**RESEARCH ARTICLE**



# **Precision of T1‑relaxation time measurements in the hepatic portal vein: infuence of measurement technique and sequence parameters**

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#### **Abstract**

**Objective** To investigate the effects of a range of parameter settings on  $T_1$  measurement stability in the portal vein using the T1-mapping sequences Look-Locker (LL) and Modifed Look-Locker inversion recovery (MOLLI).

**Materials and methods** Ten diferent versions of LL and MOLLI sequences were tested and compared to a reference sequence provided by the MR manufacturer. Ten healthy volunteers were imaged multiple times on two separate scan days at 3T. The mean  $T_1$  values and coefficient of variation (CoV) were calculated for each of the ten sequences and compared to the reference sequence.

**Results** Six of the tested sequences had  $T_1$  values close to the reference sequence; among those, three sequences achieved lower CoV than the reference sequence. Lowest CoV was achieved using a non-triggered LL sequence with 5 beat readout and a 45<sup>o</sup> flip angle (mean  $T_1$  1733 ms  $\pm 89$  ms, CoV 1.3%  $\pm 0.58$ %).

**Conclusion**  $T_1$ -measurements in the hepatic portal vein can be performed with high precision using either MOLLI or LL sequences provided that LL sampling duration is sufficiently long and flip angle sufficiently high. The advantage of constant timing outweighed the advantage of ECG-triggering.

**Keywords** Relaxometry  $\cdot$  T<sub>1</sub> measurements  $\cdot$  Blood

# **Introduction**

The hepatic portal vein contributes for two-thirds of the total hepatic blood fow and receives blood from the stomach, spleen, pancreas, small intestine, and the colon [[1](#page-9-0)].

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Per orally administered substances are transported from the gastrointestinal tract through the portal vein to the liver [\[1](#page-9-0)]. The portal vein, therefore, offers a location for detection and evaluation of uptake of substances from the gastrointestinal tract.

The proton  $T_1$  relaxation rate  $R_1$  (= 1/ $T_1$ ), of a solution is linearly dependent on concentration of dissolved **Electronic supplementary material** The online version of this<br>
article (bttps://doi.org/10.1007/s10334-018-00731-1) contains<br>
paramagnetic ions [[2](#page-9-1)]. Hence, repeated measurements of

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 $T_1$  in the portal vein before and after ingestion may offer a non-invasive quantitative method for studying uptake of per orally administered substances. The use of  $T_1$  measurements for quantifcation of substance uptake is challenging. Apart from physiologically induced variation in T1, the estimated  $T_1$  ( $\hat{T_1}$ ) may also depend on image noise, heart frequency, sequence-related parameters, and

type of  $T_1$  mapping technique used [[3–](#page-9-2)[7](#page-9-3)]. Accurate  $(\hat{T}_1)$ can be obtained using time-consuming saturation recovery methods; however, the length of these examinations makes them unsuited for in vivo applications, as the long scan times result in unacceptable length of breath hold. Therefore, faster alternative T1 mapping sequences have been

<span id="page-2-0"></span>**Fig. 1 a** Simplifed schematic sequence diagram of a Look-Locker ◂(LL) sequence. A 180° inversion pulse used to invert the magnetization from Mz<sup>+</sup> to Mz<sup>-</sup>. Following a waiting time of Ti<sub>min</sub> a readout commences with an excitation pulse "α", subsequent readout points are performed at  $Ti<sub>inc</sub>$  intervals. In the above case, a total of seven images are acquired with inversion times of  $Ti_{min} + (n-1 \times Ti_{inc})$ , where *n* is the image number. Stationary tissue experiences a saturation effect as evident by the solid relaxation curve, while ROIs assessing infowing blood do not experience this efect, visualised by the dotted line. To change the readout duration the repetition time, in this study given in number of beats, are either increased or decreased. If  $Ti<sub>inc</sub>$  is kept constant, an increase in readout duration also results in an increase in number of sampling points. Increasing the flip angle " $\alpha$ " would increase the saturation efect of stationary tissue, but not that of fast fowing blood. **b** Simplifed sequence diagram of the Modifed Look-Locker inversion recovery (MOLLI) sequence. Following a non-selective 180° inversion pulse, the first single shot balanced-SSFP readout is performed at a predefned trigger delay. The resulting image has an inversion time of  $Ti_{\text{min}}$ . For each subsequent cardiac cycle, one image is acquired with the same trigger delay.  $Ti<sub>inc</sub>$  is then defned by the lengths of the following heart cycles. Following the readout of the frst inversion pulse (frst cycle), a recovery period is allowed, in this example, 2 beats before a second cycle is performed and so on. For each new cycle, position of the inversion pulse is shifted relative to the trigger pulse spreading the sampling points on the recovery curve. In this example, fve readouts are performed in the frst cycle, followed by a recovery period of 2 beats and then three readouts in the second cycle. This results in the MOLLI scheme of 5(2)3 which has a total scan duration of 10 heartbeats and produces eight images with varying Ti. Increasing the number of beats in the longest cycle effectively increases the readout duration. The density of Ti can be increased, by adding additional cycles and the time allotted for free recovery can be increased by increasing the duration of the recovery period

introduced, where Look-Locker (LL) [[8](#page-9-4)] and Modifed Look-Locker inversion recovery (MOLLI) sequences [[9\]](#page-9-5) are currently the most established alternatives for in vivo use.

MOLLI was originally developed for cardiac applications, where cardiac triggering and breath holding are required to avoid severe motion artefacts. The need for cardiac triggering and breath hold put stringent restrictions on sequence parameters and optimal parameters for cardiac  $T_1$  mapping which are unlikely to be optimal for non-cardiac applications. Some work has been reported in the literature on optimizing  $T_1$  measurements in vessels, like jugular vein [\[10](#page-9-6)], carotid artery  $[11]$  $[11]$ , and sagittal sinus  $[6]$  $[6]$  $[6]$ . In these studies, the effects of sequence parameters such as readout duration, heart rate  $[5, 9, 12]$  $[5, 9, 12]$  $[5, 9, 12]$  $[5, 9, 12]$  $[5, 9, 12]$  $[5, 9, 12]$ , and flip angle  $[10, 11]$  $[10, 11]$  $[10, 11]$  have been evaluated.

To our knowledge, there is no literature on  $T_1$  measurements in the hepatic portal vein. Such measurements are subject to conditions that differ from cardiac  $T_1$  mapping in several aspects. In particular, cardiac triggering may not be critical in this region, which may allow for further improvement on measurement precision, in this study defned as reduced coefficient of variation  $(CoV)$ , by adjusting relevant acquisition parameters.

The aim of this study was to investigate the effect of acquisition techniques, and relevant sequence parameters on the accuracy and precision of native  $T_1$  measurements in the portal vein. We specifcally address parameters related to motion and fow, since limitations on these may be less stringent compared to conventional cardiac T1-assessments. We also address parameters related to sampling duration, as precise measurements of the long T1 of native blood may require longer read-out times than for myocardial tissue.

#### **Theory**

In both LL (Fig. [1](#page-2-0)a) and MOLLI (Fig. [1](#page-2-0)b), one or more inversion pulses are followed by a train of read-out segments, sampling the magnetization recovery curves. In LL, a series of low fip angle echo-planar-imaging (EPI) acquisitions is commonly used for readout following an initial IR pulse. Typically, high EPI acceleration and low fip angle are used to avoid saturation of the magnetization recovery. Studies have shown, however, that in fowing blood, the use of higher fip angles can be benefcial due to continuous inflow of unsaturated blood [[10](#page-9-6), [11](#page-9-7)].

MOLLI was initially developed to allow for  $T_1$  mapping of the heart. The readout is based on the fow-insensitive single-shot-balanced SSFP technique [[13](#page-9-11)] and is fxed in a predefned cardiac phase to allow for pixelwise relaxation assessment. The time between subsequent measurements in MOLLI is then given by the duration of the cardiac cycle. Density of measurements may be increased by adding MOLLI cycles, i.e., repeating the inversion recovery experiment while shifting the timing of the inversion pulse with respect to cardiac phase. The MOLLI scheme 5(3)3 then describes a sequence with fve images (one per heart beat) in the frst cycle, followed by a pause of 3 heartbeats for magnetization recovery and then a second inversion pulse followed by three images; giving a total of eight inversion recovery sample points, and a breath-hold duration of 11 heartbeats.

Common for both LL and MOLLI sequences is that the magnetization recovery trajectory is infuenced by the readout process and this needs to be corrected for to obtain correct  $\hat{T}_1$  [\[8](#page-9-4), [9,](#page-9-5) [14](#page-9-12)]. In addition, the use of cardiac triggering in both LL and MOLLI can result in a diference in readout duration and timing of the sampling points based on the patient's heart rate. For applications outside the heart, cardiac triggering may be less critical, thereby enabling a more fexible timing scheme. Based on the characteristics of the LL and MOLLI sequences, we identify readout duration, sequence timing, and fip angle to be the three main parameters (readily modifed by the user) most afecting the quality and accuracy of the resulting  $T_1$ -maps. These three parameters in the context of  $T_1$ -mapping sequences will, therefore, be briefy discussed.

#### **Readout duration**

In both LL and MOLLI, one or more inversion pulses are followed by a train of readout segments sampling the magnetization recovery curves. The readout duration is defned by the number of triggering beats in LL and the longest cycle in the MOLLI scheme, as described in Fig. [1](#page-2-0)a, b. Readout duration should not be confused with; the time between inversions, which in LL is close to the readout duration, but higher in MOLLI due to the recovery phase, or the breathhold time, which in this study is equal to the total scan time of the sequence and dependent on the number of k-space segments in LL and the MOLLI scheme. Increasing the readout duration, and hence number of readout points, gives additional information on the trajectory of the relaxation curve following the inversion pulse, potentially allowing for a more accurate assessment of  $T_1$ , at the cost of longer breath holds. It is commonly accepted that the readout duration should ideally be in the order of five times  $T_1$  and that insufficient delay between inversion pulses could result in a  $\hat{T}_1$  bias [[15](#page-9-13)].

#### **Triggering**

The MOLLI Native 5(3)3 sequence is a breath-hold technique developed for  $T_1$  quantification of non-gadolinium enhanced cardiac tissue. It is based on a fow-insensitive single-shot true-FISP readout using electrocardiogram (ECG) triggering to lock readout to a predefned cardiac phase [\[9\]](#page-9-5).

Since the time between measurements in MOLLI, and readout duration for both MOLLI and LL is given by the cardiac frequency, a shift in the frequency will change both the timing of sampling points and read-out duration. Such variation has been reported to make  $\hat{T}_1$  heart rate dependent [[5,](#page-9-9) [9,](#page-9-5) [15](#page-9-13)]. Deactivating ECG-triggering may prevent this dependency in  $\hat{T}_1$  when imaging in areas without cardiac motion.

## **Flip angle**

To minimize saturation efects in LL, a series of low fip angle echo-planar imaging (EPI) acquisitions is commonly used. The low fip angle mitigates the saturation efect, but does result in a lower signal-to-noise ratio (SNR) [[10\]](#page-9-6). Studies have shown, however, that in veins and arteries, the use of higher fip angles to increase signal-to-noise ratio is pos-sible due to continuous inflow of unsaturated blood [\[6](#page-9-8), [10,](#page-9-6) [11](#page-9-7)].

# **Materials and methods**

Ten male volunteers average age 28.8 (range 24–33 years) were recruited with the following inclusion criteria: male, age 20–35 years, no known liver or blood disease, no contraindications against MRI, and no medications. A written informed consent was signed by the volunteers, which subsequently attended two MRI sessions with a minimum of 1 week between. No dietary restrictions were given.

Examinations were performed using a Philips 3T Ingenia MR system (Philips Medical systems, Best, The Netherlands) with a 16-channel dStream Torso coil, and an embedded 16-channel posterior coil. Following T2-weighted breath-hold localizers the hepatic portal vein was identifed, and the image plane was angulated perpendicular to the portal vein. During MR acquisitions, the volunteers were asked to hold their breath during the expiratory phase.

Initially seven variants of LL (Fig. [1](#page-2-0)a) and MOLLI (Fig. [1b](#page-2-0)) were included in the study. In lack of a gold standard, the vendor recommended MOLLI Native [MOLLI scheme 5(3)3] sequence was used as a reference, as this sequence has a proven reproducibility [[16](#page-9-14)]. MOLLI variants were constructed by varying the length of the cycles in the MOLLI scheme while keeping scan duration short for maximum 30 s breath hold. LL variations included comparing readout durations of 3 and 5 beats and flip angles of  $7^\circ$ and 45°. Non-ECG-triggered variants of both MOLLI and LL sequences were implemented by simulating the ECG signal at a rate of 60 beats per min. Table [1](#page-4-0) provides an overview of the sequences evaluated. After the ffth volunteer preliminary data were analysed. Four of the sequences were excluded from the study at this time due to resulting  $\hat{T}_1$ values being signifcantly lower than those obtained with the reference sequence as well as values previously reported in the literature. Three additional sequences based on further optimization of the remaining sequences were then added (Table [1\)](#page-4-0).

Phase-sensitive flow measurements (q-flow) were acquired at the start and end of all but fve examinations giving a total of 35 fow measurements. Scan geometry was identical to that of the T1-mapping sequences.

#### **Data analysis**

All  $T_1$  relaxation time calculations were performed in nordicICE (NordicNeuroLab, Bergen, Norway). Elliptical ROIs were placed in the portal vein in the raw images. The average signal from three ROI placements was used as input to the curve fit function in nordicICE to obtain  $\widehat{T}_{1s}$  for each scan. Figure [2](#page-4-1) shows an example raw image with an ROI placed in the portal vein. When performing ROI placement as much as possible of the vein was included in the ROI, while visual artefacts and vessel wall were avoided.

Flow measurements were analysed using SEGMENT version 2.1 R 6078 (Medviso, Lund Sweden).



<span id="page-4-0"></span>

Fixed triggering indicates that the sequence was acquired with a simulated ECG of 60 bpm

a Sequence removed after the ffth volunteer

<sup>b</sup>Sequence added after the fifth volunteer



**Fig. 2** Representative image of an example ROI placed in the hepatic portal vein. ROIs were placed in the vessel to include as much as possible of the vessel lumen while avoiding vessel wall, and visual artefacts

#### <span id="page-4-1"></span>**Statistical analysis**

For each sequence s and examination e, an average of estimated  $T_1$  was calculated by the expression:

$$
\overline{T_{1\mathrm{e},\mathrm{s}}} = \frac{1}{n} \sum_{i=0}^{n} \widehat{T_{1\mathrm{e},\mathrm{s},i}},
$$

where i denotes the ith repetition of a given sequence, and *n* is the number of repetitions of a given sequence in an examination. Over all mean  $T_{1s}$  was then calculated as the average of  $\overline{T_{\text{les}}}$  over all examinations. For all but one examination each sequence was repeated six times, in the fnal examination, only fve repetitions were acquired.

To compare the longitudinal stability in the  $T_1$  measurements across all volunteers the mean, standard deviation (SD), and standard error of mean (SEM) for all sequences in all sessions were calculated. CoV was calculated for each sequence at each examination by

$$
CoV_{\rm e,s} = \frac{SD_{\rm e,s}}{T_{\rm 1e,s}},
$$

where  $s =$  sequence,  $e =$  examination.

The mean  $CoV_s(\overline{CoV_s})$  and SEM across all volunteers and examinations were then calculated for all sequences.

In cases where the veins could not be positively recognized, due to low SNR or artefacts in the image, the data point was defned as non-readable and removed from the data set.

Paired samples *t* test with Bonferroni correction was used to compare  $T_{1s}$  and  $CoV_s$  of the test sequences against the reference MOLLI Native sequence with the null hypothesis being that there are no signifcant diferences between the test sequences and the reference sequence. The adjusted level of signifcance from Bonferroni correction was 0.005 for mean *T*1 and 0.01 for CoV. In addition, paired samples *t* test were performed to evaluate the efect of each parameter on CoV.

Flow curves were averaged over all scans to give average minimum, maximum, and mean velocity.

## **Results**

Mean  $T_1$  values for all sequences analysed are shown in Fig. [3](#page-5-0) and are summarized in Table [2.](#page-6-0) Six of the tested sequence variants reported  $T_1$  values that were not signifcantly diferent from the reference sequence (1759 ms,  $SEM = 26$  ms). All variants of the LL with 3 beat readout sequences and one MOLLI sequence reported a signifcantly lower  $T_1$  than the reference sequence.

*CoV*s for all sequence variants are shown in Fig. [4](#page-6-1) and are sum-marized in Table [2.](#page-6-0) Among the sequences with  $T_1$  values close to the reference sequence, three sequence estimated  $\overline{CoV_s}$  which was lower than the reference sequence CoV of 2% (SEM=0.171%)

<span id="page-5-0"></span>**Fig.** 3 Plot of mean  $T_1$  values measured in volunteer's hepatic portal vein for the ten sequences tested. Mean of the reference sequence with SEM is shown as solid grey line and dotted, respectively. Five sequences had  $\hat{T}_1$  values similar to that of the reference sequence: LL 5 beat ECG-triggered 7° flip angle 1757 ms  $(\pm 26, p=0.95)$ , LL  $5$  beat ECG-triggered  $45^{\rm o}$  flip angle 1720 ms  $(\pm 21, p=0.14)$ , LL 5 beat non-triggered 45° flip angle 1734 ms  $(\pm 28, p=0.92)$ , MOLLI 10(5)5; 1756 ms  $(\pm 19,$ *p*=0.035), and MOLLI 10(5)5 non-triggered; 1771 ms  $(\pm 17,$  $p=0.018$ ). The other five sequences had lower  $T_1$  than the reference sequence. LL 3 beat ECG-triggered 7<sup>°</sup> flip angle 1357 ms  $(\pm 19) p < 0.001$ , LL 3 beat ECG-triggered  $45^{\circ}$  flip angle 1340 ms  $(\pm 15, p < 0.001)$ , LL 3 beat non-triggered 7° flip angle 1553 ms (±15, *p*<0.001), and LL 3 beat non-triggered 45 $^{\circ}$  flip angle 1525 ms ( $\pm$ 16, *p*<0.001). And MOLLI 10(1)1 ECG-triggered;  $1702 \text{ ms } (\pm 25,$ 

*p*=0.0006)

(Table [2\)](#page-6-0), but the diference was not statistically signifcant 29. The lowest CoV was achieved using a non-triggered LL with 5 beat readout and 45° flip angle, with a CoV of 1.29%.

Comparing the LL sequences with 3 beat readout to those with [5](#page-7-0) beat readout (Fig. 5a), we found a lower  $T_{1s}$  (1408 ms) vs. 1737 ms,  $p < 0.001$ ) and a non-significant increase in  $CoV<sub>s</sub>$  (3.24% vs. 2.83%,  $p=0.22$ ). Non-triggered sequences performed better in terms of precision than corresponding sequences with ECG-triggering with a signifcant increase in  $T_{1s}$  (1643 ms vs. 1580 ms, respectively,  $p < 0.001$ ) and a reduction in  $CoV_s$  (2.2% vs. 2.9%, respectively,  $p=0.0023$ ) (Fig. [5b](#page-7-0)). Finally, increasing the flip angle from  $7^{\circ}$  to  $45^{\circ}$ resulted in an increase in precision with a  $CoV<sub>s</sub>$  reduction from 4.35 to 2.5%. ( $p < 0.001$ ) (Fig. [5c](#page-7-0)) and a decrease in  $T_1$ from 1593 to 1574 ms,  $(p=0.01)$ .

A total of 13 scans were removed from the study due to artefacts caused by breath-hold challenges.

The flow results revealed a large range in flow measurements both at minimum 12 cm/s (range 8–21 cm/s) and at maximum 18 cm/s (range 13–26 cm/s) and at mean velocity 15.43 cm/s (range 10–24 cm/s). Paired samples *t* test among the examinations, where two flow measurements were acquired showed no signifcant diferences in Vmin ( $p = 0.33$ ), Vmax ( $p = 0.15$ ), or Vmean ( $p = 0.23$ ) between the two measurements.



<span id="page-6-0"></span>



Bonferroni corrected levels of significance  $0.005$  for  $T_1$  measurements and  $0.01$  for CoV calculations

CoV comparisons where only performed for sequences which had  $T_1$  values equal to reference

\*Indicates signifcant diferences from the reference sequence [MOLLI 5(3)3]

<span id="page-6-1"></span>**Fig. 4** Plot of the average coefficient of variation (CoV) in % of the sequences. Mean of the reference sequence are presented as solid grey line with dotted lines representing SD. Among the sequences that had  $T_1$  similar to the reference sequence,  $CoV<sub>s</sub>$  of three sequences were lower than the reference, but the diference was not statistically signifcant. These were: LL 5 beat nontriggered 45° flip angle; 1.29% (±0.183%, *p*=0.0597), MOLLI 10(5)5 ECG-triggered; 1.88% (±0.245%, *p*=0.73), MOLLI 10(5)5 non-triggered, and  $1.86\%$  ( $\pm 0.262\%$ ,  $p = 0.75$ )



## **Discussion**

The results of the present study suggest that  $T_1$  of hepatic portal vein blood can be determined with high precision  $(\overline{CoV_s}$  < 1.3%). MOLLI acquisitions with longer cycles, and an optimized LL sequence performed slightly better than the reference sequence. We further found that improvements in precision can be made by increasing fip angle in LL, and by not using ECG-triggering in both LL and MOLLI.

If  $T_1$ -mapping is to be used to quantify absorbed agents in portal vein, it is of importance that the  $T<sub>1</sub>$  values measured are correct and reproducible. The measured  $T_1$  in blood of our reference sequence falls well within the range reported by the previous studies of  $T_1$  in blood at 3T (range 1618–1878 ms, average 1746 ms) [\[6,](#page-9-8) [10,](#page-9-6) [11,](#page-9-7) [17–](#page-9-15)[20\]](#page-9-16).





<span id="page-7-0"></span>**Fig. 5** Comparison of the CoV by individual parameters. The difference in CoV caused by readout duration **a** timing method, **b** fip angle, **c** shows a signifcant decrease in CoV for timing method (CoV 2.9% vs. 2.2%, paired samples *t* test  $p = 0.0023$ ) and flip angle (CoV

The shorter  $T_1$  estimates obtained with the LL sequences with 3 beat readout make these sequences unsuited for evaluation of  $T_1$  in blood. From visual inspection of the relaxation curves (data not shown), it is probable that the shorter  $T_1$ measured with 3 beats is a result of insufficient sampling time in relation with the target  $T_1$  relaxation times. As stated by both McRobbie et al. and Taylor et al., the readout duration should be in the order of 5 times  $T_1$  [\[15](#page-9-13), [21\]](#page-9-17). It should also be recognized that the time between inversions in our LL sequences are close to equal to the readout duration and thus not allowing for a substantial time of free recovery, also in the MOLLI sequence the time allotted for free recovery is only equal to that of the recovery duration.

With the measured  $T_{1s}$  from our reference sequence of 1759 ms, this results in a suggested readout duration of 8795 ms. A change from 5(3)3 in the reference sequence to 10(5)5 results in a doubling of the readout duration and an increase in free recovery from 3 to 5 heart beats. Assuming non-triggered acquisition with a simulated heart rate of 60 bpm, the time between inversion pulses will be 8 s for

4.35% vs.  $2.5\% p < 0.001$ ). The decrease caused by increasing readout duration was, however, not signifcant (CoV 3.24% vs. 2.83%, paired samples *t* test  $p=0.22$ )

the benchmark sequence and 15 s for the  $10(5)$ 5 scheme. Especially, for long  $T_1$ , values and faster heart rates, there will be a bias in  $T_1$  due to an incomplete relaxation recovery between the inversion pulses [[15](#page-9-13)]. The sequences used in this study had scan durations ranging from 10.8 to 20.2 s and readout durations ranging from 3 to 10 s. It appears from the results that an increase in both scan and readout duration resulted in an increase of  $T_1$ . Increasing the readout duration, and thus the scan duration, allows for a higher number of sampling points in both LL and MOLLI sequences, which is likely to increase the precision of the T1 measurements. In a clinical setting, however, increasing scan time may provide increasing motion artefacts due to prolonged breath-hold requirements, which could ofset the beneft of increasing readout duration observed in healthy volunteers. One may also argue that the through plane fow situation ensure-free recovery also during imaging and hence the need for additional time after readout only afect scans with short read-out durations. This may allow further improvement in scan efficiency when measurements are restricted to fowing blood.

With the slice thickness of 5 mm in our LL sequences, the observed fow velocities indicate a necessary spacing of approximately 40 ms in average and 62 ms for the lowest flow measured to allow for sufficient inflow of fresh blood to allow for the use of high fip angles.

As can be seen from both Fig. [4](#page-6-1) and the statistical analysis, the LL with 5 beat readout and  $45^{\circ}$  flip angle trended towards a lower  $\overline{CoV_s}$  than the reference sequence. The lower  $\overline{CoV_s}$  of this sequence indicates a higher precision, making it more adept for longitudinal studies in the hepatic portal vein.

Some general assumptions on  $T_1$  measurements might be drawn from our data. Sufficient sampling duration is reported to be of great importance to avoid bias in  $\hat{T}_1$ . Nontriggered acquisitions result in both a fxed readout duration and a fxed spacing between readout timepoints. As shown by others [\[5](#page-9-9), [9](#page-9-5), [12](#page-9-10)],  $\hat{T}_1$  is to some degree dependent on heart rate. Changes in patient heart rate during acquisition may, therefore, result in a change in  $\hat{T}_1$  that could obscure, or mimic, real changes in  $T<sub>1</sub>$ . Patient compliance could potentially also be increased by not using ECG-triggering, since potential challenges with triggering signal are avoided. This does, however, not allow for a compensation of pulsatile flow and motion from the cardiac cycle. According to Gallix et al. [\[22](#page-9-18)], there are some uncertainties whether or not portal vein flow is continuous or pulsatile among healthy volunteers. As evident from our results, when performing  $T_1$  mapping in hepatic portal vein, the potential efect of pulsatile flow and motion is mitigated by the positive effects of fixed readout duration and timing. If LL is used, then increasing the flip angle appears to reduce  $CoV<sub>s</sub>$ . This is in agreement with other studies such as Qin et al. [\[10](#page-9-6)] that uses a flip angle of 90° in their study. A prerequisite for using the higher flip angle is necessarily that the spacing of readout segments is higher than the time required for through slice fow. The spacing between readout segments in our LL protocols was 120 ms for the LL 3 beat and 200 ms for the LL 5 beat which is higher than the required 62 ms from the volunteer with the lowest fow velocity justifying the use of high fip angles.

Although not addressed directly in this study, there are some limitations when using a high fip angle for LL imaging. While this gave a significant reduction in  $CoV<sub>s</sub>$  in our study, it must be stressed that the high fip angle will result in increased saturation of stationary tissue and this sequence might, therefore, not be applicable if  $T_1$  measurements of both stationary tissue and blood are of interest. In this study, ROI measurements and subsequent  $T_1$  estimations were based on the raw data images and not parametric maps. We have not performed a stringent analysis of the diferences in  $\hat{T}_1$  and CoV depending on the method of analysis; however, a parametric map-based analysis was also performed on these data and a scatterplot comparing the two methods can be seen in Fig. 1 in supplementary materials.

#### **Limitations**

Although the scan protocol was repeated in each volunteer several times in succession, and on two separate days, only ten volunteers were included, limiting the statistical power. Furthermore, the volunteers were young and healthy and may not be directly comparable to older patients, where, e.g., breath-hold capacity may be an issue. In addition, the volunteers were not asked to follow any dietary restrictions. This may result in a change of the true  $T_1$  during scanning; however, such an effect should be equal for all sequences and, therefore, not signifcantly alter the results of our variation analysis. A further limitation is the absence of a gold standard sequence for validation of the accuracy of the  $T_1$ measurements. We do, however, believe that the choice of using the vendor supplied MOLLI  $5(3)3$  as a benchmark sequence for comparison is a good compromise as this sequence is well established in clinical practice and has been thoroughly validated [\[16\]](#page-9-14).

# **Conclusion**

In conclusion, T1-measurements in the hepatic portal vein may be performed with comparable accuracy and precision using either the MOLLI sequences or LL sequences described in this study. Portal vein blood flow is sufficiently fast with little pulsation to allow the use of high fip angle readout without cardiac triggering. Increasing read-out duration beyond that of the standard MOLLI showed only a small effect on both  $\hat{T}_1$  and CoV. Among the sequences tested in this study, a non-triggered Look-Locker with 5 beat readout, 45° flip angle had the lowest CoV.

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#### **Compliance with ethical standards**

**Conflict of interest** The authors declare that they have no confict of interest.

**Ethical approval** All procedures performed in studies involving human participants were in accordance with the ethical standards of the institution and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

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