# Parasites: Small Players with Crucial Roles in the Ecological Theater

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**Abstract:** Effective management of our natural resources requires an understanding of ecosystem structure and function; effectively, an ecosystem-based approach to management. Parasites occur, albeit cryptically, in almost all ecosystems, yet they are usually neglected in studies on populations and communties of organisms. Parasites can have pronounced or subtle effects on hosts affecting host behavior, growth, fecundity, and mortality. Furthermore, parasites may regulate host population dynamics and influence community structure. Many parasites have complex life cycles and depend for transmission on the presence of a variety of invertebrate and vertebrate intermediate hosts. Often transmission involves predator–prey interactions. Thus, parasites reflect the host's position in the food web and are indicative of changes in ecosystem structure and function. Parasites can provide information on population structure, evolutionary hypotheses, environmental stressors, trophic interactions, biodiversity, and climatic conditions. I use examples from diverse freshwater and marine systems to demonstrate that parasites should be incorporated into research and monitoring programs to maximize information gathered in ecosystem-based studies and resource management.

Key words: parasitism, stress, parasite-induced host mortality, food webs, ecosystems, fresh water, marine, indicators

#### INTRODUCTION

Parasitic organisms are often neglected in the management and conservation of biological resources and ecosystems. They are analagous to "extras" in a theatrical production who do not have speaking parts, yet are crucial to a deeper comprehension of the ongoing scene. They are most often small, short-lived, and rarely observed in the external envi-

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ronment, or more commonly hidden within organisms during their parasitic phase. Their effects on their hosts may be obvious and profound, or more subtle, reflecting principal characters or supporting players on the ecosystem stage. Typically, they attract attention only when they cause pathology and disease, or somehow degrade biological products, thus reducing production yields and economic benefits. This is a role where they are only temporarily prominent on the scene and always panned by their critics. Yet, virtually all species are host to at least one parasite species, with the remitting probability that parasitic organisms outnumber freeliving species (Price, 1980). Thus parasites comprise an important component of the cast of organisms in ecological theaters throughout the world, including freshwater and

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Supplementary Tables 3, 4, 5, 6 and additional references for this article are available for viewing by subscribers only at http://www.springerlink.com

marine, whose effects can be manifested in the evolutionary play of life (with apologies to Hutchinson, 1965).

Research on parasites can provide a great deal of information about their host organisms and habitats. A population or ecosystem-scale approach to parasitology can be applied not only to the control of disease, but to the proper management and conservation of aquatic resources, be they species targeted for harvest or areas designated as protected. In this article, I summarize basic parasitic lifestyles and transmission patterns and review the various applications of this information to species management in aquatic ecosystems. I outline a holistic approach whereby knowledge of parasites can be applied to conservation of multispecies systems to aid in conservation management. Examples are given from freshwater and marine ecosystems including plants, invertebrates, and vertebrates, for the implications of parasitism extend beyond commercially exploited species to include all trophic levels and organisms within habitats. The goal is to foster an appreciation for the role played by parasites as the drama of life unfolds on the global ecosystem stage.

# LIFE-STYLES AND TRANSMISSION PATTERNS

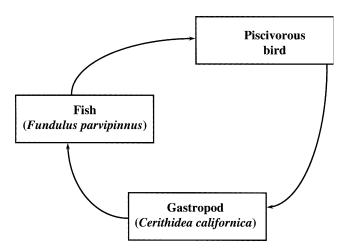
For epidemiological purposes, parasites are traditionally divided into microparasites and macroparasites (Anderson and May, 1979; May and Anderson, 1979). Microparasites consist of small organisms that are primarily unicellular, including viruses, bacteria, and protozoans, but also multicellular organisms of small size (< 50 µm) such as myxozoans. These organisms typically multiply in or on the host and are often associated with disease. Transmission is usually direct but may be indirect, involving alternate hosts (e.g., myxozoans), or vectors (e.g., many protozoans). Macroparasites are larger, multicellular organisms such as monogeneans, digeneans, cestodes, nematodes, acanthocephalans, arthropods, leeches, and others. These typically undergo sexual reproduction in or on a host, but do not normally proliferate there (the production of cercariae by digeneans in molluscan intermediate hosts is an exception). They often possess complex life cycles, with one or more intermediate hosts required for development or growth (Figs. 1 and 2). These parasites are sometimes clearly detrimental to their hosts, but in many cases their effects are more subtle and difficult to measure.

For those parasites with complex life cycles, a variety of transmission modes has evolved. Transmission may involve

one or more free-living infective stages, where the infective stage is passively ingested by the next host in the parasite's life cycle, for example, certain larvae of digeneans (metacercariae) that encyst in the external environment; freeswimming cestode larvae (coracidia) that are preyed upon by crustacean intermediate hosts; or parasite eggs of many groups. Free-living infective stages also may be transmitted actively by penetrating the next host in the life cycle, as with the larval cercariae of many digeneans in fish and other organisms, and nematode larvae in moist soil. Parasite life cycles may involve a parasitic infective stage in one precursor host that must be ingested for transmission to occur. Examples can be found among all the principal helminth groups (except monogeneans) including many digeneans, and all species of cestodes, nematodes, and acanthocepahalans. In these cases, occurrence of a parasite in a host reflects predator-prey interactions between the host and its prey and predators. The diversity of parasites within a host reflects the presence of diverse intermediate and definitive hosts in the ecosystem participating in the parasites' life cycles. Thus, by the nature of their different life cycles, the parasites in a host population provide information on that organism's role in the food web (Marcogliese and Cone, 1997a; Marcogliese, 2001b, 2002, 2003).

## Impacts on Hosts and Communities

By their very nature, parasites have a variety of impacts on their hosts. They impose energetic demands, alter behavior, affect morphology and appearance, reduce fecundity and growth, and cause mortality. These effects are well documented in numerous host-parasite systems in both freshwater and marine habitats. Behavioral alterations may lead to increased vulnerability to predation. In some cases, this is a pathological side effect, but in others it may enhance transmission to the next host in the life cycle. Thus, parasites can affect the diet of predators, influencing predatorprey dynamics (Fig. 1) and competitive interactions between that host and other organisms (Price et al., 1986; Lafferty et al., 2000). Parasites can affect sex ratio and mate choice (Minchella and Scott, 1991). Taken together with the effects listed above, it seems likely that parasites can have an impact on host fitness, and thus, play a role in natural selection of host characteristics. A list of parasites that affect commercial fish stocks through parasite-induced host mortality, reduction in fecundity, or reduction in market value or weight appears in Dobson and May (1986).



**Figure 1.** Life cycle of a typical digenean (*Euhaplorchis californiensis*) in a salt marsh. Piscivorous birds such as egrets or herons serve as definitive hosts. Eggs are passed with feces, which are ingested by the first intermediate host (horn snail, *Cerithidea californica*). Cercariae are asexually produced in the snail and released into the water, infecting a suitable second intermediate fish host (Pacific killifish, *Fundulus parvipinnus*) upon contact. The parasite then encysts as a metacercaria in the brain. The life cycle is completed when the bird eats the infected fish. The parasite impacts upon the ecosystem at multiple trophic levels. Infected snails are castrated, possibly affecting snail population levels. Infected fish display behavioral alterations such that they become 30 times more susceptible to predation. Conceivably, the presence of the parasite permits the persistence of piscivorous waterfowl by facilitating predation (Lafferty and Morris, 1996).

Pathology caused by parasitism is widespread in all organisms (Kinne, 1980-1990; Woo, 1995; Bondad-Reantaso et al., 2001), and virtually all organs and tissues can be damaged by a plethora of parasitic organisms. Examples of parasites of a range of invertebrate and vertebrate aquatic organisms that have different negative impacts on their hosts are listed in Table 1. Infection with many types of parasites causes sublethal effects in virtually all types of organisms, including ctenophores, cnidarians, molluscs, crustaceans, insects, echinoderms, chaetognaths, fish, and plants (see supplementary Table 3, available for viewing by subscribers only at http://www.springerlink.com), as well as amphibians, waterfowl, and aquatic mammals Though most parasites do not normally kill their hosts, death can result from infection. Parasites of numerous different types of organisms, from algae through vertebrates, have been shown to cause parasite-induced host mortality (see supplementary Table 4, available for viewing by subscribers only at http://www.springerlink.com). Clearly, the effects of

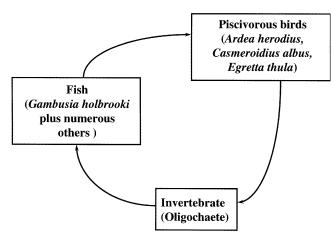


Figure 2. Life cycle of a typical nematode (Eustrongylides ignotus) in the aquatic environment. Piscivorous birds, including great blue herons (Ardea herodius), great egrets (Casmeroidius albus), and snowy egrets (Egretta thula), are definitive hosts. Eggs are passed into the water with the feces and are ingested by oligochaetes, the first intermediate hosts. Numerous fish species, including mosquitofish (Gambusia holbrooki) may function as second intermediate hosts, acquiring the parasite by eating infected oligochaetes. The avian definitive hosts become infected when they consume the fish, thus completing the life cycle. This parasite causes pathology in piscivorous fish hosts and renders smaller forage fish more susceptible to predation (Coyner et al., 2001). In addition, the parasite causes significant mortality among nestling birds. Furthermore, the parasites' abundance is amplified in eutrophic conditions, probably via effects on oligochaete populations. Thus, this parasite has impacts at different trophic levels, and these impacts are substantially amplified by anthropogenic nutrient enrichment.

parasites are manifest throughout the food webs of aquatic systems, and not confined to well-studied commercial species. These lists of host–parasite associations represent a variety of invertebrate and vertebrate taxa at different trophic levels, along with different types of impacts. Virtually all types of parasites manifest some sort of effect and all types of organisms are impacted in some way, no matter where they occur within a food web.

Parasites also function as ecosystem engineers by directly or indirectly modifying the environment of other organisms (i.e., the host phenotype). A parasite may alter host biology such that new habitat for other species is formed (Thomas et al., 1999; Lafferty et al., 2000).

If prevalence and abundance are high, parasites can have a significant impact on the host population, and regulate its numbers (Anderson and May, 1979; May and Anderson, 1979). If that particular host population is a dominant species in an ecosystem, then the presence of the parasite may have consequences for the entire food web

Table 1. Selected Ex	xamples of Subletha	Selected Examples of Sublethal and Lethal Impacts on Ac	quatic Hosts by Paras	on Aquatic Hosts by Parasites in Freshwater and Marine Environments	1arine Environments	
Host species	Common name	Parasite species	Parasite group	Ecosystem	Impact	Reference <sup>a</sup>
Mollusca Hydrobia ulvae	Gastropod	Microphallids	Digeneans	Intertidal	MHId	Jensen and Mouritsen, 1992
Illyanassa obsoleta	Gastropod	Numerous species	Digeneans	mudflats Intertidal	Castration; alter	Curtis, 2002
• • •	4		)	mud flats	vertical distribution	
Arthropoda Cyclops strenuus	Copepod	Triaenophorus spp.	Cestodes	Lab (freshwater)	Alter activity, decrease depth, reduce fecundity, feeding, PIVP	Pasternak et al., 1999; Pulkkinen et al., 2000
Daphnia spp.	Cladocerans	Ordospora colligata (= Pleistophora intestinalis), Flabelliforma magnivora, Octosporea baveri	Microsporidians	Freshwater pools, lab	Reduce fecundity, reduce competitive ability, PIHM	Ebert, 1994; Ebert et al., 2000; Salathé and Ebert, 2003
Gammarus pulex	Amphipod	Pomphorhynchus laevis	Acanthocephalan	Freshwater rivers, lab	Reduce respiration, altered drift, increase phototactism and activity, alter appearance, PIVP, increase sensitivity to acid conditions and cadmium, decrease feeding, reduce lipid in gravid females	Rumpus and Kennedy, 1974; Brown and Pascoe, 1989; McCahon and Poulton, 1991; McCahon et al., 1991; Bakker et al., 1997; Plaistow et al., 2001
Echinodermata Strongylocentrotus droebachiensis	Sea urchin	Paramaoba invadens	Amoeba	Nova Scotia coast	PIHM	Hagan, 1996
Pisces		Echinomermella matsi	Nematode	Norwegian coast	PIHM, castration	
Clupea harengus	Herring	Ichthyophonus hofferi Scolex pleuronectis	Fungus Cestode	North Sea Lab (marine), marine	PIHM PIHM, reduce larval feeding	Patterson, 1996 Rosenthal, 1967; Heath and Nicoll, 1991

Lab (marine) PIHM Balbuena et al., 2000	e) PIHM Rosenthal, 1967 PIHM Pennycuick, 1971	labReduce growth, affectPennycuick, 1971; Pascoe, and diet choice andCram, 1977; Pascoe, and 	River PIHM Fischer and Kelso, 1988	pond, lab PIHM Lemly and Esch, 1984	ater) Reduce respiration Kumaragura et al., 1995 and swimming	Deformities, abnormal Densmore et al., 2001 swimming, lower leuckocyte and lymphocyte counts	I	Reduce hemoglobin K	and hematocrit
Lab (marine) Freshwater Freshwater, lab	Freshwater, lab		Mississippi River	Freshwater pond, lab	Lab (freshwater)	Freshwater	Lab (freshwater)	Lab (freshwater)	Lab (freshwater)
	Copepod Digenean	Cestode	Digenean	Digenean	Hemoflagellate	Myxozoan	Monogenean	Digenean	Digenean
aduncum	Lernaeocera sp. Diplostomum vasterostei	Schistocephalus solidus	Allacanthochasmus sp.	Uvulifer ambloplitis	Cryptobia salmositica	Myxobolus cerebralis	Gyrodactylus spp.	Crepidostomum farionis	Nanophyetis salmincola
	Threespine stickleback		Bluegill	Sunfish	Rainbow trout				
	Gasterosteus aculeatus		Lepomis macrochirus		Oncorhynchus mykiss				

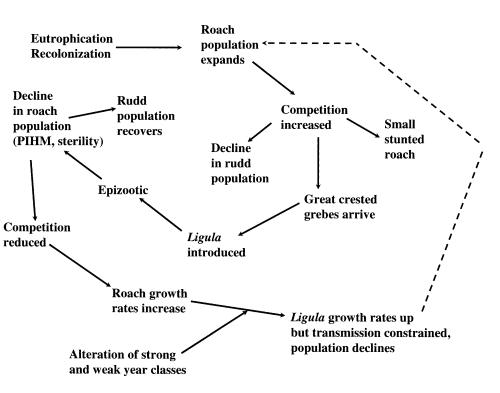


Figure 3. Schematic displaying role of the cestode Ligula intestinalis in population cycles of the roach, Rutilus rutilus (direct interactions) and rudd, Scardinius erythrophthalmus (indirect interactions) in Slapton Ley, United Kingdom. In the first phase of the epizootic cycle, roach population dynamics are controlled by the parasite population. In the second phase, roach population dynamics determine infection levels of L. intestinalis. Initiation of a new cycle (dashed line) may depend on local conditions and is not obligatory (after Kennedy et al., 2001). PIHM, parasite-induced host mortality.

and ecosystem structure (Dobson and Hudson, 1986; Minchella and Scott, 1991; McCallum and Dobson, 1995). Such parasites are termed "keystone parasites" by Minchella and Scott (1991). For example, two different parasites are known to control local populations of the green sea urchin (Strongylocentrotus droebachiensis) which can overgraze kelp beds and completely alter coastal ecosystems. In Norwegian waters, populations of sea urchins are limited by the nematode Echinomermella matsi, while in Nova Scotia by the protozoan Paramoeba invadens (reviewed in Hagen, 1996). Outbreaks of the microsporidian Cougourdella sp. reduce populations of the dominant grazer in the system, the caddisfly Glossosoma nigrior, thus permeating increases in the production of periphyton and the abundance of other invertebrate grazers in Michigan streams (Kohler and Wiley, 1997). The parasitic plant Cuscuta salina preferentially infects the dominant competitor Salicornia virginiana over three other subordinate salt marsh plants, thus affecting community composition and dynamics (Pennings and Callaway, 2002). Parasites may actually drive plant succession, with very small effects on competitive ability being translated into communitywide consequences (Dobson and Crawley, 1994). These few examples illustrate the broad range of communities that can be influenced by parasitic infections in key species.

Depending on the location within the food web of a species infected by parasites, impacts may permeate through

a bottom-up or a top-down cascade on the rest of the web. It should be stressed that organisms found throughout the food web are subjected to the impacts of parasitism. These impacts strongly suggest that it is imperative to consider parasites in management and conservation of their hosts.

Parasites may exert more subtle effects on host communities. For example, epizootics of plerocercoids of the cestode *Ligula intestinalis* in their intermediate host, the roach (*Rutilus rutilus*), clearly determine not only the population structure of the roach via parasite-induced host mortality and sterility but of another sympatric cyprinid fish, the rudd (*Scardinius erythrophthalmus*) in Slapton Ley via roach–rudd interactions (Fig. 3; Kennedy et al., 2001). Such interactions only became apparent after collection of long-term data and may generally be more common than previously assumed in other host–parasite systems.

In terms of conservation, the introduction or elimination of a parasite may affect interactions between a variety of species within a community (Dobson and Hudson, 1986; Kennedy et al., 2001). Furthermore, parasites and disease are a major threat to endangered species (McCallum and Dobson, 1995), especially if those species are maintained at high densities in small, fragmented areas that promote transmission and parasite exchange across species (Scott, 1988; McCallum and Dobson, 1995; Holmes, 1996). In addition, introductions and other emerging infectious diseases, including parasites, are a potentially serious threat to endangered species and biodiversity at large (Cunningham et al., 2003; Daszak and Cunningham, 2003).

# Parasites As Indicators of Host Biology

Numerous studies in aquatic systems have effectively used parasites as indicators of host stocks and their ontogenetic or seasonal migrations. For some recent reviews, consult in Williams et al. (1992), Arthur (1997), and MacKenzie and Abaunza (1998) for rationale, examples, and guidelines. Most of the studies to date involve marine species of fish, but some work in fresh waters has successfully used parasites to discriminate among stocks (Marcogliese et al., 2001, and references therein). Because many parasites are transmitted via predator-prey interactions, and parasites possess a variety of complex life cycles with different intermediate hosts, parasites are excellent indicators of host diet (Williams et al., 1992; Marcogliese and Cone, 1997a). In fact, for numerous reasons, parasites provide complementary information to that obtained through stomach content analysis. Parasites reflect trophic interactions over weeks or months, whereas gut contents provide details of the animal's diet only over the last 24 hours or less (Williams et al., 1992; Curtis, 1995). They can indicate ontogenetic shifts in diet, whether hosts feed on more than one trophic level, niche shifts due to competition or other factors, individual feeding specializations within a population, seasonal changes in diet, and temporary links in a food web such as periodic migrants into a system (reviewed in Williams et al., 1992; Marcogliese and Cone, 1997a; Marcogliese, 2003). When combined with other techniques used in fisheries science such as meristics and population genetics, research managers have at their disposal a very powerful array of tools to study the biology of virtually any organism. Moreover, and not insignificantly, parasites are used as indicators of historical biogeography and phylogenetics of their hosts (Brooks and Hoberg, 2000).

# Parasites As Indicators of Ecosystem Stress, Food Webs, and Biodiversity

A number of reviews summarize existing knowledge of the relationship between parasites and pollution, and parasites as indicators of stress (Overstreet and Howse, 1977; Overstreet, 1988, 1993, 1997; Khan and Thulin, 1991; Poulin, 1992; MacKenzie 1993, 1999; MacKenzie et al. 1995; Lafferty, 1997; Lafferty and Kuris, 1999). Basically, parasites can be used as indicators of environmental stress in a way analogous to their employment to differentiate among host populations or stocks. Many parasites possess complex life cycles and depend on the presence of one or more intermediate or paratenic hosts for transmission. Should the abundance of any of these hosts decline, for example, by exposure to chemical contaminants, then transmission of the parasite may be impaired. Similar results occur if infected hosts are more sensitive to effects of contaminants and are selectively removed (MacKenzie et al., 1995). Furthermore, free-living stages of parasites or those inhabiting the external surface or gastrointestinal tract are directly exposed to toxicants, and those parasites may be directly susceptible to pollution (Poulin, 1992; Overstreet, 1997; MacKenzie, 1999). See Table 1 in Pietrock and Marcogliese (2003) for a list of various endohelminths where survival and infectivity of their free-living stages are susceptible to toxicological effects caused by environmental contaminants. Parasites demonstrate different types of sensitivity to contaminants and environmental stress in aquatic hosts and ecosystems (see supplementary Table 5, available for viewing by subscribers only at http:// www.springerlink.com). In terms of other relationships with pollutants, intestinal parasites appear to be more sensitive bioaccumulators of heavy metals than their fish hosts, and may serve as excellent indicators of heavy metal pollution (Sures et al., 1999; Sures, 2001, 2003). Alternatively, if a host's immune response is compromised by toxin exposure, its parasite burden may increase. Such a situation is often observed for monogeneans and protozoans that proliferate in hosts that inhabit polluted habitats (Overstreet, 1997). Commonly parasites that increase in abundance in contaminated habitats often possess direct life cycles (see Table 1 in MacKenzie et al., 1995, for numerous examples). Interpreted another way, abundance of infections with endoparasitic helminths tends to decrease, while those of ectoparasites tend to increase with pollution (MacKenzie, 1999). Moreover, many facultative parasites such as pathogenic viruses and bacteria also proliferate under these circumstances. Guidelines for selecting the most appropriate host-parasite combinations and the most vulnerable stages as indicators are provided in MacKenzie (1993, 1999) and MacKenzie et al. (1995).

Just as entire communities of free-living organisms are affected by environmental stress, so are the communities of parasites that infect any particular host species. Diversity

<b>Table 2.</b> Studies that Indicate Reductions or Increases in Paras to Various Types of Environmental Stress in Aquatic Habitats <sup>a</sup>	Indicate Reductions o vironmental Stress ir	or Increases in Parasite Species Ri n Aquatic Habitats <sup>a</sup>	ichness and Abundance, or C	Studies that Indicate Reductions or Increases in Parasite Species Richness and Abundance, or Changes in Species Composition for Parasite Communities in Hosts Exposed Types of Environmental Stress in Aquatic Habitats <sup>a</sup>	asite Communities in Hosts Exposed
Host species	Common name	Ecosystem	Contaminants or stress	Observations	References
Buccinum undatum	Common whelk	Firth of Clyde, UK	Sewage	Decline in prevalence of digeneans toward source	Siddall et al., 1993
Stagnicola emarginata	Snail	Lakes, northern Michigan	Human development	Decline in species richness and total prevalence	Cort et al., 1960; Keas and Blankespoor, 1997
Anguilla rostrata	American eel	Rivers, Nova Scotia, Canada	Acidification	Decline in species richness, loss of digeneans, change in species composition (acanthocephalans), with acidity	Cone et al., 1993; Marcogliese and Cone, 1996, 1997b
Bairdiella chrysura	Silver perch	Estuaries, Florida	Contaminants	Decrease in prevalence of crustaceans and parasites with indirect life cycles	Landsberg et al., 1998
Gambusia affinis	Western mosquitofish	Mississippi and Texas	Organic toxicants, heavy metals	Decline in species richness (heteroxenous species)	Overstreet, 1997
Gambusia holbrooki	Eastern mosquitofish	Florida	Sewage	Increase in prevalence of Eustrongylides ignotus	Coyner et al., 2003
Leuciscus cephalus	Chub	Moravia River, Czech Republic; rivers, Italy Moravia River, Czech Republic	Eutrophication, domestic and industrial Industrial and domestic	Change in abundance of acanthocephalans, decline in species richness Decline in species richness downstream of source	Dusek et al., 1998; Galli et al., 1998, 2001 Gelnar et al., 1997
Limanda limanda	Common dab	Firth of Forth, UK	Sewage	Change in prevalence and abundance of certain species	Siddall et al., 1994
Notropis hudsonius	Spottail shiner	St. Lawrence River, Ouebec. Canada	Sewage	Increase in myxozoan prevalence and species richness	Marcogliese and Cone, 2001
Perca fluviatilis	Perch	Lakes, Finland	Pulp and paper, eutrophication Acidification	Decline in species richness, variable effects on species Decline in species richness	Valtonen et al., 1997 Halmetoja et al., 2000
Platichthys flesus	Flounder	SE Baltic; North Sea	Domestic and industrial	and digeneans Decline in species richness	Sulgostowska et al., 1987; Broeg et al., 1999
Gobies	Fish	Baltic Sea	Eutrophication	Decline in species richness	Zander, 1998

<sup>&</sup>lt;sup>a</sup>For further examples, please consult Tables 2 and 3 in MacKenzie et al. (1995).

and species richness may increase under stressful conditions, but more often a decrease occurs, at least for endoparasites with indirect life cycles. Reductions in parasite species richness have been observed following acidification, eutrophication, and chemical contamination (Table 2; see also supplementary Table 6 for a more comprehensive list, available for viewing by subscribers only at http:// www.springerlink.com). These reductions in parasite diversity are believed to parallel diversity loss in free-living species, because populations of intermediate hosts are impacted by environmental changes. Furthermore, parasite communities may recover concomitantly with free-living communities when conditions improve (Cone et al., 1993).

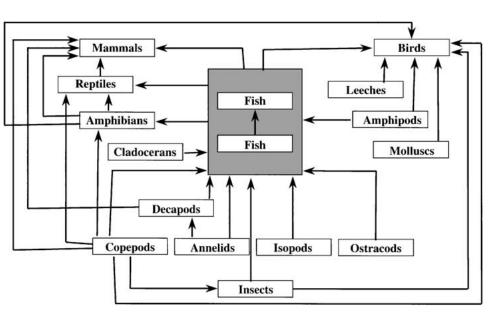
However, resource managers must be aware that parasite taxa respond differently to environmental perturbations (Curtis, 1995; Lafferty, 1997; Marcogliese and Cone, 1997b; Lafferty and Kuris, 1999; MacKenzie, 1999), depending on the life cycle of the parasite, the concentration and type of contaminant, and the exposure time (Overstreet and Howse, 1977; Khan and Thulin, 1991; Poulin, 1992). Thus, generalizations about the relationship between parasitism and pollution cannot be made without taking into account the biology of individual species. Kennedy (1997) further highlights the intrinsic difficulties to using parasites as indicators of pollution, but concludes that they can be excellent, nonspecific, early-warning indicators of environmental change.

Because many parasites depend on predator-prey relationships for transmission, parasites are sentinels for food web interactions. One species of parasite may depend on the presence of only one or a few intermediate and paratenic hosts for transmission, however, the total parasite diversity within a host represents a diversity of life cycles that utilize numerous different organisms as hosts at some point in their life cycles (Figs. 4 and 5). Not only do parasites provide information on a host's diet, but this information is complementary, and in many ways superior, to gut content analysis (see above). Furthermore, information on a host's predators can also be derived from a host's parasites. Thus, the parasite fauna within a host population provides information about the role of the host in the food web, and the variety of predator-prey relationships in which it participates (Marcogliese and Cone, 1997a, b; Marcogliese, 2002, 2003).

Parasite life cycles evolve in tandem with the evolution of their hosts' life history traits, and have adapted to longstanding predator–prey interactions. Thus, the presence of particular parasites in a host may also tell us something about the stability of the ecosystem (George-Nascimento, 1987; Marcogliese and Cone, 1997a; Marcogliese, 2003). Moreover, parasites may actually maintain the stability and integrity of ecosystems (Brooks and Hoberg, 2001). Environmental changes resulting from global warming, for example, may disrupt synchronous population cycles of predators and their prey in aquatic habitats, interfering with parasite transmission between those organisms (Marcogliese, 2001a). The climate change-related environmental perturbations that can affect parasitism include alterations in temperature regimes, precipitation, host range, water levels and flow rates, eutrophication, stratification, extent of ice cover, acidification, oceanic circulation patterns, and UV radiation (Marcogliese, 2001a).

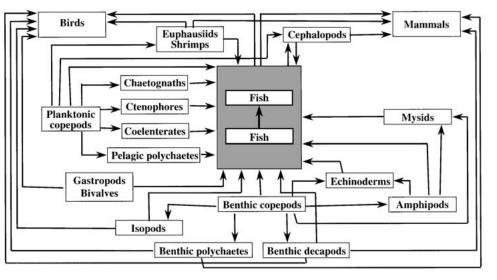
Parasites may be excellent, economical, early-warning indicators of changes to environmental conditions and ecosytem health (MacKenzie, 1993, 1999; Overstreet, 1993, 1997; Lafferty, 1997; Marcogliese and Cone, 1997b; Marcogliese, 2003). This concept can be expanded to changes in biodiversity by taking into account the nature of parasites, their life cycles, and transmission. Biodiversity and its measurement have been an increasing concern for research managers and conservationists. By definition, biodiversity includes not only species diversity, but the ecological complexes of which they are a part (Glowka et al., 1994). Developing appropriate indicators for biodiversity has been a difficult task. The idea that certain taxa can be used as surrogates is popular, but few indicators have proven reliable, especially across different ecosystems. Because parasites respond to environmental stressors and track food webs via their transmission processes, it is logical to extend their use to indicators of biodiversity. They have the further advantage that they belong to many distinct and unrelated taxa. Thus, there are no phylogenetic constraints such as those imposed when specific taxa are used as indicators. In addition, because they infect hosts on different trophic levels, their usage is not trophically constrained (Marcogliese and Cone, 1997b; Marcoglies, 2003).

Those organisms in the middle of the food web such as small fish may be best suited as biodiversity indicators. They tend to be more heavily infected than top piscivores, because they prey upon intermediate and paratenic hosts and thus acquire parasites that may or may not mature in them (George-Nascimento, 1987). They, in turn, are preyed upon by larger piscivores and pass on larval parasites to these organisms, where they may or may not mature. In addition, predators on small fish include not only larger fish, but other vertebrates such as birds and mammals.



**Figure 4.** Potential transmission pathways involving predator–prey interactions for helminth parasites in freshwater environments. In this figure and in Figure 5, other types of parasites are not shown for simplicity's sake, nor are interactions involving free-living infective stages depicted (e.g., cestode coracidia and digenean miracidia and cercariae). For both this figure and Figure 5, specificity for the intermediate and definitive hosts within any one life cycle (and any one compartment in the diagrams) will vary with individual parasite

species. Routes of trophic pathways will also vary with parasite species, with some being obligate and others facultative, depending on the nature of the host–parasite interaction. In addition, within each life cycle, parasites may follow more than one path to reach the definitive host, again depending on the specificity of the host– parasite interaction. Reprinted in adapted form from Marcogliese and Cone (1997a), copyright 1997, with permission from Elsevier Science.



**Figure 5.** Potential transmission pathways involving predator–prey interactions for helminth parasites in marine environments (see Fig. 4 for details about the food web diagram). Note that the marine food web appears more complex than the freshwater web. This is a result of the presence of an additional trophic level in marine

systems, that of large invertebrate predators, which play a relatively greater role in marine food chains than in freshwater food chains. Reprinted in adapted form from Marcogliese and Cone (1997a), copyright 1997, with permission from Elsevier Science. Thus, it is possible to obtain information on food web pathways linking the aquatic and terrestrial milieux.

In summary, parasites are ubiquitous in the aquatic environment. They have impacts ranging from the subtle, to the sublethal, to the lethal. Their impacts on hosts are propagated up and down food webs and thus are manifested throughout entire communities. Like free-living organisms, they are affected by biotic and abiotic changes to the environment. Parasites are effective indicators of many aspects of host biology and thus extremely useful as management and conservation tools. Moreover, they are uniquely situated within food webs, and their transmission processes may permit their usage as indicators of environmental stress, food-web structure, and biodiversity. Indeed, far from being mere extras without speaking parts in the ecological theater, parasites may be bit players but with incredibly important roles who should step forward and take a bow as the curtain goes down on the ecosystem stage. Critics must acknowledge the significance of their wonderfully complex roles that are intricately woven into the scripts of virtually all the principal players in the theater of life.

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### References

- Anderson RM, May RM (1979) Population biology of infectious diseases: part I. *Nature* 280:361–367
- Arthur JR (1997) Recent advances in the use of parasites as biological tags for marine fish. In: *Diseases in Asian Aquaculture III*, Flegel TW, MacRae IH (editors), Manila: Fish Health Section, Asian Fisheries Society, pp 141–154
- Bakker TCM, Mazzi D, Zala S (1997) Parasite-induced changes in behavior and color make *Gammarus pulex* more prone to fish predation. *Ecology* 78:1098–1104
- Balbuena JA, Karlsbakk E, Kvenseth AM, Saksvik M, Nylund A (2000) Growth and emigration of third-stage larvae of *Hysterothylacium aduncum* (Nematoda: Anisakidae) in larval herring (*Clupea harengus*). Journal of Parasitology 86:1271–1275

- Barber I, Huntingford FA (1995) The effects of *Schistocephalus solidus* (Cestoda: Pseudophyllidea) on the foraging and shoaling behaviour of three-spined sticklebacks, *Gasterosteus aculeatus*. *Behaviour* 132:1223–1240
- Bondad-Reantaso MG, McGladdery SE, East I, Subasinghe RP (2001) Asian diagnostic guide to aquatic animal diseases. FAO Fisheries Technical Paper 402/2, FAO, Rome, Italy
- Broeg K, Zander S, Diamant A, Korting W, Kruner G, Paperna I, et al. (1999) The use of fish metabolic, pathological and parasitological indices in pollution monitoring. I. North Sea. *Helgoländer Marine Research* 53:171–194
- Brooks DR, Hoberg EP (2000) Triage for the biosphere: the need and rationale for taxonomic inventoried and phylogenetic studies of parasites. *Comparative Parasitology* 67:1–25
- Brooks DR, Hoberg EP (2001) Parasite systematics in the 21st century: opportunities and obstacles. *Trends in Parasitology* 17:273–275
- Brown AF, Pascoe D (1989) Parasitism and host sensitivity to cadmium: an acanthocephalan infection of the freshwater amphipod *Gammarus pulex*. Journal of Applied Ecology 26:473–487
- Butler JA, Millemann RE (1971) Effect of the "salmon poisoning" trematode, *Nanophyetus salmincola*, on the swimming ability of juvenile salmonid fishes. *Journal of Parasitology* 57:860–865
- Cone DK, Marcogliese DJ, Watt WD (1993) Metazoan parasite communities of yellow eels (*Anguilla rostrata*) in acidic and limed rivers of Nova Scotia. *Canadian Journal of Zoology* 71:177–184
- Cort WW, Hussey KL, Ameel DJ (1960) Seasonal fluctuations in larval trematode infections in *Stagnicola emarginata angulata* from Phragmites Flats on Douglas Lake. *Journal of the Helminthological Society of Washington* 27:11–13
- Coyner DF, Schaack SR, Spalding MG, Forrester DJ (2001) Altered predation susceptibility of mosquitofish infected with *Eustrongylides ignotus. Journal of Wildlife Diseases* 37:556–560
- Coyner DF, Spalding MG, Forrester DJ (2003) Influence of treated sewage on infections of *Eustrongylides ignotus* (Nematoda: Dioctophymatoidea) in eastern mosquitofish (*Gambusia holbrooki*) in an urban watershed. *Comparative Parasitology* 70:205–210
- Cunningham AA, Daszak P, Rodriguez JP (2003) Pathogen pollution: defining a parasitological threat to biodiversity conservation. *Journal of Parasitology* 89(Suppl):S78–S83
- Curtis LA (2002) Ecology of larval trematodes in three marine gastropods. *Parasitology* 124(Suppl):S43–S56
- Curtis M (1995) The ecological parasitology of charrs: relationships between parasites and food web structure in northern lakes. *Nordic Journal of Freshwater Research* 71:92–101
- Daszak P, Cunningham AA (2003) Anthropogenic change, biodiversity loss, and a new agenda for emerging diseases. *Journal of Parasitology* 89(Suppl):S37–S41
- Densmore CL, Blazer VS, Waldrop TB, Pooler PS (2001) Effects of whirling disease on selected hematological parameters in rainbow trout. *Journal of Wildlife Diseases* 37:375–378
- Dobson A, Crawley M (1994) Pathogens and the structure of plant communities. Trends in Ecology & Evolution 9:393–398
- Dobson A, Hudson PJ (1986) Parasites, disease and the structure of ecological communities. *Trends in Ecology & Evolution* 1:11–15
- Dobson A, May RM (1986) The effects of parasites on fish populations—theoretical aspects. In: *Parasitology—quo vadit*?

*Proceedings of ICOPA VI*, Howell MJ (editor), Canberra, Australia: Australian Academy of Science, pp 363–370

- Dusek L, Gelnar M, Sebelová S (1998) Biodiversity of parasites in a freshwater environment with respect to pollution: metazoan parasites of chub (*Leuciscus cephalus* L.) as a model for statistical evaluation. *International Journal for Parasitology* 28:1555–1571
- Ebert D (1994) Genetic differences in the interactions of a microsporidian parasite and four clones of its cyclically parthenogenetic host. *Parasitology* 108:11–16
- Ebert D, Lipsitch M, Mangin KL (2000) The effect of parasites on host population density and extinction: experimental epidemiology with *Daphnia* and six microparasites. *American Naturalist* 156:459–477
- Fischer SA, Kelso WE (1988) Potential parasite-induced mortality in age-0 bluegills in a floodplain pond of the lower Mississippi River. *Transactions of the American Fisheries Society* 117:565–573
- Galli P, Mariniello L, Crosa G, Ortis M, Ambrogi AQ, D'Amelio S (1998) Populations of *Acanthocephalus anguillae* and *Pomphorhynchus laevis* in rivers with different pollution levels. *Journal of Helminthology* 72:331–335
- Galli P, Crosa G, Mariniello L, Ortis M, D'Amelio S (2001) Water quality as a determinant of the composition of fish parasite communities. *Hydrobiologia* 452:173–179
- Gelnar M, Sebelová S, Dusek L, Koubková B, Jurajda P, Zahrádková S (1997) Biodiversity of parasites in freshwater environment in relation to pollution. *Parassitologia* 39:189–199
- George-Nascimento MA (1987) Ecological helminthology of wildlife animal hosts from South America: a literature review and a search for patterns in marine food webs. *Revista Chilena de Historia Natural* 60:181–202
- Glowka L, Burhanne-Guilmin F, Synge H, McNeeley JA, Gundling L (1994) *A guide to the Convention on Biological Diversity*, Gland, Switzerland: IUCN—The World Conservation Union
- Hagen NT (1996) Sea urchin outbreaks and epizootic disease as regulating mechanisms in coastal ecosystems. In: *Biology and Ecology of Shallow Coastal Waters*, Eleftheriou A, Ansell AD, Smith CJ (editors), Fredensborg, Denmark: Olsen & Olsen, pp 303–308
- Halmetoja A, Valtonen ET, Koskenniemi E (2000) Perch (*Perca fluviatilis* L.) parasites reflect ecosystem conditions: a comparison of a natural lake and two acidic reservoirs in Finland. *International Journal for Parasitology* 30:1437–1444
- Heath M, Nicoll N (1991) Infection of larval herring by helminth parasites in the North Sea and the effect on feeding incidence. *Continental Shelf Research* 11:1477–1489
- Holmes JC (1996) Parasites as threats to biodiversity in shrinking ecosystems. *Biodiversity and Conservation* 5:975–983
- Hutchinson GE (1965) *The Ecological Theatre and the Evolutionary Play*, New Haven, CT: Yale University Press
- Jacobson KC, Arkoosh MR, Kagley AN, Clemons ER, Collier TK, Casillas E (2003) Cumulative effects of natural and anthropogenic stress on immune function and disease resistance in juvenile Chinook salmon. *Journal of Aquatic Animal Health* 15:1–12
- Jensen KT, Mouritsen KN (1992) Mass mortality in two common soft-bottom invertebrates, *Hydrobia ulvae* and *Corophium volutator*—the possible role of trematodes. *Helgoländer Meeresuntersuchungen* 46:329–339
- Keas BE, Blankespoor HD (1997) The prevalence of cercariae from *Stagnicola emarginata* (Lymnaeidae) over 50 years in northern Michigan. *Journal of Parasitology* 83:536–540

- Kennedy CR (1997) Freshwater fish parasites and environmental quality: an overview and caution. *Parassitologia* 39:249–254
- Kennedy CR, Shears PC, Shears JA (2001) Long-term dynamics of Ligula intestinalis and roach Rutilus rutilus: a study of three epizootic cycles over thirty-one years. Parasitology 123:257–269
- Khan RA, Thulin J (1991) Influence of pollution on parasites of aquatic animals. *Advances in Parasitology* 30:201–238
- Kinne O (1980–1990) Diseases of Marine Animals, Vol 1–14, Chichester, UK and Hamburg, Germamny: John Wiley and Biologische Anstalt Helgoland
- Klein WD, Olsen OW, Bowden DC (1969) Effects of intestinal fluke, *Crepidostomum farionis*, on rainbow trout, *Salmo gairdneri*. *Transactions of the American Fisheries Society* 98:1–6
- Kohler SL, Wiley MJ (1997) Pathogen outbreaks reveal large-scale effects of competition in stream communities. *Ecology* 78:2164–2176
- Kumaraguru AK, Beamish FWH, Woo PTK (1995) Impact of a pathogenic haemoflagellate, *Cryptobia salmositica* Katz, on the metabolism and swimming performance of rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Journal of Fish Disease* 18:297–305
- Lafferty KD (1997) Environmental parasitology: what can parasites tell us about human impacts on the environment? *Parasitology Today* 13:251–255
- Lafferty KD, Kuris AM (1999) How environmental stress affects the impacts of parasites. *Limnology and Oceanography* 44:925– 931
- Lafferty KD, Morris AK (1996) Altered behavior of parasitized killifish increases susceptibility to predation by bird final hosts. *Ecology* 77:1390–1397
- Lafferty KD, Thomas F, Poulin R (2000) Evolution of host phenotype manipulation by parasites and its consequences. In: *Evolutionary Biology of Host-parasite Relationships: Theory Meets Reality*, Poulin R, Morand S, Skorping A (editors), Amsterdam: Elsevier, pp 117–127
- Landsberg JH, Blakesley BA, Reese RO, McRae G, Forstchen PR (1998) Parasites of fish as indicators of environmental stress. *Environmental Monitoring and Assessment* 51:211–232
- Lemly D, Esch GW (1984) Effects of the trematode Uvulifer ambloplitis on juvenile bluegill sunfish, Lepomis macrochirus: ecological implications. Journal of Parasitology 70:475–492
- MacKenzie K (1993) Parasites as biological indicators. *Bulletin of* the Scandinavian Society for Parasitology 1:1–10
- MacKenzie K (1999) Parasites as pollution indicators in marine ecosystems: a proposed early warning system. *Marine Pollution Bulletin* 38:955–959
- MacKenzie K, Abaunza P (1998) Parasites as biological tags for stock discrimination of marine fish: a guide to procedures and methods. *Fisheries Research* 38:45–56
- MacKenzie K, Williams HH, Williams B, McVicar AH, Siddall R (1995) Parasites as indicators of water quality and the potential use of helminth transmission in marine pollution studies. *Advances in Parasitology* 35:85–144
- Marcogliese DJ (2001a) Implications of climate change for parasitism of animals in the aquatic environment. *Canadian Journal* of Zoology 79:1331–1352
- Marcogliese DJ (2001b) Pursuing parasites up the food chain: implications of food web structure and function on parasite communities in aquatic systems. *Acta Parasitologica* 46:82–93
- Marcogliese DJ (2002) Food webs and the transmission of parasites to marine fish. *Parasitology* 124(Suppl):S83–S99

- Marcogliese DJ (2003) Food webs and biodiversity: are parasites the missing link? *Journal of Parasitology* 89(Suppl):S106–S113
- Marcogliese DJ, Cone DK (1996) On the distribution and abundance of eel parasites in Nova Scotia: influence of pH. *Journal of Parasitology* 82:389–399
- Marcogliese DJ, Cone DK (1997a) Food webs: a plea for parasites. Trends in Ecology & Evolution 12:320-325
- Marcogliese DJ, Cone DK (1997b) Parasite communities as indicators of ecosystem stress. *Parassitologia* 39:227–232
- Marcogliese DJ, Cone DK (2001) Myxozoan communities parasitizing *Notropis hudsonius* (Cyprinidae) at selected localities on the St. Lawrence River, Quebec: possible effects of urban effluents. *Journal of Parasitology* 87:951–956
- Marcogliese DJ, Dumont P, Gendron A, Mailhot Y, Bergeron E, McLaughlin JD (2001) Spatial and temporal variation in abundance of *Diplostomum* spp. in walleye (*Stizostedion vitreum*) and white suckers (*Catostomus commersoni*) from the St. Lawrence River. *Canadian Journal of Zoology* 79:355–369
- May RM, Anderson RM (1979) Population biology of infectious diseases: part II. *Nature* 280:455–461
- McCahon CP, Poulton MJ (1991) Lethal and sub-lethal effects of acid, aluminum and lime on *Gammarus pulex* during repeated simulated episodes in a Welsh stream. *Freshwater Biology* 25:169–178
- McCahon CP, Maund SJ, Poulton MJ (1991) The effect of the acanthocephalan parasite (*Pomphorhynchus laevis*) on the drift of its intermediate host (*Gammarus pulex*). *Freshwater Biology* 25:507–513
- McCallum H, Dobson A (1995) Detecting disease and parasite threats to endangered species and ecosystems. *Trends in Ecology* & *Evolution* 10:190–194
- Minchella DJ, Scott ME (1991) Parasitism: a cryptic determinant of animal community structure. *Trends in Ecology & Evolution* 6:250–254
- Overstreet RM (1988) Aquatic pollution problems, southeastern U.S. coasts: histopathological indicators. *Aquatic Toxicology* 11:213–239
- Overstreet RM (1993) Parasitic diseases of fishes and their relationship with toxicants and other environmental factors. In: *Pathobiology of Marine and Estuarine Organisms*, Couch JA, Fournie JW (editors), Boca Raton, FL: CRC Press, pp 111–156
- Overstreet RM (1997) Parasitological data as monitors of environmental health. *Parassitologia* 39:169–175
- Overstreet RM, Howse HD (1977) Some parasites and diseases of estuarine fishes in polluted habitats of Mississippi. *Annals of the New York Acadamy of Science* 298:427–462
- Pascoe D, Cram P (1977) The effect of parasitism on the toxicity of cadmium to the three-spined stickleback, *Gasterosteus aculeatus* L. *Journal of Fish Biology* 10:467–472
- Pasternak AF, Pulkkinen K, Mikheev VN, Hasu T, Valtonen ET (1999) Factors affecting abundance of *Triaenophorus* infection in *Cyclops strenuus*, and parasite-induced changes in host fitness. *International Journal for Parasitology* 29:1793–1801
- Patterson KR (1996) Modelling the impact of disease-induced mortality in an exploited population: the outbreak of the fungal parasite *Ichthyophonus hoferi* in the North Sea herring (*Clupea harengus*). *Canadian Journal of Fisheries and Aquatic Sciences* 53:2870–2887
- Pennings SC, Callaway RM (2002) Parasitic plants: parallels and contrasts with herbivores. *Oecologia* 131:479–489

- Pennycuick L (1971) Quantitative effects of three species of parasites on a population of three-spined sticklebacks, *Gasterosteus aculeatus*. *Journal of Zoology, London* 165:143–162
- Pietrock M, Marcogliese DJ (2003) Free-living endohelminth stages: at the mercy of environmental conditions. *Trends in Parasitology* 19:293–299
- Plaistow SJ, Troussard J-P, Cézilly F (2001) The effect of the acanthocephalan parasite *Pomphorhynchus laevis* on the lipid and glycogen content of its intermediate host *Gammarus pulex*. *International Journal for Parasitology* 31:346–351
- Poulin R (1992) Toxic pollution and parasitism in freshwater fish. Parasitology Today 8:58–61
- Price PW (1980) *Evolutionary Biology of Parasites*, Princeton, NJ: Princeton University Press
- Price PW, Westoby M, Rice B, Atsatt PR, Fritz RS, Thompson JN, et al.(1986) Parasite mediation in ecological interactions. *Annual Reviews in Ecology and Systematics* 17:487–505
- Pulkkinen K, Pasternak AF, Hasu T, Valtonen ET (2000) Effect of *Triaenophorus crassus* (Cestoda) infection on behavior and susceptibility to predation of the first intermediate host *Cyclops strenuus* (Copepoda). *Journal of Parasitology* 86:664– 667
- Rosenthal H (1967) Parasites in larvae of the herring (*Clupea* harengus L.) fed with wild plankton. Marine Biology 1:10–15
- Rumpus AE, Kennedy CR (1974) The effect of the acanthocephalan Pomphorhynchus laevis upon the respiration of its intermediate host, Gammarus pulex. Parasitology 68:271–284
- Salathé P, Ebert D (2003) The effects of parasitism and inbreeding on the competitive ability in *Daphnia magna*: evidence for synergistic epistasis. *Journal of Evolutionary Biology* 16:976– 985
- Scott ME (1988) The impact of infection and disease on animal populations: implications for conservation biology. *Conservation Biology* 2:40–56
- Siddall R, Pike AW, McVicar AH (1993) Parasites of Buccinum undatum (Mollusca: Prosobranchia) as biological indicators of sewage-sludge dispersal. Journal of the Marine Biological Association of the United Kingdom 73:931–948
- Siddall R, Pike AW, McVicar AH (1994) Parasites of flatfish in relation to sewage sludge dumping. *Journal of Fish Biology* 45:193–209
- Stoltze K, Buchmann K (2001) Effect of *Gyrodactylus derjavini* infections on cortisol production in rainbow trout fry. *Journal of Helminthology* 75:291–294
- Sulgostowska T, Banaczyk G, Grabda-Kazubska B (1987) Helminth fauna of flatfish (Pleuronectiformes) from Gdansk Bay and adjacent areas (south-east Baltic). *Acta Parasitologica Polonica* 31:231–240
- Sures B (2001) The use of fish parasites as bioindicators of heavy metals in aquatic ecosystems: a review. *Aquatic Ecology* 35:245–255
- Sures B (2003) Accumulation of heavy metals by intestinal helminths in fish: an overview and perspective. *Parasitology* 126 (Suppl):S53–S60
- Sures B, Siddall R, Taraschewski H (1999) Parasites as accumulation indicators of heavy metal pollution. *Parasitology Today* 15:16–21
- Thomas F, Poulin R, de Meeüs T, Guégan J-F, Renaud F (1999) Parasites and ecosystem engineering: what roles could they play? *Oikos* 84:167–171

- Valtonen ET, Holmes JC, Koskivaara M (1997) Eutrophication, pollution, and fragmentation: effects on parasite communities in roach (*Rutilus ruitlus*) and perch (*Perca fluviatilis*) in four lakes in central Finland. *Canadian Journal of Fisheries and Aquatic Sciences* 54:572–585
- Williams HH, MacKenzie K, McCarthy AM (1992) Parasites as biological indicators of the population biology, migrations, diet,

and phylogenetics of fish. Reviews in Fish Biology and Fisheries 2:144-176

- Woo PTK (1995) Fish Diseases and Disorders, Vol 1. Protozoan and Metazoan Infections, Wallingford, CT: CABI Publishing
- Zander CD (1998) Ecology of host parasite relationships in the Baltic Sea. *Naturwissenschaften* 85:426–436