RESEARCH

Anticoagulant rodenticides exposure status among wild pit vipers (*Protobothrops flavoviridis***) and green anoles (***Anolis carolinensis***) in two Japanese Islands**

Yoshiya Yamamura¹ • Shintaro Nakagawa² • Mitsuki Kondo³ • So Shinya⁴ • Rio Doya² • Masashi Koide² • **Yared Beyene Yohannes2 · Yoshinori Ikenaka2,5,6,7 · Mayumi Ishizuka2 · Shouta M. M. Nakayama2,8**

Received: 22 January 2024 / Revised: 5 May 2024 / Accepted: 7 May 2024 / Published online: 17 May 2024 © The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2024

Abstract

To investigate the exposure of wild reptiles to anticoagulant rodenticides (ARs), liver ARs concentrations were analyzed using LCMS in individuals captured within ARs application areas. A total of 185 pit vipers (*Protobothrops flavoviridis*) from Amami Oshima Island and 89 green anoles (*Anolis carolinensis*) from Ogasawara Island were captured. Among the pit vipers, approximately 9% of the samples showed detection of three ARs (warfarin (WF), diphacinone (DP), and coumatetralyl (CT), with a maximum concentration of 436.5 ng/g. Additionally, the WF metabolite 10-OH WF was detected in 4 out of 10 individuals where WF was detected. No correlation was found between ARs detection rates and sex or sampling seasons, and no hemorrhaging signs were observed in pit vipers with ARs detection. In green anoles, DP was detected in only one individual at a concentration of 51.9 ng/g. Although high ARs accumulation was not observed, careful monitoring is needed as lizards may serve as exposure resources for higher−tier consumers.

Keywords Anticoagulant rodenticides · Reptiles · Secondary poisoning

Introduction

Anticoagulant rodenticides (ARs), which are also known as vitamin K antagonists, are widely used to control pests such as invasive rodents. They cause chronic bleeding by inhibiting vitamin K epoxide reductase. Depending on their

Yoshiya Yamamura and Shintaro Nakagawa contributed equally to this work.

 \boxtimes Shouta M. M. Nakayama shouta-nakayama@vetmed.hokudai.ac.jp; shoutanakayama0219@gmail.com

- ¹ The Institute of Environmental Toxicology (IET), 4321, Uchimoriya-machi, Joso-shi, Ibaraki 303-0043, Japan
- Laboratory of Toxicology, Department of Environmental Veterinary Sciences, Faculty of Veterinary Medicine, Hokkaido University, Kita 18 Nishi 9, Kita-ku, Sapporo 060-0818, Japan
- ³ National Institute for Environmental Studies, 16-2 Onogawa City of Tsukuba, Ibaraki 305-8506, Japan

toxicities, ARs are classified into two main groups: first− generation (FGARs) and second−generation (SGARs). While SGARs, such as bromadiolone (BM), difenacoum (DF), and brodifacoum (BF), cause lethal effects in a single intake due to their high toxicity and long−acting time, the FGARs (warfarin (WF), diphacinone (DP),

- ⁴ Amami Wildlife Medical Center, JP, 11-26, Naze Ishibashicho, Amami-City, Kagoshima, Japan
- ⁵ Water Research Group, Unit for Environmental Sciences and Management, North-West University, 11 Hoffman Street, Potchefstroom 2531, South Africa
- Translational Research Unit, Veterinary Teaching Hospital, Faculty of Veterinary Medicine, Hokkaido University, Sapporo, Japan
- ⁷ One Health Research Center, Hokkaido University, Sapporo, Japan
- School of Veterinary Medicine, The University of Zambia, Great East Road, P.O. Box 32379, Lusaka 10101, Zambia

and chlorophacinone) need multiple ingestions for killing rodents (Damin-Pernik et al. [2017.](#page-3-0) Although ARs are recognized as an effective tool for controlling pests, concerns regarding their detrimental impacts on non-target wild species are growing. For example, according to 30 studies from 1998 to 2015, ARs were detected in 55% of liver samples from non-target species (2694/4891 samples) (Nakayama et al. [2019\)](#page-3-1). However, the target organisms of these field surveys are primarily mammals and avian species, and there are few reports on the exposure status of reptiles in ARs application areas. Therefore, this study aimed to assess the ARs exposure status of wild reptiles living in ARs application sites.

Amami Oshima Island, located approximately 1,400 km southeast of Tokyo, was treated with toxic DP baits in 2017 and 2018 to eradicate the invasive Philippine mongoose (*Herpestes auropunctatus*) (Abe [2022\)](#page-3-2). Pit vipers (*Protobothrops flavoviridis*) are common reptiles on the island and prey on rodents in the wild (Koba [1963](#page-3-3); Mori and Moriguchi [1988\)](#page-3-4); hence we chose them as ARs exposure models.

Ogasawara Islands, volcanic islands located approximately 1,000 km south of Tokyo, have implemented an ARs application program to combat the population of invasive black rats (*Rattus rattus*) or Norway rats (*Rattus norvegicus*). This program involves the aerial dispersion of DP via helicopters and the placement of toxic DP bait stations across the islands (Kawakami [2019\)](#page-3-5). Green anoles (*Anolis carolinensis*), small to medium−sized lizards, are also famous alien species on the islands. These anoles can be exposed to ARs through accidental ingestion of toxic baits or secondary exposure via the consumption of ARs−contaminated insects. Moreover, they could potentially serve as a source of exposure for higher−tier predators. Therefore, we chose anoles as the second ARs exposure model for reptiles.

Materials and methods

Detailed information on the analysis was presented in supplementary information. In brief, 185 pit vipers (male: 114, female: 71) captured in the Amami Oshima Island under the Habu Capture Encouragement and Purchase Project from February to October 2023 were obtained from the Naze Public Health Center (Kagoshima, Japan). 89 green anoles (male: 53, female: 32, unknown: 4) captured in the Ogasawara islands (Ani-jima, Chichi-jima, and Haha-jima islands) from July 2022 to September 2023 were obtained from the Kanto Regional Environment Office and the Ogasawara Wildlife Research Society. For both species, body length (from cloaca to snout tip) and body weight as well as head length for pit vipers were measured, and sex was determined. Livers, kidneys, intestines, muscles, and stomach and intestine contents were collected and frozen before being sent to Hokkaido University (Sapporo, Japan). Samples were kept at 20 ºC until use. ARs were extracted from livers using a combination of the QuEChERS approach and dispersive solid phase extraction method described by Lawal et al. ([2018\)](#page-3-6). Quantifications of ARs were performed by liquid chromatography coupled to mass spectrometry (1290 Infinity II LC system, 6495B Triple Quad LC/MS; Agilent Technologies, Santa Clara, CA, USA) using a Poroshell 120 column (EC-C18, φ1.9 μm, 3.0×100 mm; Agilent Technologies, Santa Clara, CA, USA).

Results

Table [1](#page-1-0) shows the number of pit vipers in which ARs were detected. Of the 145 pit vipers surveyed, 13 individuals (approximately 9%) showed ARs residue levels above the limit of quantification (LOQ) (1.6 ng/g) . the detected ARs were all FGARs (WF, DP, and CT), but no SGARs were found. In order to understand the metabolic profiles of

Table 1 ARs residual status in wild pit viper livers on the Amami Oshima Island [LOQ: limit of quantification]

2023		$>$ LOO number / Sample number									
		February	March	April	May	June	July	August	September	October	Total
FGARs	WF	0/25	2/20	1/20	0/20	1/20	0/20	3/20	2/20	1/20	10/145
	DP	1/25	1/20	0/20	0/20	0/20	0/20	0/20	0/20	0/20	2/145
	CT	0/25	0/20	0/20	0/20	1/20	0/20	0/20	0/20	0/20	1/145
SGARs	BF	0/25	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/145
	ΒM	0/25	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/145
	DF	0/25	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/145
Total ARs		1/25	3/20	1/20	0/20	2/20	0/20	3/20	2/20	1/20	13/145
WF metabolites	$4'$ -OH	0/25	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/145
	$6-OH$	0/25	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/145
	$7-OH$	0/25	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/145
	$8-OH$	0/25	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/20	0/145
	$10-OH$	0/25	0/20	0/20	0/20	1/20	0/20	2/20	1/20	0/20	4/145

ARs in reptiles, we also analyzed the concentrations of five hydroxylated forms of WF (4'-OH, 6-OH, 7-OH, 8-OH, and 10-OH WF). One of the warfarin metabolites i.e., 10-OH WF was also detected from certain pit vipers. There was no correlation between ARs detection rate and sampling seasons (month (individual numbers in which ARs were detected); February (1), March (3), April (1), May (0), June (2) , July (0) , August (3) , September (2) , and October (1)). In green anoles, only one (out of 89 individuals) showed a DP residue level above LOQ.

Table [2](#page-2-0) provides detailed information about the 13 pit vipers and one green anole in which ARs were detected. In pit vipers, no correlation was observed between sex and ARs residue status (6 males and 7 females). WF was detected in 10 pit vipers at concentrations ranging from 2.19 to 436.48 ng/g, with 10-OH WF also found in 4 of them at concentrations ranging from 14.75 to 167.37 ng/g. DP and CT were also detected in 2 and 1 individual, respectively (concentrations of 32.20 and 36.97 ng/g, 30.3 ng/g, respectively). One male green anole showed a DP residue level of 51.9 ng/g.

Discussion

In this study, various concentrations of FGARs were detected in livers of pit vipers regardless of season, indicating a year−round application of ARs on the island. Although no hemorrhaging signs were observed in individuals where ARs were detected, it remains uncertain whether this absence of symptoms was due to low toxicity or the potential disappearance of signs caused by the prolonged freezing time. Weir et al. [\(2016](#page-3-7)) investigated the toxicity of ARs on western fence lizards (*Sceloporus occidentalis*) and reported low toxicity since LD_{50} values were near 1750 mg/ kg for DP and above 1750 mg/kg for CT. Moreover, Mauldin et al. ([2020\)](#page-3-8) observed no death or hemorrhaging signs in boa constrictors (*Boa constrictor*) despite the liver residual level of DP reaching 890 ng/g. Since this concentration is higher than the maximum level observed in our study (436.5) ng/g of WF), it is likely that none of the 13 pit vipers with ARs residue in livers experienced acute adverse effects. The variation of blood coagulation chemistry between reptiles and poikilotherms may be a key factor in determining susceptibility to ARs. AROCHA-PISIANGO et al. ([1981\)](#page-3-9) reported very high concentrations of plasma fibrinogen in *Caiman crocodilus*. This high fibrinogen level in reptiles could potentially lead to resistance against ARs.

Pit vipers eat not only frogs or lizards but also rodents such as house mice in the wild (Koba [1963;](#page-3-3) Mori and Moriguchi [1988](#page-3-4)). Therefore, secondary ARs poisoning may occur throughout the food chain. In our study, fur believed to be from rodents was found in the intestine of one pit viper (individual number: 9–12, WF concentration: 320.4 ng/g). Watanabe et al. [\(2010](#page-3-10)) demonstrated in vitro experiment using liver microsomes and reported that 4'-OH WF was the most dominant metabolite (70–80%) in avians, while in rats, it accounted for 50% of all metabolites. In addition, Khidkhan et al. ([2024\)](#page-3-11) confirmed that birds administered WF via gavage predominantly exhibited 4'-OH WF in their blood, while 10-OH WF was also detected in some avian species. In turtles, 4'-OH WF seemed to be the major metabolite (Yamamura et al. [2021\)](#page-3-12). Interestingly, our study found that the sole metabolite detected in the livers of pit vipers was 10-OH WF, suggesting a unique WF metabolic system in pit vipers which is distinct from mammals, birds and some reptilian species.

In New Zealand, Wedding et al. [\(2010](#page-3-13)) recorded the first observation of native lizards eating baits containing ARs. Although only one specimen in our study showed a DP level above LOQ, it is plausible that green anoles in the

Table 2 Individual information of pit vipers and a green anole in which ARs were detected

Species	Individual ID	Body weight (g)	Sex	Concentration (ng/g)				
				WF	10-OH WF	DP	CT	
Pit viper	$2 - 4$	546	F			32.2		
	$3 - 4$	701	M	12.9				
	$3 - 10$	583	$\boldsymbol{\mathrm{F}}$	23.9				
	$3 - 19$	463	F			37.0		
	$4 - 5$	690	F	5.8				
	$6 - 5$	635	${\rm F}$				30.3	
	$6 - 15$	792	M	48.5	20.4			
	$8 - 4$	637	M	45.6	14.8			
	$8 - 8$	703	M	436.5	149.4			
	$8 - 20$	867	${\bf F}$	2.8				
	$9 - 11$	425	${\rm F}$	9.4				
	$9 - 12$	722	M	320.4	167.4			
	$10 - 10$	771	M	2.2				
Green anole	$2 - 37$	5.3	M			51.9		

LOQ were 1.6 ng/g for pit vipers and 3.0 ng/g for green anole

Ogasawara islands could inadvertently ingest ARs through toxic baits. Several studies have shown that invertebrates such as insects are exposed to ARs in their natural environments (Alomar et al. [2018](#page-3-14); Dowding et al. [2010\)](#page-3-15). Given that green anoles are insectivores (Stehle et al. [2017](#page-3-16)), food chain transfer could be another possible route for ARs exposure. The concentration of DP in our study was 51.9 ng/g in one green anole and it is lower than 127 ng/g observed in giant ameivas (*Ameiva ameiva*), which showed no clinical signs (Mauldin et al. [2020\)](#page-3-8). Therefore, it is highly probable that this anole experienced no acute toxic effects. According to Kato and Suzuki [\(2005](#page-3-17)), Ogasawara buzzards, an endemic subspecies in the Ogasawara islands, primarily prey mainly on rodents (50.9%) and green anoles (32.7%). Although no significant accumulation of ARs in green anoles was observed in this study, continued monitoring of ARs exposure in anoles is crucial as they may serve as exposure sources for higher−tier consumers.

Supplementary Information The online version contains supplementary material available at [https://doi.org/10.1007/s10344-](https://doi.org/10.1007/s10344-024-01812-4) [024-01812-4.](https://doi.org/10.1007/s10344-024-01812-4)

Author contributions Y.Y and S.N. made research concenpt, analyzed data and wrote the main manuscript text. M.K., S.S., R.D., M.K., Y.B.Y., and Y.I. analyzed data. M.I. and S.M.M.N. made research concenpt, acquired fundings and supervised. All authors reviewed the manuscript.

Funding This study was supported by Grant-in-Aid for Scientific Research, 23H03545 (SMMN) and Nos. 18KK0287, 21H04919, 22KK0163 (MI), as well as by Hokkaido University SOUSEI-TOKU-TEI Specific Research Projects (MI).

Data availability The data is available upon requested to the corresponding author.

Declarations

Ethical approval All the animal samples used in this manuscript were collected under the permission obtained from the Kanto Regional Environment Office and the Ogasawara Wildlife Research Society.

Competing interests There is no Competing Interests in this manuscript.

References

- Abe S (2022) Final stage of invasive alien mongoose eradication from Amami-Oshima Island by the Amami Mongoose Busters. J Japanese Inst Landsc Archit 86:110–113
- Alomar H, Chabert A, Coeurdassier M, Vey D, Berny P (2018) Accumulation of anticoagulant rodenticides (chlorophacinone, bromadiolone and brodifacoum) in a non-target invertebrate, the slug, Deroceras reticulatum. Sci Total Environ 610:576–582
- Arocha-Pisiango C, Ojeda Gorzulas (1981) A., The blood clotting mechanism of spectacled caiman caiman crocodilus
- Damin-Pernik M, Espana B, Lefebvre S, Fourel I, Caruel H, Benoit E, Lattard V (2017) Management of rodent populations by anticoagulant rodenticides: toward third-generation anticoagulant rodenticides. Drug Metab Dispos 45:160–165
- Dowding CV, Shore RF, Worgan A, Baker PJ, Harris S (2010) Accumulation of anticoagulant rodenticides in a non-target insectivore, the European hedgehog (Erinaceus europaeus). Environ Pollut 158:161–166
- Kato Y, Suzuki T (2005) Introduced animals in the diets of the Ogasawara Buzzard, an endemic insular raptor in the Pacific Ocean. J Raptor Res 39:12
- Kawakami K (2019) The history of anthropogenic disturbance and invasive alien species impact on the indigenous avifauna of the Ogasawara Islands, southern Japan. Japanese J Ornithol 68:237–262
- Khidkhan K, Yasuhira F, Saengtienchai A, Kasorndorkbua C, Sitdhibutr R, Ogasawara K, Adachi H, Watanabe Y, Saito K, Sakai H, Horikoshi K, Suzuki H, Kawai YK, Takeda K, Yohannes YB, Ikenaka Y, Rattner BA, Ishizuka M, Nakayama SMM (2024) Evaluation of anticoagulant rodenticide sensitivity by examining in vivo and in vitro responses in avian species, focusing on raptors. Environ Pollut 341:122837
- Koba K (1963) Food of Trimeresurus flavoviridis Flavoviridis and T. okinavensis in the Amami Group of the Loo Choo Islands (supplementary notes, I). Mem Fac Edu Kumamoto Univ Nat Sci 11:35–40
- Lawal A, Wong RCS, Tan GH, Abdulra'uf LB, Alsharif AMA (2018) Recent modifications and validation of QuEChERS-dSPE coupled to LC–MS and GC–MS instruments for determination of Pesticide/Agrochemical residues in fruits and vegetables: review. J Chromatogr Sci 56:656–669
- Mauldin RE, Witmer GW, Shriner SA, Moulton RS, Horak KE (2020) Effects of brodifacoum and diphacinone exposure on four species of reptiles: tissue residue levels and survivorship. Pest Manage Sci 76:1958–1966
- Mori A, Moriguchi H (1988) Food habits of the snakes in Japan: a critical review. Snake 20:98–113
- Nakayama SM, Morita A, Ikenaka Y, Mizukawa H, Ishizuka M (2019) A review: poisoning by anticoagulant rodenticides in non-target animals globally. J Vet Med Sci 81:298–313
- Stehle CM, Battles AC, Sparks MN, Johnson MA (2017) Prey availability affects territory size, but not territorial display behavior, in green anole lizards. Acta Oecol 84:41–47
- Watanabe KP, Saengtienchai A, Tanaka KD, Ikenaka Y, Ishizuka M (2010) Comparison of warfarin sensitivity between rat and bird species. Comp Biochem Physiol C: Toxicol Pharmacol 152:114–119
- Wedding CJ, Ji W, Brunton DH (2010) Implications of visitations by shore skinks Oligosoma Smithi to bait stations containing brodifacoum in a dune system in New Zealand. Pac Conserv Biol 16:86–91
- Weir SM, Yu S, Knox A, Talent LG, Monks JM, Salice CJ (2016) Acute toxicity and risk to lizards of rodenticides and herbicides commonly used in New Zealand. N. Z. J Ecol 40:342–350
- Yamamura Y, Takeda K, Kawai YK, Ikenaka Y, Kitayama C, Kondo S, Kezuka C, Taniguchi M, Ishizuka M, Nakayama SM (2021) Sensitivity of turtles to anticoagulant rodenticides: risk assessment for green sea turtles (Chelonia mydas) in the Ogasawara Islands and comparison of warfarin sensitivity among turtle species. Aquat Toxicol 233:105792

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.