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A New Composite of O-aminobenzene Sulfonic Acid Self-Doped Polyaniline and Multi-Walled Carbon Nanotubes as a Fiber Coating for Solid-Phase Microextraction Gas Chromatography

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Abstract

A new composite of o-aminobenzene sulfonic acid self-doped polyaniline (SPAN) and multi-walled carbon nanotubes was prepared on a stainless steel wire by electrochemical method as a headspace solid-phase microextraction fiber for gas chromatography. Preparation conditions were investigated, and the resulting composite coating was characterized via scanning electron microscopy, Brunauer–Emmett–Teller (BET) analysis and thermogravimetric analysis. Introduction of MWCNTs and o-aminobenzene sulfonic acid into the composite leads to formation of plenty of microglobules and mesopores and makes BET surface area and thermal decomposition temperature of the composite increase by ~ 340% and ~ 130 °C, respectively, compared to common polyaniline coating. Extraction conditions were optimized, and chromatographic analysis of 2,4-dichlorophenol was conducted. Extraction efficiency of the new composite coated fiber increases by 320% compared to polyaniline-coated fiber. Relative standard deviations of peak areas are 4.3%, 6.3%, 7.1% (n=5) for identical fiber, fibers prepared in identical batch and different batches, respectively, exhibiting good repeatability and reproducibility. Detection limit is 1.30 ng L⁻¹ (S/N=3), and recoveries are in the ranges from 82.3 to 110.2%, from 91.5 to 109.4% and from 94.2 to 113.0% for three different water samples spiked at three levels of 500, 100 and 10 ng mL⁻¹, respectively.

Keywords Multi-walled carbon nanotubes · O-Aminobenzene sulfonic acid · Polyaniline · Solid-phase microextraction · 2,4-Dichlorophenol

Introduction

Gas chromatograph (GC) is widely used in the analysis of environmental pollutants [1]. However, due to the complex nature of sample matrices reportedly containing low level of analyte concentration, preconcentration is usually required prior to their quantitation via GC [2]. Among all the preconcentration methods, solid-phase microextraction (SPME) is one of the most popular techniques owning to its simplicity, environmentally friendliness, and high enrichment coefficient [3–6].

The reported SPME techniques include fiber SPME [5, 6], in-needle SPME [7], in-tip SPME [8], in-tube SPME [9, 10] and thin-film SPME [11, 12]. Among them, the fiber

☑ Jinxiang Li jxli@lnnu.edu.cn SPME is commonly used, and its extraction performance depends mainly on the fiber coatings [4, 13–15]. Nowadays, many commercial coated fibers have been applied to SPME, such as polydimethylsiloxane, divinylbenzene, polyethylene glycol and carbowax [3, 13]. However, the availability of the commercial coated fibers is limited and cannot fully meet the needs for various pollutants and their metabolites in environment. Also, some of these fibers have some shortcomings, such as insufficient thermal or/and solvent stability and high cost [5, 14]. Therefore, varieties of in-house made fiber coatings have been reported [15-18]. Conductive polymers have been extensively used as SPME fiber coatings [19]. Among them, polyaniline (PANI) and its derivatives are attracting since they have multifunctional properties such as electroactivity, hydrophobicity, polar functional groups, and $\pi - \pi$ interaction [15–18].

One of the drawbacks of PANI coating, however, is the poor thermal stability [20]. Using different substituted aniline monomers or dopants, the thermal stability of synthesized polymers can be, to some extent, improved under

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controlled electrochemical conditions [20–22]. Mehdinia et al. [22] prepared a aminobenzene sulfonic acid self-doped polyaniline (SPAN) fiber coating, and this coating was proved to be still stable at the high temperatures up to $350 \degree$ C.

Another drawback of PANI coating is its insufficient extraction efficiency [16, 23–25]. Several published reports suggest that carbonaceous materials [5, 15, 26], inorganic materials [24, 25], metal–organic framework [17, 27], and so on contribute to improving the porosity of the PANI-based coatings and thus enhance the extraction efficiency. Among the carbonaceous materials, multi-walled carbon nanotubes (MWCNTs) are the most attractive since they can be homogeneously distributed in aniline by forming donor–acceptor complexes and increase specific surface areas of the PANIbased coatings [28, 29]. Du et al. [29] prepared a MWCNTsdoped PANI coating. This coating exhibited a higher extraction efficiency than common PANI coating.

In the present work, in order to simultaneously overcome the two drawbacks of common PANI coatings, we prepared a new composite coating of o-aminobenzene sulfonic acid self-doped polyaniline and multi-walled carbon nanotubes (MWCNTs-SPAN) by electrochemical method for headspace solid-phase microextraction GC (HS-SPME-GC). Preparation conditions were investigated, and the resulting composite coating was characterized via scanning electron microscopy (SEM), Brunauer–Emmett–Teller (BET) analysis and thermogravimetric analysis (TGA). Extraction and desorption conditions were optimized, and the performance in HS-SPME-GC was examined using 2,4-dichlorophenol (2,4-DCP) as the test analyte. 2,4-DCP was selected because it is a typical polar pollutant and has been used for evaluation of SPME fibers by some other authors [32–36].

Experimental

Chemicals and Materials

Methanol, 2,4-DCP, aniline, sodium chloride and sulfuric acid were from Comio Chemical Reagents Co., Ltd. (Tianjin, China). O-aminobenzene sulfonic acid was provided by Aladdin Industries (Shanghai, China). MWCNTs (10–20 nm O.D., 5–10 nm I.D., 2–8 μ m length) were from Yuechuang Technology Co., Ltd. (Shenzhen, China). Stainless steel wire (0.2 mm diameter) was from local shops. Double-distilled water was prepared using a Yarong SZ-93 Water Distiller (Shanghai, China) and used throughout this work. All reagents were of analytical grade.

An aqueous standard solution of 2,4-DCP at 500 μ g L⁻¹ was prepared and used in the present work unless otherwise noted. Tap, mineral and sea water samples were collected from the city waterworks, Nongfu Spring Drinking Water

Company and the sea near the coast of Xinghai Square, Dalian, China, respectively.

Apparatus

Electrochemical experiments were carried out on a CHL660A instrument (Shanghai Chenhua Instrument Co., Ltd., China). GC experiments were performed on a GC-126 instrument with a built-in flame ionization detector (FID) (Shanghai Instrument Co., Ltd., China). Chromatographic data were obtained using a FJ3000 Chromatography Workstation (Shanghai Precision Instrument Co., Ltd., China). Separations were accomplished with a PEG-20-M capillary column (30 m×0.32 mm×0.5 μ m, Shanghai Kexiao Instrument Co., Ltd., China). SEM images were taken by a SU8010 SEM instrument (Hitachi Ltd., Japan). BET analysis and TGA were conducted on a JW-BK122W analyzer (Beijing JWGB Sci. & Tech. Co. Ltd., China) and an Setaram SETSYS thermogravimetric analyzer (TGA Setaram, France).

Preparation of MWCNTs-SPAN Coating

A section of 7-cm stainless steel wire was ultrasonically cleaned in acetone and water sequentially for 15 min. MWC-NTs and self-doped polyaniline were simultaneously electrochemically deposited onto one end of the stainless steel wire for the length of 3 cm in a 20 mL solution containing MWCNTs (0.0012 g), aniline (0.1 M), o-aminobenzene sulfonic acid (0.1 M) and H_2SO_4 (0.5 M). Electrodeposition was achieved by potential cycling between – 0.3 and 1.0 V at a scan rate of 0.025 V s⁻¹ for 80 cycles. After being flushed with water, the coated wire was inserted into a homemade holder and used as a SPME fiber assembly (see SI 1). Prior to use, the coating was aged for 1 h in GC injection port at 280 °C until the baseline was stable.

For evaluation of the new MWCNTs-SPAN composite coating by comparing with other coatings reported previously, three PANI-based coatings, including PANI, MWCNTs-PANI and SPAN coatings, were prepared referring to Bagheri's [18], Du's [29] and Mehdinia's [22] work, respectively.

Headspace Solid-Phase Microextraction

A 10-mL volume of aqueous 2,4-DCP solution was transferred into a 20 mL extraction bottle with a rubber cap. After adding appropriate amount of sodium chloride into the solution, the bottle was capped tightly. The SPME fiber was inserted into the bottle through the holder for HS-SPME. After extraction, the fiber carrying the analyte was withdrawn from the extraction bottle and inserted into GC injection port for performing thermal desorption process.

GC Calibration

Considering that the sensitivity might be different on different days, and the used GC was calibrated every day. The calibration method was as follows. A solution of 2,4-DCP $(1 \text{ g } \text{L}^{-1})$ was prepared and used as the reference solution. Three repeated injections (1 µL/injection) of the reference solution were performed on the first day, and the average of the peak areas was set as the permanent reference peak area. On the following each working day, the above injections were reperformed, the obtained average of peak areas was divided by the reference peak area, and the result was used as the GC calibration factor for the working day. All of the other peak areas obtained on the identical working day multiplied the calibration factor, and the results were used as the peak area values presented in the text unless otherwise noted. The calibration factor was measured at least once every day.

Results and Discussion

Preparation of the MWCNTs-SPAN Coating

As described in the earlier section, PANI is one of the most commonly used fiber coatings. Its application, however, is limited by poor thermal stability and insufficient extraction efficiency. Du et al. [29] and Mehdinia et al. [22] increased extraction efficiency of the MWCNTs-PANI coating and improved thermal stability of SPAN coating through introducing MWCNTs and - SO₃⁻ groups, respectively. In this study, in order to simultaneously resolve the above two problems of the PANI coating, a new PANI-based composite coating containing both MWCNTs and - SO₃⁻ groups was designed referring to Du's and Mehdinia's work, and its preparation conditions were investigated.

The molar ratio of aniline to o-aminobenzene sulfonic acid has an effect on the thermal stability. Mehdinia et al. [22] adopted a ratio of 1:1 for the preparation of SPAN coating and obtained a good improvement of the thermal stability. Therefore, the ratio of 1:1 was also used for the preparation of the MWCNTs-SPAN coating in the present study. The effect of amount of MWCNTs on extraction efficiency was investigated in the range of 0.0002-0.0022 g in 20 mL polymerization solutions containing o-aminobenzene sulfonic acid and aniline (each 0.1 M). The composite coating fibers prepared at different amounts of MWCNTs were used for the HS-SPME-GC of 2,4-DCP in the standard solution under same experimental conditions. The curve of peak area to MWCNTs amount was obtained (see SI 2). Peak area increases rapidly with the increase of MWCNTs amount until 0.0012 g and then remains almost unchanged. The amount of 0.0012 g was used in the present work.

Cyclic voltammetry (CV) is commonly chosen for the preparation of fiber coatings since the parameters, including potential scanning range, rate and cycles, can be precisely controlled, and thus, coatings of different thickness and morphology can be obtained in a reproducible manner [18, 30]. For the preparation of PANI coatings, the suitable potential scanning range is between - 0.3 and 0.9 V [25, 29, 31]. For the preparation of SPAN coatings, the upper potential limit should be increased to 1.0 V since o-aminobenzene sulfonic acid is hard to be electropolymerized due to its higher oxidation potential [22]. Therefore, in the present study, the scanning range was set between -0.3 and 1.0 V. The scanning cycles have a significant effect on the thickness and morphology of coatings [29]. The effect was investigated from 10 to 200 cycles. The maximal peak area was obtained at the cycles of 80 (see SI 3). The cycles of 80 were used in the following experiments.

Surface Morphology and Pore Structure

The morphology of the MWCNTs-SPAN composite coating was observed using SEM (Fig. 1A). For comparison, SEM images were also taken for the PANI (Fig. 1B), MWCNTs-PANI (Fig. 1C) and SPAN (Fig. 1D) coatings. By comparison of Fig. 1A and C with Fig. 1B and D, it can be seen that when MWCNTs are present in the coatings (Fig. 1A and C), the composite firstly forms in the form of microglobules, these globules then aggregate together and form many clusters, and these clusters interconnect and finally form a porous monolithic structure. By comparison of Fig. 1A with Fig. C, it can be found that the sizes of the globules, clusters and pores of the MWCNTs-SPAN coating (Fig. 1A) are remarkably smaller and more uniform than those of the MWCNTs-PANI coating (Fig. 1C). BET analysis of the four coating powders was carried out. The BET surface areas of the PANI, SPAN, MWCNTs-PANI and MWCNTs-SPAN coatings are measured to be 23.3, 22.8, 82.5 and 102.3 m² g⁻¹, respectively. Compared to the PANI and SPAN coatings, the BET surface areas of the MWCNTs-PANI and MWCNTs-SPAN composites increase by ~ 340%. Du et al. [29] prepared a MWCNTs-doped PANI coating and achieved a higher extraction efficiency than common PANI coatings, but they did not measure the specific surface area of the coating. The BET cumulative absorption pore volumes are 0.11, 0.9, 0.58 and 0.28 cm³ g⁻¹ for the PANI, SPAN, MWCNTs-PANI and MWCNTs-SPAN coatings, respectively. The pore diameter distribution curves are shown in Fig. 2. For the MWCNTs-SPAN composite, overwhelmingly majority of the pore diameters are in the range from 2 to 80 nm with the maximal dV/dlogD value being at ~ 20 nm. For the MWCNTs-PANI composite, the most of the pore diameters are above 100 nm, and no maximal dV/dlogD value can be obtained from the curve. The Fig. 1 SEM images for comparison of surface morphology among the PANI, SPAN, MWCNTS-PANI and MWC-NTS-SPAN coatings. Magnified rate:10,000





Fig. 2 BJH pore diameter distribution curves for comparison of pore structure among the PANI, SPAN, MWCNTs-PANI and MWCNTs-SPAN coatings. Sample activation temperature and time: 120° C and 8 h, respectively; *P/P*₀: 0–0.99

maximal value is estimated to be in the diameter range of macropores, or maybe even in the diameter range of megapores according to the observation from the SEM images of Fig. 1. For the PANI and SPAN coatings, it is difficult to measure their pore diameters because of their insignificant and almost unchanged dV/dlogD values in the whole test diameter range. The results of SEM and BET analysis unitedly demonstrate that MWCNTs can greatly modify the surface morphology and pore structure and thus increase the surface areas of the MWCNTs-SPAN and MWCNTs-PANI composites.

Selection of HS-SPME Conditions

Many authors had performed the HS-SPME of 2,4-DCP in aqueous samples using different fiber coatings [32–36]. For phenolic pollutants in aqueous samples, the presence of NaCl and low pH values can reduce their solubilities, increase the distribution ratios (gas phase/liquid phase), and thus improve extraction efficiency. Referring to their work, the NaCl concentration and the pH value were set at 0.35 g mL⁻¹ and 2, respectively. Extraction temperature and time were investigated in the ranges of 50–90 °C and 40–60 min, respectively. The maximal peak area was obtained at 80 °C and 50 min (see SI 4). Therefore, the temperature of 80 °C and the time of 50 min were selected.

Selection of Desorption Conditions

The desorption temperature and time in GC injection port were investigated from 240 to 300 °C and from 20 to 300 s, respectively. The curves of peak area to desorption time show that when GC injection port was set at temperatures higher than 280 °C, 2,4-DCP was desorbed completely from the fiber coating after 3 min (see SI 5). Considering that higher temperatures and longer times are more harmful to coating lifetime, the desorption temperature of 280 °C and the desorption time of 3 min were adopted. No memory effect was observed in blank injections under the selected desorption conditions.

Reproducibility of the MWCNTs-SPAN Coating

The reproducibility of the MWCNTs-SPAN coating was assessed through HS-SPME-GC of 2,4-DCP in the standard solution using five coated fibers prepared in identical batch and five coated fibers from five different batches, respectively. The relative standard deviations (RSDs, %) of the peak areas are 6.3% (n=5) and 7.1% (n=5), respectively, and are comparable or superior to those of other fiber coatings reported previously (2.1%–13%) [29, 32–36] (see SI 6).

Thermal Stability and Lifetime of the MWCNTs-SPAN Coating

Thermal stabilities of the PANI, MWCNTs-PANI, SPAN and MWCNTs-SPAN coatings were tested by TGA (Fig. 3). For the four coatings, the weight loss in the range of 20–100 °C can be attributed to evaporation of the water moisture in the coatings [23, 29]. For the SPAN and MWCNTs-SPAN coatings, in the range of 100–350 °C, the weights remain almost unchanged, indicating that the SPAN and MWCNTs-SPAN coatings are stable up to ~ 350 °C. By contrast, the PANI and MWCNTs-PANI coatings start to thermally decompose at ~ 220 °C as reported by other authors [23, 37]. With the introduction of $-SO_3^-$ groups, the thermal decomposition temperatures of SPAN and MWCNTs-SPAN coating increase by ~ 130 °C. This is consistent with that reported by Mehdinia et al. [22].

The thermal stability was further examined through comparing chromatograms (Fig. 4). The chromatograms were obtained by leaving the coating fibers in GC injection port at 320 °C and simultaneously monitoring signal changes. No effluent was found for the SPAN and MWCNTs-SPAN-coated fibers within 12 min, whereas some effluents were detected after 3 min for the PANI and MWCNTs-PANI-coated fibers owing to decomposition of the coating polymers.

In order to evaluate the lifetime of the MWCNTs-SPAN coating, the dependance of relative peak area on the cumulative time for which the fiber was left in GC injection port was investigated at 280 °C, and the plot is given in Fig. 5, together with those obtained using the MWCNTs-PANI, SPAN and PANI coatings for comparison. For the SPAN and MWCNTs-SPAN coatings, the peak areas remain almost unchanged within the time of ~900 min, during which 30 absorption-desorption cycles have been carried out. In contrast, for the PANI and MWCNTs-PANI coatings, the areas gradually decline by ~ 20% within 540 min, during which just 18 absorption-desorption cycles have been performed. This result implies that the SPAN and MWCNTs-SPAN-coated fibers have more longer lifetimes than the PANI and MWCNTs-PANI-coated fibers. Undoubtedly, the longer lifetimes of the SPAN and MWC-NTs-SPAN coating fibers should be ascribed to their better thermal stabilities as demonstrated above.





Fig. 3 TGA curves for comparison of thermal decomposition temperature among the PANI, SPAN, MWCNTs-PANI and MWCNTs-SPAN coatings. Conducted in nitrogen gas atmosphere; Heat rate: $10 \,^{\circ}\mathrm{C} \,\mathrm{min}^{-1}$

Fig. 4 Chromatograms for comparison of thermal stability among the PANI, SPAN, MWCNTs-PANI and MWCNTs-SPAN coatings. Injection port temperature: 320 °C; column oven temperature: 160 °C; detector temperature: 280 °C; carrier gas: N₂; carrier gas pressure: 0.22 MPa



Fig. 5 Curves of relative peak area to cumulative time for comparison of lifetime among the PANI, SPAN, MWCNTs-PANI and MWCNTs-SPAN coatings. 2,4-DCP concentration: 500 μ g L⁻¹; extraction temperature, 80 °C; extraction time, 50 min; salting strength: 0.35 g mL⁻¹ NaCl; pH value: 2; desorption temperature/injection port temperature: 280 °C; desorption time, 3 min; other conditions, as in Fig. 4

Extraction Efficiency

The extraction efficiency of the MWCNTs-SPAN coating was compared with those of the PANI, SPAN and MWCNTs-PANI coatings. In the present work, the extraction efficiencies were measured in peak areas referring to other authors' work [18, 21, 29, 31, 33]. The peak areas were obtained through HS-SPME-GC of 2,4-DCP in the standard solution using the four coating fibers at 280 °C. The extraction efficiencies of the MWCNTs-SPAN and MWCNTs-PANI coatings are about fourfold higher than those of the PANI and SPAN coatings (Fig. 6). Undoubtedly, this is attributed to the larger specific surface areas of the coatings as demonstrated in the earlier section. Du et al. [29] had also reported that the MWCNTs-doped PANI coating provided a higher extraction efficiency than the PANI coating in their work. It also should be noted that the extraction efficiencies of the MWCNTs-SPAN and SPAN coatings are a little higher than those of the MWCNTs-PANI and PANI, respectively. This probably results from the $-SO_3^-$ groups in the MWCNTs-SPAN and SPAN coatings. The presence of $-SO_3^-$ groups, to some extent, modifies the surface morphology and pore structure and enlarges the specific surface areas of the coatings, just as demonstrated in the earlier section. Additionally, the intermolecular interaction between the highly polar – SO_3^- groups and the polar 2,4-DCP molecules may



Fig. 6 Bar chart for comparison of extraction efficiency among the PANI, SPAN, MWCNTs-PANI and MWCNTs-SPAN coatings. Experimental conditions, as in Fig. 5

have some contribution to the extraction efficiencies of the MWCNTs-SPAN and SPAN coatings.

Performance of the MWCNTs-SPAN Fiber Coating in HS-SPME-GC

The curve of peak area to concentration was plotted for 2,4-DCP in purified water in the concentration span from 10 ng L⁻¹ to 600 μ g L⁻¹ (SI 7). Up to ~ 500 μ g L⁻¹, the peak area increases with the increase in concentration and then remains almost unchanged. Although no good linearity exists in the whole test concentration span, two calibration curves were separately made in the ranges from 50 ng L⁻¹ to 10 μ g L⁻¹ and from 10 to 500 μ g L⁻¹. The correlation coefficients (R²) are 0.9990 and 0.9487, and the slopes are 118.1 and 4.6 $\text{cm}^2 \text{ L ng}^{-1}$ for the two calibration curves, respectively (SI 8). There are a much better linearity and a higher sensitivity/slope in the lower concentration range. This is more beneficial for determination of the analytes at low concentration levels. The limit of detection (LOD, N/S = 3) and the precision (RSD, %) are 1.3 ng L⁻¹ and 4.3% (n=5), respectively. The results are summarized in Table 1, together with some data reported by other authors for comparison. Overall, the MWCNTs-SPAN coating is comparable or superior to the others in the measured indexes. In particular, it should be mentioned that LOD is dependent on not only extraction efficiency but also other factors in HS-SPME-GC. The relatively low LOD achieved by Li et al. [32], for an example, perhaps results mainly from the mass spectrometric detector used in their work. Mass spectrometers are highly sensitive

Table 1Data for comparisonof the performance inHS-SPME-GC among theMWCNTs-SPAN coatingand several others reportedpreviously using 2,4-DCP as thetypical analyte

Coatings	Linear range $(ng L^{-1})$	LOD^a (ng L ⁻¹)	Correlation coefficient	RSD (%)	Refer- ences
MWCNTs-SPAN	50-10,000	1.30	0.9990	4.3 (<i>n</i> =5)	This study
	10,000-500,000		0.9487		
N6	7-1000	2	0.999	7.4(n=3)	[24]
MWCNTs-PANI	25-35,000	1.05	0.996	3.9(n=6)	[29]
Co-MONTs	1-1000	0.18	0.9994	6.32(n=5)	[32]
GO/POE	5-1000	0.31	0.9997	7.8(n=6)	[33]
PAMAM	4-1000	1.0	0.9988	6(n=3)	[34]
PA-PEG	8-800	2.0	0.9937	2.1 (n=3)	[35]
poly (MAA-EGDMA)	200-50,000	10	0.9964	5.2(n=5)	[36]

^aLODs were calculated based on S/N = 3

 Table 2
 Results of determination of 2,4-DCP in environmental water samples by HS-SPME-GC for examination of applicability of the MWCNTs-SPAN coating

Water sample	2,4-DCP	Add (ng mL ⁻¹)	Found (ng mL^{-1})	Recovery $(n = 3\%)$
Mineral water	nd ^a	500	442.5 ± 7.5^{b}	88.5±1.5
		100	91.5 ± 3.2	91.5 ± 3.2
		10	11.3 ± 0.5	113.0 ± 4.8
Tap water	nd	500	411.6 ± 21.5	82.3 ± 4.3
		100	92.8 ± 6.4	92.8 ± 6.4
		10	9.4 ± 0.8	94.2 ± 7.9
Sea water	nd	500	551.0 ± 28.0	110.2 ± 5.6
		100	109.4 ± 7.1	109.4 ± 7.1
		10	11.2 ± 1.0	112.1±9.8

^and not detected

^bMean value ± standard deviation

to highly electronegative compounds such as 2,4-DCP, whereas the situation is just the opposite for FIDs. Therefore, if the MWCNTs-SPAN coating is combined with a mass spectrometer, a much lower LOD should be expected.

The reliability of the MWCNT-SPAN-coated fiber in practical use was assessed through determination of 2,4-DCP in three real water samples (mineral water, tap water and sea water) by HS-SPME-GC (Table 2). No the target analyte was found. Therefore, spiked samples were used as no natural matrix standard sample was available. Several typical chromatograms are given as supporting information (SI 9). The recoveries range from 82.3 to 110.2%, from 91.5 to 109.4% and from 94.2 to 113.0% with the relative standard deviations (RSD, %) between 1.5% and 9.8% (n=3) for the samples spiked at three levels of 500, 100 and 10 ng mL⁻¹, respectively. The accuracy and precision are acceptable for the analysis of environmental water samples.

Conclusions

A new composite of MWCNTs-SPAN has been presented for HS-SPME-GC of polar pollutants. SEM and BET analysis reveals that MWCNTs can greatly modify the surface morphology and pore structure and thus remarkably increase the surface areas of the MWCNTs-SPAN and MWCNTs-PANI coatings compared to the SPAN and PANI coatings. The result of TGA demonstrates that $-SO_3^-$ groups can notably enhance the thermal stabilities of SPAN and MWCNTs-SPAN coatings compared to PANI and MWCNTs-PANI coatings. Owing to the larger specific area and higher thermal stability, MWCNTs-SPAN coating exhibits a higher extraction efficiency and a longer lifetime than PANI, SPAN and MWCNTs-PANI coatings. The performance of the MWCNTs-SPAN coating in HS-SPME-GC is assessed to be comparable or superior to those of some fiber coatings reported previously. All the merits unitedly suggest that the MWCNTs-SPAN coating has potential applications in HS-SPME-GC of a variety of polar compounds such as phenol, chlorophenols and aromatic amines in water samples.

Declarations

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