $p < 0.001$), lymph node metastasis (N1 vs. N0: RR: 1.81; 95 % CI 1.43–2.29; $p \lt 0.001$), distant metastasis (M1 vs. M0: RR: 1.94; 95 % CI 1.56–2.41; $P \lt 0.001$), TNM stage (III/VI vs. I/II: RR: 1.81; 95 % CI: 1.48–2.23; $P < 0.001$ (Table 2, Figure S1).

Group	Studies (n)	Patients (n)		RR (95 % CI)	P	I^2 (%)	$P_{\text{het}}^{\text{a}}$
		$VM+$	$VM-$				
Sex (female vs. male)	17	592	1560	$0.91(0.74 - 1.12)$	0.38	11.4	0.32
Differentiation (moderate/poor vs. well)	14	791	1241	$1.08(1.04-1.12)$	< 0.001	85.5	< 0.001
Lymph metastasis (N1 vs. N0)	10	458	1277	$1.81(1.43 - 2.29)$	< 0.001	75.4	< 0.001
Distant metastasis (M1 vs. M0)	13	356	1260	$1.94(1.56-2.41)$	< 0.001	44.2	0.04
TNM stage (III/VI vs. I/II)	13	557	1358	$1.81(1.48-2.23)$	< 0.001	79.0	< 0.001

Table 2 Subgroup differences in a meta-analysis of the association between vasculogenic mimicry and disease features in patients with cancer

VM+ patients with positive vasculogenic mimicry, VM- patients without vasculogenic mimicry, RR risk ratio, $P_{\text{het}} P$ values for heterogeneity from O test

^a A random-effect model was used when the P value for heterogeneity test was <0.05; otherwise, a fixed-effect model was used. I² the percentage of variability in RR attributable to heterogeneity

Vascular mimicry and overall survival

The meta-analysis revealed that the risk of mortality was significantly higher for cancer patients with VM than without VM (HR, 2.16; 95 %CI 1.95–2.38; $P < 0.001$) (Table 3, Fig. [2](#page-5-0)). There was moderate heterogeneity among studies ($P_{\text{het}} = 0.055$ and $I^2 = 29.4$ %) (Table 3), so we conducted subgroup analyses according to confounders, such as cancer types, ethnicity, VM detection methods, patient number, and quality score. After stratifying by cancer type, VM was still associated with statistically significantly poorer overall survival from melanoma, gastric cancer, sarcomas, ovarian tumor, hepatocellular carcinoma, breast cancer, head and neck cancer, colorectal cancer, glioma, lung cancer, and other types of cancer (Table 3). After stratifying by ethnicity, the pooled HRs of Asians and non-Asians were 2.13 and 3.45, respectively (Table 3). In the subgroup analysis based on VM detection methods, the pooled HR was 2.25 for PAS staining, 2.11 for CD31/PAS staining, 2.15 for CD34/PAS

Table 3 Estimated effect of vasculogenic mimicry on overall survival in a meta-analysis of 36 studies of cancer patients by subgroup

Subgroup	Studies (n)	Patients (n)		Overall survival, HR (95 % CI)	P^a	I^2 (%)	$P_{\rm het}$
		$VM+$	$VM-$				
Total	36	964	2645	$2.16(1.95 - 2.38)$	< 0.001	29.4	0.055
Cancer type							
Melanoma	5	234	495	$2.14(1.47-2.82)$	< 0.001	12.6	0.33
Gastric cancer	5	169	469	$1.79(1.37-2.20)$	< 0.001	31.3	0.21
Sarcomas	4	59	173	$2.08(1.23 - 2.94)$	< 0.001	$0.0\,$	0.59
Ovarian tumor	3	105	119	$1.91(0.63 - 3.18)$	0.003	0.0	0.49
HCC	$\sqrt{2}$	36	151	$2.18(1.36-3.00)$	< 0.001	0.0	0.55
Breast cancer	\overline{c}	52	369	$2.40(0.85 - 3.95)$	0.002	77.1	0.04
HNC	\overline{c}	85	230	$3.06(1.85-4.27)$	< 0.001	0.0	0.73
Colorectal cancer	\overline{c}	62	258	$2.57(1.72 - 3.42)$	< 0.001	61.8	0.11
Glioma	\overline{c}	36	151	$1.80(1.22 - 2.39)$	< 0.001	0.0	0.36
Lung cancer	$\boldsymbol{2}$	126	230	$2.05(1.26-2.85)$	< 0.001	0.0	0.46
Other ^a	6	187	384	$3.05(2.45-3.64)$	< 0.001	44.6	0.11
Ethnicity							
Asian	31	965	2746	$2.13(1.91 - 2.35)$	< 0.001	28.8	0.07
Non-Asian	4	193	339	$3.45(2.13 - 4.77)$	< 0.001	$0.0\,$	0.52
Assay methods							
PAS	τ	285	801	$2.25(1.68-2.82)$	< 0.001	13.7	0.33
CD31/PAS	14	341	1044	$2.11(1.67-2.54)$	< 0.001	25.0	0.18
CD34/PAS	13	491	1169	$2.15(1.87-2.43)$	< 0.001	48.1	0.03
Pan-cytokeratin/CD34	$\mathbf{1}$	41	71	2.84 (1.09-4.59)	0.002	NA	NA
Sample size (n)							
≤ 100	16	756	2170	$2.01(1.74 - 2.29)$	< 0.001	1.6	0.43
>100	19	402	915	$2.43(2.07-2.78)$	< 0.001	39.3	0.04
Study quality ^c							
Low quality	17	506	1535	$2.47(2.13 - 2.81)$	< 0.001	42.4	0.03
High quality	18	652	1550	$1.95(1.67-2.23)$	< 0.001	0.0	0.60

 VM patients with positive vasculogenic mimicry, VM patients without vasculogenic mimicry, HR hazard ratio, I^2 the percentage of variability in attributable to heterogeneity, P_{het} P values for heterogeneity from Q test, HCC hepatocellular carcinoma, HNC head and neck cancer, PAS Periodic acid–Schiff staining, NA not available

 $^{\circ}$ A random-effect model was used when the P value for heterogeneity was <0.05; otherwise, a fixed-effect model was used

^b Other included testicular germ cell malignant tumors, gallbladder cancer, prostate cancer, esophageal cancer, renal carcinoma, and osteosarcoma

 \degree Low-quality studies had Newcastle–Ottawa quality scores of 0–4, and high-quality studies had scores of 5–9

Study		%
ID	ES (95% CI)	Weight
Maniotis AJ (1999)	3.83(2.23, 6.57)	1.00
Shirakawa K (2002)	2.07(1.03, 4.18)	1.88
Sun BC (2004)	5.13 (1.54, 17.08)	0.08
Sun BC (2004)	2.94(1.67, 5.17)	1.53
Sun BC (2004)	1.74 (0.82, 3.70)	2.27
Sun BC (2004)	3.46(1.87, 6.43)	0.90
Sun BC (2006)	2.55(1.48, 4.41)	2.18
Sun BC (2008)	5.82 (2.65, 12.80)	0.18
Liu SY (2008)	2.84 (1.58, 5.09)	1.52
Hillen F (2008)	1.64(0.44, 6.12)	0.58
Gao Y (2009)	1.13(0.32, 3.92)	1.45
Zhang SW (2009)	1.79 (1.17, 2.73)	7.68
Baeten CI (2009)	4.27 (2.59, 7.05)	0.94
Wang L (2009)	1.77 (0.87, 3.59)	2.53
Wang XY (2009)	4.04 (3.16, 4.93)	5.95
Li M (2010)	1.57(1.06, 2.31)	12.05
Wang W (2010)	3.26(1.99, 5.34)	1.67
Liu WB (2011)	2.01 (1.25, 3.23)	4.81
Liu XM (2011)	2.46(1.37, 4.42)	2.01
Sun W (2012)	2.68 (1.04, 6.88)	0.55
Liu RL (2012)	2.12 (0.88, 5.12)	1.04
Liu ZY (2012)	2.28 (1.54, 3.38)	5.54
Van Beurden A (2012)	2.82 (0.68, 11.58)	0.16
Wu S (2012)	1.91 (1.23, 2.98)	6.10
Chai DM (2013)	2.18(1.03, 4.61)	1.47
Wang SY (2013)	1.69(1.17, 2.44)	11.69
Zhang Y (2013)	2.08(1.15, 3.75)	2.77
Liu T (2014)	10.75 (5.40, 21.38)	0.07
Ren K (2014)	2.49 (1.28, 4.85)	1.47
Song YY (2014)	3.02(1.61, 5.66)	1.14
Zhou L (2015)	1.71(1.21, 2.42)	12.93
Yu, L. (2012)	2.63(1.12, 6.17)	0.73
Qi, H. (2014)	3.02(1.61, 5.66)	1.14
Sun, Q. (2014)	2.76 (1.19, 6.38)	0.70
Li, Y. (2015)	2.71 (1.40, 5.24)	1.27
Overall (I-squared = 29.4% , $p = 0.055$)	2.16 (1.95, 2.38)	100.00
-21.4 0	21.4	

Fig. 2 Forest plots of the effects of vasculogenic mimicry on survival in patients with cancer

staining, and 2.84 for pan-cytokeratin/CD34 staining (Table [3](#page-4-0)). In studies with larger samples ($n \ge 100$) and smaller samples ($n < 100$), the pooled HRs were 2.01 and 2.43, respectively (Table [3\)](#page-4-0). Stratification by quality scores revealed significant associations for both high- and lowquality studies (Table [3\)](#page-4-0).

Test of heterogeneity

We created a Galbraith plot to identify potential sources of heterogeneity. The main contributors to heterogeneity were the studies by Liu and Wang $[25, 36]$ $[25, 36]$ $[25, 36]$ $[25, 36]$ (Fig. [3](#page-6-0)). When these two studies were omitted, I^2 % decreased from 29.4 to 0 %, and the statistical significance of the combined HRs in the overall and subgroup analyses was not substantially altered (data not shown.)

Sensitivity analysis, cumulative analyses, and publication bias

The pooled HRs remained the same in a sensitivity analysis in which each study was sequentially excluded (Figure S2). Moreover, cumulative meta-analyses suggested that the association has been significant since the first study was published in 1999 (Figure S3). These results indicate stabile and reliable findings. No indication of publication bias was found using Egger's test ($P = 0.82$), and the funnel plot seemed symmetrical (Fig. [4](#page-6-0)), indicating no marked publication bias.

Fig. 3 Galbraith plot analysis of the amount of heterogeneity from all the included studies. The Y-axis shows the ratio of the log HR to its standard error (SE), and the X-axis shows the reciprocal of the SE. At a 2 standard deviation distance parallel to the regression line, the 2 lines create an interval. Studies lacking in heterogeneity would lie within the 95 % confidence interval. Studies by Liu and Wang were identified as potential sources of heterogeneity in a meta-analysis of the effects of vasculogenic mimicry on cancer survival

Discussion

Our meta-analysis summarized the results of 36 clinical studies representing 3609 patients. Overall, positive VM status strongly predicted lower overall survival in patients with malignant cancers. Moreover, similar, significant results were found in the subgroup analyses by cancer types, ethnicity, VM detection methods, sample size, and study quality. Additionally, VM was also significantly associated with cancer differentiation, lymph node metastasis, distant metastasis, and TNM stage, which might adversely affect cancer survival.

Tumor growth and hematogenous dissemination depend on adequate blood supply. Tumor vasculature is highly complex and possibly derived from a variety of sources, including angiogenesis [[58\]](#page-9-0), vasculogenesis [\[59–61](#page-9-0)], cooption [[62\]](#page-9-0), and vasculogenic mimicry [\[8](#page-7-0), [63\]](#page-9-0). In VM, highly aggressive cancer cells form vessel-like structures by their high plasticity, which enables them to function as endothelial-like cells.

Several recent studies [\[7](#page-7-0), [10](#page-7-0), [12](#page-8-0), [14–31,](#page-8-0) [34–43,](#page-8-0) [57\]](#page-9-0) have reported that VM exists in various malignant tumors and is associated with tumor progression, metastasis, treatment failure, and increases death in many $[6, 9, 18-23, 38]$ $[6, 9, 18-23, 38]$ $[6, 9, 18-23, 38]$ $[6, 9, 18-23, 38]$ $[6, 9, 18-23, 38]$ $[6, 9, 18-23, 38]$.

Although one meta-analysis [\[64\]](#page-9-0) has reported on the prognostic value of VM in cancer, it has some shortcomings. Data from one study were published in two articles $[10]$, one of which should have been excluded from the analysis. Despite evidence of remarkable heterogeneity and publication bias, there was no attempt to explore the sources of heterogeneity, which might have compromised the interpretation. Time-to-event outcomes were assessed with relative risks, rather than with hazards ratios. Additionally, 12 new studies [[12,](#page-8-0) [16](#page-8-0), [17,](#page-8-0) [21,](#page-8-0) [25](#page-8-0), [31,](#page-8-0) [35](#page-8-0), [39–42,](#page-8-0) [57\]](#page-9-0) evaluating the relationship between VM and survival outcomes have been published since September 2013. Thus, it was important to update the meta-analyses. The pooled results of our meta-analysis showed that VM was significantly associated with poor OS in cancer patients. What was more, the presence of VM was also significantly associated with cancer differentiation, lymph node metastasis, distant metastasis, and TNM stage.

We found an association of VM with a more aggressive tumor phenotype. Several underlying mechanisms are possible. First, VM could provide a functional perfusion pathway for rapidly growing tumors and possibly a metastatic escape route by transporting blood from leaky vessels or by connecting with the endothelial-lined vasculature [\[65](#page-9-0)]. Second, accumulating evidence [\[66](#page-9-0)] indicates that cancer stem cells are involved in VM formation, have been associated with tumor invasion and metastasis, and are therefore important in tumor progression. Third, the formation of VM involves signaling pathways and some factors related to tumor cell migration, invasion, and matrix remodeling, including Notch [[67\]](#page-9-0) and Nodal [\[68](#page-9-0)] signaling pathways, VE-cadherin [\[69](#page-9-0), [70](#page-9-0)], EPH receptor A2(EphA2) [\[71](#page-9-0), [72](#page-9-0)], matrix metalloproteinases [[20,](#page-8-0) [22](#page-8-0), [73](#page-9-0)], and hypoxia-inducible factor-1alpha (HIF-1(alpha)) [\[37](#page-8-0), [39,](#page-8-0) [69,](#page-9-0) [74,](#page-9-0) [75](#page-9-0)]. Forth, hypoxia can modulate the expression of genes essential for cell viability, tumor growth, metastasis, and VM in cancer [\[76–78](#page-9-0)]. These hypoxia-induced VM and VM-associated genes highlight the critical importance of hypoxia in tumor progression. Finally, VM is lined with tumor-derived endothelial-like cells that differ from conventional endothelial cells. Therefore, VM might present with a native resistance to anti-angiogenic compounds, such as bevacizumab, sorafenib, and sunitinib.

Our study has some limitations. The moderate heterogeneity among the studies was a potential problem when interpreting the results, although when the studies by Liu et al. and Wang et al. [[25,](#page-8-0) [36](#page-8-0)] were deleted from the analysis, our results remained the same. A more accurate pooled HRs analysis with sufficient data was required to adjust for covariates, such as age, sex, histological type, tumor differentiation, TNM stage, lymphatic metastasis, and distant metastasis, if these data were available. Finally, although publication bias was not detected using Begg's funnel plot

and Egger's test, it always remains a possibility. However, several studies with contradictory results would be required to refute or reverse our findings. Our meta-analysis also has some strengths. Sensitivity analysis revealed no significant difference when any individual article was omitted. Moreover, cumulative meta-analyses demonstrated that inclinations toward the significant association have been evident since the first study in 1999. These analyses indicate that our results are stable and reliable.

We found that VM is associated with a worse prognosis in several different tumor types. Vasculogenic mimicry is also significantly associated with cancer differentiation, lymph metastasis, distant metastasis, and TNM stage. These results suggest that developing strategies against the VM would be a promising therapeutic approach to solid tumors.

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References

- 1. Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. Cell 144:646–674
- 2. Stacker SA, Achen MG (2013) The VEGF signaling pathway in cancer: the road ahead. Chin J Cancer 32:297–302
- 3. Liu J, Huang J, Yao WY, Ben QW, Chen DF et al (2012) The origins of vacularization in tumors. Front Biosci (Landmark Ed) 17:2559–2565
- 4. Qin L, Bromberg-White JL, Qian CN (2012) Opportunities and challenges in tumor angiogenesis research: back and forth between bench and bed. Adv Cancer Res 113:191–239
- 5. Qian CN (2013) Hijacking the vasculature in ccRCC–co-option, remodelling and angiogenesis. Nat Rev Urol 10:300–304
- 6. Qian CN, Tan MH, Yang JP, Cao Y (2016) Revisiting tumor angiogenesis: vessel co-option, vessel remodeling, and cancer cell-derived vasculature formation. Chin J Cancer 35:10
- 7. Maniotis AJ, Folberg R, Hess A, Seftor EA, Gardner LM et al (1999) Vascular channel formation by human melanoma cells in vivo and in vitro: vasculogenic mimicry. Am J Pathol 155:739–752
- 8. Qiao L, Liang N, Zhang J, Xie J, Liu F et al (2015) Advanced research on vasculogenic mimicry in cancer. J Cell Mol Med 19:315–326
- 9. Chen YS, Chen ZP (2014) Vasculogenic mimicry: a novel target for glioma therapy. Chin J Cancer 33:74–79
- 10. Sun B, Zhang S, Zhao X, Zhang W, Hao X (2004) Vasculogenic mimicry is associated with poor survival in patients with mesothelial sarcomas and alveolar rhabdomyosarcomas. Int J Oncol 25:1609–1614
- 11. Zhang S, Li M, Zhang D, Xu S, Wang X et al (2009) Hypoxia influences linearly patterned programmed cell necrosis and tumor blood supply patterns formation in melanoma. Lab Invest 89:575–586
- 13. Dunleavey JM, Xiao L, Thompson J, Kim MM, Shields JM et al (2014) Vascular channels formed by subpopulations of PECAM1+ melanoma cells. Nat Commun 5:5200
- 14. Sun B, Qie S, Zhang S, Sun T, Zhao X et al (2008) Role and mechanism of vasculogenic mimicry in gastrointestinal stromal tumors. Hum Pathol 39:444–451
- 15. Li M, Gu Y, Zhang Z, Zhang S, Zhang D et al (2010) Vasculogenic mimicry: a new prognostic sign of gastric adenocarcinoma. Pathol Oncol Res 16:259–266
- 16. Song YY, Sun LD, Liu ML, Liu ZL, Chen F et al (2014) STAT3, p-STAT3 and HIF-1alpha are associated with vasculogenic mimicry and impact on survival in gastric adenocarcinoma. Oncol Lett 8:431–437
- 17. Zhou L, Yu L, Feng ZZ, Gong XM, Cheng ZN et al (2015) Aberrant expression of markers of cancer stem cells in gastric adenocarcinoma and their relationship to vasculogenic mimicry. Asian Pac J Cancer Prev 16:4177–4183
- 18. Wang LGY, Zhang S, Sun B (2009) Pilot study and clinical prognosis significance of vasculogenic mimicry in leiomyosarcoma. Tianjin Med J 37:161
- 19. Gao Y, Zhao XL, Gu Q, Wang JY, Zhang SW et al (2009) Correlation of vasculogenic mimicry with clinicopathologic features and prognosis of ovarian carcinoma. Zhonghua Bing Li Xue Za Zhi 38:585–589
- 20. Yu L, Wu SW, Zhou L, Song WQ (2012) Correlation between bacterial L-form infection, expression of HIF-1alpha/MMP-9 and vasculogenic mimicry in epithelial ovarian cancer. Sheng Li Xue Bao 64:657–665
- 21. Sun Q, Zou X, Zhang T, Shen J, Yin Y et al (2014) The role of miR-200a in vasculogenic mimicry and its clinical significance in ovarian cancer. Gynecol Oncol 132:730–738
- 22. Sun B, Zhang S, Zhang D, Du J, Guo H et al (2006) Vasculogenic mimicry is associated with high tumor grade, invasion and metastasis, and short survival in patients with hepatocellular carcinoma. Oncol Rep 16:693–698
- 23. Liu WB, Xu GL, Jia WD, Li JS, Ma JL et al (2011) Prognostic significance and mechanisms of patterned matrix vasculogenic mimicry in hepatocellular carcinoma. Med Oncol 28(Suppl 1):S228–S238
- 24. Shirakawa K, Wakasugi H, Heike Y, Watanabe I, Yamada S et al (2002) Vasculogenic mimicry and pseudo-comedo formation in breast cancer. Int J Cancer 99:821–828
- 25. Liu T, Sun B, Zhao X, Li Y, Gu Q et al (2014) OCT4 expression and vasculogenic mimicry formation positively correlate with poor prognosis in human breast cancer. Int J Mol Sci 15:19634–19649
- 26. Liu SY, Chang LC, Pan LF, Hung YJ, Lee CH et al (2008) Clinicopathologic significance of tumor cell-lined vessel and microenvironment in oral squamous cell carcinoma. Oral Oncol 44:277–285
- 27. Wang W, Lin P, Han C, Cai W, Zhao X et al (2010) Vasculogenic mimicry contributes to lymph node metastasis of laryngeal squamous cell carcinoma. J Exp Clin Cancer Res 29:60
- 28. Baeten CI, Hillen F, Pauwels P, de Bruine AP, Baeten CG (2009) Prognostic role of vasculogenic mimicry in colorectal cancer. Dis Colon Rectum 52:2028–2035
- 29. Liu Z, Sun B, Qi L, Li H, Gao J et al (2012) Zinc finger E-box binding homeobox 1 promotes vasculogenic mimicry in colorectal cancer through induction of epithelial-to-mesenchymal transition. Cancer Sci 103:813–820
- 30. Liu XM, Zhang QP, Mu YG, Zhang XH, Sai K et al (2011) Clinical significance of vasculogenic mimicry in human gliomas. J Neurooncol 105:173–179
- 31. Wang SY, Ke YQ, Lu GH, Song ZH, Yu L et al (2013) Vasculogenic mimicry is a prognostic factor for postoperative survival in patients with glioblastoma. J Neurooncol 112:339–345
- 32. Cheng L, Huang Z, Zhou W, Wu Q, Donnola S et al (2013) Glioblastoma stem cells generate vascular pericytes to support vessel function and tumor growth. Cell 153:139–152
- 33. Wang R, Chadalavada K, Wilshire J, Kowalik U, Hovinga KE et al (2010) Glioblastoma stem-like cells give rise to tumour endothelium. Nature 468:829–833
- 34. Wu S, Yu L, Cheng Z, Song W, Zhou L et al (2012) Expression of maspin in non-small cell lung cancer and its relationship to vasculogenic mimicry. J Huazhong Univ Sci Technol Med Sci 32:346–352
- 35. Li Y, Sun B, Zhao X, Zhang D, Wang X et al (2015) Subpopulations of uPAR+ contribute to vasculogenic mimicry and metastasis in large cell lung cancer. Exp Mol Pathol 98:136–144
- 36. Wang X, Wang L, Gu Y, Zhang S, Zhao X et al (2009) Vasculogenic mimicry in testicular germ cell malignant tumor and its significance for prognosis. Chin J Clin Oncol 36:78–82
- 37. Sun W, Shen ZY, Zhang H, Fan YZ, Zhang WZ et al (2012) Overexpression of HIF-1alpha in primary gallbladder carcinoma and its relation to vasculogenic mimicry and unfavourable prognosis. Oncol Rep 27:1990–2002
- 38. Liu R, Yang K, Meng C, Zhang Z, Xu Y (2012) Vasculogenic mimicry is a marker of poor prognosis in prostate cancer. Cancer Biol Ther 13:527–533
- 39. Chai DM, Bao ZQ, Hu JG, Ma L, Feng ZZ et al (2013) Vasculogenic mimicry and aberrant expression of HIF-lalpha/E-cad are associated with worse prognosis of esophageal squamous cell carcinoma. J Huazhong Univ Sci Technol Med Sci 33:385–391
- 40. Zhang Y, Sun B, Zhao X, Liu Z, Wang X et al (2013) Clinical significances and prognostic value of cancer stem-like cells markers and vasculogenic mimicry in renal cell carcinoma. J Surg Oncol 108:414–419
- 41. Ren K, Yao N, Wang G, Tian L, Ma J et al (2014) Vasculogenic mimicry: a new prognostic sign of human osteosarcoma. Hum Pathol 45:2120–2129
- 42. Zhang X, Liu C, Luo L, Cai X (2013) Vasculogenic mimicry in tongue squamous cell carcinoma. Nan Fang Yi Ke Da Xue Xue Bao 33:593–597
- 43. Hillen F, Baeten CI, van de Winkel A, Creytens D, van der Schaft DW et al (2008) Leukocyte infiltration and tumor cell plasticity are parameters of aggressiveness in primary cutaneous melanoma. Cancer Immunol Immunother 57:97–106
- 44. Jang TL, Bekelman JE, Liu Y, Bach PB, Basch EM et al (2010) Physician visits prior to treatment for clinically localized prostate cancer. Arch Int Med 170:440–450
- 45. Knobloch K, Yoon U, Vogt PM (2011) Preferred reporting items for systematic reviews and meta-analyses (PRISMA) statement and publication bias. J Craniomaxillofac Surg 39:91–92
- 46. Panic N, Leoncini E, de Belvis G, Ricciardi W, Boccia S (2013) Evaluation of the endorsement of the preferred reporting items for systematic reviews and meta-analysis (PRISMA) statement on the quality of published systematic review and meta-analyses. PLoS ONE 8:e83138
- 47. Sun B, Zhang S, Ni C (2005) The clinical significance study of vasculogenetic mimicry in 337 cases of bi-directional differential malignant tumors. Chin J Clin Oncol 32:64–67
- 48. Jadad AR, Moore RA, Carroll D, Jenkinson C, Reynolds DJ et al (1996) Assessing the quality of reports of randomized clinical trials: is blinding necessary? Control Clin Trials 17:1–12
- 49. DerSimonian R, Laird N (1986) Meta-analysis in clinical trials. Control Clin Trials 7:177–188
- 50. Whitehead A, Whitehead J (1991) A general parametric approach to the meta-analysis of randomized clinical trials. Stat Med 10:1665–1677
- 51. Higgins JP, Thompson SG (2002) Quantifying heterogeneity in a meta-analysis. Stat Med 21:1539–1558
- 52. Galbraith RF (1988) A note on graphical presentation of estimated odds ratios from several clinical trials. Stat Med 7:889–894
- 53. Patsopoulos NA, Evangelou E, Ioannidis JP (2008) Sensitivity of between-study heterogeneity in meta-analysis: proposed metrics and empirical evaluation. Int J Epidemiol 37:1148–1157
- 54. Mullen B, Muellerleile P, Bryant B (2001) Cumulative metaanalysis: a consideration of indicators of sufficiency and stability. Pers Soc Psychol Bull 27:1450–1462
- 55. Soeken KL, Sripusanapan A (2003) Assessing publication bias in meta-analysis. Nurs Res 52:57–60
- 56. Egger M, Davey Smith G, Schneider M, Minder C (1997) Bias in meta-analysis detected by a simple, graphical test. BMJ 315:629–634
- 57. Qi H, Sun B, Zhao X, Du J, Gu Q et al (2014) Wnt5a promotes vasculogenic mimicry and epithelial-mesenchymal transition via protein kinase Calpha in epithelial ovarian cancer. Oncol Rep 32:771–779
- 58. Cao Y (2009) Angiogenesis in malignancy. Semin Cancer Biol 19:277–278
- 59. Silvan U, Diez-Torre A, Bonilla Z, Moreno P, Diaz-Nunez M et al (2015) Vasculogenesis and angiogenesis in nonseminomatous testicular germ cell tumors. Urol Oncol 33(268):e217–e268
- 60. Brown JM (2014) Vasculogenesis: a crucial player in the resistance of solid tumours to radiotherapy. Br J Radiol 87:20130686
- 61. Qing YL, Sha TH, Ji FY, Qin HK, Jun CX et al (2009) Angiogenesis, vasculogenesis, and vasculogenic mimicry in ovarian cancer. Int J Gynecol Obstet 107:S242
- 62. Pezzella F, Harris AL (2014) When cancer co-opts the vasculature. N Engl J Med 370:2146–2147
- 63. Seftor RE, Hess AR, Seftor EA, Kirschmann DA, Hardy KM et al (2012) Tumor cell vasculogenic mimicry: from controversy to therapeutic promise. Am J Pathol 181:1115–1125
- 64. Cao Z, Bao M, Miele L, Sarkar FH, Wang Z et al (2013) Tumour vasculogenic mimicry is associated with poor prognosis of human cancer patients: a systemic review and meta-analysis. Eur J Cancer 49:3914–3923
- 65. Wagenblast E, Soto M, Gutierrez-Angel S, Hartl CA, Gable AL et al (2015) A model of breast cancer heterogeneity reveals vascular mimicry as a driver of metastasis. Nature 520:358–362
- 66. Yao XH, Ping YF, Bian XW (2011) Contribution of cancer stem cells to tumor vasculogenic mimicry. Protein Cell 2:266–272
- 67. Vartanian A, Gatsina G, Grigorieva I, Solomko E, Dombrovsky V et al (2013) The involvement of Notch signaling in melanoma vasculogenic mimicry. Clin Exp Med 13:201–209
- 68. McAllister JC, Zhan Q, Weishaupt C, Hsu MY, Murphy GF (2010) The embryonic morphogen, Nodal, is associated with channel-like structures in human malignant melanoma xenografts. J Cutan Pathol 37(Suppl 1):19–25
- 69. Tang N, Shi H, Zhang J (2013) HIF-1alpha induces expression of VE-cadherin and modulates vasculogenic mimicry in ESCC cells. J Gastroenterol Hepatol 28:863
- 70. Hess AR, Seftor EA, Gruman LM, Kinch MS, Seftor REB et al (2006) VE-cadherin regulates EphA2 in aggressive melanoma cells through a novel signaling pathway: implications for vasculogenic mimicry. Cancer Biol Ther 5:228–233
- 71. Wang W, Lin P, Sun B, Zhang S, Cai W et al (2014) Epithelialmesenchymal transition regulated by EphA2 contributes to vasculogenic mimicry formation of head and neck squamous cell carcinoma. Biomed Res Int 2014:803914
- 72. Margaryan NV, Strizzi L, Abbott DE, Seftor EA, Rao MS et al (2009) EphA2 as a promoter of melanoma tumorigenicity. Cancer Biol Ther 8:275–284
- 73. Lu XS, Sun W, Ge CY, Zhang WZ, Fan YZ (2013) Contribution of the PI3 K/MMPs/Ln-5gamma2 and EphA2/FAK/Paxillin signaling pathways to tumor growth and vasculogenic mimicry of gallbladder carcinomas. Int J Oncol 42:2103–2115
- 74. Shi R, Jin HL, Zhang HJ (2011) Effect of RNA interference targeting for HIF-1(alpha) on vasculogenic mimicry in esophageal squamous cell carcinoma. Gastroenterology 140:S827
- 75. Comito G, Calvani M, Giannoni E, Bianchini F, Calorini L et al (2011) HIF-1alpha stabilization by mitochondrial ROS promotes Met-dependent invasive growth and vasculogenic mimicry in melanoma cells. Free Radic Biol Med 51:893–904
- 76. Du J, Sun B, Zhao X, Gu Q, Dong X et al (2014) Hypoxia promotes vasculogenic mimicry formation by inducing epithelialmesenchymal transition in ovarian carcinoma. Gynecol Oncol 133:575–583
- 77. Osinsky S, Zavelevich M, Vaupel P (2009) Tumor hypoxia and malignant progression. Exp Oncol 31:80–86
- 78. Sun B, Zhang D, Zhang S, Zhang W, Guo H et al (2007) Hypoxia influences vasculogenic mimicry channel formation and tumor invasion-related protein expression in melanoma. Cancer Lett 249:188–197