

Understanding the key ecological traits of cyanobacteria as a basis for their management and control in changing lakes

Evanthia Mantzouki · Petra M. Visser · Myriam Bormans · Bas W. Ibelings

Received: 30 September 2014/Accepted: 1 June 2015/Published online: 16 June 2015 © Springer Science+Business Media Dordrecht 2015

Abstract Anticipated climatic changes combined with eutrophication are predicted to enhance the dominance of several notorious cyanobacterial taxa. Cyanobacteria have many key ecological traits that may allow them to thrive under foreseen scenarios of environmental change. Understanding the ecophysiological traits of harmful species has proven important for their successful control and management. Indeed, if the links between key cyanobacterial traits and the specific environmental conditions that allow expression of these traits can be disrupted, we could identify (novel) means for operational control and mitigate or prevent water quality problems. A good example is artificial mixing of a lake that breaks down the water column stability on which fast floating, buoyant cyanobacteria depend. Based upon Reynolds' functional phytoplankton

Guest editors: Petra M. Visser, Bas W. Ibelings, Jutta Fastner & Myriam Bormans/Cyanobacterial blooms. Ecology, prevention, mitigation and control.

E. Mantzouki (⊠) · B. W. Ibelings Institut F.A. Forel, University of Geneva, Geneva, Switzerland e-mail: evanthia.mantzouki@unige.ch

P. M. Visser

Institute for Biodiversity and Ecosystem Dynamics, University of Amsterdam, Amsterdam, The Netherlands

M. Bormans

UMR CNRS 6553 ECOBIO, Université de Rennes 1, Rennes, France

classification, we focused on five groups of cyanobacteria that from a management point of view can be seen as homogeneous and have comparable environmental sensitivities. For each group, we present (1) its key traits, (2) how these characteristics will maintain their function under future environmental change, (3) explanation of how understanding the function of these traits can reveal the "Achilles heel" of the particular functional group and (4) which (combination of) control measures is most likely to be successful. Despite looking for specific environmental sensitivities of individual groups, we maintain that controlling nutrients remains the basis for managing blooms, no matter which functional type dominates. Providing further ecological knowledge to lake management could be the key to effective bloom control and healthier, sustainable freshwater ecosystems even in a warmer future.

Keywords Climate change · Cyanobacterial blooms · Eutrophication · Functional traits · Lake management · Microcystins

Introduction

This publication is the first paper in a special issue of Aquatic Ecology on "Cyanobacterial blooms: Ecology, prevention, mitigation and control." In this special issue, the various contributions discuss a range of control measures available to lake management to prevent and control nuisance blooms of cyanobacteria. In this contribution, we review what is known about those key traits of cyanobacteria, like buoyancy or nitrogen fixation that define their ecological niche and are key to their successful management. Our title, however, also speaks of "changing lakes," and we cannot properly address cyanobacterial problems without discussing the—changing—environment they live in. So this publication also outlines the main consequences of eutrophication and climate change, as two major drivers of bloom formation.

Phosphorus and nitrogen pollution is the biggest threat to water quality worldwide (Schindler and Vallentyne 2008). Anthropogenic eutrophication from sewage, fertilizers and detergents is responsible for massive algal blooms that capitalize on the excess of nutrients. Cyanobacterial blooms are the most conspicuous sign of eutrophication, mostly resulting in a strong smell, discoloration of the water and accumulation of floating cyanobacteria at the lake surface (Ibelings and Chorus 2007). Highly eutrophic waters can support excessive cyanobacterial biomass that increases turbidity, limits the penetration of light to deeper layers and increases anoxia in the hypolimnion (Schindler et al. 2008). Managers have worked hard to control eutrophication. The International Lake Environment Committee's (ILEC) review on 217 lakes showed that water management in industrialized areas succeeded in reducing nutrient input in many lakes (e.g., Lake Biwa—Japan, Lake Constance—Germany and Switzerland, Lake Balaton-Hungary, the Great Lakes in North America), but the nutrient concentrations did not yet return to the levels of the 1950s, before the peak in the eutrophication, and in many lakes with high historical loads, internal loading will continue to fuel cyanobacterial blooms for many years after external nutrient inputs have been reduced. In many cases, for instance, Lake Veluwe-the Netherlands-lakes showed severe hysteresis to restoration measures and remained in a turbid state for decades, despite tackling external nutrient loading (Ibelings et al. 2007).

An important question is which nutrient, phosphorus or nitrogen, is chiefly responsible to promote cyanobacterial biomass in an ecosystem. Schindler et al. (2008) proved experimentally that algal blooms in the eutrophic lakes they studied could not be controlled by reducing nitrogen input, but only by managing phosphorus. According to their wholeecosystem experiment, reducing nitrogen inputs increasingly favored nitrogen-fixing cyanobacteria that outcompeted the other phytoplankton. Nitrogen fixation could sustain cyanobacterial biomass and the lake remained highly eutrophic, despite showing indications of extreme nitrogen limitation seasonally (Schindler et al. 2008). But this shift to nitrogen fixers does not occur in most N-limited lakes of low to moderate fertility, and where it does, lakes typically remain N limited despite the presence of nitrogen fixers (Paerl et al. 2011). N₂ fixation can only sustain <50 % of primary production, even when P supplies are sufficient (Scott and McCarthy 2010). Furthermore, there are records of eutrophic systems dominated by non-N2 fixers that are N and P co-limited or even N limited (Paerl et al. 2011). Rigosi et al. (2014) showed that both nutrients are dominant factors promoting blooms. Thus, the feasibility of focusing only on P deficiency must be evaluated on a case-bycase basis (Lewis and Wurtsbaugh 2008) since it is likely that both N and P input controls are needed for long-term cyanobacterial bloom management. Indeed, we emphasize that nutrient control always needs to be the basis of lake restoration measures, aimed to reduce or prevent problems caused by algal blooms.

There is mounting evidence that the current worldwide increase in cyanobacterial blooms is not just the result of anthropogenic eutrophication. Climate change, through direct and indirect effects of warming, is seen to play an increasing role in changing the properties of the aquatic ecosystems (Elliott 2012; Kosten et al. 2012; Paerl and Huisman 2008). Global warming has the potential to directly affect phytoplankton community composition as the growth rates of different phytoplankton taxa exhibit different optimal ranges (Butterwick et al. 2005). Apart from the direct consequences of warming, the increase in surface water temperature in peri-alpine lakes, for instance, has resulted in up to 20 % higher water column stability (Livingstone 2003). As a consequence of global warming, the thermocline of deep stratifying lakes remains stable for longer periods (earlier onset in spring, later and incomplete overturn in autumn) and becomes positioned at shallower depths, potentially enhancing the growth of certain cyanobacterial taxa (Paerl and Huisman 2008). Intensified vertical density stratification reduces vertical mixing resulting in enhanced scope for buoyancycontrolled vertical positioning of cyanobacteria (Paerl et al. 2011). Under stable water column conditions,

positively buoyant cyanobacteria will be able to regulate their position in the water column and remain in the euphotic zone, while non-motile, negatively buoyant algae that can only remain suspended in the water column under mixing conditions will tend to sink (Reynolds 1984; Bormans and Condie 1998). Moreover, being closer to the lake surface, buoyant cyanobacteria are closer to the source of CO₂, diffusing in from the atmosphere (Paerl et al. 2011). Higher atmospheric carbon dioxide concentrations will result in higher dissolved carbon dioxide concentrations that may (partly) relieve the potential C limitation of the cyanobacteria with higher cyanobacterial biomass as a consequence (Verspagen et al. 2014). Microelectrode studies, however, demonstrated that the local demand for carbon in cyanobacterial surface waterblooms far outstrips the supply, resulting in severe C limitation, despite being at the surface (Ibelings and Maberly 1998).

Furthermore, climate change may affect the occurrence of cyanobacterial blooms in other, perhaps more subtle ways. We give a few examples. Changes in precipitation are expected due to global warming (IPCC 2007). Intensified precipitation will result in higher terrestrial runoff, increasing nutrient input and stimulating blooms. Increased flushing rates, on the other hand, may prevent blooms when lake residence times are reduced to very low values (Verspagen et al. 2006), but ultimately enhanced nutrient loads always carry a risk of promoting blooms (Paerl and Huisman 2008). Droughts may reduce the flow rate of major rivers to values low enough to support cyanobacteria as found in many Australian rivers (Bormans et al. 2005; Viney et al. 2007), and this may increase their presence in the potamoplankton, even in temperate regions, as demonstrated for the rivers Rhine and Meuse in extremely warm and dry summers (Ibelings et al. 1998). According to the World Meteorological Organization's 2013 report, the occurrence of some extreme events (e.g., hurricane Katrina in 2005 or floods in Pakistan in 2010) can be attributed to climate change (www.cgd.ucar.edu/cas/ace/). The impact of extreme events, like summer heat waves as occurred in Europe in 2003 on lake ecosystems, is likely to be different from the effects of long-term gradual warming (Anneville et al. 2010; Gallina et al. 2011; Straile et al. 2010), but not all summer heat waves, for instance, were shown to promote blooms of cyanobacteria (cf. Jöhnk et al. 2008 vs. Anneville et al. 2015).

We have thus identified eutrophication and climate change as two major anthropogenic stressors that may favor cyanobacterial blooms. These two factors have a clear impact on algal blooms by themselves, but they do not necessarily work only independently. A high algal biomass in eutrophic systems, for instance, can increase the water temperature at a local scale due to strong light absorption (Ibelings et al. 2003). Consequently, eutrophication may promote warmer water and provide additional competitive advantage of buoyant over non-buoyant phytoplankton (Paerl and Huisman 2008; Fig. 1). The underwater light climate as experienced by the phytoplankton is determined by two factors: light attenuation and mixing depth. Steep light attenuation in eutrophic lakes limits the availability of light to a shallow euphotic zone. Climate warming decreases the mixing depth, shifting the advantage to buoyant cells that can float up into the illuminated near-surface mixed layer (Ibelings et al. 1991; Fig. 1). These surface blooms will further increase the underwater light extinction coefficient (Fig. 1). These examples demonstrate the potential for eutrophication to ally with climate warming in promoting the conditions for cyanobacterial growth and dominance (Moss et al. 2011).

The general expectation is that ongoing climate change will amplify eutrophication problems, but much uncertainty remains about the extent to which both stressors will interact. These two stressors may interact in different modes, additive, antagonistic or synergistic. Specifically, it remains an open question whether nutrient and temperature will interact synergistically, so that the combined effects of eutrophication and global warming will be worse than the effect of each single factor alone. Several authors, either through the use of long-term monitoring data or in modeling simulations, have studied the interaction between eutrophication and climate warming as important drivers of blooms (Elliott et al. 2005; Wagner and Adrian 2009). Studies by Kosten et al. (2012), Posch et al. (2012) and Paerl and Huisman (2008) point to higher temperatures combined with nutrients inputs as being important for cyanobacterial bloom formation. However, Jeppesen et al. (2005), using long-term data from 35 lakes located from the subtropical to the temperate zone in North America and Europe, argued that phytoplankton composition is primarily driven by nutrient loading, with global warming effects less detectable. Recent work by

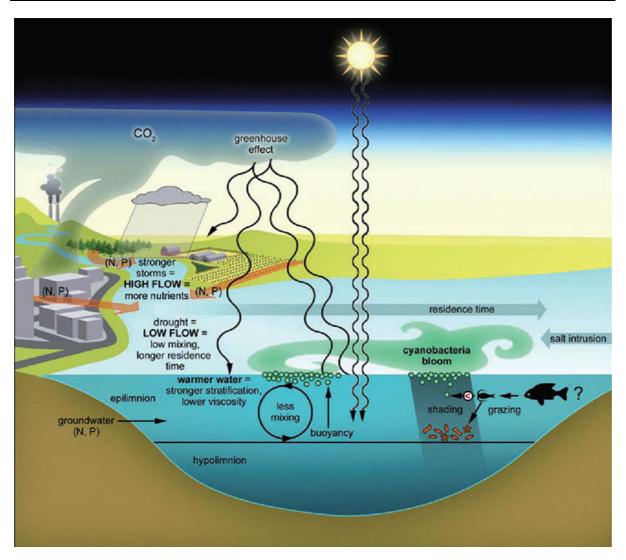


Fig. 1 Conceptual figure, illustrating the environmental controls of cyanobacterial bloom dynamics, and the direct and indirect effects of climate change on these dynamics in

Rigosi et al. (2014) on ca. 1000 American lakes showed that both nutrients and temperature are main determinants of variation in cyanobacterial occurrence between lakes, but they found no overall support for a significant synergistic interaction between nutrients and temperature on cyanobacterial biomass. Moreover, the relative importance of these two factors and their interaction was clearly dependent on lake trophic state and dominant cyanobacterial taxa. Apparently, there is no scientific consensus on the importance and mode of interaction between nutrients and temperature and further investigations are needed. Cyanobacterial blooms are very persistent, and we need to work on combination with external nutrient loading. Figure taken from Paerl and Huisman (2009) with permission by Wiley Journals

better solutions to limit their occurrence and to control their impacts on lake ecosystems and the crucial ecosystem services they provide (Cardinale et al. 2011).

Cyanobacteria and their key traits

Ecology, as a predominantly biological discipline, is concerned with the distributions of organisms and their interrelationships as well as the interactions with their environment (Reynolds et al. 2002). Phytoplankton possesses a high diversity of well-defined ecological traits that can vary predictably over different environmental gradients. Trait-based approaches in ecology demonstrate the importance of functional diversity in determining key properties of ecosystems and their functioning (Litchman and Klausmeier 2008). Typically, planktonic food webs are characterized by a high degree of redundancy indicating a strong functional similarity among species, with a small proportion of species exerting disproportionally strong effects on lake ecosystem functioning (Woodward 2009). Therefore, an important but not yet fully resolved question in the study of biodiversity and functioning of freshwater ecosystems is not what species are (species identity), but what species do (functional role) (Hillebrand and Matthiessen 2009; Suding et al. 2008). This implies that assemblages of organisms should be characterized based on phenotypic functional traits that respond directly to environmental changes (response traits) and determine the effects on aggregated processes at the community and ecosystem level (effect traits). Several studies have shown how key traits can favor the occurrence of trait holders under defined sets of environmental conditions. This includes blooms of nuisance cyanobacteria, where decades of study have resulted in a fair understanding of how key traits determine the occurrence of the main bloom forming taxa (Dokulil and Teubner 2000). Therefore, lake management needs to explore ways to disrupt the conditions where cyanobacteria benefit from these traits, in order to better control bloom dynamics. Understanding the ecology of harmful species and their key traits proved already successful for their control and management in cases like the hypertrophic Lake Nieuwe Meer in the Netherlands (Visser et al. 1996), where artificial mixing removed the water column stability on which buoyant Microcystis depended. Ecology can provide the right guidelines for controlling nuisance harmful blooms and inform new technological measures.

Cyanobacteria, formerly known as blue-green algae, are the Earth's oldest known oxygenic photoautotrophs, and they have a global distribution (Cheung et al. 2013). Cyanobacteria are prokaryotic organisms that may occur as large colonies or filaments in lakes giving them a greater resistance to grazing (Reynolds 2006). They contain chlorophyll a like all other phytoplankton taxa and specific accessory pigments, such as phycocyanin or phycoerythrin (Glazer 1984), that can help them to adapt to the available light conditions (Tandeau de Marsac 1991). Cyanobacteria have several more ecophysiological adaptations that allow them to dominate aquatic systems under changing environmental conditions (Carey et al. 2012). For example, some cyanobacteria produce gas vesicles (Walsby 1994) that allow them to regulate their buoyancy (Ganf and Oliver 1982; Hudnell and Dortch 2008; Huisman et al. 2006). Under the increased thermally stratified conditions, which are anticipated with global warming, these cyanobacterial taxa might be able to migrate between well-illuminated surface layers and nutrient-rich hypolimnetic waters (Ganf and Oliver 1982), escaping the increasingly nutrient-depleted epilimnia of lakes during extended stratification periods (Livingstone 2003). See, however, Bormans et al. (1999) reporting that no studies provide support for migration to sufficient depths to access nutrients in stratified systems. Cyanobacteria may also take direct advantage of warming because their growth rate will continue to increase with temperature, while the growth rates of many other phytoplankton taxa decline over 20 °C (Litchman et al. 2010; Reynolds 2006). See Lürling et al. (2013), however, who found no significant differences in optimum growth temperatures between cyanobacteria and green algae. Other key traits such as the ability to fix nitrogen (Oliver and Ganf 2000; Reynolds 2006) and to produce dormant cells as a means to survive unfavorable conditions (Bartram and Chorus 2002) can provide cyanobacteria with a competitive advantage over other phytoplankton (Carey et al. 2012; Litchman et al. 2010), especially under increasingly unpredictable future climate conditions (IPCC 2007). See further discussion of these traits when we list the various cyanobacterial functional groups.

Cyanobacteria are responsible for numerous problems with high environmental and socioeconomical impacts. Cyanobacterial blooms, like all high algal crops, have considerable negative effects on aquatic food webs and ecosystem functioning by turning lakes anoxic, inducing fish mortality, reducing diversity and increasing habitat loss (Chorus and Bartram 1999; Havens 2008; Paerl et al. 2011). There is a growing concern that the frequency and magnitude of cyanobacterial blooms will be enhanced globally (Huber et al. 2012; Jöhnk et al. 2008) and that the geographic range of some cyanobacterial species will expand (Briand et al. 2004; Ryan et al. 2004; Sinha et al. 2012). Many cyanobacterial taxa produce various types of toxins, such as hepatotoxins, neurotoxins and dermatotoxins (Carmichael 1997). These toxins present a considerable risk to drinking water (Codd et al. 2005) and pose a substantial economic cost (Dodds et al. 2009; Steffensen 2008). The World Health Organization (WHO 1996) has been gathering information on toxic cyanobacteria including issues of human health, safe water practices, management, prevention and remediation. Toxic water blooms in many eutrophic to hypereutrophic freshwater ecosystems (sometimes also in low nutrient levels) worldwide are responsible for sporadic but recurrent episodes of wild and domestic animal illness and death as well as for occasional human poisonings from municipal and recreational water supplies (Carmichael 2001). Toxic and non-toxic strains can coexist within a population (Kurmayer et al. 2002; Kardinaal et al. 2007; Briand et al. 2008). Experiments with Microcystis showed that increased temperatures independently and in interaction with elevated phosphorus concentrations can yield more toxic strains of Microcystis than non-toxic ones suggesting that nutrient loading and global warming may additively promote blooms with higher microcystin content (Davis et al. 2009). However, previous experiments with Planktothrix agardhii have shown that increased temperatures do not result in an increase in toxin production, while high nitrogen concentrations affected the increase in both toxic and non-toxic strains (Sivonen 1990). In these experiments though, the interaction between temperature and nutrients was not tested.

Although cyanobacteria share certain key traits, they are a heterogeneous group, in which each taxon possesses different ecophysiological adaptations. For example, the size range of a cyanobacterial unit spans nearly five orders of magnitude (from the smallest single-celled cyanobacterial picoplankton to the multi-cellular filaments and colonies); the photosynthetic rates and growth rates of different taxa vary widely (Reynolds et al. 2002). Different combinations of these morphological and physiological traits might result in varied responses to changes in temperature and nutrients. Plasticity of these traits arises from differential gene expression within species and results in higher diversity of trait values (Litchman et al. 2010). Plastic responses reflected in morphological traits occur on a range of timescales from sub-second (e.g., re-arrangements in the light-harvesting antenna) to days (e.g., augmentation of the gas vesicle volume) in response to the prevailing environmental situation (Ibelings et al. 1994; de Tezanos Pinto and Litchman 2010a). While phenotypic plasticity can be advantageous under changing environmental conditions, it is dependent on a match between timescales of environmental fluctuations and plastic responses (Stomp et al. 2008). Thus, if an environmental disturbance occurs too fast, the cyanobacterial species present at the moment might not be as quick as necessary to adapt to the new conditions. This functional plasticity and its variation between cyanobacterial taxa have to be taken into account if we want to properly understand their behavior and achieve an effective control of toxic cyanobacterial blooms.

Functional classification of cyanobacteria

Trait-based approaches are gaining in popularity, given their opportunity to explain and predict group distributions along environmental gradients. It has been argued that patterns of functional diversity may afford better insights into processes of ecosystem change and responses (Reiss et al. 2009). Recognizing how key traits can enable the proliferation of certain species or functional groups can help to predict which cyanobacterial taxa are most probably going to increase disproportionally under eutrophication and global warming (Litchman et al. 2010). Trait-based phytoplankton ecology arguably started with Margalef (1978) (however also read Lund, 1965) who predicted the occurrence of different taxonomic/functional groups of phytoplankton along gradients of nutrients and turbulence. A more detailed trait-based approach was developed by Reynolds (2002, 2006) who catalogued the natural assemblages of phytoplankton species, of contrasted water bodies with specific trophic status, according to their morphological and physiological key traits. Litchman and Klausmeier (2008) proposed the use of ecologically relevant traits of phytoplankton (morphology/cell size, temperature and reproductionrelated traits) as fundamental data to be integrated in mathematical models for predicting community structure under changing environmental conditions. Kruk et al. (2011) tested a morphology-based functional groups classification against environmental conditions, for a large dataset (211 lakes) covering a wide range of conditions, while Fraisse et al. (2013) explained differences in phytoplankton community structure in rivers using a morphofunctional approach. Carey et al. (2012) gathered information about specific ecophysiological key cyanobacterial traits (ability to grow in higher temperatures; buoyancy; high affinity for and ability to store phosphorus; nitrogen fixation; akinete production; efficient light harvesting) that are predicted to enhance cyanobacterial dominance under future climate scenarios. In this paper, we will use Reynolds (2006) functional classification, which is an update to the Reynolds et al. (2002) publication, specifically for the cyanobacterial taxa in there. We will focus on five selected functional groups that include taxa of concern such as P. agardhii, Cylindrospermopsis raciborskii, Anabaena flos-aquae (now Dolichospermum flosaquae; Wacklin et al. 2009), Microcystis aeruginosa and Planktothrix rubescens, and show how trait-based cyanobacterial ecology can be used for our benefit in the management of these harmful blooms.

Reynolds (2006) treated his phytoplankton speciescounts as phytosociological relevés (small areas of uniform vegetation according to Tüxen 1955) and diagnosed the species that were co-occurring frequently, rarely or not at all. The different clusters were categorized in an alphanumeric system that conveniently suits the patterns and periodic sequences of the natural assemblages in a given lake. Reynolds' Table (2006) (presented in a shortened and revised version in Table 1) enlisted the typical representatives of each particular phytoplankton association, which demonstrate similar morphologies and the habitat type where they are typically encountered. Additionally, the environmental tolerances and the related key functional traits of each group are enlisted, providing an idea on what to expect in response to environmental change. For the purpose of this paper, the table identifies the environmental sensitivities of each group, which can in theory be exploited to compose dedicated management solutions (referred to by us as the "Achilles heel" of each group). In Table 1, we enlist only eight out of the 31 groups of Reynolds' phytoplankton associations (those containing the major bloom forming taxa), from which several were combined to form three groups (S1/S2; H1/H2; L0/ L_{M} ; additionally, we retain S_{N} and R as separate groups. We combined the aforementioned associations into three groups, based upon the conviction that from a management/control point of view these can be treated as homogeneous groups, with comparable environmental sensitivities. Further down we discuss the five groups in Table 1 by choosing one main representative per group, akin a holotype in taxonomy, that is of specific interest from an ecological and management point of view. For each group, we first discuss its key traits, then how the use of these key traits may be altered by environmental (climate) change, then whether and how understanding the key traits opens up opportunities for the management and control of the functional group in question and finally which—combination—of control measures may actually be most likely to successful.

S₁/S₂: Turbid mixed layers—*Planktothrix agardhii*

Key traits

The representative species in S_1/S_2 (Table 1) are efficient light harvesters that have the ability to grow successfully under light-limited conditions by adjusting their pigment composition effectively in response to the ambient irradiance (Fietz and Nicklish 2002). In this paper, we focus on P. agardhii that dominates very turbid lakes. The populations of P. agardhii can be maintained throughout the year when the winter temperatures are mild (Sas and Ahlgren 1989). Filamentous cyanobacterial species such as P. agardhii are superior in light harvesting compared to larger colonial species, mostly due to morphological adaptations such as their slender, attenuated form (Reynolds 2006). P. agardhii requires little energy to maintain cell function and structure; thus, it is capable of maintaining its biomass under light-limiting conditions (Van Liere and Mur 1979). P. agardhii may produce microcystins (e.g., Tonk et al. 2005).

Key traits under eutrophication and climate change

Climate warming can favor the dominance of *P. agardhii* since experiments have shown that its growth can increase steeply in a temperature range from 10 to 30 °C (Foy et al. 1976). Eutrophication, on the other hand, stimulates dense blooms that reduce light availability in the water column and support *P. agardhii* populations (Carey et al. 2012). In highly enriched shallow waters, *P. agardhii* can outcompete other phytoplankton species—including other cyanobacteria—by diminishing the opportunities for

Group	Habitat	Typical representatives	Tolerances	Key traits	Sensitivities/management action
S_1/S_2 (non- N ₂ -fixing filaments)	Deep/shallow Turbid mixed layers	Planktothrix agardhii Arthrospira	Light deficiency	Efficient light harvesting	Short residence time/ flushing
S_N (filaments)	Warm mixed layers	Cylindrospermopsis raciborskii Anabaena minutissima	Light, N deficiency	N ₂ fixation Optimal growth rate at elevated temperatures Resting cells	Short residence time/ flushing
H ₁ /H ₂ (filaments)	Small/large Mesotrophic lakes	Dolichospermum flos-aquae Gleotrichia echinulata	N, C deficiency	N ₂ fixation Buoyancy Resting cells	Light deficiency/artifical mixing P deficiency/control of phosphorus loading
L _O /L _M (colony)	Mesotrophic/ eutrophic Summer epilimnia	Woronichinia Microcystis	C deficiency Stratification	Buoyancy	Light deficiency/artificial mixing
R (non-N ₂ - fixing filaments)	Metalimnia of mesotrophic stratified	Planktothrix rubescens P.mougeotii	Light deficiency Strong segregation of light and	Buoyancy Efficient light harvesting	Physical instability/artificial mixing, stringent nutrient control

Table 1 Different cyanobacterial functional groups, with their preferable habitats, typical representatives, tolerances, key traits, sensitivities and the possible management actions, are displayed

In the S_1/S_2 , H_1/H_2 and L_0/L_M combined groups, several habitat types and typical representatives are enlisted to facilitate the association of each group to certain key traits and possible management actions. The highlighted typical representative in each group is used further in the text to describe how certain key traits are affected by environmental changes and propose certain solutions for the management of their occurrence

nutrients

light harvesting for those which are less well adapted to light deficiency (Reynolds 2006). Under nutrient-rich conditions, the excess of *P. agardhii* biomass promotes the conditions for its own growth by creating appropriate light-limiting conditions (Reynolds 2006; Scheffer et al. 1997). The occurrence of P. agardhii blooms as an exponent of eutrophication in shallow lake ecosystems follows the concept of alternative stable states (Scheffer et al. 1997). At low P concentrations, the dominance of P. agardhii is absent, since there are insufficient nutrients to create the turbid conditions on which this cyanobacterium depends. With ongoing eutrophication, the conditions for P. agardhii gradually improve, until at a critical P value the mixed phytoplankton community collapses and only P. agardhii remains. The water is now so turbid that only P. agardhii can maintain growth and sustain its population. Upon lake restoration (reduction in P loading), initially nothing will happen, since also the turbid state is stable and maintains its own prolongation, helped for instance by high biomass of benthivorous fish species, like bream which stir up the sediment, thereby increasing the turbidity too (Lammens et al. 2004; Roozen et al. 2003). A long period of hysteresis may follow (Ibelings et al. 2007). At some point during the restoration process, however, the P concentration becomes too low to support a high *P. agardhii* biomass, in particular when external factors like cold winters break up the bloom (Berger 1975; Scheffer et al. 1997) or restoration measures like biomanipulation are carried out. At this point, the underwater light climate has improved sufficiently to allow a return to a mixed phytoplankton community and *P. agardhii* will disappear. *P. agardhii* has a tendency to show biomodality in its distribution, either it is dominant or it is nearly absent, a hallmark of alternative stable states (Scheffer et al. 1997).

Management actions

Controlling the nutrient input, thus improving the lake's light environment by diminishing turbidity, is the most fundamental solution to avoid the occurrence of filamentous cyanobacteria with efficient lightharvesting abilities (Van Liere and Walsby 1982). However, several attempts to restore the original phytoplankton community in shallow lakes by reducing the nutrient loading were not always successful, in particular in the short term (Sas and Ahlgren 1989). Increased flushing rates could be effective for the management of P. agardhii. Observations from 27 case studies of lakes undergoing restoration (reduction in P loading) in Denmark demonstrated that in cases where the hydraulic retention time was long, lakes recovered more slowly from cyanobacterial dominance than lakes with low retention times (<0.5 years), where the recovery was 10-300 times faster (Jeppesen et al. 1991). This can be explained by the fact that the maximum growth rate of the filamentous P. agardhii (as in many cases of several cyanobacterial taxa) is typically ca. only half of the maximum growth rate of green algae ($r = 0.6 \text{ day}^{-1} \text{ vs. } r = 1.2 \text{ day}^{-1}$, respectively, calculated by Scheffer et al. 1997 based on Mur et al. 1977). Consequently, longer retention time favors P. agardhii, while short retention time may remove it from the system, as P. agardhii no longer has the capacity to outcompete fast-growing green algae. Scheffer et al. (1997) prediction model confirmed that P. agardhii would be absent independently of the nutrient level if the flushing rate was ≈ 18 % of the lake volume per day. Increased flushing rates along with increased water clarity can reduce hysteresis of lakes dominated by P. agardhii and lead to a faster return to a mixed phytoplankton community. Crucial for the long-term restoration of lakes formerly dominated by P. agardhii is the return of an abundant macrophyte community which competes for nutrients with the cyanobacterium, prevents sediment re-suspension and provides shelter for large-bodied zooplankton against fish predation (Gulati et al. 2008).

S_N: Warm mixed layers—*Cylindrospermopsis* raciborskii

Key traits

According to Reynolds classification (Reynolds 2006), the main representatives of the S_N group proliferate in warm mixed layers. In this paper, we will focus mainly on *C. raciborskii* whose occurrence is expanding. *C. raciborskii* blooms can negatively

affect water quality since they can produce toxins such as the neurotoxin saxitoxin (Castro et al. 2004) and the alkaloid hepatotoxin, cylindrospermopsin (Li et al. 2001). The C. raciborskii distribution range used to be restricted to tropical and subtropical ecosystems, but since the 1930s, it has progressively colonized more temperate environments, spreading from Greece and Hungary toward higher latitudes by the end of the twentieth century (Padisák 1997). These expanding C. raciborskii strains have been reported as not toxic (Fastner et al. 2003; Yilmaz et al. 2008). However, a recent study by Gkelis and Zaoutsos (2014) revealed saxitoxin production in freshwater samples from Greece dominated by C. raciborskii. The expansion of C. raciborskii is supported by the existence of different physiological strains or ecotypes, which vary in their temperature tolerance with tropical strains to be tolerant in a temperature range of 15 to 35 °C (Chonudomkul et al. 2004).

C. raciborskii occurs mainly in shallow eutrophic waters with relatively high turbidity (Reynolds 2006). It holds common phenotypic traits with P. agardhii such as tolerance to continuous mixing and shaded water columns (Stüken et al. 2006). Another similarity is its high affinity for and ability to store phosphorous under limiting conditions (Istvanovics et al. 2000) that allows C. raciborskii to proliferate in warm surface mixed layers of stratified systems (Bormans et al. 2004). These shared key traits between P. agardhii and C. raciborskii, which are also depicted in their similar morphological characteristics, indicate functional equivalence and a probability of occupying comparable niches (Kruk et al. 2010). For example, in eutrophic lakes in Eastern Germany, cyanobacteria assemblages consist mainly of members of the S_1 group (Reynolds 2006), but are often accompanied by C. raciborskii (Nixdorf et al. 2003). There are specific details, however, which differentiate the niches in which each of these cyanobacterial species proliferate, for example, C. raciborskii, has higher light requirements for growth than P. agardhii (Briand et al. 2004). Additionally, C. raciborskii, as a Nostocales genus, has the capacity to fix atmospheric nitrogen in its heterocysts (Komárek and Mareš 2012).

Key traits under eutrophication and climate change

Experiments under different temperature regimes showed a positive net growth in a wide range of

temperatures from 15 to 35 °C and light intensities ranging from 30 to 400 μ mol photons m⁻² s⁻¹ of strains of both tropical and temperate origin (Briand et al. 2004). The colonization of C. raciborskii of midlatitudinal environments hence may be a result of a high adaptability to a range of light and temperature gradients under perpetual environmental change induced by eutrophication and global warming (Briand et al. 2004). The characteristic of N_2 fixation promotes its competitive strength under nitrogendepleted environments, enabling it to effectively compete against other species like P. agardhii that is not able to fix nitrogen (Anagnostidis and Komárek 1988). Experiments, however, showed that C. raciborskii is capable of dominating over a mixed phytoplankton community of green algae and other cyanobacteria species regardless of N/P stoichiometry (Chislock et al. 2014), which reduces the capability to control its occurrence under eutrophic conditions in freshwater ecosystems. Moreover, C. raciborskii has the advantage of forming akinetes (Kaplan-Levy et al. 2010) as an effective means to survive unfavorable periods such as changes in light, nutrients, temperature or desiccation. These akinetes can also capitalize on the excess availability of P in the sediment upon germination (Istvanovics et al. 2000).

Management actions

Controlling the nutrient load of both N and P may be necessary to control the blooms of this harmful cyanobacterium (Chislock et al. 2014). Additionally and similar to the previous group (S_1/S_2), Reynolds (2006) suggests that problems with *C. raciborskii* can be solved if the retention time in basins is shortened sufficiently under increased flushing rates. In a large impounded tropical river, Bormans et al. (2005) showed that flushing was the only management option to terminate blooms of *C. raciborskii*.

H₁/H₂: Dinitrogen-fixing nostocaleans— Dolichospermum flos-aquae (formerly Anabaena flos-aquae)

Key traits

In the H_1/H_2 group, we have dinitrogen-fixing nostocaleans that occur mainly in small or large mesotrophic lakes (Table 1). Here, we will focus on *D. flos-aquae* in order to show how the key traits that characterize the H_1/H_2 group support their tolerance to limiting nitrogen availability and how we can exploit their sensitivities to eliminate their presence in freshwater ecosystems. *D. flos-aquae* is a filamentous cyanobacterium that can be highly toxic when present in water resources (Agnihotri 2013) producing harmful neurotoxins such as anatoxin-a, anatoxin-a(s) and homoanatoxin-a (Gallon et al. 1990). *D. flos-aquae* is a good competitor under low nitrogen supplies as it can fix nitrogen while it can escape poor light conditions by regulating its buoyancy (Kinsman et al. 1991; Reynolds 2006).

Nitrogen fixation is an energetically costly process that depends on light as an energy source. The rates of N₂ fixation increase with increasing light levels (Agawin et al. 2007). Low N/P ratios stimulate heterocyst development (Agawin et al. 2007), resulting in an increase in the N-fixers' biomass either in mesocosm experiments (Levine and Schindler 1999; Vrede et al. 2009) or in whole-lake experiments (Schindler et al. 2008). Experimental studies also point out that promotion of nitrogen fixers occurs only when sufficient light is provided (de Tezanos Pinto and Litchman 2010a). Light regime has a primary significance for the success of D. flos-aquae as under low photon fluxes N/P ratios appear not to have a significant effect on its abundance (de Tezanos Pinto and Litchman 2010a). Under low light intensities, which stimulate gas vesicle production (Kinsman et al. 1991), D. flos-aquae may take advantage of its gas vesicles to regulate its buoyancy, float up and in this way may manage to enhance access to light and sustain its biomass (Carey et al. 2012).

Key traits under eutrophication and climate change

Anticipated changes in the environment can stimulate the occurrence of *D. flos-aquae* in different ways. Increased temperatures induce stronger stratification and shallower mixing depths, resulting in increased light availability for floating cyanobacteria like *D. flos-aquae*, which would benefit N₂ fixation under low N availability. High temperatures also have a direct effect on optimizing N₂ fixation by enhancing the rate of gas diffusion into the heterocyst (Bauersachs et al. 2014). Staal et al. (2003) found that Q10 (acceleration of a process over a 10 °C step, generally 10–20 °C) values for N₂ fixation of heterocystous strains of *Anabaena sp.* were approximately equal to cyanobacterial growth rate responses to temperature (Q10 \geq 1.8, Reynolds 2006).

Management actions

A potential remedy against blooms of N2-fixing cyanobacteria would be to reduce light availability. Deep mixing can have a negative effect on the growth of D. flos-aquae and its capacity to fix nitrogen, as it forces the population down to poorly illuminated waters with low energy supply. In general, N₂ fixation is also restrained through the availability of phosphorus and micronutrients (molybdenum, iron) as the N₂fixing enzyme, nitrogenase, requires high levels of these nutrients to operate (Vitousek et al. 2002). Hence, for several N₂ fixers, Schindler (1977) puts forward that P is the ultimate limiting factor in freshwater ecosystems of moderate to high productivity, so that control of P could effectively control H_1/H_2 (see, however, Chislock et al. 2014 who showed that stoichiometric manipulation of N and P will not be enough as experiments on C. raciborskii showed its dominance under high or low N/P ratios). Reynolds (2006) in this case also points out that the H_1/H_2 group does not perform well under low phosphorus concentrations.

Lo/L_M: Summer epilimnia—*Microcystis* aeruginosa

Key traits

Lo/L_M are encountered when lake summer epilimnia are formed in mesotrophic and eutrophic lakes (Table 1). Our main concern here will be the genus *Microcystis*. *Microcystis* is a cosmopolitan genus which can generate dense cell accumulations (scums) on the surface of freshwater ecosystems, threatening human and ecosystem health (Visser et al. 2005). Several strains of *Microcystis* spp. are toxigenic producing a group of cyclic peptide hepatotoxins called microcystins and consist a risk for both public health and ecosystems (Carmichael 1997). *Microcystis* spp. forms large colonies that put it at a disadvantage under nutrient limited conditions as its low surface area-to-volume ratio decelerates nutrient uptake rates (Reynolds 1997). Large colonies will move faster compared with smaller colonies or single filaments, following Stokes' law (Smayda 1971). Microcystis has been referred to as the champion of buoyancy regulation. Microcystis efficiently regulates its buoyancy by varying its dense cellular constituents (mainly sugars) on a diel timescale, and buoyant colonies float up rapidly during periods of (partial) water column stability (Ibelings et al. 1991). Therefore, forming large colonies that can float fast is a key advantage for Microcystis (Ibelings et al. 1991). In experiments, *M. aeruginosa* was able to migrate 12 m within 2 h to gain access to both light and nutrients, defeating any density barriers (Ganf and Oliver 1982). Observations in the Dutch Lake Vinkeveen have demonstrated (Ibelings et al. 1991) that Microcystis spp. fast flotation velocity (Walsby 1991) allows it to efficiently track the near-surface mixed layer (Humphries and Lyne 1988) floating up until it reaches the well-mixed layer near the lake surface where even large colonies will remain entrained in turbulent flow.

Key traits under eutrophication and climate change

Increased temperatures are positively related to M. aeruginosa's growth rate as it reaches a maximum replication rate at 28 °C while its Q10-here the increase in growth rate with a 10 °C increase in temperature-reputedly is close to 9.6, one of the highest values recorded in prokaryotic and eukaryotic phytoplankton species (Reynolds 2006; Carey et al. 2012). However, Lürling et al. (2013) demonstrated experimentally that the optimum growth temperature for several cyanobacterial and green algal taxa was not significantly different. Although Lürling et al. (2013) suggest that M. aeruginosa might not benefit from higher water temperatures, Paerl and Huisman (2009) do support the notion that M. aeruginosa's blooms will be promoted albeit through the indirect effects of climate warming. Increased temperatures lead to intensified density differences enhancing lake stratification. When the stratification is well developed, shallow wind-induced mixing in the surface layer cannot be transferred to deeper layers, denying other phytoplankton the opportunity to become entrained in turbulent flow. Under stable stratification and reduced turbulence, M. aeruginosa colonies will be larger and their buoyancy will be further enhanced during vertical migration cycles (O'Brien et al. 2004).

Consequently, *M. aeruginosa's* ability to efficiently control its movement offers a clear advantage over non-migrating phytoplankton species under the expected prolonged and enhanced stratification (Carey et al. 2012; Paerl and Huisman 2009).

Management actions

Reynolds (2006) proposes a prolonged deep mixing regime in order to control the occurrence of M. aeruginosa (Table 1). The management of Microcystis in the Dutch Lake Nieuwe Meer showed that artificial mixing using bubble plumes can effectively prevent nuisance blooms and promote a mixed community of flagellates, green algae and diatoms (Visser et al. 1996). Non-buoyant algae were favored since their sedimentation rates were reduced due to the artificial mixing, while the turbulent flow diminished the advantage of M. aeruginosa to float up into the illuminated near-surface mixed layer (Visser et al. 1996). Under mixing, cells face rapid changes in light regime and competition leans toward those eukaryotic competitors that are better adapted to fluctuating irradiance levels compared to M. aeruginosa (Ibelings et al. 1994).

R: Metalimnia in stratified lakes—*Planktothrix rubescens*

Key traits

The representatives of group \mathbf{R} (Table 1) are encountered in the metalimnion of mesotrophic, stratified lakes (Reynolds 2006). P. rubescens is a filamentous cyanobacterium that may produce the hepatotoxin microcystin (Anagnostidis and Komárek 1988). It contains gas vesicles that allow it to adjust its buoyancy and regulate its position in the water column. In contrast to Microcystis, however, it positions its populations away from the saturated irradiance levels at the lake surface (Jacquet et al. 2005). It belongs to the same genus as P. agardhii from the S_1/S_2 group, and both species are able to tolerate low light intensities as both share the same slender filamentous form and auxiliary pigmentation (Reynolds 1997). Nevertheless, the niches of the two congeners are clearly different. Whereas P. agardhii prefers a shady (turbid) environment, P. rubescens requires a relatively clear epilimnion. *P. rubescens* forms metalimnetic layers in deep, stratified mesoeutrophic lakes where it can completely monopolize resources and dominate the phytoplankton community (Anneville et al. 2004). At metalimnetic depths, the light quantity and light quality are different compared to the upper mixed layers and *P. rubescens* is well adapted to these light conditions due to the abundance of the red pigment phycoerythrin.

The morphology of gas vesicles has evolved to withstand the combined pressures in the lake environment (Walsby 1994). Strains of P. rubescens have narrow gas vesicles (diameter = 50 nm) with critical collapse pressure (pc) ≈ 1.1 MPa compared to M. aeruginosa (d = 65 nm)pc ≈ 0.85 MPa) as response to withstand increased hydrostatic pressure that comes with increased depth (Visser et al. 2005). The P. rubescens gas vesicles will survive during deep winter mixing, and cells maintain their buoyancy (Carey et al. 2012). Moreover, following Stokes' law in non-turbulent conditions, filaments with a small diameter will have a low sinking or flotation velocity, compared to large colony-forming taxa (Walsby 2005) and will find it easier to maintain position in the metalimnion. The intimate interactions between carbohydrate accumulation and gas vesicle-mediated buoyancy will support close-to-neutral buoyancy and will maintain filaments at a given depth (Walsby et al. 2004). Overall, P. rubescens is able to adjust the depth of its stratifying population in response to changes in the underwater light regime and will take up different positions in the metalimnion if this is to its advantage (Jacquet et al. 2005).

Key traits under eutrophication and climate change

P. rubescens is present in high biomass during summer, when the phosphorus concentrations are very low in the epilimnion, but still remain available in deeper layers of lakes (Anneville et al. 2002). *P. rubescens* can even keep developing during autumn when there is lower irradiance due to increased mixing, forcing the thermocline to greater depth (Anneville et al. 2004). *P. rubescens* can be successful under different trophic states of the same lake; this has been observed in Lake Zurich (Switzerland) where *P. rubescens* has been found in phytoplankton assemblages during both the eutrophic and the meso-eutrophic stages of the lake (Anneville et al. 2004).

In Lake Bourget (France), however, the long-term dynamics show a different pattern. *P. rubescens* was absent prior to eutrophication in the 1950s, while also the very nutrient-rich conditions during the peak of the eutrophication prevented proliferation of this filamentous species. *P. rubescens* came to dominate Lake Bourget in the mid-1990s when lake restoration measures began to take hold. Lake restoration has been successful, even to the point that since 2009 *P. rubescens* once more is absent from the lake. Hence, the relationship between trophic state and proliferation of *P. rubescens* follows a hump-shaped pattern, where the peak of the hump is positioned at mesotrophic to meso-eutrophic conditions (Dokulil and Teubner 2012).

Recent studies show the importance of the seasonal effects of climate change on the deep, meso-eutrophic lakes in the peri-alpine region where P. rubescens typically dominates (Anneville et al. 2015; Posch et al. 2012). Contrary to the common belief that blooms prefer hot summers (Jöhnk et al. 2008), extremely warm summers (like the one in 2003) do not promote blooms of P. rubescens, and they may reduce its population size. Apparently, P. rubescens does not benefit from extremely long and stable stratification during heat wave summers, despite the ongoing depletion of epilimnetic nutrient concentrations. Warm conditions in autumn and winter, however, do promote P. rubescens in the following year, indicating the vital role of carrying on a large enough inoculum for growth through the winter to the next spring period.

Management actions

The return of *P. rubescens* in Lake Bourget in the mid-1990s upon a return to mesotrophic conditions is attributed to increased transparency following the restriction of other phytoplankton species under increasingly reduced phosphorus availability (Jacquet et al. 2005). Ernst et al. (2009) suggested that the effects of climate change strengthen the dominance of *P. rubescens* in peri-alpine lakes that undergo reoligotrophication, because this cyanobacterium can take advantage of the earlier onset of stratification. Reduced mixing in warmer winters (incomplete overturn of the water column) will stimulate survival of intact gas vesicles in *P. rubescens* (Walsby et al. 1998), aiding to achieve a large size of the inoculum. For these reasons, Reynolds suggested to alter the stability of the water column as an effective means to disrupt the development of *P. rubescens* (Reynolds 2006).

Lake managers worldwide, for instance in the Netherlands, are aware of the increasing risks of toxic cyanobacteria during heat waves, resulting in lakes being closed for recreation on an extensive scale (Ibelings et al. 2014). In contrast to the peri-alpine region, however, lakes in the Netherlands tend to be dominated by cyanobacteria from the $\#\#L_O/L_M$ group. These "subtle" differences between effects of warming in summer versus winter upon different functional groups—**R** versus L_O/L_M —underline once more the necessity to study and control cyanobacterial blooms upon the right—ecological—basis. Cyanobacteria with different key traits will respond differently to changes in the environment and require a different management approach.

Conclusions and research perspectives

Cyanobacterial blooms are still a major problem in freshwater ecosystems in spite of major nutrient reductions over the last 40 years. Lake restoration has had mixed success in controlling their development. The anticipated changes of our climate have the capacity to worsen the problem, demanding appropriate and timely measures from lake managers. In this review, we demonstrate how management actions should be tailored depending on the dominant cyanobacterial functional group. Identifying the key traits of dominant cyanobacteria and the environmental conditions under which these traits succeed can reveal the Achilles heel of cyanobacterial functional groups and provide lake managers with the most appropriate tools for their effective management and control. A comprehensive view of the many adaptive strategies of cyanobacteria to a wide range of environmental conditions is essential as controlling one species has also the potential to let another (may be even more problematic) species proliferate. This knowledge on the cyanobacterial key traits should be more incorporated in various models developed and used to predict bloom occurrences. Yet, crucially all taxa ultimately can and should be controlled by managing the external and internal nutrient loading of lakes. This special issue in Aquatic Ecology presents the state of the art in cyanobacterial bloom management and control, continuously keeping the key ecophysiological traits of the culprits in mind.

Acknowledgments We would like to acknowledge two EU COST Actions; ES 1105 "CYANOCOST—Cyanobacterial blooms and toxins in water resources: Occurrence, impacts and management" and ES1201 "NETLAKE—Networking Lake Observatories in Europe" that offer us the possibility to develop the idea of this manuscript through numerous discussions with experts and researchers on cyanobacterial blooms. We would also like to thank all the anonymous reviewers for helping us improve our manuscript with all their constructive comments.

References

- Agawin N, Rabouille S, Veldhuis M, Servatius L, Hol S, van Overzee HM, Huisman J (2007) Competition and facilitation between unicellular nitrogen-fixing cyanobacteria and non-nitrogen-fixing phytoplankton species. Limnol Oceanogr 52:2233–2248
- Agnihotri VK (2013) Anabaena flos-aquae. Crit Rev Environ Sci Technol 44:1995–2037
- Anagnostidis K, Komárek J (1988) Modern approach to the classification system of cyanophytes. 3-Oscillatoriales. Algol Stud/Archiv für Hydrobiologie, Supplement Volumes 50–53, pp 327–472
- Anneville O, Souissi S, Ibanez F, Ginot V, Druart JC, Angeli N (2002) Temporal mapping of phytoplankton assemblages in Lake Geneva: annual and interannual changes in their patterns of succession. Limnol Oceanogr 47:1355–1366
- Anneville O, Souissi S, Gammeter S, Straile D (2004) Seasonal and inter-annual scales of variability in phytoplankton assemblages: comparison of phytoplankton dynamics in three peri-alpine lakes over a period of 28 years. Freshw Biol 49:98–115
- Anneville O, Molinero JC, Souissi S, Gerdeaux D (2010) Seasonal and interannual variability of cladoceran communities in two peri-alpine lakes: uncoupled response to the 2003 heat wave. J Plankton Res 32:913–925
- Anneville O, Domaizon I, Kerimoglu O, Rimet F, Jacquet S (2015) Blue-green algae in a "greenhouse century"? New insights from field data on climate change impacts on cyanobacteria abundance. Ecosystems 18(3):441–458
- Bartram J, Chorus I (eds) (2002) Toxic Cyanobacteria in water: a guide to their public health consequences, monitoring and management. CRC Press
- Bauersachs T, Stal LJ, Grego M, Schwark L (2014) Temperature induced changes in the heterocyst glycolipid composition of N-2 fixing heterocystous cyanobacteria. Org Geochem 69:98–105
- Berger C (1975) Occurrence of Oscillatoria agardhii Gom. in some shallow eutrophic lakes. Verh Int Ver Theor Angew Limnol 19:2689–2697
- Bormans M, Condie S (1998) Modelling the distribution of Anabaena and Melosira in a stratified river weir pool. Hydrobiologia 364:3–13
- Bormans M, Sherman BS, Webster IT (1999) Is buoyancy regulation in cyanobacteria an adaptation to exploit separation of light and nutrients? Mar Freshw Res 50:897–906

- Bormans M, Ford PW, Fabbro L, Hancock G (2004) Onset and persistence of cyanobacterial blooms in a large impounded tropical river, Australia. Mar Freshw Res 55:1–15
- Bormans M, Ford PW, Fabbro L (2005) Spatial and temporal variability in cyanobacterial populations controlled by physical processes. J Plankton Res 27(1):61–70
- Briand JF, Leboulanger C, Humbert JF, Bernard C, Dufour P (2004) Cylindrospermopsis raciborskii (Cyanobacteria) invasion at mid-latitudes: selection, wide physiological tolerance, or global warming? J Phycol 40:231–238
- Briand E et al (2008) Temporal variations in the dynamics of potentially microcystin-producing strains in a bloomforming *Planktothrix agardhii* (Cyanobacterium) population. Appl Environ Microbiol 74(12):3839–3848
- Butterwick C, Heaney SI, Talling JF (2005) Diversity in the influence of temperature on the growth rates of freshwater algae, and its ecological relevance. Freshw Biol 50:291–300
- Cardinale BJ et al (2011) The functional role of producer diversity in ecosystems. Am J Bot 98:572–592
- Carey CC, Ibelings BW, Hoffmann EP, Hamilton DP, Brookes JD (2012) Eco-physiological adaptations that favour freshwater cyanobacteria in a changing climate. Water Res 46:1394–1407
- Carmichael W (1997) The cyanotoxins. Adv Bot Res 27: 211–256
- Carmichael WW (2001) Health effects of toxin-producing cyanobacteria: "The CyanoHABs". Hum Ecol Risk Assess 7:1393–1407
- Castro D, Vera D, Lagos N, García C, Vásquez M (2004) The effect of temperature on growth and production of paralytic shellfish poisoning toxins by the cyanobacterium *Cylindrospermopsis raciborskii*. Toxicon 44:483–489
- Cheung MY, Liang S, Lee J (2013) Toxin-producing cyanobacteria in freshwater: a review of the problems, impact on drinking water safety, and efforts for protecting public health. J Microbiol 51:1–10
- Chislock MF, Sharp KL, Wilson AE (2014) *Cylindrospermopsis* raciborskii dominates under very low and high nitrogen-tophosphorus ratios. Water Res 49:207–214
- Chonudomkul D, Yongmanitchai W, Theeragool G, Kawachi M, Kasai F, Kaya K, Watanabe MM (2004) Morphology, genetic diversity, temperature tolerance and toxicity of *Cylindrospermopsis raciborskii* (Nostocales, Cyanobacteria) strains from Thailand and Japan. FEMS Microbiol Ecol 48:345–355
- Chorus I, Bartram J (1999) Toxic cyanobacteria in water: a guide to their public health consequences, monitoring and management. Spon Press, New York
- Codd GA, Morrison LF, Metcalf JS (2005) Cyanobacterial toxins: risk management for health protection. Toxicol Appl Pharmacol 203:264–272
- Davis TW, Berry DL, Boyer GL, Gobler CJ (2009) The effects of temperature and nutrients on the growth and dynamics of toxic and non-toxic strains of Microcystis during cyanobacteria blooms. Harmful Algae 8:715–725
- de Tezanos Pinto P, Litchman E (2010a) Eco-physiological responses of nitrogen-fixing cyanobacteria to light. Hydrobiologia 639:63–68
- Dodds WK et al (2009) Eutrophication of US Freshwaters: analysis of Potential Economic Damages. Environ Sci Technol 43:12–19

- Dokulil MT, Teubner K (2000) Cyanobacterial dominance in lakes. Hydrobiologia 438(1–3):1–12
- Dokulil MT, Teubner K (2012) Deep living *Planktothrix rubescens* modulated by environmental constraints and climate forcing. Hydrobiologia 698:29–46
- Elliott JA (2012) Is the future blue-green? A review of the current model predictions of how climate change could affect pelagic freshwater cyanobacteria. Water Res 46: 1364–1371
- Elliott JA, Thackeray SJ, Huntingford C, Jones RG (2005) Combining a regional climate model with a phytoplankton community model to predict future changes in phytoplankton in lakes. Freshw Biol 50:1404–1411
- Ernst B, Hoeger SJ, O'Brien E, Dietrich DR (2009) Abundance and toxicity of *Planktothrix rubescens* in the pre-alpine Lake Ammersee, Germany. Harmful Algae 8:329–342
- Fastner J et al (2003) Cylindrospermopsin occurrence in two German lakes and preliminary assessment of toxicity and toxin production of *Cylindrospermopsis raciborskii* Cyanobacteria) isolates. Toxicon 42(3):313–321
- Fietz S, Nicklish A (2002) Acclimation of the diatom Stephanodiscus neoastraea and the cyanobacterium Planktothrix agardhii to simulated natural light fluctuations. Photosynth Res 72(1):95–106
- Foy RH, Gibson CE, Smith RV (1976) The influence of day length, light intensity and temperature on the growth rates of planktonic blue-green algae. Br Phycol J 11:151–163
- Fraisse et al (2013) Morphofunctional traits reflect differences in phytoplankton community between rivers of contrasting flow regime. Aquat Ecol 47:315–327
- Gallina N, Anneville O, Beniston M (2011) Impacts of extreme air temperatures on cyanobacteria in five deep peri-Alpine lakes. Limnol Oceanogr 70:186–196
- Gallon JR, Chit KN, Brown EG (1990) Biosynthesis of the tropane-related cyanobacterial toxin anatoxin-a: role of ornithine decarboxylase. Phytochemistry 29:1107–1111
- Ganf GG, Oliver RL (1982) Vertical separation of light and available nutrients as a factor causing replacement of green-algae by blue-green-algae in the plankton of a stratified lake. J Ecol 70:829–844
- Gkelis S, Zaoutsos N (2014) Cyanotoxin occurrence and potentially toxin producing cyanobacteria in freshwaters of Greece: a multi-disciplinary approach. Toxicon 78:1–9
- Glazer AN (1984) Phycobilisome a macromolecular complex optimized for light energy transfer. Biochimica et Biophysica Acta (BBA)-Rev Bioenerg 768:29–51
- Gulati RD, Pires LMD, Van Donk E (2008) Lake restoration studies: failures, bottlenecks and prospects of new ecotechnological measures. Limnol Ecol Manag Inland Waters 38(3):233–247
- Havens KE (2008) Cyanobacteria blooms: effects on aquatic ecosystems. In: Hudnell HK (ed) Cyanobacterial harmful algal blooms: state of the science and research needs, vol 619. Advances in experimental medicine and biology pp 733–747
- Hillebrand H, Matthiessen B (2009) Biodiversity in a complex world: consolidation and progress in functional biodiversity research. Ecol Lett 12:1405–1419
- Huber V, Wagner C, Gerten D, Adrian R (2012) To bloom or not to bloom: contrasting responses of cyanobacteria to recent

heat waves explained by critical thresholds of abiotic drivers. Oecologia 169:245–256

- Hudnell HK, Dortch Q (2008) Chapter 2: a synopsis of research needs identified at the interagency, international symposium on cyanobacterial harmful algal blooms (ISOC-HAB). In: Hudnell HK (ed) Cyanobacterial harmful algal blooms: state of the science and research needs advances in experimental medicine and biology 619:17–43
- Huisman J, Matthijs HC, Visser PM (eds) (2006) Harmful cyanobacteria, vol 3. Springer
- Humphries SE, Lyne VD (1988) Cyanophyte blooms: the role of cell buoyancy. Limnol Oceanogr 33:79–91
- Ibelings BW, Chorus I (2007) Accumulation of cyanobacterial toxins in freshwater "seafood" and its consequences for public health: a review. Environ Pollut 150:177–192
- Ibelings BW, Maberly SC (1998) Photoinhibition and the availability of inorganic carbon restrict photosynthesis by surface blooms of cyanobacteria. Limnol Oceanogr 43:408–419
- Ibelings BW, Mur LR, Kinsman R, Walsby A (1991) Microcystis changes its buoyancy in response to the average irradiance in the surface mixed layer. Archiv für Hydrobiologie 120:385–401
- Ibelings BW, Kroon BMA, Mur LR (1994) Acclimation Of Photosystem-Ii in a cyanobacterium and a eukaryotic green-alga to high and fluctuating photosynthetic photon flux densities, simulating light regimes induced by mixing in lakes. New Phytol 128:407–424
- Ibelings B, Admiraal W, Bijkerk R, Ietswaart T, Prins H (1998) Monitoring of algae in Dutch rivers: does it meet its goals? J Appl Phycol 10:171–181
- Ibelings BW, Vonk M, Los HFJ, van der Molen DT, Mooij WM (2003) Fuzzy modeling of cyanobacterial surface waterblooms: validation with NOAA–AVHRR satellite images. Ecol Appl 13:1456–1472
- Ibelings BW, Portielje R, Lammens EH, Noordhuis R, van den Berg MS, Joosse W, Meijer ML (2007) Resilience of alternative stable states during the recovery of shallow lakes from eutrophication: lake Veluwe as a case study. Ecosystems 10:4–16
- Ibelings BW, Backer LC, Kardinaal WEA, Chorus I (2014) Current approaches to cyanotoxin risk assessment and risk management around the globe. Harmful Algae 40:63–74
- IPCC Climate Change (2007) Impacts, adaptation and vulnerability. Contribution of Working Group II to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change Cambridge University Press
- Istvanovics V, Shafik H, Presing M (2000) Growth and phosphate uptake kinetics of the cyanobacterium *Cylindrospermopsis raciborskii* (Cyanophyceae) in throughflow cultures. Freshw Biol 43:257–275
- Jacquet S et al (2005) The proliferation of the toxic cyanobacterium *Planktothrix rubescens* following restoration of the largest natural French lake (Lac du Bourget). Harmful Algae 4:651–672
- Jeppesen E, Kristensen P, Jensen JP, Søndergaard M, Mortensen E, Lauridsen T (1991) Recovery resilience following a reduction in external phosphorus loading of shallow, eutrophic Danish lakes: duration, regulating factors and methods for overcoming resilience. Mem Ist Ital Idrobiol 48:127–148

- Jeppesen E et al (2005) Lake responses to reduced nutrient loading—an analysis of contemporary long-term data from 35 case studies. Freshw Biol 50:1747–1771
- Jöhnk KD, Huisman J, Sharples J, Sommeijer B, Visser PM, Stroom JM (2008) Summer heatwaves promote blooms of harmful cyanobacteria. Glob Change Biol 14:495–512
- Kaplan-Levy RN, Hadas O, Summers ML, Rücker J, Sukenik A (2010) Akinetes: dormant cells of cyanobacteria. In: Lubzens E, Cerda J, Clark M (eds) Dormancy and resistance in harsh environments. Springer, Berlin, 5–27
- Kardinaal WEA et al. (2007) Microcystis genotype succession in relation to microcystin concentrations in freshwater lakes. Aquat Microb Ecol 48:1–12
- Kinsman R, Ibelings BW, Walsby A (1991) Gas vesicle collapse by turgor pressure and its role in buoyancy regulation by *Anabaena flos-aquae*. J Gen Microbiol 137:1171–1178
- Komárek J, Mareš J (2012) An update to modern taxonomy (2011) of freshwater planktic heterocytous cyanobacteria. Hydrobiologia 698:327–351
- Kosten S et al (2012) Warmer climates boost cyanobacterial dominance in shallow lakes. Glob Change Biol 18: 118–126
- Kruk C et al (2010) A morphological classification capturing functional variation in phytoplankton. Freshw Biol 55:614–627
- Kruk C et al (2011) Phytoplankton community composition can be predicted best in terms of morphological groups. Limnol Oceanogr 56:110–118
- Kurmayer R, Dittmann E, Fastner J, Chorus I (2002) Diversity of microcystin genes within a population of the toxic cyanobacterium *Microcystis* spp. in Lake Wannsee (Berlin, Germany). Microb Ecol 43:107–118
- Lammens EH, van Nes EH, Meijer M-L, van den Berg MS (2004) Effects of commercial fishery on the bream population and the expansion of *Chara aspera* in Lake Veluwe. Ecol Model 177:233–244
- Levine S, Schindler D (1999) Influence of nitrogen to phosphorus supply ratios and physicochemical conditions on cyanobacteria and phytoplankton species composition in the Experimental Lakes Area, Canada. Can J Fish Aquat Sci 56:451–466
- Lewis WM Jr, Wurtsbaugh WA (2008) Control of lacustrine phytoplankton by nutrients: erosion of the phosphorus paradigm. Int Rev Hydrobiol 93:446–465
- Li R et al (2001) Isolation and identification of the cyanotoxin cylindrospermopsin and deoxy-cylindrospermopsin from a Thailand strain of *Cylindrospermopsis raciborskii* (Cyanobacteria). Toxicon 39:973–980
- Litchman E, Klausmeier CA (2008) Trait-based community ecology of phytoplankton. Annu Rev Ecol Evol Syst 39:615–639
- Litchman E, Pinto PD, Klausmeier CA, Thomas MK, Yoshiyama K (2010) Linking traits to species diversity and community structure in phytoplankton. Hydrobiologia 653:15–28
- Livingstone DM (2003) Impact of secular climate change on the thermal structure of a large temperate central European lake. Clim Change 57:205–225
- Lund JWG (1965) The ecology of the freshwater phytoplankton. Biol Rev 40(2):231–290

- Lürling M, Eshetu F, Faassen EJ, Kosten S, Huszar VL (2013) Comparison of cyanobacterial and green algal growth rates at different temperatures. Freshw Biol 58:552–559
- Margalef R (1978) Life-forms of phytoplankton as survival alternatives in an unstable environment. Oceanol Acta 1:493–509
- Moss B et al (2011) Allied attack: climate change and eutrophication. Inland Waters 1:101–105
- Mur LR, Gons H, Van Liere L (1977) Some experiments on the competition between green algae and blue-green bacteria in light-limited environments. FEMS Microbiol Lett 1:335–338
- Nixdorf B, Mischke U, Rücker J (2003) Phytoplankton assemblages and steady state in deep and shallow eutrophic lakes—an approach to differentiate the habitat properties of Oscillatoriales. In: Naselli-Flores L, Padisák J, Dokulil MT (eds) Phytoplankton and equilibrium concept: the ecology of steady-state assemblages. Springer, Netherlands, pp 111–121
- O'Brien KR, Meyer DL, Waite AM, Ivey GN, Hamilton DP (2004) Disaggregation of *Microcystis* aeruginosa colonies under turbulent mixing: laboratory experiments in a gridstirred tank. Hydrobiologia 519:143–152
- Oliver RL, Ganf GG (2000) Freshwater blooms. In: Whitton B and Malcolm P (eds) The Ecology of Cyanobacteria. Kluwer Academic Publishers, pp 149–186
- Padisák J (1997) *Cylindrospermopsis raciborskii* (Woloszynska) Seenayya et Subba Raju, an expanding, highly adaptive cyanobacterium: worldwide distribution and review of its ecology. Archiv Für Hydrobiologie Supplementband Monographische Beitrage 107:563–593
- Paerl HW, Huisman J (2008) Climate—blooms like it hot. SCIENCE-NEW YORK THEN WASHINGTON 320(5872):57
- Paerl HW, Huisman J (2009) Climate change: a catalyst for global expansion of harmful cyanobacterial blooms. Environ Microbiol Rep 1:27–37
- Paerl HW, Hall NS, Calandrino ES (2011) Controlling harmful cyanobacterial blooms in a world experiencing anthropogenic and climatic-induced change. Sci Total Environ 409:1739–1745
- Posch T, Koester O, Salcher MM, Pernthaler J (2012) Harmful filamentous cyanobacteria favoured by reduced water turnover with lake warming. Nat Clim Change 2:809–813
- Reiss J, Bridle JR, Montoya JM, Woodward G (2009) Emerging horizons in biodiversity and ecosystem functioning research. Trends Ecol Evol 24:505–514
- Reynolds CS (1984) Phytoplankton periodicity—the interactions of form, function and environmental variability. Freshw Biol 14:111–142
- Reynolds CS (1997) Excellence in ecology: vegetation processes in the pelagic: a model for ecosystem theory. Ecology Institute, Germany
- Reynolds CS (2006) The ecology of phytoplankton. Cambridge University Press, Cambridge
- Reynolds CS, Huszar V, Kruk C, Naselli-Flores L, Melo S (2002) Towards a functional classification of the freshwater phytoplankton. J Plankton Res 24:417–428
- Rigosi A, Carey CC, Ibelings BW, Brookes JD (2014) The interaction between climate warming and eutrophication to

promote cyanobacteria is dependent on trophic state and varies among taxa. Limnol Oceanogr 59:99–114

- Roozen FC et al (2003) Lake age and water level affect the turbidity of floodplain lakes along the lower Rhine. Freshw Biol 48(3):519–531
- Ryan EF, Hamilton DP, Barnes GP (2004) Recent occurrence of *Cylindrospermopsis raciborskii* in Waikato lakes of New Zealand. NZ J Mar Freshw Res 37:829–836
- Sas H, Ahlgren I (1989) Lake restoration by reduction of nutrient loading: expectations, experiences, and extrapolations. Academic Verlag, St. Augustin, p 497
- Scheffer M, Rinaldi S, Gragnani A, Mur LR, vanNes EH (1997) On the dominance of filamentous cyanobacteria in shallow, turbid lakes. Ecology 78:272–282
- Schindler D (1977) Evolution of phosphorus limitation in lakes. Science 195:260–262
- Schindler DW, Vallentyne JR (2008) The algal bowl, overfertilization of the world's freshwaters and estuaries. University of Alberta Press
- Schindler DW et al (2008) Eutrophication of lakes cannot be controlled by reducing nitrogen input: results of a 37-year whole-ecosystem experiment. Proc Natl Acad Sci USA 105:11254–11258
- Scott JT, McCarthy MJ (2010) Nitrogen fixation may not balance the nitrogen pool in lakes over timescales relevant to eutrophication management. Limnol Oceanogr 55:1265–1270
- Sinha R, Pearson LA, Davis TW, Burford MA, Orr PT, Neilan BA (2012) Increased incidence of *Cylindrospermopsis* raciborskii in temperate zones—is climate change responsible? Water Res 46:1408–1419
- Sivonen K (1990) Effects of light, temperature, nitrate, orthophosphate, and bacteria on growth of and hepatotoxin production by Oscillatoria agardhii strains. Appl Environ Micorbiol 56(9):2658–2666
- Smayda TJ (1971) Normal and accelerated sinking of phytoplankton in the sea. Mar Geol 11(2):105–122
- Staal J, Hlobil H, Van Tulder M, Waddell G, Burton AK, Koes B, Van Mechelen W (2003) Occupational health guidelines for the management of low back pain: an international comparison. Occup Environ Med 60:618–626
- Steffensen DA (2008) Economic cost of cyanobacterial blooms. In: Cyanobacterial harmful algal blooms: state of the science and research needs advances in experimental medicine and biology 619:855–865
- Stomp M, van Dijk MA, van Overzee HMJ, Wortel MT, Sigon CAM, Egas M, Hoogvelt H, Gons HJ, Huisman J (2008) The timescale of phenotypic plasticity and its impact on competition in fluctuating environments. Am Nat 172(5):169–185
- Straile D, Kerimoglu O, Peeters F, Jochimsen MC, Kümmerlin R, Rinke K, Rothhaupt KO (2010) Effects of a half a millennium winter on a deep lake—a shape of things to come? Glob Change Biol 16:2844–2856
- Stüken A et al (2006) Distribution of three alien cyanobacterial species (Nostocales) in northeast Germany: Cylindrospermopsis raciborskii, Anabaena bergii and Aphanizomenon aphanizomenoides. Phycologia 45:696–703
- Suding KN et al (2008) Scaling environmental change through the community-level: a trait-based response-and-effect framework for plants. Glob Change Biol 14:1125–1140

- Tandeau de Marsac N (1991) Chromatic adaptation by cyanobacteria. In: Bogorad L, Vasil IK (eds) The photosynthetic apparatus: molecular biology and operation. Academic Press, San Diego, pp 417–444
- Tonk L et al (2005) The microcystin composition of the cyanobacterium *Planktothrix agardhii* changes toward a more toxic variant with increasing light intensity. Appl Environ Microbiol 71(9):5177–5181
- Tüxen R (1955) Das system der nordwestdeutschen. Pflanzengesellschaften. Mitt. Flor.-soz. Arbeitsgem. N.F. Stolzenau/Weser 5:155–176
- Van Liere L, Mur LR (1979) Growth-kinetics of Oscillatoria agardhii Gomont in continuous culture, limited in its growth by the light energy supply. J Gen Microbiol 115: 153–160
- Van Liere L, Walsby AE (1982) Interactions of cyanobacteria with light. In: Carr NG, Whitton BA (eds) The Biology of Cyanobacteria. Blackwell Scientific Publications, Oxford, pp 9–45
- Verspagen JM et al (2006) Water management strategies against toxic Microcystis blooms in the Dutch delta. Ecol Appl 16:313–327
- Verspagen JM et al (2014) Rising CO₂ levels will intensify phytoplankton blooms in eutrophic and hypertrophic lakes. PLoS One 9(8):e104325
- Viney NR et al (2007) Modelling adaptive management strategies for coping with the impacts of climate variability and change on riverine algal blooms. Glob Change Biol 13:1–13
- Visser P, Ibelings B, van der Veer B, Koedood J, Mur R (1996) Artificial mixing prevents nuisance blooms of the cyanobacterium *Microcystis* in Lake Nieuwe Meer, the Netherlands. Freshw Biol 36:435–450
- Visser PM, Ibelings BW, Mur LR, Walsby AE (2005) The ecophysiology of the harmful cyanobacterium *Microcystis* In: Huisman J, Matthijs HCP, Visser PM (eds) Harmful cyanobacteria. Springer, Netherlands, pp 109–142
- Vitousek PM et al (2002) Towards an ecological understanding of biological nitrogen fixation. Biogeochemistry 57:1–45
- Vrede T, Ballantyne A, Mille-Lindblom C, Algesten G, Gudasz C, Lindahl S, Brunberg AK (2009) Effects of N:P loading ratios on phytoplankton community composition, primary production and N fixation in a eutrophic lake. Freshw Biol 54:331–344
- Wacklin P, Hoffmann L, Komárek J (2009) Nomenclatural validation of the genetically revised cyanobacterial genus *Dolichospermum* (Ralfs ex Bornet et Flahault) comb. nova. Fottea 9(1):59–64
- Wagner C, Adrian R (2009) Cyanobacteria dominance: quantifying the effects of climate change. Limnol Oceanogr 54:2460–2468
- Walsby AE (1991) The mechanical properties of the *Microcystis* gas vesicle. J Gen Microbiol 137:2401–2408
- Walsby AE (1994) Gas vesicles. Microbiol Rev 58:94
- Walsby AE (2005) Stratification by cyanobacteria in lakes: a dynamic buoyancy model indicates size limitations met by *Planktothrix rubescens* filaments. New Phytol 168: 365–376
- Walsby AE, Avery A, Schanz F (1998) The critical pressures of gas vesicles in *Planktorhrix rubescens* in relation to he

depth of winter mixing in Lake Zürich, Switzerland. J Plankton Res 20:1357–1375

- Walsby AE, Ng G, Dunn C, Davis PA (2004) Comparison of the depth where *Planktothrix rubescens* stratifies and the depth where the daily insolation supports its neutral buoyancy. New Phytol 162:133–145
- WHO (1996) Guidelines for drinking water quality-health criteria and other supporting information, 2nd edn, vol 2. World Health Organization, Geneva
- Woodward G (2009) Biodiversity, ecosystem functioning and food webs in fresh waters: assembling the jigsaw puzzle. Freshw Biol 54:2171–2187
- Yılmaz M, Phlips EJ, Szabo NJ, Badylak S (2008) A comparative study of Florida strains of Cylindrospermopsis and Aphanizomenon for cylindrospermopsin production. Toxicon 51(1):130–139