

## Rheumatoid arthritis: links with cardiovascular disease and the receptor for advanced glycation end products

Lisa Carroll<sup>1</sup>, Suad Hannawi<sup>1</sup>, Thomas Marwick<sup>2</sup> and Ranjeny Thomas<sup>1</sup>

<sup>1</sup>Centre for Immunology and Cancer Research, Princess Alexandra Hospital, University of Queensland, Queensland, Australia

<sup>2</sup>School of Medicine, Southern Clinical Division, Princess Alexandra Hospital, University of Queensland, Queensland, Australia

Received October 15, 2005; accepted November 2, 2005

© Springer-Verlag 2006

### Rheumatoide Arthritis: Zusammenhänge zwischen kardiovaskulären Erkrankungen und dem „Advanced Glycation End Product Receptor“

**Zusammenfassung.** Kardiovaskuläre Erkrankungen treten bei Patienten mit chronischen entzündlichen Erkrankungen wie zum Beispiel der Rheumatoiden Arthritis (RA) gehäuft auf. Aus der Sicht der Pathophysiologie hat die Atherosklerose mit Autoimmunerkrankungen verblüffende Ähnlichkeiten. Diese Erkenntnis wurde zu einer Zeit des Paradigmenwechsels im Management der chronischen Polyarthritiden mit Biologika gemacht. Diese Übersicht beschreibt mögliche Ursachen für das gehäufte Auftreten von Gefäßerkrankungen der Rheumatoiden Arthritis einschließlich der Rolle von traditionellen kardiovaskulären Risikofaktoren. Potentielle Mechanismen wie das C-Reaktive Protein (CRP), Veränderungen im Gerinnungssystem und Cyclooxygenase (COX)-2 Inhibitoren werden kurz behandelt. Der Advanced Glycation End Product Receptor (RAGE) wurde als Kandidatenmolekül, welches laufende Entzündungen und Autoimmunprozesse beeinflusst, identifiziert. Ein Schwerpunkt dieser Übersichtsarbeit ist die Rolle von RAGE bei kardiovaskulären Erkrankungen und der chronischen Polyarthritiden. Wie es bei vielen neuen Molekülen der Fall ist, wird angenommen, dass funktionelle Polymorphismen die Krankheitsexpression verändern und zur Aufklärung der biologischen Aktivitäten des ursprünglichen Moleküls beitragen können. Diese Übersichtsarbeit schließt mit einer Diskussion der möglichen Rolle des RAGE Glycine 82 Serine Polymorphismus.

**Schlüsselwörter:** Rheumatoide Arthritis, kardiovaskuläre Erkrankungen, Advanced Glycation End Product Receptor, Polymorphismus, Genetik, Entzündung, C-Reaktives Protein, Hyperlipidämie, Blutgerinnung.

Correspondence: Prof Ranjeny Thomas, Centre for Immunology and Cancer Research, University of Queensland, Princess Alexandra Hospital, Brisbane, QLD 4102, Australia.  
Fax: ++61-7-32405946  
E-mail: rthomas@cicr.uq.edu.au

**Summary.** Cardiovascular (CV) disease is increased in patients with chronic inflammatory disease, including rheumatoid arthritis (RA). Furthermore it has become clear at a pathophysiological level, that atherosclerosis has striking similarities with autoimmune disease. This realization has come at a time of paradigm shift in how rheumatologists manage RA, with the availability of biological agents targeting key inflammatory cytokines. This review will focus on the possible causes of increased vascular disease in RA, including the role of traditional CV risk factors. Mechanisms potentially at play, such as C-reactive protein (CRP), altered coagulation, and cyclooxygenase (COX)-2 inhibitors will be covered in brief. The Receptor for Advanced Glycation End Products (RAGE) has been identified as a candidate molecule influencing response to ongoing inflammation and autoimmunity. There will be a focus on the role of RAGE in CV disease and RA. As has been the case with many novel molecules, functional polymorphisms are thought to alter disease expression and assist us in coming to terms with the biological activities of the parent molecule. The review will conclude with a discussion of the potential role of the RAGE Glycine 82 Serine polymorphism.

**Key words:** Arthritis Rheumatoid, Cardiovascular Diseases, Advanced Glycation End-Product Receptor, Polymorphism, Genetic, Inflammation, C-Reactive Protein, Hyperlipidaemia, Coagulation.

The mortality of patients with RA is increased compared to the general population [1, 2], with much of this increase attributable to excess CV deaths [2–5]. It is also known that CV disease accounts for much of the comorbidity of RA [6–9]. The known risk factors have been shown to account for about 50% of CV events in the general population [10, 11]. Many novel factors contributing to vascular disease pathogenesis have been described, but their clinical utility is still under investigation. These include hyperhomocysteinaemia [12, 13] (which occurs in RA patients taking methotrexate [14]), various prothrom-

botic factors (many of which are elevated in RA [15]) and serological markers of inflammation such as C-reactive protein (CRP), interleukin (IL)-6 and serum amyloid A [16–25].

Traditional CV risk factors have been studied in RA cohorts. However, the incidence of CV disease is high in RA, even when accounting for traditional risk factors [6, 26, 27]. Furthermore three studies, so far reported only in abstract form, have shown that RA is an independent predictor of ischemic heart disease (IHD) with half of patients having clinically silent disease, and many not even presenting with pain when suffering an acute coronary syndrome [28–30]. This suggests that IHD may be far more widespread in RA than is clinically apparent. In keeping with these clinical findings, silent ischemia has also been noted on 24 hour Holter monitoring in RA patients [31]. Furthermore echocardiography and Doppler have shown a high frequency of sub-clinical left ventricular diastolic dysfunction [32].

Patients with RA have signs of vascular dysfunction even when newly diagnosed [33, 34]. Across RA patients of all disease durations, endothelial dysfunction inversely correlates with markers of inflammation [35]. In a small study, vascular dysfunction could be reversed, at least in the short term, with successful RA therapy [34]. Even accounting for adverse effects of anti-rheumatic drugs present in long term RA, the implication of these studies is that the pathogenesis of RA and of atherosclerotic vascular disease may be similar. Very recently the results of a ten year follow up study of inflammatory polyarthritis patients, has revealed that 38.5% of patients died of CV causes. Baseline level of inflammation (CRP) was found to be an independent predictor of future CV death, supporting the hypothesis that inflammation occurring in patients with RA may play a direct role in CV disease pathogenesis [36]. Results from laboratory, clinical, and epidemiological studies all support the hypothesis that immune dysregulation as well as ongoing systemic inflammation in RA are likely to be pivotal factors in the early vascular disease of RA [37, 38]. The response to an insult is remarkably similar whether it is in the lung (pulmonary fibrosis), kidneys (glomerulosclerosis), liver (cirrhosis), pancreas (pancreatitis), joints (pannus) or arteries (atherosclerosis) [39].

### **Dyslipidaemia**

There is known to be an inverse relationship between lipid levels and any form of inflammation as measured by the acute phase response [40, 41]. It has been known for some time that RA sufferers have low levels of total cholesterol [40, 42]. Closer analysis of lipid profiles reveals that high density lipoprotein (HDL) is disproportionately lowered, giving rise to an adverse lipid profile [43–45].

The mechanisms at play that induce lipid changes in RA and other proinflammatory states have been the object of some research. It has been shown that interleukin-1 receptor antagonist (IL-1ra) gene deficiency and associated IL-1 excess promotes autoimmunity and joint-specific inflammation in animal models [46, 47], and it is clear that relative IL-1 excess exists in human RA

[48–50]. Animal models have also shown that IL-1ra gene deficiency leads to pro-atherogenic changes in lipid metabolism with elevated levels of total cholesterol and reduction of HDL under inflammatory conditions [51]. The implication is that relative IL-1 excess, be it constitutional or induced by the inflammatory state, promotes the pro-atherogenic lipid profile in RA, due to IL-1-mediated induction of enzymes critical for cholesterol metabolism. There is good evidence implicating proinflammatory effects of IL-6 on progression of atherosclerosis, with hepatic synthesis of CRP largely regulated by this cytokine [52], however knockout animal models suggest that IL-6 is important in regulating body composition, probably via hypothalamic mechanisms, and treatment with agents that block IL-6 leads to hyperlipidaemia in animals and humans [53, 54]. Circulating levels of IL-6 are increased by Tumour Necrosis Factor (TNF)  $\alpha$  and IL-1, but IL-6 also regulates the level of TNF $\alpha$ , and of IL-6 indirectly, via inducing the release of IL-1ra [52]. This plethora of interactions is somehow likely to be responsible for the dyslipidaemia of inflammatory diseases, however the changes in lipid profile seen in RA, are not in themselves sufficient to account for the increased risk of CV disease.

There is some evidence that controlling disease activity with conventional disease modifying agents, including corticosteroids and hydroxychloroquine, may reverse some of the abnormalities in lipid profile [44, 55–58]. TNF $\alpha$  blockers may also improve the CV risk profile of patients with RA [59] however not all reports have been favourable [60]. A humanized anti-IL-6 receptor antibody has been recently trialled for the treatment of refractory RA, with significant reduction of disease severity. However almost half of patients had an increase in blood cholesterol at both treatment doses, an effect that has been seen in other human trials, and there was significant elevation of triglycerides at the higher dose. HDL cholesterol also increased, but to a lesser degree suggesting an overall adverse effect [53]. Of interest, mice deficient in IL-6 develop mature onset obesity and insulin-resistance, associated with reduced energy expenditure [54]. The clinical observations provide further evidence of positive and negative roles of inflammatory cytokines on CV disease risk, and highlight an important area to scrutinize with the increasing use of anti-cytokine therapy.

### *Smoking*

Smoking dose-dependently increases the risk of RA [61–63]. Smokers with RA are more likely to be rheumatoid factor (RF) seropositive [64]. RF is generally associated with more severe rheumatoid disease and also with a greater incidence of extra-articular disease, thus it has been hypothesized that smoking may be a co-morbid dependent risk factor for CV disease. However, at least one study indicated that smoking was not a predictor of CV events or mortality in a cohort of seropositive RA patients [58].

### *Hypertension*

Hypertension is common in RA [8, 65] and diastolic BP levels are elevated compared to controls [27]. It seems

likely that the widespread use of non-steroidal anti-inflammatory drugs (NSAIDs) may be partly to blame for the high incidence of co-morbid hypertension in RA patients, and perhaps some of the increase in CV mortality [66–73]. Both traditional NSAIDs and the newer cyclooxygenase (COX)-2 inhibitors have been found to aggravate blood pressure through their effects on reduced renal function and fluid retention [74]. Clinically the elevation is likely to be important, as even small decreases of 5–6 mmHg in diastolic blood pressure were shown to decrease the risk of CV events by 20–25% and cerebrovascular events by 35–40% in a non-RA population [75].

#### *Diabetes mellitus*

The prevalence of diabetes was not found to be increased in RA [27] and even when present, failed to predict CV events [58]. However, insulin resistance is prevalent in RA [76] and increasingly it is believed that this prediabetic phase has similar pathophysiological consequences to diabetes itself [77, 78].

#### *C-Reactive protein*

CRP is the major acute phase protein which rises rapidly in response to inflammatory stimuli, and may remain elevated for prolonged periods in chronic inflammation. Smoking, aging, abdominal obesity, steato-hepatitis and subclinical vascular disease also increase levels [52, 79]. The biologic activities of CRP include activation of the classical complement cascade, enhancing phagocytosis and binding Fcγ receptors (FcγR), resulting in cytokine production [80, 81]. In addition to recognition of microbial antigens, CRP and the related molecule serum amyloid P (SAP), react with and clear dying and apoptotic cells from sites of tissue injury [82, 83] indicating that CRP is involved in regulation of the immune system [84].

The highly sensitive CRP (hsCRP) test detects small elevations in the CRP which are not usually detectable by standard assays. There is accumulated evidence that minor elevations in CRP are identified in almost every step of atherogenesis and predict poor outcome [16–19, 21–23, 85, 86] and CRP has been found to correlate with intima-media thickness (IMT) a marker of vascular disease [87–91]. Thus, CRP is an independent determinant of the risk of atherosclerotic disease. The relative risk of future events in both men and women for those with values of CRP in the highest quartile compared with the lowest quartile has been shown to be between 2 and 4 [21, 23, 86]. The clinical utility of monitoring CRP levels to establish vascular disease risk in patients with RA has been difficult to clarify, with high levels as a result of arthritis clouding the picture. However, Goodson et al have provided evidence that even a single elevated CRP, early in the inflammatory polyarthritis disease course, is predictive of future CV death [36].

CRP may actively participate in promoting the pro-inflammatory component of atherosclerosis. It induces expression of cellular adhesion molecules (CAMs), IL-6, and endothelin-1 by endothelial cells [92, 93] and has been shown to mediate monocyte chemoattractant protein (MCP)-1 induction [94] and uptake of low density lipoprotein (LDL) by macrophages [95].

In addition to this, smooth muscle cells and macrophages in arterial tissue have been shown to produce CRP, a process that is substantially upregulated in the atherosclerotic plaque [96]. Given the direct activation of complement by CRP, and that activated complement is pathogenic to cells in plaques, suggests that there is a self-sustaining autotoxic mechanism operating within the plaques as a precursor to thrombotic events [96]. As further evidence of a direct role of CRP in angiogenesis, there are data to suggest that CRP is responsible for a reduction of NO [97], which may result in inhibition of angiogenesis, an important compensatory mechanism in chronic ischemia [98]. Whether it does in fact have a direct role remains a subject of debate.

In RA, due to the high levels of systemic inflammation, it has been difficult to use CRP alone for the detection and monitoring of vascular disease. However, other potentially useful markers of vascular activation are currently being investigated [99]. Obviously if CRP plays a causative role, this has major implications for risk of vascular disease in sub-optimally treated autoimmune diseases. However, since a single elevated CRP at baseline presentation is predictive of CV death, CRP may have a direct role in CV disease pathogenesis in patients with inflammatory disease and may be a useful marker even if levels are higher in RA than in atherosclerosis in the general population [36].

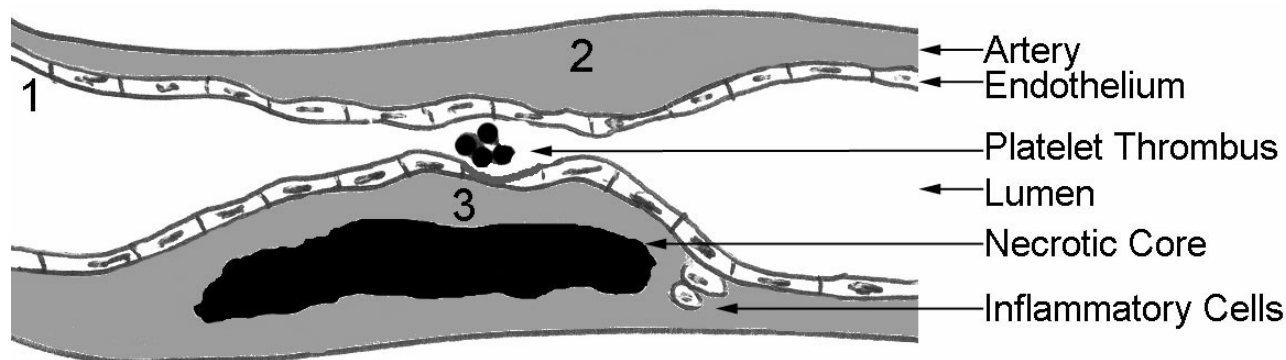
#### *Other novel risk factors*

CAMs and selectins are essential in mediation of adhesion and transendothelial migration of leukocytes, one of the earliest processes in atheroma formation, the absence of which results in a reduction of disease in murine models [39]. Local expression of intercellular adhesion molecule-1 (ICAM-1), vascular cellular adhesion molecule-1 (VCAM-1), and E-selectin has been described in both rheumatoid synovium and atherosclerotic lesions [39, 100, 101], suggesting a similar pathogenic mechanism for both. Furthermore biomarkers of endothelial dysfunction, including VCAM-1, ICAM-1 and endothelial leukocyte adhesion molecule (ELAM)-1, are higher in RA than in control subjects and correlate with markers of inflammation [102].

#### *Coagulation and cardiovascular disease in RA*

Inflammation in under-treated RA may have significant prothrombotic effects, contributing to the severity of CV disease. Fibrinogen, tissue plasminogen activator (tPA), D-dimers and Von Willebrand factor (vWF) have all been found to be elevated in RA compared to controls [27], and tPA and plasminogen activator inhibitor (PAI)-1, may be associated with CV events in RA [103].

Evidence has accumulated that highly COX-2 specific inhibitors (such as rofecoxib and celecoxib) increase CV risk [67, 69–73], however these agents have only been in use relatively recently, and the contribution by selective COX blockade is unlikely to explain much of the excess morbidity seen in previous years, despite some degree of selectivity by conventional NSAIDs. COX-2 specific inhibitors impair endothelium-dependent vasodilation.



**Fig. 1.** Stages in Vascular Inflammation

1. Endothelial dysfunction, induced by factors such as oxidized LDL and smoking, results in increased permeability, leukocyte and platelet adhesion and a procoagulant tendency.
2. An increase in smooth muscle cell volume results in an intermediate lesion. At this stage compensatory dilatation can occur.
3. A fibrous cap overlies a core of lipid and necrotic tissue. This can further erode and lead to thrombosis and complete vessel occlusion.

Modified from Adams et al [114].

ilatation [104] primarily by negatively shifting the balance of prostacyclin (PGI<sub>2</sub>) to thromboxane (TXA<sub>2</sub>), which has pro-thrombotic effects. Specific inhibition of the COX-2 enzyme decreases systemic PGI<sub>2</sub> [105] resulting in the loss of a natural inhibitor of platelet activation [106]. The effects of COX-1 and COX-2 selective inhibitors on thrombosis are almost certainly related to their divergent effects on the balance between PGI<sub>2</sub> and TXA<sub>2</sub>. The release of PGI<sub>2</sub> by ischemic tissue, contributing to protective reactive hyperaemia was first demonstrated by McGiff in 1970 [107]. Animal studies have confirmed that this occurs in the coronary vasculature [108] and that the COX-2 enzyme plays an essential role in cardio protection post ischemia [109]. The CV risk of these drugs may also be present for less COX-2 selective NSAIDs [70].

Clinically it is imperative to know whether the co-administration of a COX-1 selective inhibitor (such as aspirin) may dampen these negative vascular effects. A previously observed increase in time to occlusion with aspirin in a coronary thrombosis model was abolished with celecoxib, suggesting that co-treatment with these agents neutralizes the protective effects of COX-1 selective inhibitors [110]. Such studies have important clinical implications, given the widespread use of these agents in RA, particularly for those patients with prior vascular events or multiple risk factors who require aspirin for coronary heart disease and stroke prophylaxis.

#### *The role of inflammation in cardiovascular disease*

Atherosclerosis is now acknowledged to be an inflammatory disease [39]. Endothelial dysfunction is the initial step in atheroma formation, and may be caused by elevated and oxidized LDL, free radicals (generated for instance, by smoking [111]), hypertension, diabetes mellitus, elevated plasma homocysteine, and perhaps infectious micro-organisms [39]. This dysfunction is accompanied by increased permeability and adhesiveness (with respect to leukocytes and platelets) and a procoagulant

state as the endothelium is bathed in cytokines, chemokines, and growth factors. Within the enlarging inflammatory focus, smooth-muscle cell volume increases to form an intermediate lesion, whilst the artery thickens and compensates by gradual dilation [112] (remodelling). Continued inflammation recruits greater numbers of monocyte-derived macrophages and T-lymphocytes, and release of hydrolytic enzymes may eventually induce focal necrosis. With continuation of the process, a fibrous cap is formed over a core of lipid and necrotic tissue (advanced, complicated lesion). At some time point the artery loses its ability to compensate by dilation, the lesion intrudes into the lumen and the flow of blood is impaired. Plaque rupture and thrombosis can occur, resulting in acute coronary syndromes [39] (Fig. 1).

In atherosclerosis, most of the normal anticoagulant defences are impaired. There is loss of the vasodilator NO [113], abnormal heparin formation, local thrombin activation, relative lack of tPA and thrombomodulin, of all which favour local coagulation. In addition, increased tissue PAI and lack of tPA impair clot lysis. Thrombin activation, membrane bound platelet adhesion molecule expression, exposure of collagen, increased production of tissue factor (TF), and loss of NO also favour increased platelet adhesion and aggregation. Coagulation is greatly accelerated with endothelial erosion or plaque fissure [114].

Monocyte-derived macrophages and the related dendritic cells are scavenging and antigen-presenting cells that secrete cytokines, chemokines, growth-regulating molecules, metalloproteinases and other hydrolytic enzymes. Activated macrophages and smooth muscle cells express MHC class II antigens, allowing them to present antigens to T lymphocytes [115, 116]. Activated T-cells secrete cytokines; including interferon  $\gamma$ , TNF $\alpha$  and lymphotoxin; all of which amplify the inflammatory response [115]. One possible antigen may be oxidized LDL, which can be produced by macrophages [117]. Cell-mediated immune responses are likely to be involved in atheroge-

nesis, since both CD4 and CD8 T-cells are present in the lesions at all stages of the process.

Given the pathogenesis of atherosclerosis as described, research has delved into the possibility of an HLA-associated genetic predisposition to accelerated CV disease; however findings have been less than overwhelming. Although HLA- associations have been reported in patients with atherosclerosis [118–120], it is not clear whether the HLA tissue antigens are involved directly in pathogenesis or whether they are markers for some other genetic factor. Lipoprotein (a) has been found to be independently associated with early atherosclerosis and its sequelae, and a link between inherited high levels of lipoprotein (a) and certain HLA class II genotypes has been suggested but not proven [121, 122].

#### *The Receptor for advanced glycation end products*

RAGE is a 35-kDa polypeptide whose gene is located at the junction of the class II and III HLA regions on chromosome 6 [123]. It is a multiligand member of the immunoglobulin super-family of cell surface receptor molecules [124, 125]. This cell surface receptor is known as RAGE as it was initially found to bind AGEs (advanced glycation end products) which are nonenzymatically glycosylated adducts of proteins and lipids [126]. In addition to AGEs, RAGE was found to bind EN-RAGEs (extracellular newly identified RAGE-binding proteins), or RAGE-ligands. These include the S100/calgranulin chemokine family [127],  $\beta$  pleated sheet fibrils of amyloid [128], and amphoterin, also known as high mobility group box chromosomal protein 1 (HMGB-1) [129].

AGEs accumulate in disorders such as diabetes [130], amyloidoses [131], Alzheimer's disease [128] and renal failure [132], as well as at sites of oxidative stress, and in normal aging [133]. In rat models, RAGE expression is high in the developing nervous system (in association with amphoterin), at a time when little AGE formation would be occurring, however levels tend to decrease as the animals mature. This suggests that RAGE may have a physiological role in development [129].

The proximity of cells expressing RAGE to lesions rich in AGEs, and the activation that results, suggests that the AGE-RAGE interaction may trigger intracellular signal transduction mechanisms that alter properties of vascular and inflammatory effector cells. This may then contribute to dysfunctional repair in AGE rich tissues, as occurs in diabetes. Given that RAGE also serves as a cell surface receptor for amyloid- $\beta$  peptide [128], which accumulates in Alzheimer's disease, it has been hypothesized that the RAGE receptor converts proteinaceous deposits into bioactive substrate [127].

AGE ligation of RAGE has been shown to activate p21ras and mitogen-activated protein (MAP) kinase, and to stimulate nuclear translocation of the transcription factor NF- $\kappa$ B thereby resulting in the transcription of target genes [134]. Ligation is believed to enhance receptor expression and to initiate a positive feedback loop, in which receptor occupancy triggers increased RAGE expression, followed by further cellular activation. Ongoing expression of RAGE on endothelium, smooth muscle cells, mononuclear phagocytes, and neurons when in close

proximity to ligands, thus may induce chronic cellular activation and tissue damage [135].

#### *RAGE and cardiovascular disease*

Increased expression of RAGE and the presence of AGEs have been described at sites of vascular lesions and in blood vessel walls [135, 136]. In diabetic retinopathy and renal glomerulosclerosis, microvascular lesions correlate with the accumulation of AGEs [137, 138]. In a murine model, after the induction of neointimal expansion of smooth muscle, the process was strikingly suppressed by blockade of RAGE by using a truncated soluble form of RAGE (sRAGE), RAGE antibodies or in RAGE-/- mice [136]. Recently the importance of endogenous sRAGE in human health has been shown, with low levels independently associated with the presence of coronary artery disease in nondiabetic men [139].

Ligation of RAGE expressed by endothelial cells diminishes vascular barrier function [140], enhances expression of VCAM-1 [141], quenches NO [142], induces the expression of cytokines such as IL-6 and MCP-1 [135], and changes the balance of coagulation, partly by induction of procoagulant TF [143]. RAGE has also been shown to upregulate the COX-2 enzyme, resulting in monocyte activation and vascular cell dysfunction [144].

Diabetes provides a model of widespread chronic vascular disease. As further evidence of the role of RAGE in atherosclerosis, sRAGE has been shown to prevent the accelerated progression of atherosclerosis usually seen in diabetic apoE null mice [145], and diabetes associated vascular dysfunction in vivo can be prevented by blockade of RAGE [146]. A murine model implicated RAGE as central to diabetic neuropathy, with prevention and even reversal demonstrated with RAGE blockade [147].

#### *RAGE and rheumatoid arthritis*

Given that rheumatoid arthritis generates AGEs, as a by-product of oxidative stress, in addition to other RAGE-ligands as a result of neutrophil activation, the role of RAGE and its ligands in the pathogenesis of inflammatory joint disease has been studied. In a murine model of inflammatory arthritis, blockade of RAGE suppressed clinical and histological evidence of arthritis [148].

AGEs have been shown to accumulate in inflamed RA synovial tissue, with RAGE antigen expression overlapping with the distribution of AGE epitopes [101]. However, in a small study looking at RAGE staining patterns of synovial tissue, there were no differences in staining pattern between those patients with RA and those with osteoarthritis [149].

Members of the calgranulin family of proteins are key molecules involved in signal transduction, differentiation, and cell cycle control, and are ligands of RAGE [150]. S100A12 (calgranulin C) is one of several RAGE ligands that is expressed and secreted by granulocytes [151]. Murine models of collagen-induced arthritis demonstrate the ability of S100A12 to trigger synovial inflammation [127]. Serum levels of this cytokine have been shown to be raised in several autoimmune diseases including adults with RA, seronegative and psoriatic arthritis, inflammatory bowel disease, and in children and

adolescents with Kawasaki disease and juvenile RA (JRA) [151–153]. In systemic onset JRA, levels of S100A12 are higher than in health. There is a gradient in serum levels, those with polyarticular joint involvement having higher levels than those with oligoarticular JRA. Levels decrease in response to treatment of inflammation and increase prior to clinical flares. High concentrations in synovial fluid indicate release of the cytokine at this site [151]. Other calgranulins include S100A8 and S100A9. These RAGE ligands are also known as macrophage migration inhibitory factor-related proteins 8 and 9 (MRP8 and MRP9) and like S100A12, have been shown to accumulate at sites of inflammation, including the synovial membrane lining layer adjacent to the cartilage-pannus junction of rheumatoid arthritis [154]. Serum levels are also representative of inflammation in JRA [155].

The last of the currently known ligands for RAGE, HMGB-1 / amphoterin, has been implicated in the pathogenesis of arthritis. This molecule was first thought to function only as a nuclear factor that enhanced transcription, but it is now known to be a crucial cytokine mediating tissue response to infection, injury and inflammation [156]. Murine models have demonstrated high levels of immunohistochemical staining in collagen- or adjuvant- induced arthritis [157], a five fold elevation in the synovial fluid of rheumatoid compared to osteoarthritis patients [158], and increased serum levels during active RA [157]. Taken together, the available data strongly suggest that various RAGE ligands are proinflammatory, mediated by the RAGE receptor.

Functional polymorphisms have been described in coding and non coding regions of the RAGE gene, several may be associated with vascular disease [159–162], however most are beyond the scope of this review, which will cover only one.

#### *RAGE: Glycine 82 Serine polymorphism*

The location of the RAGE gene in the HLA region on chromosome 6 [123] has implications for RA. Like many HLA genes implicated in genetic control of the immune response, the RAGE gene has a number of polymorphisms described in the recognition coding regions [161] one of which, a Glycine to Serine substitution at position 82 (82Ser; SNP: rs2070600), has a significant prevalence and results in a functional amino acid change in the RAGE molecule [163]. This amino acid shift occurs at a predicted N-linked glycosylation site in the same immunoglobulin variable domain as the AGE binding site [163]. The tertiary structure of the RAGE molecule however, is unknown, and thus structural effects of this polymorphism are unable to be predicted [163]. However Hofmann et al have presented evidence that 82Ser may upregulate the immune response. Amongst other effects, they demonstrated human mononuclear phagocytes of healthy subjects bearing 82Ser, when exposed to S100A12, secreted significantly greater levels of TNF $\alpha$  than subjects bearing the Glycine82 allele [148].

The allele frequency of 82Ser varies between 0 to 29%, with significant differences between ethnic groups [163–165]. Allele frequencies do not vary in type I or II

diabetics compared with non-diabetic controls [161, 163], however 82Ser has been shown to be increased in subjects with RA [148]. The location of the gene within the HLA suggests that linkage disequilibrium with known RA-associated DR genes is likely. When Hofmann et al sub-analysed their cohort for the presence or absence of DRB1\*0401, they were unable to demonstrate a statistically significant association between the 82Ser allele and RA [148], suggesting the RAGE polymorphism was in linkage disequilibrium with DRB1\*0401, and thus accounting for its prevalence in the RA population. The TNF $\alpha$  gene is also located in the MHC class III region, and likewise variant alleles have been demonstrated to be in strong linkage disequilibrium with certain RA associated haplotypes, making it highly unlikely that individual polymorphisms in the TNF $\alpha$  gene would be of primary importance in RA [166, 167]. Of interest, given the close proximity of RAGE to TNF $\alpha$ , these two genes may be linked. However, given the ability of RAGE signalling to control TNF production, they may also be linked functionally.

Several studies have addressed whether the polymorphism is associated with risk of micro- or macro-vascular disease in type I or type II diabetes [160, 161, 163, 164, 168, 169]. Prevost et al found that 82Ser is significantly associated with risk of developing advanced nephropathy in Caucasian type I diabetic patients [160], whilst Kumaramanickavel et al found it was significantly associated with a low risk of developing diabetic retinopathy in Asian Indian patients who have type II diabetes [169]. Since DR4 is also associated with development of type I but not type II diabetes, it is possible that other genes in the same HLA region may influence the response of RAGE.

#### **Summary**

Patients with RA have a considerably increased risk of cardiovascular morbidity and mortality, which is likely to be at least partly explained by inflammation and altered immune mechanisms, although RA patients may be carriers of a higher load of conventional risk factors. RA and cardiovascular disease have some striking similarities. The RAGE receptor has a role in sustaining chronic inflammation and in autoimmune disease. A polymorphism of RAGE that is enriched in RA could potentially have a role in CV disease expression, as well as on RA itself. As yet, the exact role of RAGE in perpetuation of rheumatoid arthritis, vascular inflammation or perturbed immunity is unclear, but is the subject of ongoing research.

#### **References**

1. Wolfe F, et al. (1994) The mortality of rheumatoid arthritis. *Arthritis Rheum* 37(4): 481–494
2. Bjornadal L, et al. (2002) Decreasing mortality in patients with rheumatoid arthritis: results from a large population based cohort in Sweden, 1964–95. *J Rheumatol* 29(5): 906–912
3. Symmons DP, et al. (1998) Longterm mortality outcome in patients with rheumatoid arthritis: early presenters continue to do well. *J Rheumatol* 25(6): 1072–1077

4. Goodson NJ, et al. (2002) Mortality in early inflammatory polyarthritis: cardiovascular mortality is increased in seropositive patients. *Arthritis Rheum* 46(8): 2010–2019
5. Mutru O, et al. (1989) Cardiovascular mortality in patients with rheumatoid arthritis. *Cardiology* 76(1): 71–77
6. Maradit-Kremers H, et al. (2005) Cardiovascular death in rheumatoid arthritis: a population-based study. *Arthritis Rheum* 52(3): 722–732
7. Mikuls TR, Saag KG (2001) Comorbidity in rheumatoid arthritis. *Rheum Dis Clin North Am* 27(2): 283–303
8. Kroot EJ, et al. (2001) Chronic comorbidity in patients with early rheumatoid arthritis: a descriptive study. *J Rheumatol* 28(7): 1511–1517
9. Wallberg-Jonsson S, ML Ohman, SR Dahlqvist (1997) Cardiovascular morbidity and mortality in patients with seropositive rheumatoid arthritis in Northern Sweden. *J Rheumatol* 24(3): 445–451
10. Heller RF, et al. (1984) How well can we predict coronary heart disease? Findings in the United Kingdom heart disease prevention project. *Br Med J (Clin Res Ed)* 288(6428): 1409–1411
11. Braunwald E (1997) Shattuck lecture – cardiovascular medicine at the turn of the millennium: triumphs, concerns, and opportunities. *N Engl J Med* 337(19): 1360–1369
12. Oliveira GH (2005) Novel serologic markers of cardiovascular risk. *Curr Atheroscler Rep* 7(2): 148–154
13. Nygard O, et al. (1997) Plasma homocysteine levels and mortality in patients with coronary artery disease. *N Engl J Med* 337(4): 230–236
14. Whittle SL, RA Hughes (2004) Folate supplementation and methotrexate treatment in rheumatoid arthritis: a review. *Rheumatology (Oxford)* 43(3): 267–271
15. Busso N, JA Hamilton (2002) Extravascular coagulation and the plasminogen activator/plasmin system in rheumatoid arthritis. *Arthritis Rheum* 46(9): 2268–2279
16. Biasucci LM, et al. (1999) Elevated levels of C-reactive protein at discharge in patients with unstable angina predict recurrent instability. *Circulation* 99(7): 855–860
17. Buffon A, et al. (1999) Preprocedural serum levels of C-reactive protein predict early complications and late restenosis after coronary angioplasty. *J Am Coll Cardiol* 34(5): 1512–1521
18. Haverkate F, et al. (1997) Production of C-reactive protein and risk of coronary events in stable and unstable angina. European concerted action on thrombosis and disabilities angina pectoris study group. *Lancet* 349(9050): 462–466
19. Liuzzo G, et al. (1994) The prognostic value of C-reactive protein and serum amyloid A protein in severe unstable angina. *N Engl J Med* 331(7): 417–424
20. Mendall MA, et al. (1996) C reactive protein and its relation to cardiovascular risk factors: a population based cross sectional study. *Bmj* 312(7038): 1061–1065
21. Ridker PM, et al. (2000) C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. *N Engl J Med* 342(12): 836–843
22. Ridker PM, et al. (2001) Measurement of C-reactive protein for the targeting of statin therapy in the primary prevention of acute coronary events. *N Engl J Med* 344(26): 1959–1965
23. Rost NS, et al. (2001) Plasma concentration of C-reactive protein and risk of ischemic stroke and transient ischemic attack: the Framingham study. *Stroke* 32(11): 2575–2579
24. Wang TJ, et al. (2002) Association of C-reactive protein with carotid atherosclerosis in men and women: the Framingham Heart Study. *Arterioscler Thromb Vasc Biol* 22(10): 1662–1667
25. Biasucci LM, et al. (1996) Elevated levels of interleukin-6 in unstable angina. *Circulation* 94(5): 874–877
26. del Rincon ID, et al. (2001) High incidence of cardiovascular events in a rheumatoid arthritis cohort not explained by traditional cardiac risk factors. *Arthritis Rheum* 44(12): 2737–2745
27. McEntegart A, et al. (2001) Cardiovascular risk factors, including thrombotic variables, in a population with rheumatoid arthritis. *Rheumatology (Oxford)* 40(6): 640–644
28. Banks M, Flint EJ, Bacon PA, et al. (1998) Expression and prevalence of ischemic heart disease in rheumatoid arthritis {abstract}. *Arthritis and Rheumatism* 41(s20)
29. Banks M, Flint EJ, Bacon PA, et al. (2000) Rheumatoid arthritis is an independent risk factor for ischaemic heart disease {abstract}. *Arthritis and Rheumatism* 43(Suppl 9): S385
30. Banks M., Pace A, Kitis GD (2001) Acute coronary syndromes present atypically and recur more frequently in rheumatoid arthritis patients than matched controls {abstract}. *Arthritis and Rheumatism* 44(Suppl): S54
31. Wislowska M, Sypula S, Kowalik I (1998) Echocardiographic findings, 24-hour electrocardiographic Holter monitoring in patients with rheumatoid arthritis according to Steinbrocker's criteria, functional index, value of Waaler-Rose titre and duration of disease. *Clin Rheumatol* 17(5): 369–377
32. Gonzalez-Juanatey C, et al. (2004) Echocardiographic and Doppler findings in long-term treated rheumatoid arthritis patients without clinically evident cardiovascular disease. *Semin Arthritis Rheum* 33(4): 231–238
33. Wong M, et al. (2003) Reduced arterial elasticity in rheumatoid arthritis and the relationship to vascular disease risk factors and inflammation. *Arthritis Rheum* 48(1): 81–89
34. Bergholm R, et al. (2002) Impaired responsiveness to NO in newly diagnosed patients with rheumatoid arthritis. *Arterioscler Thromb Vasc Biol* 22(10): 1637–1641
35. Van Doornum S, et al. (2003) Screening for atherosclerosis in patients with rheumatoid arthritis: comparison of two in vivo tests of vascular function. *Arthritis Rheum* 48(1): 72–80
36. Goodson NJ, et al. (2005) Baseline levels of C-reactive protein and prediction of death from cardiovascular disease in patients with inflammatory polyarthritis: a ten-year follow-up study of a primary care-based inception cohort. *Arthritis Rheum* 52(8): 2293–2299
37. Kaplan MJ, WJ McCune (2003) New evidence for vascular disease in patients with early rheumatoid arthritis. *Lancet* 361(9363): 1068–1069
38. Wang L, Feng G (2004) Rheumatoid arthritis increases the risk of coronary heart disease via vascular endothelial injuries. *Med Hypotheses* 63(3): 442–445
39. Ross R (1999) Atherosclerosis – an inflammatory disease. *N Engl J Med* 340(2): 115–26
40. Park YB, et al. (1999) Lipid profiles in untreated patients with rheumatoid arthritis. *J Rheumatol* 26(8): 1701–1704
41. Situnayake RD, Kitis G (1997) Dyslipidemia and rheumatoid arthritis. *Ann Rheum Dis* 56(6): 341–342
42. Rantapaa-Dahlqvist S, Wallberg-Jonsson S, Dahlen G (1991) Lipoprotein (a), lipids, and lipoproteins in patients

- with rheumatoid arthritis. *Ann Rheum Dis* 50(6): 366–368
43. Lakatos J, Harsanyi A (1988) Serum total, HDL, LDL cholesterol, and triglyceride levels in patients with rheumatoid arthritis. *Clin Biochem* 21(2): 93–96
  44. Boers M, et al. (2003) Influence of glucocorticoids and disease activity on total and high density lipoprotein cholesterol in patients with rheumatoid arthritis. *Ann Rheum Dis* 62(9): 842–845
  45. Yoo WH (2004) Dyslipoproteinemia in patients with active rheumatoid arthritis: effects of disease activity, sex, and menopausal status on lipid profiles. *J Rheumatol* 31(9): 1746–1753
  46. Horai R, et al. (2000) Development of chronic inflammatory arthropathy resembling rheumatoid arthritis in interleukin 1 receptor antagonist-deficient mice. *J Exp Med* 191(2) 313–320
  47. Ji H, et al. (2002) Critical roles for interleukin 1 and tumor necrosis factor alpha in antibody-induced arthritis. *J Exp Med* 196(1) 77–85
  48. Firestein GS, et al. (1994) Synovial interleukin-1 receptor antagonist and interleukin-1 balance in rheumatoid arthritis. *Arthritis Rheum* 37(5): 644–652
  49. Chomarat P, et al. (1995) Balance of IL-1 receptor antagonist/IL-1 beta in rheumatoid synovium and its regulation by IL-4 and IL-10. *J Immunol* 154(3): 1432–1439
  50. Chikanza IC, et al. (1995) Dysregulation of the in vivo production of interleukin-1 receptor antagonist in patients with rheumatoid arthritis. Pathogenetic implications. *Arthritis Rheum* 38(5): 642–648
  51. Isoda K, et al. (2005) Deficiency of interleukin-1 receptor antagonist deteriorates fatty liver and cholesterol metabolism in hypercholesterolemic mice. *J Biol Chem* 280(8): 7002–7009
  52. Yudkin JS, et al. (2000) Inflammation, obesity, stress and coronary heart disease: is interleukin-6 the link? *Atherosclerosis* 148(2): 209–214
  53. Nishimoto N, et al. (2004) Treatment of rheumatoid arthritis with humanized anti-interleukin-6 receptor antibody: a multicenter, double-blind, placebo-controlled trial. *Arthritis Rheum* 50(6): 1761–1769
  54. Wallenius V, et al. (2002) Interleukin-6-deficient mice develop mature-onset obesity. *Nat Med* 8(1): 75–79
  55. Munro R, et al. (1997) Effect of disease modifying agents on the lipid profiles of patients with rheumatoid arthritis. *Ann Rheum Dis* 56(6): 374–377
  56. Nurmohamed MT, van Halm VP, Dijkmans BA (2002) Cardiovascular risk profile of antirheumatic agents in patients with osteoarthritis and rheumatoid arthritis. *Drugs* 62(11): 1599–1609
  57. Park YB, et al. (2002) Effects of antirheumatic therapy on serum lipid levels in patients with rheumatoid arthritis: a prospective study. *Am J Med* 113(3): 188–193
  58. Wallberg-Jonsson S, et al. (1999) Extent of inflammation predicts cardiovascular disease and overall mortality in seropositive rheumatoid arthritis. A retrospective cohort study from disease onset. *J Rheumatol* 26(12): 2562–2571
  59. Popa C, et al. (2005) Influence of anti-tumour necrosis factor therapy on cardiovascular risk factors in patients with active rheumatoid arthritis. *Ann Rheum Dis* 64(2): 303–305
  60. Vis M, et al. (2005) Short term effects of infliximab on the lipid profile in patients with rheumatoid arthritis. *J Rheumatol* 32(2): 252–255
  61. Heliövaara M, et al. (1993) Smoking and risk of rheumatoid arthritis. *J Rheumatol* 20(11): 1830–1835
  62. Silman AJ, Newman J, MacGregor AJ (1996) Cigarette smoking increases the risk of rheumatoid arthritis. Results from a nationwide study of disease-discordant twins. *Arthritis Rheum* 39(5): 732–735
  63. Hutchinson D, et al. (2001) Heavy cigarette smoking is strongly associated with rheumatoid arthritis (RA), particularly in patients without a family history of RA. *Ann Rheum Dis* 60(3): 223–227
  64. Wolfe F (2000) The effect of smoking on clinical, laboratory, and radiographic status in rheumatoid arthritis. *J Rheumatol* 27(3): 630–637
  65. Kitas GD, Erb N (2003) Tackling ischaemic heart disease in rheumatoid arthritis. *Rheumatology (Oxford)* 42(5): 607–613
  66. Sowers JR, et al. (2005) The Effects of cyclooxygenase-2 inhibitors and nonsteroidal anti-inflammatory therapy on 24-hour blood pressure in patients with hypertension, osteoarthritis, and type 2 diabetes mellitus. *Arch Intern Med* 165(2): 161–168
  67. Bombardier C, et al. (2000) Comparison of upper gastrointestinal toxicity of rofecoxib and naproxen in patients with rheumatoid arthritis. VIGOR Study Group. *N Engl J Med* 343(21): 1520–1528, 2 p following 1528
  68. Juni P, Reichenbach S, Egger M (2005) COX 2 inhibitors, traditional NSAIDs, and the heart. *Bmj* 330(7504): 1342–1343
  69. Crofford LJ, et al. (2000) Thrombosis in patients with connective tissue diseases treated with specific cyclooxygenase 2 inhibitors. A report of four cases. *Arthritis Rheum* 43(8): 1891–1896
  70. Johnsen SP, et al. (2005) Risk of hospitalization for myocardial infarction among users of rofecoxib, celecoxib, and other NSAIDs: a population-based case-control study. *Arch Intern Med* 165(9): 978–984
  71. Levesque LE, Brophy JM, Zhang B (2005) The risk for myocardial infarction with cyclooxygenase-2 inhibitors: a population study of elderly adults. *Ann Intern Med* 142(7): 481–489
  72. Solomon SD, et al. (2005) Cardiovascular risk associated with celecoxib in a clinical trial for colorectal adenoma prevention. *N Engl J Med* 352(11): 1071–1080
  73. Bresalier RS, et al. (2005) Cardiovascular events associated with rofecoxib in a colorectal adenoma chemoprevention trial. *N Engl J Med* 352(11): 1092–1102
  74. Whelton A, et al. (2001) Cyclooxygenase-2 – specific inhibitors and cardiorenal function: a randomized, controlled trial of celecoxib and rofecoxib in older hypertensive osteoarthritis patients. *Am J Ther* 8(2): 85–95
  75. Collins R, et al. (1990) Blood pressure, stroke, and coronary heart disease. Part 2, Short-term reductions in blood pressure: overview of randomised drug trials in their epidemiological context. *Lancet* 335(8693): 827–838
  76. Dessein PH, Stanwix AE, Joffe BI (2002) Cardiovascular risk in rheumatoid arthritis versus osteoarthritis: acute phase response related decreased insulin sensitivity and high-density lipoprotein cholesterol as well as clustering of metabolic syndrome features in rheumatoid arthritis. *Arthritis Res* 4(5): R5
  77. Kernan WN, Inzucchi SE (2004) Type 2 Diabetes Mellitus and Insulin Resistance: Stroke Prevention and Management. *Curr Treat Options Neurol* 6(6): 443–450
  78. Reaven GM (1988) Banting lecture 1988. Role of insulin resistance in human disease. *Diabetes* 37(12): 1595–1607
  79. Yudkin JS, et al. (1999) C-reactive protein in healthy subjects: associations with obesity, insulin resistance, and



- endothelial dysfunction: a potential role for cytokines originating from adipose tissue? *Arterioscler Thromb Vasc Biol* 19(4): 972–978
80. Kaplan MH, Volanakis JE (1974) Interaction of C-reactive protein complexes with the complement system. I. Consumption of human complement associated with the reaction of C-reactive protein with pneumococcal C-polysaccharide and with the choline phosphatides, lecithin and sphingomyelin. *J Immunol* 112(6): 2135–2147
  81. Ballou SP, Lozanski G (1992) Induction of inflammatory cytokine release from cultured human monocytes by C-reactive protein. *Cytokine* 4(5): 361–368
  82. Gershov D, et al. (2000) C-Reactive protein binds to apoptotic cells, protects the cells from assembly of the terminal complement components, and sustains an anti-inflammatory innate immune response: implications for systemic autoimmunity. *J Exp Med* 192(9): 1353–1364
  83. Familian A, et al. (2001) Chromatin-independent binding of serum amyloid P component to apoptotic cells. *J Immunol* 167(2): 647–654
  84. Du Clos TW (2003) C-reactive protein as a regulator of autoimmunity and inflammation. *Arthritis Rheum* 48(6):1475–1477
  85. Danesh J, et al. (2000) Low grade inflammation and coronary heart disease: prospective study and updated meta-analyses. *Bmj* 321(7255): P 199–204
  86. Ridker PM, et al. (1997) Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men. *N Engl J Med* 336(14): 973–979
  87. O’Leary DH, et al. (1999) Carotid-artery intima and media thickness as a risk factor for myocardial infarction and stroke in older adults. Cardiovascular Health Study Collaborative Research Group. *N Engl J Med* 340(1): 14–22
  88. Kawamori R, et al. (1992) Prevalence of carotid atherosclerosis in diabetic patients. Ultrasound high-resolution B-mode imaging on carotid arteries. *Diabetes Care* 15(10): 1290–1294
  89. Wofford JL, et al. (1991) Relation of extent of extracranial carotid artery atherosclerosis as measured by B-mode ultrasound to the extent of coronary atherosclerosis. *Arterioscler Thromb* 11(6): 1786–1794
  90. Yamasaki Y, et al. (1994) Atherosclerosis in carotid artery of young IDDM patients monitored by ultrasound high-resolution B-mode imaging. *Diabetes* 43(5): 634–639
  91. van der Meer IM, et al. (2002) Inflammatory mediators and cell adhesion molecules as indicators of severity of atherosclerosis: the Rotterdam Study. *Arterioscler Thromb Vasc Biol* 22(5): 838–842
  92. Pasceri V, Willerson JT, Yeh ET (2000) Direct proinflammatory effect of C-reactive protein on human endothelial cells. *Circulation* 102(18): 2165–2168
  93. Verma S, et al. (2002) Endothelin antagonism and interleukin-6 inhibition attenuate the proatherogenic effects of C-reactive protein. *Circulation* 105(16): 1890–1896
  94. Pasceri V, et al. (2001) Modulation of C-reactive protein-mediated monocyte chemoattractant protein-1 induction in human endothelial cells by anti-atherosclerosis drugs. *Circulation* 103(21): 2531–2534
  95. Zwaka TP, Hombach V, Torzewski J (2001) C-reactive protein-mediated low density lipoprotein uptake by macrophages: implications for atherosclerosis. *Circulation* 103(9): 1194–1197
  96. Yasojima K, et al. (2001) Generation of C-reactive protein and complement components in atherosclerotic plaques. *Am J Pathol* 158(3): 1039–1051
  97. Verma S, et al. (2002) A self-fulfilling prophecy: C-reactive protein attenuates nitric oxide production and inhibits angiogenesis. *Circulation* 106(8): 913–919
  98. Blake GJ, Ridker PM (2002) C-reactive protein, subclinical atherosclerosis, and risk of cardiovascular events. *Arterioscler Thromb Vasc Biol* 22(10): 1512–1513
  99. Maksimowicz-McKinnon K, Bhatt DL, Calabrese LH (2004) Recent advances in vascular inflammation: C-reactive protein and other inflammatory biomarkers. *Curr Opin Rheumatol* 16(1): 18–24
  100. Oppenheimer-Marks N, Lipsky PE (1998) Adhesion molecules in rheumatoid arthritis. *Springer Semin Immunopathol* 20(1–2): 95–114
  101. Basta G, et al. (2002) Advanced glycation end products activate endothelium through signal-transduction receptor RAGE: a mechanism for amplification of inflammatory responses. *Circulation* 105(7): 816–822
  102. Dessein PH, Joffe BI, Singh S (2005) Biomarkers of endothelial dysfunction, cardiovascular risk factors and atherosclerosis in rheumatoid arthritis. *Arthritis Res Ther* 7(3): R634–643
  103. Wallberg-Jonsson S, Cederfelt M, Rantapaa Dahlqvist S (2000) Hemostatic factors and cardiovascular disease in active rheumatoid arthritis: an 8 year follow-up study. *J Rheumatol* 27(1): 71–75
  104. Bulut D, et al. (2003) Selective cyclo-oxygenase-2 inhibition with parecoxib acutely impairs endothelium-dependent vasodilatation in patients with essential hypertension. *J Hypertens* 21(9): 1663–1667
  105. McAdam BF, et al. (1999) Systemic biosynthesis of prostacyclin by cyclooxygenase (COX)-2: the human pharmacology of a selective inhibitor of COX-2. *Proc Natl Acad Sci U S A* 96(1): 272–277
  106. Moncada S, Higgs EA, Vane JR (1977) Human arterial and venous tissues generate prostacyclin (prostaglandin x), a potent inhibitor of platelet aggregation. *Lancet* 1(8001): 18–20
  107. McGiff JC, et al. (1970) Prostaglandin-like substances appearing in canine renal venous blood during renal ischemia. Their partial characterization by pharmacologic and chromatographic procedures. *Circ Res* 27(5): 765–782
  108. Yamamoto T, et al. (1999) Production of prostanoids and nitric oxide by infarcted heart in situ and the effect of aspirin. *Biochem Biophys Res Commun* 257(2): 488–493
  109. Shimura K, et al. (2000) Cyclooxygenase-2 mediates the cardioprotective effects of the late phase of ischemic preconditioning in conscious rabbits. *Proc Natl Acad Sci U S A* 97(18): 10197–10202
  110. Hennen JK, et al. (2001) Effects of selective cyclooxygenase-2 inhibition on vascular responses and thrombosis in canine coronary arteries. *Circulation* 104(7): 820–825
  111. Burke A, Fitzgerald GA (2003) Oxidative stress and smoking-induced vascular injury. *Prog Cardiovasc Dis* 46(1): 79–90
  112. Glagov S, et al. (1987) Compensatory enlargement of human atherosclerotic coronary arteries. *N Engl J Med* 316(22): 1371–1375
  113. Freiman PC, et al. (1986) Atherosclerosis impairs endothelium-dependent vascular relaxation to acetylcholine and thrombin in primates. *Circ Res* 58(6): 783–789

114. Adams MR, et al. (2000) Atherogenic lipids and endothelial dysfunction: mechanisms in the genesis of ischemic syndromes. *Annu Rev Med* 51: 149–167
115. Hansson GK, et al. (1989) Immune mechanisms in atherosclerosis. *Arteriosclerosis* 9(5): 567–578
116. Jonasson L, et al. (1986) Regional accumulations of T cells, macrophages, and smooth muscle cells in the human atherosclerotic plaque. *Arteriosclerosis* 6(2): 131–138
117. Folcik VA, Aamir R, Cathcart MK (1997) Cytokine modulation of LDL oxidation by activated human monocytes. *Arterioscler Thromb Vasc Biol* 17(10): 1954–1961
118. Stone PH, Sherrid MV, Cohn KE (1981) Correlation of HLA types in premature coronary artery disease: an attempt to define independent genetic risk factors. *Chest* 79(4): 381–385
119. Dogan Y, et al. (2001) Relation of HLA antigens and myocardial infarction {abstract}. *Anadolu Kardiyol Derg* 1(2): 80–84, AXIII
120. Diamantopoulos EJ, et al. (2002) Association of the HLA antigens with early atheromatosis in subjects with type 2 diabetes mellitus. *Int Angiol* 21(4): 379–383
121. Jonasson L, et al. (1997) Lipoprotein (a) and HLA-DRB1 and -DQB1 genes in coronary artery disease. *Atherosclerosis* 133(1): 111–114
122. Dahlen GH, Slunga L, Lindblom B (1994) Importance of Lp(a) lipoprotein and HLA genotypes in atherosclerosis and diabetes. *Clin Genet* 46(1 Spec No): 46–51
123. Sugaya K, et al. (1994) Three genes in the human MHC class III region near the junction with the class II: gene for receptor of advanced glycosylation end products, PBX2 homeobox gene and a notch homolog, human counterpart of mouse mammary tumor gene int-3. *Genomics* 23(2): 408–419
124. Medzhitov R, Janeway C Jr (2000) Innate immunity. *N Engl J Med* 343(5): 338–344
125. Schmidt AM, et al. (2001) The multiligand receptor RAGE as a progression factor amplifying immune and inflammatory responses. *J Clin Invest* 108(7): 949–955
126. Miyata T, et al. (1996) The receptor for advanced glycation end products (RAGE) is a central mediator of the interaction of AGE-beta2microglobulin with human mononuclear phagocytes via an oxidant-sensitive pathway. Implications for the pathogenesis of dialysis-related amyloidosis. *J Clin Invest* 98(5): 1088–1094
127. Hofmann MA, et al. (1999) RAGE mediates a novel proinflammatory axis: a central cell surface receptor for S100/calgranulin polypeptides. *Cell* 97(7): 889–901
128. Yan SD, et al. (1996) RAGE and amyloid-beta peptide neurotoxicity in Alzheimer's disease. *Nature* 382(6593): 685–691
129. Hori O, et al. (1995) The receptor for advanced glycation end products (RAGE) is a cellular binding site for amphotericin. Mediation of neurite outgrowth and co-expression of rage and amphotericin in the developing nervous system. *J Biol Chem* 270(43): 25752–25761
130. King GL, Brownlee M (1996) The cellular and molecular mechanisms of diabetic complications. *Endocrinol Metab Clin North Am* 25(2): 255–270
131. Yan SD, et al. (2000) Receptor-dependent cell stress and amyloid accumulation in systemic amyloidosis. *Nat Med* 6(6): 643–651
132. Tanji N, et al. (2000) Expression of advanced glycation end products and their cellular receptor RAGE in diabetic nephropathy and nondiabetic renal disease. *J Am Soc Nephrol* 11(9): 1656–1666
133. Yan SD, et al. (1994) Enhanced cellular oxidant stress by the interaction of advanced glycation end products with their receptors/binding proteins. *J Biol Chem* 269(13): 9889–9897
134. Lander HM, et al. (1997) Activation of the receptor for advanced glycation end products triggers a p21(ras)-dependent mitogen-activated protein kinase pathway regulated by oxidant stress. *J Biol Chem* 272(28): 17810–17814
135. Schmidt AM, et al. (1999) Activation of receptor for advanced glycation end products: a mechanism for chronic vascular dysfunction in diabetic vasculopathy and atherosclerosis. *Circ Res* 84(5): 489–497
136. Sakaguchi T, et al. (2003) Central role of RAGE-dependent neointimal expansion in arterial restenosis. *J Clin Invest* 111(7): 959–972
137. Yokoi M, et al. (2005) Elevations of AGE and vascular endothelial growth factor with decreased total antioxidant status in the vitreous fluid of diabetic patients with retinopathy. *Br J Ophthalmol* 89(6): 673–675
138. Horie K, et al. (1997) Immunohistochemical colocalization of glycoxidation products and lipid peroxidation products in diabetic renal glomerular lesions. Implication for glycoxidative stress in the pathogenesis of diabetic nephropathy. *J Clin Invest* 100(12): 2995–3004
139. Falcone C, et al. (2005) Plasma levels of soluble receptor for advanced glycation end products and coronary artery disease in nondiabetic men. *Arterioscler Thromb Vasc Biol* 25(5): 1032–1037
140. Wautier JL, et al. (1996) Receptor-mediated endothelial cell dysfunction in diabetic vasculopathy. Soluble receptor for advanced glycation end products blocks hyperpermeability in diabetic rats. *J Clin Invest* 97(1): 238–243
141. Schmidt AM, et al. (1995) Advanced glycation endproducts interacting with their endothelial receptor induce expression of vascular cell adhesion molecule-1 (VCAM-1) in cultured human endothelial cells and in mice. A potential mechanism for the accelerated vasculopathy of diabetes. *J Clin Invest* 96(3): 1395–1403
142. Bucala R, Tracey KJ, Cerami A (1991) Advanced glycosylation products quench nitric oxide and mediate defective endothelium-dependent vasodilatation in experimental diabetes. *J Clin Invest* 87(2): 432–438
143. Esposito C, et al. (1989) Endothelial receptor-mediated binding of glucose-modified albumin is associated with increased monolayer permeability and modulation of cell surface coagulant properties. *J Exp Med* 170(4): 1387–1407
144. Shanmugam N, et al. (2003) Regulation of cyclooxygenase-2 expression in monocytes by ligation of the receptor for advanced glycation end products. *J Biol Chem* 278(37): 34834–34844
145. Park L, et al. (2004) Suppression of accelerated diabetic atherosclerosis by the soluble receptor for advanced glycation endproducts. *Nat Med* 4(9): 1025–1031
146. Wautier JL, Schmidt AM (2004) Protein glycation: a firm link to endothelial cell dysfunction. *Circ Res* 95(3): 233–238
147. Bierhaus A, et al. (2004) Loss of pain perception in diabetes is dependent on a receptor of the immunoglobulin superfamily. *J Clin Invest* 114(12): 1741–1751
148. Hofmann MA, et al. (2002) RAGE and arthritis: the G82S polymorphism amplifies the inflammatory response. *Genes Immun* 3(3): 123–135

149. Drinda S, et al. (2005) Identification of the receptor for advanced glycation end products in synovial tissue of patients with rheumatoid arthritis. *Rheumatol Int* 25: 411–413
150. Vogl T, et al. (1999) S100A12 is expressed exclusively by granulocytes and acts independently from MRP8 and MRP14. *J Biol Chem* 274(36): 25291–25296
151. Foell D, et al. (2004) Monitoring neutrophil activation in juvenile rheumatoid arthritis by S100A12 serum concentrations. *Arthritis Rheum* 50(4): 1286–1295
152. Foell D, et al. (2003) S100A12 (EN-RAGE) in monitoring Kawasaki disease. *Lancet* 361(9365): 1270–1272
153. Foell D, et al. (2003) Expression of the pro-inflammatory protein S100A12 (EN-RAGE) in rheumatoid and psoriatic arthritis. *Rheumatology (Oxford)* 42(11): 1383–1389
154. Youssef P, et al. (1999) Expression of myeloid related proteins (MRP) 8 and 14 and the MRP8/14 heterodimer in rheumatoid arthritis synovial membrane. *J Rheumatol* 26(12): 2523–2528
155. Frosch M, et al. (2000) Myeloid-related proteins 8 and 14 are specifically secreted during interaction of phagocytes and activated endothelium and are useful markers for monitoring disease activity in pauciarticular-onset juvenile rheumatoid arthritis. *Arthritis Rheum* 43(3): 628–637
156. Lotze MT, Tracey KJ (2005) High-mobility group box 1 protein (HMGB1): nuclear weapon in the immune arsenal. *Nat Rev Immunol* 5(4): 331–342
157. Ulloa L, et al. (2003) High mobility group box chromosomal protein 1 as a nuclear protein, cytokine, and potential therapeutic target in arthritis. *Arthritis Rheum* 48(4): 876–881
158. Taniguchi N, et al. (2003) High mobility group box chromosomal protein 1 plays a role in the pathogenesis of rheumatoid arthritis as a novel cytokine. *Arthritis Rheum* 48(4): 971–981
159. dos Santos KG, et al. (2005) The -374A allele of the receptor for advanced glycation end products gene is associated with a decreased risk of ischemic heart disease in African-Brazilians with type 2 diabetes. *Mol Genet Metab* 85(2): 149–156
160. Prevost G, et al. (2005) Polymorphisms of the receptor of advanced glycation endproducts (RAGE) and the development of nephropathy in type 1 diabetic patients. *Diabetes Metab* 31(1): 35–39
161. Prevost G, et al. (1999) Human RAGE GLY82SER dimorphism and HLA class II DRB1-DQA1-DQB1 haplotypes in type 1 diabetes. *Eur J Immunogenet* 26(5): 343–348
162. Hudson BI, et al. (2001) Study of the -429 T/C and -374 T/A receptor for advanced glycation end products promoter polymorphisms in diabetic and nondiabetic subjects with macrovascular disease. *Diabetes Care* 24(11): 2004
163. Hudson BI, Stickland MH, Grant PJ (1998) Identification of polymorphisms in the receptor for advanced glycation end products (RAGE) gene: prevalence in type 2 diabetes and ethnic groups. *Diabetes* 47(7): 1155–1157
164. Liu L, Xiang K (1999) RAGE Gly82Ser polymorphism in diabetic microangiopathy. *Diabetes Care* 22(4): 646
165. [www.hapmap.org/cgi-perl/gbrowse/gbrowse/hapmap](http://www.hapmap.org/cgi-perl/gbrowse/gbrowse/hapmap)
166. Newton J, et al. (2003) The effect of HLA-DR on susceptibility to rheumatoid arthritis is influenced by the associated lymphotoxin alpha-tumor necrosis factor haplotype. *Arthritis Rheum* 48(1): 90–96
167. Newton JL, et al (2004) Dissection of class III major histocompatibility complex haplotypes associated with rheumatoid arthritis. *Arthritis Rheum* 50(7): 2122–2129
168. Pulkkinen A, et al (2000) Gly82Ser polymorphism of the receptor of advanced glycation end product gene is not associated with coronary heart disease in Finnish nondiabetic subjects or in patients with type 2 diabetes. *Diabetes Care* 23(6): 864
169. Kumaramanickavel G, et al. (2002) Association of Gly82Ser polymorphism in the RAGE gene with diabetic retinopathy in type II diabetic Asian Indian patients. *J Diabetes Complications* 16(6): 391–394