REVIEW ARTICLE

Production and growth of microalgae in urine and wastewater: A review

Shahida Anusha Siddiqui^{1,2} • Yuan Seng Wu^{3,4} • Trideep Saikia⁵ • İlknur Ucak⁶ • Maliha Afreen⁶ • **Mohd Asif Shah7,8,11,12 [·](http://orcid.org/0000-0002-0351-9559) Raphael D. Ayivi9,10**

Received: 23 February 2023 / Accepted: 20 June 2023 / Published online: 19 July 2023 © The Author(s), under exclusive licence to Springer Nature Switzerland AG 2023

Abstract

Issues of climate change, energy demand, and natural resources depletion are calling for circular methods to produce valueadded products such as biomass, biofuel, biofertilizers, and nutraceuticals from waste. For instance, culturing photoautotrophic microalgae in wastewater and urine appears promising to recycle nutrients from waste. Here we review microalgal production with focus on wastewater treatment and urine utilization. We present photobioreactors, parameters infuencing algal growth, economical aspects, and limitations from consumer acceptance.

Keywords Microalgae production · Wastewater · Urine treatment · Nutrient recovery · Environment · Photobioreactor

Introduction

Microalgae have been cultured to produce organic matter for an extensive range of applications, involving human and animal nutrition, as biofertilizers in the agriculture sector,

 \boxtimes Shahida Anusha Siddiqui S.Siddiqui@dil-ev.de

 \boxtimes Mohd Asif Shah drmohdasifshah@kdu.edu.et

> Yuan Seng Wu sengwu_21@yahoo.com

Trideep Saikia trideep.saikia3@gmail.com

İlknur Ucak ilknurucak@ohu.edu.tr

Maliha Afreen malihaafreen120@gmail.com

Raphael D. Ayivi rdayivi@aggies.ncat.edu

- ¹ Department of Biotechnology and Sustainability, Technical University of Munich, Essigberg 3, 94315 Straubing, Germany
- ² German Institute of Food Technologies (DIL E.V.), Prof.-Von-Klitzing-Straße 7, 49610 Quakenbrück, Germany
- ³ Centre for Virus and Vaccine Research, School of Medical and Life Sciences, Sunway University, 47500 Subang Jaya, Selangor, Malaysia

in cosmetics, and the health departments. Microalgae have great potential for the manufacturing of many high-value bioactive composites, including intracellular and extracellular, like lipids, amino acids, polysaccharides, phenols, antioxidants, vitamins, minerals, and pigments that can assist

- ⁴ Department of Biological Sciences, School of Medical and Life Sciences, Sunway University, 47500 Subang Jaya, Selangor, Malaysia
- ⁵ School of Pharmaceutical Sciences, Girijananda Chowdhury University, Guwahati-17, India
- ⁶ Faculty of Agricultural Sciences and Technologies, Nigde Omer Halisdemir University, Nigde, Turkey
- ⁷ Department of Economics, Kebri Dehar University, Kebri Dehar, Somali 250, Ethiopia
- ⁸ School of Business, Woxsen University, Kamkole, Sadasivpet, Hyderabad, Telangana 502345, India
- ⁹ Department of Food and Nutritional Sciences, North Carolina A&T State University, Greensboro, NC 27411, USA
- ¹⁰ Department of Nanoscience, Joint School of Nanoscience and Nanoengineering, University of North Carolina, Greensboro, NC 27412, USA
- ¹¹ Division of Research and Development, Lovely Professional University, Phagwara, Punjab 144001, India
- 12 School of Engineering and Technology, Sharda University, Greater Noida 20131, India

to compensate the cost of biofuel production in bio refnery setup (Mastropetros et al. [2022](#page-30-0); Koutra et al. [2022](#page-29-0)). These organisms can grow well in wastewater and can be used to remove metal ions from the wastewater by combining them with biomass (Zagklis et al. [2021\)](#page-33-0). In this context aquaculture, municipal and industrial wastewater are recommended as good nutrient sources for microalgal growth (Sakarika et al. [2022\)](#page-32-0).

Scientists are trying to find new sustainable energy sources and hence pushing boundaries to research urine. It was reported that human urine is a good source of agriculture nutrients which can produce nitrogen (N), phosphorus (P), and potassium (K) as natural fertilizers (Sharma et al. [2021](#page-32-1)). Urine has also been reported as a good fertilizer that improves crop production and subsequently minimizes waste production (Ranasinghe et al. [2016\)](#page-31-0). In addition, urine comprises less than 1% of the whole volume of domestic wastewater containing 80% of the N and 40–50% of the P, making it a promising source for the recovery of nutrients. Urine can be applied to agricultural areas in its original form for fertilization. But many legal issues are arising with this method, such as surplus to the ecosystem and vaporization of nitrogen contingent on climate situations. Additionally, high volumes of liquid are required for the storage and transportation of urine, where the continuous application becomes an overdose and adversely increases the salinity of the soil. A substitute method can be the assimilation of fertilizing nutrients into phototrophic organisms, microalgae, which are much more competent at captivating these composites.

Microalgae biomass developed on different types of wastewater can be used as a slow-release fertilizer, because of its primary confguration being alike to that of plants, though practical experiments are still limited. Instead, the economic viability of this method is high because rich-value composites like pigments could be taken out from algal biomass (Cai et al. [2013](#page-26-0)). However, these days research work is carried out for nutrient recovery through the growth of microalgae in urine (Chang et al. [2013;](#page-26-1) Jaatinen et al. [2016](#page-28-0)). Presently, domestic wastewater is considered as a reserve than waste like a nutrient reserve for the fertilization of plants, involving N and P. Large portion of the fertilizing nutrients in domestic wastewater are obtained from human urine, though it comprises only less than one percent of the total stream fow. These nutrients include 90% of K, 80% of N, and 50% of P as well as organic composites involving creatine, creatinine, and uric acid, as main components (Martin et al. [2022](#page-30-1)). Additionally, other than great concentrations of K, N, and P, many other trace elements are found in urine composition. Urine also comprises additional ions like Cu²⁺, Na⁺, Mg²⁺, and Cl[−] that are essential for plant growth (Viskari et al. [2018\)](#page-33-1).

Cultivation of diferent types of microalgae was observed in previous studies by using existent and artifcial human urine. These types are known as *Chlorella sorokiniana* (Tuantet et al. [2014a](#page-33-2), [2014b\)](#page-33-3), Cyanobacterium *Spirulina platensis* and *Scenedesmus acuminatus*. Some species of microalgae grow well in diluted human urine including *Chlorella sorokiniana*. It grew well in a continuous bioreactor with a dilution ratio of 1:2–1:3 of human urine to get a high biomass production of 9.3 g L⁻¹ d⁻¹ (Tuantet et al. [2014b](#page-33-3)). There is also another recent study that shows that sourceseparated urine can be utilized to improve the growth of microalgae photoautotrophic microorganisms that recovered nutrients by using carbon dioxide utilizing an inorganic carbon source and light by means of an energy source (Tao et al. [2022](#page-32-2); Tuantet [2015](#page-33-4)). Microalgae cultivation needs nitrogen and phosphorus (N, P) nutrients, which are mainly present in urine with other several minor elements including (e.g., Zn, Cu, Fe, Mo, Co, and Mn) that are essential for the growth of microalgae. It was also observed that 97% of ammonium nitrogen, 96.5% of the total phosphorus and 85–98% of urea in diluted urine can be removed by the cultivation of microalgae, and this is a good strategy for waste management by using potential existing sources of urine (Behera et al. [2020](#page-26-2)).

Therefore, human urine appears as a promising medium for the growth of microalgae. This review evaluates algae growing in urine from a diferent perspective which includes diferent types of wastewater, their constituent and their treatment methods for the cultivation of microalgae. It also focused on diferent types of photobioreactors which are used for the combined action of wastewater treatment and cultivation of microalgae which will be helpful for future research related to the utilization of urine in algae production.

Wastewater treatment

Types of wastewaters

Wastewater that originates from home activities known as municipal sewage wastewater includes water excretions from public toilets, schools, hospitals, restaurants and hotels, which usually comprises urine and feces (Ahmed et al. [2021](#page-25-0); Rose et al. [2015\)](#page-31-1). Conservative methods of municipal sewage wastewater treatment systems contain diferent stages (Gandiglio et al. [2017](#page-27-0)). Firstly, they take in the sedimentation of solid ingredients known as the primary treatment stage. Secondly, the removal of suspended and liquefed organic ingredients occurs known as the secondary treatment stage. The fnal stage of the treatment of water, the removal of numerous liquefed inorganic composites involving nitrogen and phosphorus takes place. It is called the tertiary treatment stage (Naidoo et al. [2014\)](#page-30-2) after which water is released into the surrounding environment. If these procedures are not done properly, they can contaminate shallow and groundwater supplies (Bustillo-Lecompte and Mehrvar [2017\)](#page-26-3).

Wastewater that originates from diferent types of farm activities involving processes of animal nourishing and the handling of agricultural harvests is agricultural wastewater (Odoemena et al. [2020](#page-31-2)). Another wastewater type is industrial wastewater that originates from the processing of commercial goods such as beverages, food, clothes, paper and chemical produce that we consume. This industrial wastewater contains oils-grease, suspended solids, bacteria, pH, heavy metals and other chemical compounds. Heavy metal contaminants include chromium, cadmium, zinc, and other carbon-based chemical toxins include biocides, hydrocarbons, and surfactants (Kumar et al. [2021\)](#page-29-1). Table [1](#page-3-0) shows the types of wastewater and their efects on algae growth.

Substances in diferent types of wastewater and their purifcation

Wastewater contaminants range from steroids, antibiotics, pesticides, hydrocarbons, heavy metals, and their derivatives which eventually cause considerable environmental damage (Ahmad et al. [2020](#page-25-1); Bhatt et al. [2021;](#page-26-4) Mishra et al. [2021](#page-30-3)). Conventional wastewater treatment methods are not entirely suitable for eradicating contaminants due to high energy consumption, high cost, large area demand, and still less ability to remove contaminants (Crini and Lichtfouse [2019;](#page-26-5) Craggs et al. [2012\)](#page-26-6). Therefore, scientists are trying to fnd out more easy-to-maintain, cost-efective and eco-friendly technologies to eradicate contaminants from wastewater (Saravanan et al. [2021](#page-32-3)). Numerous methodologies have been established to minimalize wastewater releases and lessen the threats of pollutants, including physical methods (adsorption, coagulation/focculation, and membrane fltration, chemical methods (oxidation, ozonation), and biological methods (enzymes, microorganisms) (Xu et al. 2018). Wastewater treatment methods are considered to have purification requirements and effluents' characterization, overlooking their infuence on the environment and general treatment performance (Khan et al. [2020](#page-29-2)).

Adsorption is refected as one of the utmost efective methods among all the above-mentioned methods, for the treatment of wastewater because of many features, which are not so effective in other related methods (Slatni et al. [2020](#page-32-4)). Researchers have just dedicated their struggles to exploring the adsorbents with outsized surface areas, less cost, and friendliness to the surrounding environment. They found their way for efective adsorbents y using nanosized constituents, which have been noticed as an important material for confscating dyes, organic compounds, and heavy metals from wastewater (Nassar et al. [2017](#page-30-4)). Biological treatment methods can be used to remove contaminants, which bacteria, fungi, and algae are commonly used (Rodriguez-Rangel et al. [2022\)](#page-31-3). In this way, wastewater can be treated employing microbes through two types of methods, known as bioaccumulation, secondly biosorption, and biodegradation (Bilal et al. [2021;](#page-26-7) Bai and Acharya [2017;](#page-26-8) Kalra et al. [2021](#page-29-3)).

Microalgae has a signifcant role in the immobilized wastewater treatment system. However, some factors which can afect the immobilized microalgal wastewater treatment system and removal of nutrients, are mainly linked to environmental, biological and operational factors (Nie et al. [2020](#page-31-4)). The main structure of the microalgae immobilized system involves the accepted immobilization technique, matrix material, and essential microalgal species. For instance, alginates-based immobilized technique involves proportions of microalgae beads and the concentration of target contaminants (Emparan et al. [2021](#page-27-1)), bead size (Lee et al. [2020](#page-29-4)), volumetric fractions of diverse matrixes (Abu Sepian et al. [2019](#page-25-2); Rushan et al. [2019\)](#page-32-5), selection of alginate (Kube et al. [2019](#page-29-5); Zamani et al. [2012](#page-33-5)), and microalgal strain (Kube et al. [2021\)](#page-29-6). The bed reactor is another biological method used for the treatment of wastewater. There is limited research on the combined procedure of immobilized microalgae and bed-based reactors. There are three major types of bed reactors, which include fuidized bed reactors (Cheirsilp et al. [2017;](#page-26-9) Zheng and Ke [2017\)](#page-34-0), moving bed reactors (Akizuki et al. [2021\)](#page-25-3), and fxed-bed reactors (Babatsouli et al. [2015](#page-25-4); Garbowski et al. [2020](#page-27-2)).

Over the previous few eras, the importance of the treatment of wastewater has been increased for evaluating the environmental efficiency of conventional wastewater treatment found on the activated slurry procedure (Li et al. [2022a;](#page-30-5) Soares [2020](#page-32-6)). There are various issues related to the wastewater treatment process including emission, con-sumption of energy, and financial gain (Dong et al. [2017](#page-26-10)). The presence of various organics and benefcial nutrients in wastewater promotes the growth of microalgae (Yadav et al. [2021\)](#page-33-6). Consequently, microalgae have successfully aided in the recycling of carbon in the environment (Wang et al. [2016\)](#page-33-7). Thus, neutralizing carbon in the ecosystem efectively yields 1 kWh/kg of biochemical oxygen demand (Sukla et al. [2019](#page-32-7); Al-Jabri et al. [2020](#page-25-5)) which is essential for biological processes. Wastewater treatment methods through microalgae consume less power as compared to conventional methods (Khaldi et al. [2017](#page-29-7)). Apart from all these benefts, harvesting of microalgae is a critical factor, as normally microalgae cultivate at comparatively low cell density (Tan et al. [2020](#page-32-8)).

The main purpose of wastewater treatment is the removal of highly concentrated nutrients, particularly nitrogen and phosphorus, or else these nutrients can cause eutrophication by accumulating in lakes and rivers (Tang et al. [2018](#page-32-9)). Phosphorus is problematic to eradicate from wastewater; however, this challenge has been mitigated by the help of microbial action that helps extract and transform phosphorus into an insoluble solid element during

wastewater treatment. For example, metal salts including iron (Fe) and aluminum (Al) for the congealing process consequently produce Fe–P or Al–P complexes (Kajjumba et al. [2021;](#page-29-8) Korving et al. [2019\)](#page-29-9). Microalgae are efective in eradicating nitrogen, phosphorus, pesticides, toxic compounds, and heavy metals from wastewater through intracellular and extracellular mechanisms of their cell membrane and thus have the possibility to play a significant role in remediation mainly in the fnal treatment of wastewater (Zhao et al. [2019;](#page-34-1) Hussein et al. [2018](#page-28-1)).

Algae can cultivate in numerous aquatic environments, like fresh and oceanic water, municipal sewage wastewater (Zhou et al. [2012a](#page-34-2)), agricultural wastewater (Zhou et al. [2012b,](#page-34-3) [2012d;](#page-34-4) Hu et al. [2013](#page-28-2), [2012](#page-28-3)) and industrial wastewaters (Ali et al. [2021](#page-25-6)). Wastewaters are distinctive in their chemical and physical characteristics depending on whether they come from fresh or oceanic waters. Modern research showed that those algal biomasses which are produced from wastewater have the excessive potential for the production of biofuel and further applications (Wang et al. [2020\)](#page-33-8). More research has been done on the growth of microalgae in municipal sewage wastewater and agricultural wastewater because they are most accessible and are less unstable as compared to other types of wastewater including industrial wastewater (Zhou et al. [2012c](#page-34-5), [2012d](#page-34-4)). Scientists examined the growth of Chlorella species on these four diverse types of wastewater to check their capabilities to remove and utilize nitrogen, phosphorus and chemical oxygen demand, and other minor elements and determined that the growth of algae and efficacy of nutrient removal were relational to the concentration of nutrient of municipal sewage wastewaters (Nirmalakhandan et al. [2019\)](#page-31-5).

Agricultural wastewater, which is often derived from manure, contains high amounts of nutrients like nitrogen and phosphorus in comparison to municipal sewage wastewater. It was observed from studies that microalgae growing on agricultural waste are efficient at removing nitrogen and phosphorus from animal manure-based wastewater. For instance, the green alga (*Botryococcus braunii*) is well developed in piggery wastewater comprising 788 mg L−1 $NO₃$ and 80% $NO₃$ was removed at the initial growth stage. Scientists developed a cheap media by using deep seawater and agitated swine urine and cow dung water for increasing production of algal biomass, signifying the high possibility of the rich nutrient animal compost wastewater stimulating fast algae growth (Pittman et al. [2011](#page-31-6)). There are some studies on retrieval of nutrients by algae growth on animal manure wastewater which evaluated the probability of benthic freshwater algae despite planktonic algae because benthic freshwater algae have more ability of nutrients recovery as compared to planktonic species of algae (Pittman et al.

Table 2 Literature on algae production in urine and wastewater

Types of Algae	Method of cultivation	Source of cultivation	Reference
Spirulina platensis	Bubble column photobioreactor	Human urine	Chang et al. (2013)
Chlorella sorokiniana	Short light-path photobioreactor	Human urine	Tuantet et al. (2014b)
Chlorella			
sorokiniana	Batch cultures Semi-continuous cultures 6-L sequencing-batch microalgae cultivation	Concentrated unfiltered fresh urine. filtered fresh urine as	
N and P sources, non-pre- treated, non-filtered fresh urine in the photobioreactor	Zhang et al. (2014)		
algae production	Mixed algal bacterial bioreactor	Liquid portion of animal manure	Shin and Schideman (2015)
Chlorella vulgaris	Batch cultivations	Diluted human urine	Jaatinen et al. (2016)
Tetradesmus dimorphus	Open photobioreactor	Urban wastewater	Gentili and Fick (2017)
Chlorella singularis, Mic- ractinium pusillum, and Chlorella sorokiniana	Culturing incubation	Human urine, Poultry waste, Cow dung, and urine	Kumar et al. (2018)
Desmodesmus abundans	Immobilized cultivation	Diluted human urine	Piltz and Melkonian (2018)
Chlorella sorokiniana	Bench-scale panel		
photobioreactors	Both synthetic urine and real human urine	Tuantet et al. (2019)	
Chlorella vulgaris	Light-receiving-plate		
(LRP)-enhanced raceway pond	Fresh pig urine	Zou et al. (2020)	
Chlorella vulgaris	Lab-scale membrane photobioreactor	Fresh male urine	Nguyen et al. (2021)
Chlorella emersonii	Erlenmeyer flasks and bubble column reactors. Photoautotrophic biofilm	Municipal wastewater, Brewery waste- water, Dairy wastewater	Schagerl et al. (2022)

[2011](#page-31-6)). Table [2](#page-4-0) shows types of wastewater and their effects on algae growth.

Treatment of wastewater by using algal strains is economically efective and also completes the rigorous release and reusing principles (Whitton et al. [2015](#page-33-11)). Additionally, algal biomass can also be transformed into many valued products like biofuels, biogas, bio-fertilizers, and antibacterial and antiviral mediators, with the consequent treatment of wastewater (Raheem et al. [2015\)](#page-31-8). In recent decades, the cultivation of microalgae has been studied on various types of wastewater (Fig. [1](#page-5-0)) for the removal of contaminants following nutrient recovery with biomass production (Renuka et al. [2015\)](#page-31-9). The accomplishment of those studies mainly depends on the enactment of the particular strains of microalgae. Many species of microalgae including *Chlorella sp.*, (Gouveia et al. [2016\)](#page-27-4) *Micractinium sp.*, (Santiago et al. [2013](#page-32-12)), *Scenedesmus sp.*, (Wang et al. [2019a](#page-33-12), [2019b](#page-33-13)) *Heynigia sp., Actinastrum sp., Hindakia sp., Chlamydomonas sp.,* (Ghayal and Pandya [2013](#page-27-5)), *Pediastrum sp., Botryococcus sp., Dictyosphaerium sp., and Coelastrum sp. and Spirulina sp.,* (Emparan et al. [2019\)](#page-27-6) have been confrmed and were ascertained to be capable to use and remove nitrogen, phosphorus and other minor elements from the wastewater.

Application of microalgae for the treatment of wastewater can be done either by using conventional oxidation ponds or the advanced suspended algal pond schemes including high-rate algal ponds (Arashrio et al. [2018;](#page-25-7) Rao et al. [2019](#page-31-10)), which are known as shallow raceway-type oxidation ponds with automatic mixing, and these are considered as most efficient for the treatment of wastewater. Previous studies

show that microalgae can be cultivated in diferent systems including ponds and bioreactors (Fig. [2](#page-6-0)). These bioreactors involve both open and closed systems (Liyanaarachchi et al. [2021;](#page-30-7) Rezvani et al. [2022](#page-31-11)). For treating great amounts of wastewater produced from municipal sewage, agriculture, and industry, bioreactors should be certainly clambered up and activated for efective removal of nutrients. For this purpose, a raceway pond with paddle-wheel agitation and a multi-layer bioreactor can be used which were assumed as the most reasonable and cheap culture systems for wastewater treatment (Hu et al. [2013](#page-28-2)). Some large-scale assembly of algal biofuels was produced from wastewater by using highrate narrow, open raceway algal ponds, which were used for the treatment of municipal sewage wastewater. For instance, a 2000-L and 40.000-L pilot-scale multi-layer bioreactors were developed successfully for the cultivation of microalgae in wastewater of animal manure and centrate for efficient production of algal biomass by removing nutrients from wastewater (Min et al. [2014\)](#page-30-8). In recent times, an innovative system for the cultivation of microalgae was developed known as "OMEGA" system (offshore membrane enclosures for growing algae). In this system, microalgae were grown in foating photobioreactors positioned in a safe coastal marine environment where outlets of municipal wastewater would also occur (Wiley et al. [2013](#page-33-14); Peter et al. [2022](#page-31-12)).

Urine as wastewater

Composition of urine

Human urine is a liquid solution comprising urea, nitrogen, potassium, phosphorus and sodium chloride as main elements while calcium, magnesium, phosphate, and sulfate as minor elements. Some other trace elements also exist in urine including glucose, amino acid, vitamins and hormones. In general, source-separated urine involves the diversion or collection of urine from its point of production for treatment processes that helps in the maximum recovery of nutrients such as nitrogen and phosphorus (Jimenez et al. [2015](#page-28-4)). There were some methods devised for the recovery of nutrients from source-separated urine. Now, a new bioelectrochemical method is used for the production of energy joints with the recovery of nitrogen from urine found on microbial energy cells (Kuntke et al. [2012](#page-29-11)). More maintainable growth of microalgae can be attained if the nutrients are from reprocessed sources or wastewater supplies (Ahmad et al. [2014](#page-25-8); Rawat et al. [2013](#page-31-13)).

Scientists successfully grew *Spirulina platensis* with already treated urine, and *Spirulina platensis* can also be grown in nitrifed human urine which has a key composition close to the Zarrouk medium, which is an improved medium for culturing. Many studies proved that the growth of *Spirulina platensis* has the potential for the production of aquatic food. The beneft of cultivating microalgae on urine is that they clean urine by eliminating phosphorus and nitrogen from it and also harvest high production of biomass which can be used as biofuel (Chang et al. [2013\)](#page-26-1). A large amount of nitrogen consists of urea $[CO(NH₂)₂]$ in fresh urine, which will be converted into ammonia by hydrolyzation during storage, in the presence of microbes (Zhang et al. [2013](#page-33-15); Kim et al. [2014\)](#page-29-12). Based on this process, the microbial fuel cell has the benefit of handling ammonia (NH_3) in urine besides generating electricity.

The transduction of urine to operational energy for charging smartphones and remote systems by using an integrated microbial fuel cell was done by (Walter et al. [2017](#page-33-16)). The microbial fuel cell generates electrical power through the transformation of chemical energy (Sahu [2019](#page-32-13)). This infers that sources like wastewater, urine, or any other organic matter could be used as fuels for the generation of electricity by a microbial fuel cell (Ha et al. [2012](#page-28-5); Rengasamy and Berchmans [2012](#page-31-14); Samsudeen et al. [2015\)](#page-32-14). The notion of the microbial fuel cell was frst recognized in 1911 by Potter, but only recent data are published which contains pile confguration (Ledezma et al. [2013\)](#page-29-13), optimization of the constituent, and reactor design (Winfeld et al. [2015;](#page-33-17) Gajda et al. [2015](#page-27-7); Ieropoulos et al. [2013\)](#page-28-6).

Cow urine also has been applied to produce electricity by means of a clayware microbial fuel cell, which was costefective and an easy procedure (Jadhav et al. [2016](#page-28-7)). This conversion causes an increase in pH from approximately 6 to 9 followed by bicarbonate ions and ammonia (NH_3) (Larsen et al. [2021\)](#page-29-14). This increase in pH activates the precipitation of magnesium and calcium in the form of phosphate and carbonate composites, which can be easily identifed in stored urine with a decreased level of magnesium and calcium. Diverse sources of nitrogen like ammonium, urea, and nitrate have been considered for the growth of many diverse types of microalgae (Chang et al. [2013;](#page-26-1) Hulatt et al. [2012](#page-28-8)). It was observed that the growth of *Chlorella vulgaris* in a medium that contains nitrogen supply in the form of urea had a somewhat maximum level of biomass growth and harvest as compared to nitrate, whereas no growth happened in the presence of ammonium (Hulatt et al. [2012](#page-28-8)).

Annual production quantities of urine

Typically, one person produces 1.4 L of urine per day (DWA [2016\)](#page-27-8). Normally each individual gives about 2.5–4.3 kg of nitrogen, 0.9–1.0 kg of potassium and 0.7–1.0 kg of phosphorus each year through urine excretion. Additionally, some substances which improve growth also exist in human urine including glucose, amino acids, and vitamins. Though human urine encompasses less than 1% of the entire municipal wastewater by capacity, it gives considerably 69% of nitrogen, 40% of phosphorus, and 60% of potassium in wastewater and, consequently, can be a reason for pollution (Chatterjee et al. [2019\)](#page-26-11).

Urine disposal as source material and input in processes

Fresh urine can be toxic due to the presence of ammonia; therefore, stored urine is supposed to be fne for activities of aquaculture involving algae production because of the primary degradation of microbes, which is followed by detoxifcation of ammonia, deactivation of the pathogen, and stability of nutrients for its usage as nontoxic fertilizer. Human urine has great nutrients and is easily available from urineaverted desiccated eco-toilets. Human urine is considered to be one of the best fertilizers for the growth of phytoplankton in aquatic systems, which are known as important natural food for fsh in pond ecologies. In recent periods, wastewater produced from human actions has instigated many harmful efects on surface water reservoirs.

Human urine is one of the contaminants, which is a waste produced by the human body in liquid form, which comes through the digestion of endogenic wastes (food and drinks) and purifcation of blood and then released by kidneys (Patel et al. [2020\)](#page-31-15). Due to pollution problems, it needs to be considered that an ideal decentralized waste treatment system should be applied for evolving countries where the urine would be properly collected and treated for nutrient recovery purposes (Igos et al. [2017\)](#page-28-9). For example, wastewater-alienated urine has been used as fuid fertilizer for plants and soil. Environmental hygiene toilets are urine departure desiccating toilets constructed on the notion that urine, feces, and water are parts of the environmental loop supplies. Human urine is confned to various types of contaminants with high absorption quantities of nitrogen (76 to 86%), phosphorus (46 to 50%), and potassium (51 to 55%). Human feces also contain 10% nitrogen, 12% potassium, and 40% phosphorus. Consumers use 80% of these sources in their felds as fertilizer. Nutrients from urine are recuperated by cost-efective techniques including urine-diverting desiccation toilets, then precipitation of struvite then nitrifcation, and then distillation, generating valued fertilizer for native societies (Udert et al. [2015](#page-33-18); Krahenbühl et al. [2016;](#page-29-15) Tilley [2016\)](#page-33-19). These

techniques of urine separation are possible, specifcally for rural areas and small towns (Xu et al. [2019\)](#page-33-20).

The capabilities of human urine nutrients should not be ignored and undervalued. It is important to consider the use of urine for the growth of microalgae since in many countries fsh farming is a common means of income for livelihood where culturing and production of fsh are costly because of the implementation of conventional chemicals and fertilizers in aquaculture. On the other hand, the implementation of human urine in place of fertilizer would decrease the cost of production by using standard protocols. One experimental study was performed to compare the results of the primary production of phytoplankton in trial tanks nourished with fresh human urine, stored human urine, and mixed human urine. It was concluded that the prime production of phytoplankton developed in these trial tanks was impacted by the nutrient condition of the tank, specifcally the nitrogen and phosphate added to the tanks that were recovered from human urine (Jana et al. [2012\)](#page-28-10). Modifications in the production of phytoplankton were strongly reliant on the concentration of phosphate in each treatment. The production of phytoplankton occurred due to the increased concentration of phosphate to nitrogen. This shows that the higher concentration of phosphate in the stored human urine was the reason for the maximum values of prime production of phytoplankton in that trial tank (Jana et al. [2012](#page-28-10)).

Human urine comprised some soluble liquefed composites or drug elements with inhibitory efects (Fig. [3](#page-7-0)), which can be omitted from the fuid by stimulated carbon treatment. These pharmaceutical drugs commonly include non-steroidal anti-infammatory drugs (NSAIDs) such as diclofenac and ibuprofen. Non-steroidal anti-infammatory drugs (NSAIDs) have been tested for their infuence on cultures of eukaryotic algae in the axenic laboratory (Bacsi et al. [2016\)](#page-25-9). A stimulated carbon treatment system has been suggested as a feasible choice for decreasing the threat of

Fig. 3 Pharmaceutical compounds found in human urine can inhibit the growth of algae

the augmentation of pharmaceuticals such as antibiotics, non-steroidal anti-infammatory drugs, and beta blockers in a decentralized system of sanitation and recycling systems (Udert et al. [2015](#page-33-18)). Some previous studies show that 60–100% of pharmaceutical drugs like ibuprofen, diclofenac, and paracetamol possibly are removed in an algae photobioreactor, through photolysis and biodegradation (de Wilt et al. [2016\)](#page-26-12). High pH levels and free concentration of ammonia in urine could be the reason to constrain the growth of microalgae (Zhang et al. [2014;](#page-33-9) Chang et al. [2013\)](#page-26-1). At least 100% dilution of urine should be essential for the stable growth of algae (Tuantet et al. [2014a\)](#page-33-2).

The source separation of urine from gray water, brown water, and stormwater might open up several chances for the reclamation of energy, nutrients, and water from waste (Larsen et al. [2016](#page-29-16)). Even with its small volume compared to the wastewater, human urine normally gives more than two-thirds of the nitrogen and half the potassium and phosphorous present in sewage waste (Udert and Wachter [2012](#page-33-21); Chrispim et al. [2017;](#page-26-13) Ishii and Boyer [2015](#page-28-11)). Even with its rich concentration of nutrients, urine cannot be used directly in the feld as an ideal fertilizer because of its non-ideal ratio of phosphorus, nitrogen, and potassium, and its high content of ammonia and organic acids. However, if the conversion of these nutrients into valuable products is done in the right way, the possible ecological and economic benefts would be huge (Ishii and Boyer [2015](#page-28-11); Tarpeh et al. [2017](#page-32-15); Mehta et al. [2015](#page-30-9)).

Utilization of urine by microalgae

Microalgae growth in urine

To prevent eutrophication of surface waters and subsequent blooms of algae or cyanobacteria, nutrients must be removed from wastewater. The same microbes could thus provide a method for nutrient removal in contained treatment systems (Tuantet et al. [2019](#page-33-10)). Human urine contributes approximately 80% of nitrogen, 50% of phosphorous, and 90% of potassium to urban wastewaters despite having a volume of only 1–1.5 L/d per capita compared to 150–200 L/d of wastewater generated per capita (Chatterjee et al. [2019](#page-26-11)). Due to the high content of nutrients, urine can be used as a great source for microalgal cultivation rather than commercial fertilizer. Urine has the potential to provide nutrients for the growth of microalgal biomass, which can then be used as fertilizer and soil conditioner (Tuantet et al. [2019](#page-33-10)). Microalgae are the most promising biofuel feedstock due to their rapid growth rate, high lipid content, comparatively low land usage, and high carbon dioxide $(CO₂)$ absorption and uptake rate (Chang et al. [2013](#page-26-1)). Hence, microalgae-based biomass production can help solve the energy crisis and prevent eutrophication by growing microalgae in urine.

Various microalgae species like *Chlorella sp., Scenedesmus sp., Spirulina Sp. Chlamydomonas sp. Phormidium sp., Synechocystis sp., Synechococcus sp.* etc. have been grown in urine. (Piltz [2018;](#page-31-7) Abdelaziz et al. [2013](#page-25-10)). For instance, Chang et al. ([2013](#page-26-1)) cultivated *Spirulina platensis (Arthrospira platensis)* a cyanobacteria, using synthetic and human urine in a bubble column photobioreactor under autotrophic conditions for ten (10) days. Biomass production was further increased with the addition of sodium acetate as algae could efectively remove and utilize nitrogen and phosphorus*. Chlorella* species like *C. vulgaris, C. sorokiniana, and C. singularis* are widely used in cultivation by using urine as a culture media. Jaatinen et al. [\(2016\)](#page-28-0) cultured *C. vulgaris* using diluted urine and artifcial media and compared their biomass yields. The results showed that diluted urine (1:100) produced the highest biomass yield, while diluted urine $(1:300)$ was sufficient for biomass production in the pH range of 5 to 9. Ammonia production did not show an inhibitory efect on biomass yield.

C. vulgaris has also been studied by Nguyen et al. ([2021\)](#page-30-6) and Volpin et al. ([2019\)](#page-33-22)*.* Nguyen and his team studied nutrient recovery and microalgae biomass production rate using a membrane photobioreactor. Using real urine as a draw solution, algae concentration was increased fourfold in Volpin study. *C. sorokiniana* is also one of the most cultivated microalgae using urine as compared to other studies. Zhang et al. ([2014\)](#page-33-9) cultivated *C. sorokiniana* with a daily supply of fresh urine; it was found that a high amount of biomass production along with nutrient recovery was achieved. Tuantet et al. ([2014a,](#page-33-2) [b](#page-33-3)) and Tuantet et al. [\(2019\)](#page-33-10) optimized algae production and studied biomass production and nutrient removal capacity of *C. sorokiniana* using a photobioreactor. They demonstrated that increased volumetric biomass production and nutrient removal could be achieved by enriching the urine with magnesium, adjusting the nitrogen and phosphorus molar ratio, and shortening the reactor light path.

Other than that, Piltz et al. ([2018\)](#page-31-7) compared the growth of *Chlorella sp, Chlamydomonas sp, Sphaerocystis sp, Chlamydomonas sp, Scenedesmus sp, Synechococcus sp, and Phormidium sp* in urine using a Twin-Layer PSBR. They found that diluted urine provided better growth results than undiluted urine. A comparative study in terms of growth and biomass production was studied between *Secendesmus sp, C. vulgaris,* and *Chlorococcum humicola* in diluted urine in the range of 1:50 to 1:200 (Torres et al. [2014](#page-33-23)). Table [3](#page-9-0) summarizes the relevant studies about the usage of urine as a medium to grow and the associated outcome.

Table 3 Studies on microalgae grown in urine

Piltz (2018)

References

Tuantet et al. (2019) Tuantet et al. ([2019\)](#page-33-10)Torres et al. (2014)

Nutrient recovery by microalgae

Nutrient recovery is a long-term solution for wastewater treatment and the process recovers nutrients that should not be in waterways. These nutrients are then converted into high-quality fertilizers *(Nutrient recovery from wastewater, 2022)*. For concentrated waste streams, nutrient recovery may be economically feasible (Fig. [4\)](#page-11-0). Sourceseparated urine is an ideal target stream in this context because it contains most nutrients found in urban waste-water (Coppens et al. [2016](#page-26-14)). Nitrogen (N) is used in about 100 million metric tons of fertilizer each year. Phosphorus (P) reserves exist in the earth's crust, with annual use of more than 14 million metric tons in fertilizers, accounting for roughly 80% of total P production (Zhang et al. [2014](#page-33-9)). Although urine production volume is only 1–1.5 L per person per day, and total wastewater discharge can be up to 150–250 L/d, approximately 80% of N, 50% of P, and 90% of potassium (K) in domestic wastewater are originated from human urine that made up just 1% of the total wastewater. Human urine contains trace elements such as B, Cu, Zn, Mo, Fe, Co, and Mn in addition to high levels of N, P, and K (Chatterjee et al. [2019\)](#page-26-11). Recycling of nutrients leads to wastewater treatment management. Also recycling carbon (C), N, and P from wastewater and excreta for algae growth is cost-efective (Miranda et al. [2015\)](#page-30-11). Not only does microalgae cultivation promote nutrient recovery from wastewater, but it also promotes biomass production for value-added products such as biodiesel, bioethanol, bio-fertilizers, bioplastics, pharmaceuticals, and feed supplements. If microalgae can be grown in urine, it may be possible to extract nutrients from the urine (Luo et al. [2017;](#page-30-12) Li et al. [2019\)](#page-30-13).

Various innovative techniques, such as struvite precipitation, ion exchange, and ammonia stripping, using urine diversion and treatment systems, have demonstrated the possibility of recycling, and saving water by using underlying nutrients as a source of energy (Chatterjee et al. [2019;](#page-26-11) Posadas et al. [2017](#page-31-16)).

Biomass retention times have a signifcant efect on nutrient remediation and biomass productivity. *C vulgaris* shows biomass retention time-dependent nutrient recovery and biomass retention with the MBPR system (Nguyen et al. [2021](#page-30-6)). Urine concentration, source of urine and urine culture temperature also have profound efects on biomass and lipid production along with nutrient recovery. Diluted human urine has better lipid accumulation in comparison with various artifcial media (Behera et al. [2020](#page-26-2)). *S. platensis* shows great nutrient removal capacity with human urine compared to artifcial urine media (Chang et al. [2013\)](#page-26-1). Nutrient recovery and biomass production increase with a reduction in the dilution of human urine. For example, *C. sorokiniana* biomass

Name of the microalgae

Adding organic carbon sources can reduce the inhibition of NH4

Adding organic carbon sources can reduce the inhibition of NH_4^+ -N to algae

growth and gain more biomass that is high in protein and has high lipid contents

growth and gain more biomass that is high in protein and has high lipid contents

Chlorella sp, Chlamydomonas sp, Sphaerocystis sp, Chlamydomonas sp, Scened-

esmus sp, Synechococcus sp, Phormidium sp

Secendesmus sp, Chlorella vulgaris, and Chlorococcum humicola

Chorella sorokiniana

Sphaerocystis sp, Chlamydomonas sp, Scened-

like autotrophic culture

Chlorella sp, Chlamydomonas sp, like autotrophic culture

+ -N to algae

Chang et al. [\(2013](#page-26-1))

Chang et al. (2013)

hlorella sp, Chlamydomonas sp, Sphaerocystis sp, Chlamydomonas sp, Scened – Growth was satisfactory with diluted urine but not with undiluted urine Piltz ([2018](#page-31-7))
esmus sp, Synechococcus sp, Phormidium sp

Growth was satisfactory with diluted urine but not with undiluted urine

Secendesmus sp, Chlorella vulgaris, and Chlorococcum humicola Biomass production was observed with diluted urine in the range of 1:50 to 1:200 Torres et al. ([2014](#page-33-23))

Chorella sorokiniana can treat urine

efficiently and effectively

efficiently and effectively

The Twin-Layer PSBR system was used to compare the growth of strains on urine

Biomass production was observed with diluted urine in the range of 1:50 to 1:200 The Twin-Layer PSBR system was used to compare the growth of strains on urine

Photoautotrophic cultivation of microalgae on pre-diluted urine can treat urine

Name of the microalgae **References**

Outcome

Fig. 4 Nutrient recovery and biomass production from source-separated urine using a digester and bioreactor in the presence of a light source like sunlight

production improved from 1 g/L to 8 g/L with a 5-time decrease in the dilution of human urine (Tuantet et al. [2014\)](#page-33-2). A decrease in culture temperature leads to lower nutrient recovery in the case of *S. acuminatus* (Chatterjee et al.[2019\)](#page-26-11). Fresh urine as a daily nutrient stock is more efective compared to concentrated used urine for better nutrient recovery (Zhang et al. [2014\)](#page-33-9).

Taken together, it can be concluded that microalgae growth in urine does help in nutrient removal along with biomass production, and the dilution of human urine along with light intensity can play a vital role (Tuantet et al. [2014a,](#page-33-2) [b](#page-33-3)). Table [4](#page-11-1) shows a compilation of studies using urine as a growth media for various microalgae and their nutrient recovery and biomass production outcome.

Impact of urine application on the environment and socioeconomic sector

Considering the high nutrient content of urine, the treatment of source-separated urine appears to be very promising in terms of efficiently recovering nutrients in the form of fertilizers from the perspective of circular economy and energy limitations (Igos et al. [2017;](#page-28-9) Ishii [2015](#page-28-11)). P and N are essential nutrients in crop cultivation and the main component in urine, and their use in food production is becoming increasingly important as the world's population grows (Viskari et al. [2021](#page-33-25)). Despite the numerous advantages of urine separation and fertilizer efficiency demonstrated by research, several obstacles prevent or impede the use of urine nutrients as fertilizer. Photobioreactors are critical to producing microalgae, but they are expensive. Furthermore, urine-diverting toilets, separate pipelines, and maintenance are all required to collect urine as a source for microalgae cultivation. Building all this infrastructure will have a signifcant socioeconomic impact on poor and developing countries. There is still a lack of efficiency in biomass and nutrient recovery at the industrial level, as well as regulatory policies (Pathy et al. [2021](#page-31-17); Badeti et al. [2022](#page-26-15)). Environmental efects such as soil salinization and the spread of pathogens, as well as harmful substances such as heavy metals and pharmaceuticals from urines, are also of great concern. Table [5](#page-12-0) summarizes the impact of the use of urine in various sectors

Table 5 Summary of the impact of the use of urine in various sectors

Impact Sector	Possible Impact	References
Techno-economic and market factor	To use urine as media for microalgae growth and use for nutrient recovery installation of urine- diverting toilets, separate pipelines and maintenance will be necessary	Simha et al. (2017) ; Zhou et al. (2018) ; Pathy et al. (2021) ; Badeti et al. (2022)
	Above all, transportation and management necessitate additional resources means additional costs	
	To harvest microalgae algae reactors like photobioreactors are required those also come at a great price	
	Poor and developing countries where basic infrastructures are lacking will face great economic impact by upgrading the sanitization system and applying urine separation and using sepa- rated urine for microalgae production for nutrient recovery and biomass production	
	The use of urine for biomass production and nutrient recovery is highly effective on the labora- tory scale but as soon as it comes to scale up, efficiency drops significantly, and market value will go very high due to the lack of commercialization	
	Among all low regulatory policies and lack of coordination between the stakeholder will give more hard time in the market sector economically	
Environmental	The risk of spreading pathogens and harmful substances, such as heavy metals and pharmaceu- ticals, into the environment is a concern with agricultural urine use	Viskari et al. (2021) ; Giannakis et al. (2018); Arias et al. (2019); Salehi et al. (2018)
	The high salt content in urine like sulfate and chloride soil salinization poses a high risk	
	Urine contains a high concentration of biodegradable organic compounds, which causes urine to become contaminated with a variety of microorganisms such as bacteria and pathogens, leading to several infections when handled improperly	
	Bacterial presence in the urine causes spontaneous changes in urine such as ammonia volatili- zation, microbial urea hydrolysis, and mineral precipitation	
	Due to the high use of pharmaceutical products, urine contains a high number of such product and those compounds undergo several drug metabolism pathways and leads to the production of formation of endocrine-disrupting chemicals (EDC) and compounds. EDC exposure may contribute to several acute and chronic diseases, including cancer, diabetes, infertility, and brain damage	

Fig. 5 Photobioreactor mechanism for biomass production in the presence of light source and with the help of a pump module to pump media and gas mixture through the photobioreactor

such as the environment, socioeconomic, and global markets (Giannakis et al. [2018\)](#page-27-9).

Microalgae production

Design of photobioreactors

Photobioreactors (PBR) are closed systems made of glass, plastic, or other transparent materials in the form of plates, tubes, or bags. The high-power consumption and operating costs of artifcial light have traditionally been major costs associated with photobioreactors, but solar-lit photobioreactors do exist (Fig. [5](#page-13-0)). The light intensity is critical for the microalgae growth in the photobioreactor; hence, the optical properties of the materials used in photobioreactor construction are vital. Materials like glass, polyvinyl chloride, polyethylene, polycarbonate, plexiglass, fberglass, etc. are commonly used for constructing photobioreactors (Johnson et al. [2018](#page-29-17)).

The integration of capturing, transportation, distribution, and utilization of light by microalgae via photosynthesis determines the efectiveness of PBRs. In the 1940s, the frst PBR systems were developed and designed (Johnson et al. [2018](#page-29-17)). PBR design can be efective if (a) the PBR utilizes the maximum light available and process it to the PBR cultivation vessel for maximum biomass production, (b) operative and pinpointed control over operational parameters, (c) mitigate capital and operational costs, as well as energy consumption during operation with maximum light energy used for biomass production (Wang et al. [2012](#page-33-26); Fu et al. [2019](#page-27-10)).

When selecting a PBR design for a specifc manufacturing process, there are numerous factors to consider. Some PBR designs are more commonly used than others; thus, a design should be chosen on a case-by-case basis to optimize the efficiency of a microalgae biofuel production process. The following sections describe some of PBR designs.

Porous membrane photobioreactors

The porous membrane of the tubular PBRs can be either vertically or horizontally supported. The growth medium circulates within the dialysis membrane tube and seeps through pores to the membrane's outer surface (Ojanen et al. [2015](#page-31-18)). A dialysis membrane tube is placed inside the PBR tube, and microalgae can be grown outside the membrane. A $CO₂$ source is used to supply aerated $CO₂$ gas to the culture, which, along with light, can aid biomass production. The signifcant interfacial area provided by the membrane is the biggest advantage of porous membrane PBRs, but high-water vapor permeability could be a pitfall (Sarbatly et al. [2013](#page-32-18)).

Bubble column photobioreactors

Bubble column reactors are multiphase contactors that use the dispersion of a gas phase in the form of bubbles inside a cylindrical vessel containing a liquid or a suspension (Leonard et al.[2015](#page-29-18)). Bubble column PBRs have a higher surface area-to-volume ratio than other types of photobioreactors, making them more efficient in microalgae production with greater volumetric and areal productivity (Mubarak et al. 2019). The bottom of the reactor is sparged with $CO₂$, causing bubbles to rise uniformly across the cross-sectional area and exit through the reactor top (Gerdes et al. [2014](#page-27-11)). Compressors and gas flow meters are combined with controllers to control the speed and size of gas bubbles dispersed in the microalgae suspension (Saldarriaga et al. 2020). Microalgae cells can utilize $CO₂$ from the bubble for biomass production. If external illumination is used, bubble column PBRs are typically made of transparent material. Plastics such as polyvinyl chloride and plexiglass are common, and glass is also used commonly. Transparent double jackets are wrapped around PBR or the PBR is placed inside an incubator for temperature control. Stainless steel tubes as heat exchangers also can be used (López-Rosales et al. [2016\)](#page-30-15). Because microalgae are susceptible to mechanical shear force, the excessive mechanical shear force caused by over-aeration causes cell damage (Ding et al. [2021](#page-26-16)). PBR has several advantages, including high heat and mass transfer rates, no moving parts, compactness, ease of operation, and low maintenance and operational costs. Their main disadvantage is the signifcant back mixing, which can afect reaction product selectivity and conversion (Leonard et al. [2015](#page-29-18)).

Airlift photobioreactors

Airlift PBRs come under vertical tubular PBRs. Cylindrical airlift PBRs are built with distinct vertical fows known as risers and downcomers separated by a physical barrier. The barrier that separates the downcomer and riser does not extend to the airlift reactor's top or bottom. The air enters the riser via a sparger located in the PBR's riser section. The growth medium is compressed in the sparger's bubble fow and travels up the PBR to the surface, where air exits the growth medium. The growth medium cannot exit the PBR with the air, forcing it to turn laterally to the downcomer, where it turns and moves vertically down the PBR. Because the growth medium cannot exit the bottom of the PBR, it returns to the riser, from which the sparger entrains gas into the fow, and the process is repeated (Johnson et al.

[2018\)](#page-29-17). For adequate light availability in the center of an airlift PBR, the diameter of the column should not exceed 0.2 m. Furthermore, the cylinder's height is limited to about 4 m for structural reasons, due to the strength of the transparent materials used, and to reduce mutual shading in large commercial cultivations. High capital and cleaning costs are big disadvantages for an airlift PBR, but it does help with low power consumption, low shear stress, good mixing, and mass transfer (Huang et al. [2017\)](#page-28-12).

Tubular photobioreactors

The PBR is divided into two major components: the solar receiver and the airlift system. Solar receivers are designed to maximize solar radiation interception while minimizing flow resistance and occupying as little space as possible. A bubble column, on the other hand, is used for mixing, degassing, and heat exchange culture (Fernandez et al. 2014). CO₂ is also injected at the end of the airlift system and at the start of the solar receiver to increase $CO₂$ residence time in the solar receiver section. A heat exchanger also can be included in this section to improve control over the cultivation temperature (Ojanen et al. [2015\)](#page-31-18).

Vertical and horizontal tubular photobioreactors are the two most common tube orientations used in laboratories, and both can be operated continuously, semi-continuously, or discontinuously. Depending on the type of experiment, temperature, pH, and oxygen concentration can be measured at various points along the tube, while liquid and gas flow rates are measured at the $CO₂$ injection point (Solimeno et al. [2017\)](#page-32-20). The challenges associated with tubular PBRs are related to maintaining a low partial pressure of $CO₂$ along with uniform light intensity. Due to the mixing and cooling system of PBRs, tubular PBR is relatively costly to construct and maintain. The rise in $CO₂$ concentration at the injection point, and the maintenance of the same, can be a problem. The production of microalgae biomass is directly proportional to $CO₂$ concentration (Giannelli et al. [2012;](#page-27-13) Moraes et al. [2020\)](#page-30-16).

Flat plate photobioreactors

Flat plate PBRs are used for medium-scale cultivation of microalgae. PBR fat panels are made of two joined plates that are kept at a constant distance from each other. Flat plates can be placed vertically, horizontally, or at an angle depending on the method used. Because of the larger surface area, light consistency is very high, resulting in better microalgae cultivation (Li et al. 2014; Huang et al. [2015\)](#page-28-13). For aeration and cultivation, spargers, tubes, or membranes are used to mix and deliver air and CO_2 (Yan et al. [2016](#page-33-27)). CO_2 can be used for the pH balance of the microalgae suspension as the pH normally rises due to the use of $CO₂$ by the microalgae (Pfaffinger et al. [2019](#page-31-19)). Though Light-Emitting Diodes (LEDs) are commonly used as a light source, other sources like neon lamps, sodium vapor lamps, OLEDs, fuorescent, halogen lamps, etc. also have been used. The fat panel PBRs have two-sided lighting (Huang et al.[2015;](#page-28-13) Benner et al. [2022](#page-26-17)). When light distribution in flat panel PBRs is compared to the light distribution in cylindrical bubble column PBRs illuminated from the outside, the light attenuation caused by microalgae in suspension is much more pronounced than in fat panel reactors (Jacobi et al. [2012\)](#page-28-14). Transparent double jackets are used as heat exchangers for temperature control, or PBRs can be placed in water baths to maintain temperature. pH, dissolved oxygen, and temperature are measured online with sterilizable electrodes (Vogel et al. [2018\)](#page-33-28).

Bag photobioreactors

This type of reactor is specifcally important for low- and middle-income countries that require an inexpensive cultivation system, are technically simple to operate, scalable, and meet basic good manufacturing practice requirements. These requirements are fulflled by a disposable bag PBR (Cui et al. [2022](#page-26-18)). Bag PBRs are made up of three parts: plastic bags, a frame (to hold the plastic bag), and aeration systems. The size, materials, aeration types, mixing methods, and frame structure are all important design considerations (Ting et al. [2017\)](#page-33-29). Sterilization of these bags is a big disadvantage as the material used for bag production is mostly polyethylene. Although UV or gas sterilization are options, they come at a high cost. Bag PBR systems are frequently hung vertically, with growth medium pumped to the top and flowing down to the bottom. Similar to tubular PBRs, gas exchange occurs in a unit separated from the photosynthetically active area of the bag (Johnson et al. [2018\)](#page-29-17). Cost-efectiveness is the main advantage of the bag type PBR but photolimitation, bad mixing, frailty to leakage, and short life span are some disadvantages associated with it (Huang et al. [2017](#page-28-12)).

Photobioreactors are closed systems used for the cultivation of microalgae. They provide precise control of environmental conditions such as temperature, light, and nutrients, which can result in higher productivity and better quality of microalgae biomass (Sun et al. [2016\)](#page-32-21). Photobioreactors have higher productivity than open pond systems because they can provide optimal conditions for growth. In photobioreactors, microalgae receive a uniform amount of light, CO2, and nutrients, which promotes their growth and reproduction. Photobioreactors provide a sterile environment that minimizes contamination by unwanted microorganisms. This makes the cultivation of specifc strains of microalgae easier and can result in higher purity of the fnal product.

Photobioreactors allow for precise control of environmental conditions, which can be adjusted according to the requirements of diferent microalgae strains. The control of temperature, pH, and nutrient levels can optimize the growth and productivity of microalgae. Photobioreactors can be scaled up or down depending on the production needs. This makes them suitable for small-scale or large-scale production of microalgae (Chanquia et al. 2021).

However, photobioreactors also have some disadvantages that should be considered before choosing this cultivation method. Photobioreactors require a signifcant capital investment to build and maintain. The cost of the equipment and technology needed for photobioreactor cultivation is often higher than that of open pond systems. The use of artifcial lighting and heating systems in photobioreactors requires a signifcant amount of energy, which can result in high operating costs. Additionally, the energy source used to power these systems may have environmental impacts. Photobioreactors require regular maintenance to ensure their proper functioning. This includes cleaning, sterilization, and monitoring of the system components. Photobioreactors require skilled personnel to operate and maintain the system. The complexity of the equipment and technology used in photobioreactors can make the cultivation process more challenging than open pond systems (Amaral et al. [2020](#page-25-12)). In conclusion, photobioreactors offer several advantages over open pond systems to produce microalgae. They provide a controlled environment that allows for higher productivity and reduced contamination. However, they also have some disadvantages that should be considered, such as high capital cost, energy consumption, maintenance requirements, and complex operation. Ultimately, the decision to use photobioreactors for microalgae cultivation should be based on the specifc needs and goals of the production process.

Parameters for optimal microalgae growth

Strain, carbon and nutrient levels, light intensity, light/dark cycle, temperature, and pH have an impact on microalgae growth and carbon fixation efficiency. These factors are critical for providing optimal condition management. Understanding how these variables afect algae growth and biological carbon sequestration can aid in process optimization.

Carbon source and strain

Microalgae can absorb dissolved inorganic carbon (DIC) from the aquatic environment in the forms of $CO₂$, $H₂CO₃$, HCO_3^- , and CO_3^2 ⁻ from the suspension (Zhou et al. [2017](#page-34-8)). Diferent microalgae have diferent preferences for DIC form. For instance, gaseous $CO₂$ is the most suitable for *Chlorella sp.* Similarly, *Nannochloropsis sp.* can actively

transport HCO_3^- (Piltz [2018](#page-31-7); Zhou et al. [2017](#page-34-8)). A few strains of microalgae can absorb HCO_3^- , which is then converted to $CO₂$ by carbonic anhydrase. The process of converting $CO₂$ and water into organic compounds powered by ATP and NADPH generated by photosynthesis is referred to as microalgae $CO₂$ fixation. Microalgae capture $CO₂$ through the Calvin cycle, such as carboxylation, reduction, and regeneration. During the transfer process, the main limiting factors influencing $CO₂$ fixation are $CO₂$ transportation resistance and difusion (Razzak et al. [2017](#page-31-20); Gerotto et al. [2020](#page-27-14)). Low carbon concentrations promote unsustainable microalgae growth, and dissolving enough $CO₂$ in the growth suspension is expensive. Below 5% (V/V) CO₂ concentration is best for microalgae growth, which is less than 10–20% flue gas $CO₂$ concentration. Microalgae cannot always use organic carbon due to the large molecular size of carbon or to being trapped in large solid particles (Chowdury et al. [2020](#page-26-19); Zhou et al. [2017](#page-34-8)).

Light source and light intensity

Light intensity is critical in microalgae photosynthesis, and when light intensity exceeds a critical level, light saturation and photoinhibition may occur (Fig. [6\)](#page-16-0). Depending on the light stress and the length of time the microalgae are exposed to the stress, photoinhibition is either reversible or irreversible (Wang et al. [2012](#page-33-26)). Natural light is the most easily available and the cheapest option as a light source for microalgae cultivation, but it has some disadvantages due to changes in weather, diurnal cycle, and season. Although sunlight has a photon flux density (PFD) of over 2000 µmol. m^{-2} . s⁻¹, organisms that use oxygenic photosynthesis can only achieve a theoretical maximum conversion efficiency of 8%-10% solar-to-biomass energy (Huang et al. [2017](#page-28-12)).

Photosynthetic microorganisms such as microalgae can only use the 400–700 nm range, known as photosynthetically active radiation (PAR), but PAR accounts for only 50%

Fig. 6 Effect of light intensity on the growth rate of algae and rep-
Fig. 6 Effect of light intensity on the growth rate of algae and represents the requirement of optimal light intensity for optimal growth rate

of sunlight (Acien et al. [2013](#page-25-13)). Non-uniform light intensity distribution inside a PBR occurs due to the absorption and scattering of light in the culture. Radiation attenuation is dependent on the light wavelength, cell concentration, PBR geometry, and light penetration distance. If the light intensity is higher than a critical value, photoinhibition will take place and extra light will be wasted as fuorescent and heat. In case of lower light intensity, photolimitation will take place and the culture will collapse.

Depending on the cell growth due to light intensity, PBR can be divided into three zones, viz. strong illumination zone, weak illumination zone, and dark zone (Huang et al. [2017](#page-28-12)). Even during the day, microalgae in a controlled cultivation system are subjected to a diferent type of light/dark cycle. Because of the light gradient inside a PBR, algae are cyclically exposed to diferent microenvironments, from the light near the PBR surface to the dark in the center of a solar tube. As a result, algae are subjected to light/dark cycles that afect biomass production (Yen et al. [2019\)](#page-33-30).

Light-Emitting Diodes (LEDs), halogen lamps, fuorescent lamps, and incandescent bulbs are all examples of artifcial light sources that can be used for microalgal cultivation. The energy consumption, spectrum, wavelength distribution, and cost of these light sources difer. When these factors are considered, fuorescent lamps and LEDs are the most used light sources for microalgae production. When compared to fuorescent lamps, the use of LEDs allows for better light control, and the use of diferent wavelengths can be benefcial for biomass production (Chowdury et al. [2020](#page-26-19)).

Temperature

It has been reported that $20-30$ ^oC is the optimal temperature for microalgae cultivation. The growth rate becomes slow below 20^0C and when the temperature goes above 30^0C severe death and damage can occur. Marine microalgae are easily susceptible to slow growth rate at 28° C and death at 30^0 C. Freshwater microalgae can operate between 25 and 30^0 C, and at even 40^0 C also shows tolerance for a brief period (Acien et al. [2013\)](#page-25-13). PBRs are optically transparent due to the need for light. Without a temperature control unit, the inside PBR temperature can rise to $10{\text -}30^0C$ from the ambient temperature in the summertime. High temperatures can lead to photocatalytic reactions and cause disturbance in the chemical equilibrium of the species, along with the solubility of supplied gas and pH (Huang et al. [2017\)](#page-28-12). As a result, additional cooling mechanisms are frequently used to keep the culture within a safe range. Submerging the entire culture in a water pool, spraying with water, shading, or incorporating a heat exchanger with PBR for cooling is some

pH and salinity

The optimal pH range for most of the microalgae species is between 7 and 9, and it is crucial for optimal growth and target product formation (Yen et al. [2019](#page-33-30)). The solubility of $CO₂$ and carbonates in the microalgae suspension and availability are greatly dependent on the pH. The hydroxide ion (OH) accumulates in the growing medium, causing the pH to gradually rise during photosynthetic $CO₂$ fixation. This shifts the chemical equilibrium of the inorganic carbon in the medium toward the formation of carbonates (CO^{2-}_{3}) (Chowdury et al. 2020). Thus, $CO₂$ availability at high pH becomes a trouble for microalgae growth. Nutrient uptake by cells, intracellular metabolite production, and metal toxicity can be caused by acidic pH, whereas alkaline pH decreases the affinity of microalgae for $CO₂$ and interferes with cell division (Chang et al. [2017](#page-26-20)). pH also can interfere with photosynthetic activity. The pH of the culture is also afected by nitrogen addition. When nitrogen is supplied in the form of ammonium, the pH of the solution rises due to a decrease in the concentration of nitrogen in microalgae. Elevated pH can also infuence the concentration of phosphorus in a culture medium by causing phosphate precipitation. Microalgae, bacteria, and cyanobacteria can generally withstand salt concentrations of up to 1.7 M. (Cai et al. [2013](#page-26-0)). pH control mechanism should be employed with the PBR, with sodium bicarbonate widely used to control the pH (Huang et al. [2017\)](#page-28-12). *Chlorella sp.* KR-1, for example, grew well in a pH range of 4–7, but its growth was signifcantly inhibited at a pH below 3.5 (Zhou et al. [2017\)](#page-34-8).

Oxygen removal

Microalgae photosynthesis uses light energy and $CO₂$ to produce oxygen (O_2) as a byproduct. In the rubisco enzyme, microalgae have a lower binding affinity for $CO₂$ than $O₂$. Excessive dissolved O_2 competes with CO_2 for the rubisco binding site, resulting in photorespiration that reduces photosynthetic activity (Chang et al*.* [2017\)](#page-26-20). The presence of excess oxygen and light can lead to the formation of reactive oxygen species (ROS), thus leading to cellular damage. The dissolved O_2 concentration in the medium should not be higher than 400% of the air saturation level, and algae cannot tolerate elevated $CO₂$ levels for more than 2 or 3 h. Only the degasser removes accumulated dissolved O_2 via mass transfer because dissolved O_2 not only oxidizes one or more enzymes but also afects the electron transmission chain and suppresses photosynthesis. Thus, dissolved $O₂$ accumulation should be avoided, and effective mixing is an excellent way to reduce dissolved O_2 (Huang et al. [2017\)](#page-28-12).

Mixing

Mixing is directly related to biomass production because it is the most limiting parameter for optimal microalgae growth and biomass production (Yen et al*.* [2019\)](#page-33-30). Microalgae density can reach $10⁹$ cells, resulting in reduced light transmission, increased rates of dissolved $CO₂$ consumption and dissolved $O₂$ accumulation, and a rapid increase in culture temperature (Wang et al. [2012](#page-33-26)).

Besides, mixing ensures uniform nutrient distribution to cells and efficient light utilization, as well as facilitating heat transfer, promoting gas exchange, keeping cells suspended, and preventing cell clumping and attachment to PBR walls. Furthermore, mixing is also important in PBRs because it moves cells from the photic zone to the dark zone, artifcially providing light and dark cycles to boost productivity.

A mixing or agitation system is classifed into two types, namely nonmechanical and mechanical. Non-mechanical mixing systems include airlift, bubble columns, and tubular PBR, while mechanical mixing systems include stirred tanks and raceway ponds (Chang et al. 2017).

Sterility and cleanability

Slow-growing microalgae species are hard to cultivate as they take a longer period for growth, biofuel production, and $CO₂$ sequestration, thus leading to impurity formation in the PBR (Wang et al. 2012). Bag PBR has issues with sterilization as they are made up of polyethylene (Johnson et al. [2018](#page-29-17)). Cleanability is essential for a PBR to reduce the formation of bioflm on the surface and to ensure the sterility of the PBR. In this situation, PBR should be designed with a smooth internal surface that helps the prevention of bioflm formation and other sedimentation in the wall along with a large enough surface area for conventional cleaning (Wang et al. 2012).

Nutrient

For microalgae growth, major nutrient requirements are N, P, and mineral elements such as magnesium (Mg), potassium (K), iron (Fe), cobalt (Co), and vitamins. Optimal N and P levels, and $C: N: P$ ratios, promote rapid growth and efficient CO_2 fixation (Zhou et al. [2017\)](#page-34-8). Urine as a microalgae growth media is highly suitable because of the high presence of N and P along with other essential nutrients. According to the molecular formula of biomass, approximately 50% of the biomass is composed of C (Chowdhury et al. [2020\)](#page-26-19). C is essential as the building block for carbohydrates, proteins, nucleic acids, vitamins, and lipids. A high autotrophic production rate requires a supply of CO_2 and HCO_3^- . N can be found in 1–14% of the dry weight of microalgae biomass and is responsible for proteins, nucleic acids, vitamins, and

photosynthetic pigment formation. The percentage of P is in the range of 0.05 to 3.3% in dry mass of microalgae biomass and is essential for the formation of nucleic acid and DNA along with energy transfer (de Alva et al. [2018](#page-26-21)). Nutrients such as ammonium, ammonia, and other substances found in urine, including heavy metals, drugs, and disinfectant byproducts are all very toxic to microbial growth (Collos et al. [2014\)](#page-26-22). To overcome this situation, microalgae-based nutrient removal can be efective.

Nutrient content

Several nutrients in microalgae can potentially be used as the best substitute for traditional foods. Microalgae comprise many nutritional composites which can generally be considered superfoods. Spirulina is known as "the future food" as confrmed by the Food and Agriculture Organization (Ahsan et al. [2008\)](#page-25-14). Microalgae have compressed nutrients that can be used in space missions; therefore, the National Aeronautics and Space Administration of the USA built some algal species into astronauts' diets as they fundamentally require a great intake of nutrients in dense volumes (Kainer [2018](#page-29-19)). Microalgae also have bioactive composites including antioxidant, anti-infammatory, anti-carcinogenic, antihypertensive, anticoagulant, anti-diabetic, immunomodulatory, cardiovascular protective, antiviral, and antimicrobial (Elaya Perumal and Sundararaj [2020](#page-27-15); Nova et al. [2020\)](#page-31-21). The study on the use of algae having nutritional and nutraceutical characteristics allowed their inclusion into the "foodomics" (Dvir et al. [2021](#page-27-16)). There is a description of nutrient contents that are present in algae, and both extrinsic and intrinsic

factors such as nutrients, light, and CO2 generally enhance algal biomass production (Fig. [7\)](#page-18-0).

Proteins and peptides The main concern of microalgae nutrient contents is the protein that is abundantly present in algae; for instance, *Spirulina platensis* have up to 70% of their dry weight as protein. It was observed that half of the customary protein will be substituted by proteins from algal or insect-based diets in the coming year 2054 (Khanra et al. [2018](#page-29-20)). Moreover, some species of microalgae like *Cholera vulgaris* and *Arthrospira platensis* have similar quality of amino acids as in soybean (Grossmann et al. [2019\)](#page-28-15). Microalgae proteins can be well digested as compared to other food products like oats, wheat, and beans, and help to reduce causes of hypertension, cardiovascular diseases, and oxidative stress (Levasseur et al. [2020](#page-30-17)). Species of microalgae that provides useful peptides in food are *Chlorella* spp., such as *C. vulgaris, C. ellipsiodea*, and *Spirulina platensis, Nannochloropsis oculata* (Ejike et al. [2016\)](#page-27-17).

Lipids and essential fatty acids Microalgae also have essential fatty acid and polyunsaturated fatty acids including omega-3 polyunsaturated fatty acids like docosahexaenoic acid and eicosapentaenoic acid and (DHA). These bioactive composites have many benefts for human health involving the control of high blood pressure, and stroke, which decreased the risk of cardiac arrest, and helped in resolving asthma, and depression (Adarme-Vega et al. [2012\)](#page-25-15). These are important factors involved in brain development, especially at the early stages of childhood (Yeiser et al. [2016](#page-33-27); Charles et al. [2019\)](#page-26-23). Microalgae are a good substitute for

High Light, Low Light, **Strong Light** CO₂ and CO₂ and and Nutrient **Nutrient Nutrient Starvation** Light Concentration **Biomass** Concentration **Lipid and Biomass** Content **Dissolve CO2** Nutrient Adaptation Exponential **Stationary** Phase Phase Phase

fish for the provision of these polyunsaturated fatty acids (Yeiser et al. [2016\)](#page-33-27), and the main species with the status of Generally Recognized As Safe involve *Arthrospira, Haematococcus, Chlorella, Crypthecodinium cohnii, Dunaliella,* and *Porphyridium cruentum* (Caporgno and Mathys [2018\)](#page-26-24).

Carbohydrates and polysaccharides Carbohydrates also present a major part of microalgae contents; however, the content of carbohydrates can vary in diferent developmental conditions. For instance, *Chlorella vulgaris* can yield 9–41% carbohydrates in its dry mass and S*cenedesmus obliquu*s provides 10 to 47% of carbohydrates. The foremost species of microalgae used to provide polysaccharides involve *Chlorella* sp., *Porphyridium purpureum, Tetraselmis* sp., *Isochrysis* sp., *Porphyridium cruentum,* and *Rhodella reticulate* (Levasseur et al. [2020\)](#page-30-17). Polysaccharides are the most ample sort of carbohydrate in microalgae, which were used for congealing and gelling properties (Bernaerts et al. [2019](#page-26-25)). For instance, beta-glucans might control the immune response, while sulfated polysaccharides can prevent a variety of cancer cell lines by having antioxidant properties (Nazih and Bard [2018](#page-30-18)).

Carotenoids Carotenoids are regular pigments that have many functional properties for improving human health, including antioxidants, anticancer, anti-infammatory, and antiobesity efects, and averting ophthalmic and cardiac diseases (Cezare-Gomes et al. [2019](#page-26-26)). Many commercially signifcant carotenoids (containing β-carotene, astaxanthin, and zeaxanthin) can be obtained through microalgae. Some species like *Chlorella* spp., *Dunaliella salina and Haematococcus pluvialis* have carotenoids which are mostly used for the manufacturing of nutraceutical products due to their anti-aging and antioxidant characteristics (Mehariya et al. [2021\)](#page-30-19). Matters of astaxanthin in *Haematococcus pluvialis* can range from 3% of dry weight while β-carotene in *Dunaliella salina* can range up to 10%, of dry weight (Cezare-Gomes et al. [2019](#page-26-26)).

Vitamins Vitamins obtained from microalgae have an important role in the metabolism of all humans, animals, and microbes. A deficiency of vitamins causes night blindness, beriberi, and rickets. *Dunaliella* spp. microalgae produced pro-vitamin A (β-carotene), vitamin E (Tocopherol) which was higher than soybean oil, and vitamin C (Ascorbic acid). Though, *Euglena gracilis* was the greatest manufacturer of vitamin C as compared to lemon, for instance (Ismailkhodjaev et al. [2019\)](#page-28-16). *Arthrospira platensis* and *Chlorella* spp. Also produced Vitamin C (Andrade et al. [2018](#page-25-16)). *Dunaliella tertiolecta* has the potential to produce cobalamin (vitamin *B*-12) and ribofavin (vitamin *B-*2), while *Chlorella* spp. produce a high amount of biotin (vitamin *B-*7) (Koyande et al. [2019](#page-29-21)). Recent research showed that *Nannochloropsis* *limnetica, N. oceanica,* and *Dunaliella salina* can produce vitamin D3 after exposure to Ultraviolet-B light (Ljubic et al. [2020\)](#page-30-20).

Minerals Minerals have important roles in many physiological functions in the human body, and their scarcity can lead to disease indications. *Arthrospira* species can be signifcant sources of minerals like potassium, magnesium, phosphorus, zinc, and chromium, calcium 180% more than milk, and iron 5100% more than spinach (Alagawany et al. [2021\)](#page-25-17). *Chlorella vulgaris* also produce calcium, zinc, magnesium, manganese, copper, and iron (Alagawany et al. [2021](#page-25-17)). Manufacturing of food with microalgae can upsurge their mineral matters which is lower than minerals of traditional compounds (Pomport et al. [2021](#page-31-22)).

From the above discussion, it is confrmed that the use of urine as a nutrient source for microalgae cultivation can have a signifcant infuence on the nutrient content of microalgae-based food. Urine is a rich source of nitrogen, phosphorus, and potassium, which are essential nutrients for microalgae growth. When urine is used as a fertilizer for microalgae, it can enhance the growth rate and biomass yield of the microalgae, resulting in higher nutrient content in the fnal product. Microalgae grown with urine as a nutrient source may contain higher levels of nitrogen, phosphorus, and potassium than microalgae grown with conventional synthetic fertilizers or other organic sources of nutrients. In addition, microalgae cultivated with urine may also contain other essential micronutrients such as calcium, magnesium, and sulfur, which are also present in urine. The nutrient content of microalgae-based food can be infuenced by various factors such as the strain of microalgae used, the cultivation conditions, and the processing methods.

However, the use of urine as a nutrient source for microalgae cultivation can be an efective way to increase the nutrient content of microalgae-based food, particularly in terms of nitrogen, phosphorus, and potassium. Moreover, the use of urine as a nutrient source for microalgae production can also have environmental benefts, as it can reduce the reliance on conventional synthetic fertilizers, which can have negative impacts on soil and water quality. Urine is a readily available and renewable resource that can be collected and processed locally, reducing the carbon footprint associated with the transportation of fertilizers. In conclusion, the use of urine as a nutrient source for microalgae cultivation can significantly influence the nutrient content of microalgaebased food, particularly in terms of nitrogen, phosphorus, and potassium. However, the safety and hygiene of the fnal product must be ensured through appropriate treatment and processing methods (Chang et al. [2013;](#page-26-1) Lamminen et al. [2019](#page-29-22); Tao et al. [2022](#page-32-2)).

Capital cost, operating cost, and proftability

A PBR's rational design is critical, and the capital cost (high surface area/volume ratio), operating cost (i.e., the cost of auxiliary energy demand, cleaning, and maintenance), and lifespan of the PBR should all be considered before commercial exploitation. A good PBR should have a simple structure, simple operation, and temperature control, a low capital cost, a low operating cost (i.e., low energy consumption and easy cleaning and maintenance), and a long lifespan (Huang et al. 2017).

The three storage carbon substances (i.e., protein, carbohydrate, and lipid) are the main products from microalgae that are exploited in various market scenarios, according to the economic analysis (Sun et al*.* [2018\)](#page-32-22). Animal feed prices are calculated using a broad correlation with protein content. Protein-rich microalgae, such as *Chlorella pyrenoidosa*, are promising strains in animal feed, with a value of \$ 750 tonne−1 (Ruiz et al*.* [2016](#page-32-23)). There are numerous business opportunities for microalgae products, and the interest in investing in them is growing over time. If nutrients are supplied in sufficient quantities, production can be produced, leading to a higher yield.

Algae must be sold as high-value products to attain a reasonable proft. For instance, *Haematococcus pluvialis*, an algae species rich in the natural source of the red pigment astaxanthin, has a high market value and demand (Kaur [2021](#page-29-23)). Biodiesel also can be obtained from microalgae biomass. If the capital cost and operational cost are maintained and along with sustainability, there will be a great market valuation for microalgae-based products.

Potential hurdles

Consumer opinion on the use of urine as an input in microalgae cultivation

Consumers are very concerned about the chemical and microbial safety of algal food products, for example, the cyanobacteria, and microalgae *spirulina*, are generally consumed as food or used as a nutritional supplement in many European countries such as France, Germany, and Spain (Araújo et al. [2021](#page-25-18)). Consequently, in Europe, strict legislations such as EC 2073/2005 and EC 1441/2007 are evident for both producers and consumers as vital requirements for the safety of food products. Although no standard microbial criteria have been established for microalgae, microalgae considered for food and nutritional purposes are categorized as 'vegetable products' under the European Food Safety Authority system of classifcation (Heinsoo [2014](#page-28-17)).

As urine is considered an alternative nutrition cultivation medium for alga growth, many studies have concluded the

high content of contaminants such as pharmaceutical pollutants and the presence of heavy metals such as lead and copper. It has thus been confrmed that between 64 and 80% of undigested pharmaceutical products are excreted through urine which fnds their way as residues or metabolites of pharmaceutical compounds into mainstream wastewater treatment plants (Ronteltap et al. [2007](#page-31-23); Heinsoo [2014](#page-28-17)). Thus, an efective wastewater treatment process that can considerably eliminate most contaminants in urine will ensure maximum nutrient recovery and a safer and contaminant-free urine suitable for algae cultivation for food application purposes. Furthermore, the adoption of cost-efective industrial processes in wastewater treatment that can efficiently provide an efective concentration of treated urine for microalgal cultivation is very essential. Thus, nutritionally recovered and safety-assessed urine with enhanced yield performance for biomass production from algal growth will address the challenges of consumer perception of the safety of algal food products (Heinsoo [2014;](#page-28-17) Muluye et al. [2021\)](#page-30-21).

One of the biggest challenges in evolving countries is the requirement for food production which should be enough to feed the increasing population (Akpan-Idiok et al. [2012](#page-25-19)). Malnutrition poses a threat to those nations which majorly rely on agricultural food production (Benoit [2012\)](#page-26-27). Extensive drops in soil fertility caused due to the use of commercial chemical fertilizers in several areas have resulted in the deterioration in organic matter of the soil and hence a decreased crop yield. Conventional agriculture practices have become more problematic due to the rise in fertilizer prices for farmers in evolving countries. Human urine is a favorable product to be used in agricultural activities as it contains comparatively high amounts of nitrogen, phosphorus, and additional nutrients including potassium and micronutrients, and therefore can be used as a multi-constituent

Fig. 8 Human urine comprises water and a diversity of organic and inorganic compounds as compared to solid inorganic fertilizers and is a vital source of liquid nitrogen, and thus can be considered an inexpensive and good fertilizer system

Food production by using human urine as fertilizer	Locations	References
<i>Hordeum vulgare</i> (barley)	Germany	Mugivhisa and Olowoyo (2015)
<i>Beta vulgaris</i> (Swiss chard)	Ethiopia	Mugivhisa and Olowoyo (2015)
<i>Brassica oleracea</i> (head cabbage)	Kotebe Metropolitan University, Ethiopia	Alemayehu et al. (2020)
<i>Phaseolus vulgaris</i> (snap beans)	South Africa	Pandorf et al. (2018)
<i>Brassica rapa</i> (Turnip)	South Africa	Pandorf et al. (2018)
<i>Brassica oleracea</i> (Cauliflower)	Nepal	Kc and Shinjo (2020)
Abelmoschus esculentus (Lady finger)	South Eastern	
Nigeria,	Akpan-Idiok et al. (2012)	
Cicer arietinum (chick pea)	Comparative analysis	Ganesapillai et al. (2016)
Triticum (Wheat)	Comparative analysis	Kishor et al. (2020)
Vegetables	Zimbabwe	Parwada et al. (2021)
Conventional practice for production of food production	China	Ferguson (2014)

Table 6 Observing fertilizing effects of human urine higher than that of mineral fertilizers

inorganic fertilizer. In fresh urine, nitrogen found as urea is useful for plants and it is also found in marketable fertilizers. Urine also comprises water and a diversity of organic and inorganic compounds in comparison to solid inorganic fertilizers (Fig. [8](#page-20-0)). Human urine is a quick and cheap source of enough liquid nitrogen. The fertilizing efects of human urine are higher than that of mineral fertilizers which were observed in many diferent countries as shown in Table [6](#page-21-0).

In terms of the legal use of urine as fertilizer, legislation of the European Union does not prohibit or permit the usage of human urine as a nutrient fertilizer. Previous studies show that the application of urine as fertilizer for the cultivation of food was an ancient practice in China and was well accepted. In Southern China and Vietnam, farmers used human urine as a "valuable fertilizer." Conventionally human urine was a good alternative to costly inorganic fertilizers to enhance production in home gardens and agriculture. In many other countries, human urine is applied as a nutrient supply for agricultural drives including Mexico, USA, Germany, Denmark, Sweden, and some African countries. There is less information on the usage of human urine as fertilizer in South Africa due to the lack of urine diversion toilets that permit the sanitary separation of urine. The new implementation of urine diversion toilets in South Africa must enhance the usage of urine as fertilizer for agricultural practices (Lamichhane et al. [2013](#page-29-24)). The application of human urine for agricultural practices is still considered a foreign concept and normally is not accepted since human urine is considered a waste product and unhealthy (Benoit [2012](#page-26-27)). Personal preconceptions and moral concerns have additional efects to cause a reduction in the approval of using human urine as a fertilizer. Perceptions and approaches concerning the health risks and peoples' repulsion from human urine difer between diferent cultures and even within particular cultures universally. The common attitudes of people concerning the usage of human urine depend on many social factors linked to the use of human urine as fertilizer, such as, employment, culture, age, locality, education, religion, sex, status, class, and martial status.

In a research study, the approaches of workers and students at the Medunsa Campus of Limpopo University were questioned and discussed the implementation of human urine in the farming and production of crop plants was evaluated (Mugivhisa and Olowoyo [2015\)](#page-30-22). The purpose was to assess exactly how frequently people eat cultivated fresh vegetables, fruits, and other crops, how much knowledge and awareness they had about various types of fertilizers including manure, compost, and chemical, and which kind of fertilizers they chose for cultivating their food crops. Further objectives were to evaluate how much knowledge people had about the nutritional importance of human urine and what would they like to eat, maize or spinach, enriched with animal urine or human urine. The major concern of this evaluation was to check the willingness of people to modify their approaches after they become well aware of the concerns, they had for refusing to eat those food crops which were enriched with human urine (Mugivhisa and Olowoyo [2015](#page-30-22)). In a study of Swiss farmers, 57% of farmers believed that urine can be used as an agricultural fertilizer and 42% of farmers were inclined toward purchasing products enriched by urine (Lienert et al. [2003](#page-30-23); Lamichhane et al. [2012\)](#page-29-25).

Consumer acceptance and legal barriers for use of algae as food

It was observed that the foremost challenge for the consumption of microalgae as food products is sensory acceptance, generally concerning appearance and flavor for greater concentrations. Some species of microalgae can also act as smart elements, such as smart favor or smart color, which means that they can impart the flavor or color besides increasing nutritional value, and subsidize environmental and technological characteristics in more or fewer cases (Ashaolu and Ashaolu [2020\)](#page-25-21), though they are worthy for the consumers, the manufacturer and the globe.

Normally microalgae have nasty favors, such as moldy, grassy, earthy, or fshy, which establish major hurdles for usage in food products. These unwanted tastes are mostly created by volatile organic compounds, comprising phenolic, hydrocarbons, ketones, lactones, alcohols, aldehydes, and esters. The quantity and confguration of volatile organic compounds might change according to the species of microalgae, growth stage and cultivation circumstances (Fu et al. [2021](#page-27-20)). A study was done in Europe to explore consumers' acceptance of three altered microalgae-based food products: sushi, pasta, and jerky, all incorporated with *Arthrospira platensis* to enhance the protein content. They were served to 1035 people from 3 countries: France, Germany, and Netherlands, who would taste and categorize the food products according to sensory parameters. It was observed that Pasta was the utmost desired one, maybe due to the awareness of this type, but there were no substantial variances among the three countries examined (Grahl et al. [2018](#page-28-18)). It was concluded from this study that eating practices and knowledge about food are signifcant aspects and can afect consumer selections. So, customary food products might be possible mediums for microalgae integration. In the above study, all three examining countries were European, which perceives that results can be diferent if this study was done in Asian countries, for example, Sushi is a customary food in Japan and can be chosen in its place of pasta by Japanese customers.

In another study, smoothies were developed from fresh fruits and vegetables without heating, also added *Chlorella vulgaris* and *Dunaliela salina* microalgae*.* It was concluded that *D. salina* presented a superior effect both, on values of sensory parameters and antioxidant activity (Sahin and Ozturk [2021\)](#page-32-24). It shows that the amalgamation of diverse colors of vegetables and fruits possibly will mask the colors from microalgae and increase consumer acceptance of algaebased food products. In another case, masking of appearance and color, broccoli soups were formed by added with *Chlorella* sp., *Spirulina* sp., or *Tetraselmis* sp. and got nicely accepted food products with greater contents of antioxidant and polyphenols.

Two types of microalgae *Dunaliella* (*D. salina*) and *Spirulina* (*A. platensis*) are mostly cultured and consumed all over the world. One experiment was performed in which both types of microalgae were added in cookies, to enhance the increasing hardness and dark color of cookies. After examining, it was observed that cookies with *Spirulina* (*A. platensis*) were not chosen by panelists as compared to traditional cookies products, but *Dunaliella* (*D. salina*), had reasonable sensory acceptance even with its great quantity of antioxidant and phenolic compounds (Sahin [2020\)](#page-32-25). A survey of French consumers gives the basic causes that might avert consumers from buying algae-based foodstufs, known as a lack of consumer awareness about consuming microalgae as food, their inaccessibility in grocery stores, unpleasant taste and appearance, and hesitation of consumers (Lucas et al. [2019](#page-30-24)). Except these environmental apprehensions are relatively diverse among countries; for example, although Spanish consumers contemplate microalgae as environmentally friendly, the French have oppositely dual opinions and the Belgians do not perceive this as an encouragement to inspire microalgae consumption (Lafarga et al. [2021](#page-29-28); Moons et al. [2018;](#page-30-25) Thomas et al. [2020\)](#page-32-26). Interestingly, a survey in Germany reported that they wanted microalgae as a global sustainability biomass, but on the other hand, their preference was based on their total health, well-being and the intake of a balanced diet (Roßmann and Rösch [2020](#page-32-27)).

It was observed that consumers' perception of algae consumption had changed in the last two decades due to continuous research and the growing cultivation of algae in the European market (Enzing et al. [2014](#page-27-21); Rumin et al. [2020](#page-32-28); Ricci et al. [2018\)](#page-31-26). To overturn this inclination, research demonstrates that it is critical to abolish consumer reluctance toward exasperating novel food products and to emphasize on diferentiation of products by applying quality status labels and certifcations (Lucas et al. [2019\)](#page-30-24). The panel of the European Food Safety Authority ponders that no apprehension has been found about the noxiousness of this novel microalgae-based food, and it is harmless for use in nutritional supplements at the concentrated consumption limit of 1 g docosahexaenoic acid per day for adults, except lactating and pregnant women (EFSA [2020\)](#page-27-22). *Chlorella* and Spirulina are available in the Brazilian market as a type of food supplement. This type includes minerals, vitamins, bioactive substances and probiotics. These food supplements are used for athletes, pregnant women and particular for the pledge medications. After the booklet of RDC 240/2018 (Collegiate Board Resolution), simply those supplements which contained probiotics or enzymes need to be registered with "The Brazilian Health Surveillance Agency" (BRASIL [2018](#page-26-28)).

The Dietary Supplements Information Expert Committee in the USA assessed the safety of nutritional supplements and their ingredients and has acknowledged Spirulina algae species in The United States Pharmacopeia Resolution— National Formulary (Fu et al. [2021;](#page-27-20) García et al. [2017](#page-27-23)). According to Regulation (EU) No. 2015/2283, the European Union, with the "European food safety authority," confrms that new food and its ingredients go through a safety evaluation to care for public health. The European list of new foods are *Tetraselmis chui, Haematococcus pluvialis, Schizochytrium* sp., *Odontella aurita, and Ulkenia* sp. (EFSA [2020;](#page-27-22) Niccolai et al. [2019](#page-31-27); EU [2018,](#page-27-24) [2015](#page-27-25)). Furthermore,

Europe accredited the usage of β-carotene derivative of Dunaliella salina by means of a food stabilizer (Hachicha et al. [2022](#page-28-19); Koutsoumanis et al. [2020](#page-29-29)).

In the USA, the safety of algae food products is considered under the law of "Food and Drug Administration," which gives the standing of food presence "generally recognized as safe" to any food matter which is well thought-out to be safe for human ingestion (García et al. [2017](#page-27-23)). Canada has parallel principles to those of the USA for novel food assessment; the "Health Canada" organization supervises food safety regulations and refers to "novel foods" as any food product which is innovative or has been reformed as compared to customary food (Government of Canada [2021](#page-27-26); Health Canada [2021](#page-28-20)).

Legal situation and legal hurdles regarding the use of urine

Microalgae are recognized as a potential source for the removal of pollutants from wastewater discharges (Gupta et al. [2017a](#page-28-21)). Treatment of wastewater through the cultivation of microalgae into it is a cost-efective and efective procedure without contributing infectious constituents (Gupta et al. [2015](#page-28-22), [2016;](#page-28-23) Ansari et al. [2017\)](#page-25-22). Despite these benefts, there are several challenges, which involve the cost and efectiveness of microalgae harvesting and scaling up the production of bioenergy through microalgae from the laboratory level to the commercial level. Furthermore, the cultivation of microalgae in an ongoing wastewater treatment plant also poses many technical and economic issues that require observation, including culture instability, the requirement of land for cultivation, climatic aspects, cost of production, and analysis of the life cycle (Gupta et al. [2017b](#page-28-24)). Some of the hindrances involve the existence of growth-inhibiting factors which can be biotic (algal pathogens like bacteria, viruses, and fungi, inborn photosynthetic microbes, zooplankton, phytoplankton, and grazers), and abiotic factors (light, pH, temperature, salinity, carbon dioxide, nitrogen oxides, sulfur oxides, oxygen, ammonia, and heavy metals). These factors have the strongest infuence on algae growth in wastewater (Hwang et al. [2016](#page-28-25); Posadas et al. [2017](#page-31-16)). The level of such nutrients in wastewater, particularly those from agriculture and municipal sources, can constrain the growth of microalgae; thus, in certain cases, diluting the nutrient system is recommended. Further research is required to understand how to adjust the level of nutrients in wastewater for the efficient growth of microalgae (Acien et al. 2012).

Environmental factors like intensity, light, and temperature are important for the procedure and cost efficacy of algal treatment methods (Mohsenpour et al. [2021\)](#page-30-26). Thus, the site of the algal-based wastewater recovery plant is also considered. Climatic changes in day span and light concentrations are site specifc and have a signifcant role in algae species growth (Ferro et al. [2020](#page-27-27); Posadas et al. [2017\)](#page-31-16). The usage of site-specifc local algal species is benefcial in developing wastewater recovery plants due to their integral suitability to precise environmental situations, which signifcantly enhances maximum efficiency as less energy is required for recovery processes. Studies on algal species from Mohsen pour Finland (Jamsa et al. [2017;](#page-28-26) Lynch et al. [2015\)](#page-30-27), Sweden (Ferro et al. [2018;](#page-27-28) Lindberg et al. [2021\)](#page-30-28), and Canada (Abdelaziz et al. [2014\)](#page-25-24) have been used for their capability to treat municipal wastewater in cold weather.

One of the main hurdles for the growth of microalgae in the treatment of wastewater is the harvesting of algal biomass because harvesting normally causes more than thirty percent of the overall production charge and can be changed according to the type of harvesting method and the properties of selected species of microalgae and the desired product (Barros et al. [2015\)](#page-26-29). Mostly used harvesting processes are founded on biological, chemical, mechanical, and electrical methods including fltration, centrifugation, gravity sedimentation, and fotation (Kadir et al. [2018](#page-29-30)). However, some of these methods are costly and not eco-friendly like chemical-based methods (Lam and Lee [2012](#page-29-25)). Filtration is a simple and cost-efective method for harvesting microalgae. The major beneft of this method is that the end product of biomass has no chemical impurities, and this can be specifcally applied for the recovery of remaining nutrients (Ahmad et al. [2012\)](#page-25-25).

Urine is one of the most plentiful wastes on earth that can develop the ''carbon substance of the future'' (Chaudhari et al. [2014](#page-26-30)). In present studies, human urine is used to yield novel carbon, known as ''Urine Carbon'' for existing and upcoming energy-required applications. It was noticed from the experiment that urea is the main component of urine, which is an important carbon-based source of hydrogen, carbon, oxygen, and nitrogen. Meanwhile, human urine comprises major elements including urea, sodium, chloride, creatinine, potassium, and other organic and inorganic ions and composites, and the urine is supposed to produce a compound comprising of carbon and a combination of salts after carbonization (Lu et al. [2013](#page-30-29)). Silicon and phosphorus compounds are also excluded from urine from the human body after many metabolic processes. Therefore, the existence of these components makes urine such a vigorous antecedent for the creation of uniquely doped carbon. Though despite being recycled to provide hydrogen through urea also worked for the nourishment of microbial energy cells (Ieropoulos et al. [2012](#page-28-27); Kuntke et al. [2012](#page-29-11)), still, no work is reported where human liquid waste like urine was used as an antecedent to yield carbon or some other constituents.

Sensible usage of urine as per carbon antecedent will not simply improve cost-efectiveness in the growth of purposeful materials but also assist to decrease the ecological burden

by inhibiting the potassium, nitrogen, and rich phosphate urine contaminants from reaching the water bodies, therefore reducing the oxygen-depriving eutrophication, development of algal blooms, and development of marine ''dead zones'' in oceans and rivers. Furthermore, the human body does not digest pharmaceutical medicines entirely and ejects some vital or in a combined form besides further organic impurities in urine. These composites are carried by means of domestic waste watercourses to municipal sewage wastewater management plants or, in some circumstances, directly reach waterbodies without treatment, posing many ecological distresses. In this discovery, transforming urine to carbon will not merely increase the abundance of directly above ecological concerns, but also help in evolving valued tools for several mechanical applications. Some of these applications include the development of involving supercapacitors, fuel cells, Li-ion batteries, and many others (Zhuang et al. [2018\)](#page-34-9), which can make this technique technically, fnancially, and environmentally encouraging (Chaudhari et al. [2014](#page-26-30)). Some benefts of human urine conversion to carbon are given in Table [7.](#page-24-0)

Conclusion

Although synthetic media has been employed to support the growth and cultivation of microalgae commercially, there is, however, a growing potential for low-cost alternative nutrients to be sourced from urine for the commercial production of microalgae as they are of economic value. Valorization of urine from wastewater for secondary applications such as microalgae cultivation has added value and enhanced natural resource efficiency. Interestingly, there are several ecological principles or environmental concerns that have been linked to large-scale microalgae production via wastewater treatment. Some of these concerns are focused on how environmentally sustainable utilizing urine from wastewater treatment for large-scale microalgae cultivation is. This stems from the fact that microalgae production with urine could lead to toxic atmospheric emissions such as the release of methane and nitrous oxide gases into the environment leading to the depletion of the ozone. Also, these atmospheric emissions can be highly signifcant, especially for open system cultivation facilities than closed systems whereby emissions could be trapped by photobioreactors. In addition, a poor control system or its failure during waste urine treatment could result in a major potential environmental problem if microalgae nutrients are released which could result in a serious eutrophication crisis for the aquatic ecosystem. Another point for future evaluation should be focused on the toxicological impact of the biomass produced from microalgae cultivated with waste urine. Biomass sourced from microalgae for food and pharmaceutical applications should be devoid of traces of toxic elements, and as such a regulatory and environmental policy could support consumer interest in products sourced from such microalgae. Consequently, awareness creation for stakeholders and the public about the broad benefts of using microalgae cultivated from treated wastewater could enhance the setting up of more large-scale facilities for the production of microalgae, as this is a costefective alternative nutrient recovery process.

Acknowledgements We confrm the fnal authorship for this manuscript, and we ensure that anyone else who contributed to the manuscript but does not qualify for authorship has been acknowledged with their permission. We acknowledge that all listed authors have made a signifcant scientifc contribution to the research in the manuscript approved its claims and agreed to be an author.

Author contribution SAS involved in conceptualization, methodology, validation, formal analysis, investigation, resources, data curation, writing—original draft, writing—review & editing, visualization, supervision, project administration. YSW took part in writing—review & editing. TS took part in writing—original draft. İU involved in writing—original draft. MA took part in writing—original draft. MAS involved in resources. RDA took part in writing—original draft, writing—review & editing.

Funding This manuscript has not received any funding.

Data availability statement The data that support the fndings of this study are available from the corresponding author, upon request.

Declarations

Conflict of interest No potential confict of interest relevant to this article was reported.

Ethical approval The study does not involve any human or animal testing.

References

- Abdelaziz AE, Leite GB, Belhaj MA, Hallenbeck PC (2014) Screening microalgae native to Quebec for wastewater treatment and biodiesel production. Bioresour Technol 157:140–148. [https://](https://doi.org/10.1016/j.biortech.2014.01.114) doi.org/10.1016/j.biortech.2014.01.114
- Abdelaziz AE, Leite GB, Hallenbeck PC (2013) Addressing the challenges for sustainable production of algal biofuels Harvesting and conversion to biofuels. Environ Technol 34:1807–1836
- Abu Sepian NR, Mat Yasin NH, Zainol N, Rushan NH, Ahmad AL (2019) Fatty acid profle from immobilized Chlorella vulgaris cells in diferent matrices. Environ Technol 40(9):1110–1117. <https://doi.org/10.1080/09593330.2017.1408691>
- Acien FG, Fernandez JM, Magán JJ, Molina E (2012) Production cost of a real microalgae production plant and strategies to reduce it. Biotechnol Adv 30(6):1344–1353. [https://doi.org/](https://doi.org/10.1016/j.biotechadv.2012.02.005) [10.1016/j.biotechadv.2012.02.005](https://doi.org/10.1016/j.biotechadv.2012.02.005)
- Acién Fernández FG, Fernández Sevilla JM, Molina Grima E (2013) Photobioreactors for the production of microalgae. Rev. Environ. Sci Bio/Technology 12:131–151. [https://doi.org/10.1007/](https://doi.org/10.1007/s11157-012-9307-6) [s11157-012-9307-6](https://doi.org/10.1007/s11157-012-9307-6)
- Adarme-Vega TC, Lim DKY, Timmins M, Vernen F, Li Y, Schenk PM (2012) Microalgal biofactories: a promising approach towards sustainable omega-3 fatty acid production. Microb Cell Factories 11(1):1–10.<https://doi.org/10.1186/1475-2859-11-96>
- Ahmad AL, Yasin NM, Derek CJC, Lim JK (2012) Crossfow microfltration of microalgae biomass for biofuel production. Desalination 302:65–70.<https://doi.org/10.1016/j.desal.2012.06.026>
- Ahmad AL, Yasin NM, Derek CJC, Lim JK (2014) Comparison of harvesting methods for microalgae Chlorella sp and its potential use as a biodiesel feedstock. Environ Technol 35(17):2244–2253
- Ahmad S, Pandey A, Pathak VV, Tyagi VV, Kothari R (2020) Phycoremediation: algae as eco-friendly tools for the removal of heavy metals from wastewaters. Bioremed Industr Waste Environ Safety. https://doi.org/10.1007/978-981-13-3426-9_3
- Ahmed J, Thakur A, Goyal A (2021) Industrial wastewater and its toxic efects. 1-14 [https://doi.org/10.1039/9781839165399-00001](10.1039/9781839165399-00001)
- Ahsan, M., Habib, B., Parvin, M., Huntington, T. C., & Hasan, M. R. (2008). A review on culture, production and use of spirulina as

food for humans and feeds for domestic animals. [https://www.](https://www.fao.org/3/i0424e/i0424e00.pdf) [fao.org/3/i0424e/i0424e00.pdf](https://www.fao.org/3/i0424e/i0424e00.pdf)

- Akizuki S, Cuevas-Rodríguez G, Toda T (2021) Nitrifcation of anaerobic digestate using a consortium of microalgae and nitrifers in an open photobioreactor with moving bed carriers. Chemosphere 263:127948. <https://doi.org/10.1016/j.chemosphere.2020.127948>
- Akpan-Idiok AU, Udo IA, Braide IE (2012) The use of human urine as an organic fertilizer in the production of okra (Abelmoschus esculentus) In South Eastern Nigeria. Resour Conserv Recycl 62:14–20.<https://doi.org/10.1016/j.resconrec.2012.02.003>
- Alagawany M, Taha AE, Noreldin A, El-Tarabily KA, El-hack MEA (2021) Nutritional applications of species of Spirulina and Chlorella in farmed fsh: a review. Aquaculture 542:736841. [https://](https://doi.org/10.1016/j.aquaculture.2021.736841) doi.org/10.1016/j.aquaculture.2021.736841
- Alemayehu YA, Demoz AA, Degefu MA, Gebreeyessus GD, Demessie SF (2020) Effect of human urine application on cabbage production and soil characteristics. J Water Sanit Hyg Dev 10(2):262– 275. <https://doi.org/10.2166/washdev.2020.136>
- Ali S, Peter AP, Chew KW, Munawaroh HSH, Show PL (2021) Resource recovery from industrial effluents through the cultivation of microalgae: A review. Bioresour Technol 337:125461. <https://doi.org/10.1016/j.biortech.2021.125461>
- Al-Jabri H, Das P, Khan S, Thaher M, AbdulQuadir M (2020) Treatment of sewage by microalgae and the potential applications of the produced biomass—A review. Water 13(1):27. [https://doi.](https://doi.org/10.3390/w13010027) [org/10.3390/w13010027](https://doi.org/10.3390/w13010027)
- Amaral MS, Loures CC, Naves FL, Samanamud GL, Silva MB, Prata AM (2020) Microalgae cultivation in photobioreactors aiming at biodiesel production. In *Biotechnological* Applications of Biomass. Intech Open.<https://doi.org/10.5772/intechopen.93547>
- Andrade LM, Andrade CJ, Dias M, Nascimento CAO (2018) Chlorella and Spirulina microalgae as sources of functional foods, nutraceuticals, and food supplements; an overview. MOJ Food Process Technol 6(1):45–58. [https://doi.org/10.15406/mojfpt.](10.15406/mojfpt.2018.06.00144) [2018.06.00144](10.15406/mojfpt.2018.06.00144)
- Ansari FA, Wahal S, Gupta SK, Rawat I, Bux F (2017) A comparative study on biochemical methane potential of algal substrates: implications of biomass pre-treatment and product extraction. Bioresour Technol 234:320–326. [https://doi.org/10.1016/j.biort](https://doi.org/10.1016/j.biortech.2017.03.068) [ech.2017.03.068](https://doi.org/10.1016/j.biortech.2017.03.068)
- Arashiro LT, Montero N, Ferrer I, Acien FG, Gomez C, Garf M (2018) Life cycle assessment of high rate algal ponds for wastewater treatment and resource recovery. Sci Total Environ 622:1118– 1130.<https://doi.org/10.1016/j.scitotenv.2017.12.051>
- Araújo R, Vázquez Calderón F, Sánchez López J, Azevedo IC, Bruhn A, Fluch S, Garcia TM, Ghaderiardakani F, Ilmjärv T, Laurans M, Mac MM, Mangini S, Peteiro C, Rebours C, Stefansson T, Ullmann J (2021) Current status of the algae production industry in Europe: an emerging sector of the blue bioeconomy. Front Mar Sci 7:626389. <https://doi.org/10.3389/fmars.2020.626389>
- Arias MA, Arnold U, Goldbach H (2019) Change in estrogenic activity in stored human urine before reuse as fertilizer. Int J Recycl Org Waste Agric 8(1):195–202. [https://doi.org/10.1007/](https://doi.org/10.1007/s40093-019-0289-z) [s40093-019-0289-z](https://doi.org/10.1007/s40093-019-0289-z)
- Ashaolu TJ, Ashaolu JO (2020) Perspectives on the trends, challenges and benefts of green, smart and organic (GSO) foods. Int J Gastron Food Sci 22:100273. [https://doi.org/10.1016/j.ijgfs.2020.](https://doi.org/10.1016/j.ijgfs.2020.100273) [100273](https://doi.org/10.1016/j.ijgfs.2020.100273)
- Babatsouli P, Fodelianakis S, Paranychianakis N, Venieri D, Dialynas M, Kalogerakis N (2015) Single stage treatment of saline wastewater with marine bacterial–microalgae consortia in a fxed-bed photobioreactor. J Hazard Mater 292:155–163. [https://doi.org/](https://doi.org/10.1016/j.jhazmat.2015.02.060) [10.1016/j.jhazmat.2015.02.060](https://doi.org/10.1016/j.jhazmat.2015.02.060)
- Bacsi I, Viktoria B, Kokai Z, Gonda S, Novak Z, Nagy SA, Vasas G (2016) Efects of non-steroidal anti-infammatory drugs on cyanobacteria and algae in laboratory strains and in natural algal
- Badeti U, Jiang J, Almuntashiri A, Pathak N, Dorji U, Volpin F, Freguia S, Ang WL, Chanan A, Kumarasingham S, Shon HK, Phuntsho S (2022) Impact of source-separation of urine on treatment capacity, process design, and capital expenditure of a decentralized wastewater treatment plant. Chemosphere 300:134489. [https://](https://doi.org/10.1016/j.chemosphere.2022.134489) doi.org/10.1016/j.chemosphere.2022.134489
- Bai X, Acharya K (2017) Algae-mediated removal of selected pharmaceutical and personal care products (PPCPs) from Lake Mead water. Sci Total Environ 581:734–740. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.scitotenv.2016.12.192) [scitotenv.2016.12.192](https://doi.org/10.1016/j.scitotenv.2016.12.192)
- Barros AI, Gonçalves AL, Simoes M, Pires JC (2015) Harvesting techniques applied to microalgae: a review. Renew Sustain Energy Rev 41:1489–1500.<https://doi.org/10.1016/j.rser.2014.09.037>
- Behera B, Patra S, Balasubramanian P (2020) Biological nutrient recovery from human urine by enriching mixed microalgal consortium for biodiesel production. J Environ Manag 260:110111. <https://doi.org/10.1016/j.jenvman.2020.110111>
- Benner P, Meier L, Pfefer A, Krüger K, Oropeza Vargas JE, Weuster-Botz D (2022) Lab-scale photobioreactor systems: principles, applications, and scalability. Bioprocess Biosyst Eng. [https://doi.](https://doi.org/10.1007/s00449-022-02711-1) [org/10.1007/s00449-022-02711-1](https://doi.org/10.1007/s00449-022-02711-1)
- Benoit N (2012) Individual's perception and the potential of urine as a fertilizer in eThekwini, South Africa (Doctoral dissertation).
- Bernaerts TMM, Gheysen L, Foubert I, Hendrickx ME, Van Loey AM (2019) The potential of microalgae and their biopolymers as structuring ingredients in food: A review. Biotechnol Adv 37(8):107419.<https://doi.org/10.1016/j.biotechadv.2019.107419>
- Bhatt P, Gangola S, Bhandari G, Zhang W, Maithani D, Mishra S, Chen S (2021) New insights into the degradation of synthetic pollutants in contaminated environments. Chemosphere 268:128827. <https://doi.org/10.1016/j.chemosphere>
- Bilal M, Bagheri AR, Bhatt P, Chen S (2021) Environmental occurrence, toxicity concerns, and remediation of recalcitrant nitroaromatic compounds. J Environ Manage 291:112685. [https://doi.](https://doi.org/10.1016/j.jenvman.2021.112685) [org/10.1016/j.jenvman.2021.112685](https://doi.org/10.1016/j.jenvman.2021.112685)
- Brasil (2018) RESOLUÇÃO DA DIRETORIA COLEGIADA – RDC N° 240, DE 26 DE JULHO DE 2018. Altera a Resolução – RDC n° 27, de 6 de agosto de 2010, que dispõe sobre as categorias de alimentos e embalagens isentos e com obrigatoriedade de registro sanitário. <https://bvsms.saude.gov.br/bvs/saudelegis/> anvisa/2018/rdc0240_26_07_2018.pdf (accessed 13 March 2021).
- Bustillo-Lecompte C, Mehrvar M (2017) Slaughterhouse wastewater: treatment, management and resource recovery. Physico-chemical Wastewater Treat Resour Recover. <https://doi.org/10.5772/65499>
- Cai T, Park SY, Li Y (2013) Nutrient recovery from wastewater streams by microalgae: status and prospects. Renew Sustain Energy Rev 19:360–369. <https://doi.org/10.1016/j.rser.2012.11.030>
- Caporgno MP, Mathys A (2018) Trends in microalgae incorporation into innovative food products with potential health benefts. Front nutr.<https://doi.org/10.3389/fnut.2018.00058>
- Cezare-Gomes EA, Mejia-da-Silva L, del C, P´erez-Mora LS, Matsudo MC, Ferreira-Camargo LS, Sing AK, Carvalho JCM (2019). Potential of microalgae carotenoids for industrial application. Appl Biochem Biotechnol 188(3): 602–634 [https://doi.org/10.](https://doi.org/10.1007/s12010-018-02945-4) [1007/s12010-018-02945-4](https://doi.org/10.1007/s12010-018-02945-4)
- Chang JS, Show PL, Ling TC, Chen CY, Ho SH, Tan CH, Nagarajan D, Phong WN (2017) Photobioreactors. In: Current developments in biotechnology and bioengineering. Elsevier, pp 313–352 [https://](https://doi.org/10.1016/B978-0-444-63663-8.00011-2) doi.org/10.1016/B978-0-444-63663-8.00011-2
- Chang Y, Wu Z, Bian L, Feng D, Leung DY (2013) Cultivation of Spirulina platensis for biomass production and nutrient removal from synthetic human urine. Appl Energy 102:427–431. [https://](https://doi.org/10.1016/j.apenergy.2012.07.024) doi.org/10.1016/j.apenergy.2012.07.024
- Chanquia SN, Vernet G, Kara S (2022) Photobioreactors for cultivation and synthesis: Specifcations, challenges, and perspectives. Eng Life Sci 22(12):712–724.<https://doi.org/10.1002/elsc.202100070>
- Charles CN, Msagati T, Swai H, Chacha M (2019) Microalgae: an alternative natural source of bioavailable omega-3 DHA for promotion of mental health in East Africa. Sci Afr. [https://doi.org/](https://doi.org/10.1016/j.sciaf.2019.e00187) [10.1016/j.sciaf.2019.e00187](https://doi.org/10.1016/j.sciaf.2019.e00187)
- Chatterjee P, Granatier M, Ramasamy P, Kokko M, Lakaniemi AM, Rintala J (2019) Microalgae grow on source separated human urine in Nordic climate: Outdoor pilot-scale cultivation. J Environ Manage 237:119–127. [https://doi.org/10.1016/j.jenvman.](https://doi.org/10.1016/j.jenvman.2019.02.074) [2019.02.074](https://doi.org/10.1016/j.jenvman.2019.02.074)
- Chaudhari NK, Song MY, Yu JS (2014) Heteroatom-doped highly porous carbon from human urine. Scie Rep 4(1):1–10. [https://](https://doi.org/10.1038/srep05221) doi.org/10.1038/srep05221
- Cheirsilp B, Thawechai T, Prasertsan P (2017) Immobilized oleaginous microalgae for production of lipid and phytoremediation of secondary effluent from palm oil mill in fluidized bed photobioreactor. Bioresour Technol 241:787–794. [https://doi.org/10.](https://doi.org/10.1016/j.biortech.2017.06.016) [1016/j.biortech.2017.06.016](https://doi.org/10.1016/j.biortech.2017.06.016)
- Chowdury KH, Nahar N, Deb UK (2020) The growth factors involved in microalgae cultivation for biofuel production: a Review. CWEEE 9(4):185–215. [https://doi.org/10.4236/cweee.2020.](https://doi.org/10.4236/cweee.2020.94012) [94012](https://doi.org/10.4236/cweee.2020.94012)
- Chrispim MC, Tarpeh WA, Salinas DT, Nolasco MA (2017) The sanitation and urban agriculture nexus: urine collection and application as fertilizer in São Paulo. Brazil J Water Sanit Hyg Dev 7(3):455–465. <https://doi.org/10.2166/washdev.2017.163>
- Collos Y, Harrison PJ (2014) Acclimation and toxicity of high ammonium concentrations to unicellular algae. Mar Pollut Bull 80(1– 2):8–23. <https://doi.org/10.1016/j.marpolbul.2014.01.006>
- Coppens J, Lindeboom R, Muys M, Coessens W, Alloul A, Meerbergen K, Lievens B, Caluwaert P, Boon N, Vlaeminck SE (2016) Nitrifcation and microalgae cultivation for two-stage biological nutrient valorization from source separated urine. Bioresour Technol 211:41–50. <https://doi.org/10.1016/j.biortech.2016.03.001>
- Craggs R, Sutherland D, Campbell H (2012) Hectare-scale demonstration of high rate algal ponds for enhanced wastewater treatment and biofuel production. J Appl Phycol 24(3):329–337. [https://](https://doi.org/10.1007/s10811-012-9810-8) doi.org/10.1007/s10811-012-9810-8
- Crini G, Lichtfouse E (2019) Advantages and disadvantages of techniques used for wastewater treatment. Environ Chem Lett 17(1):145–155.<https://doi.org/10.1007/s10311-018-0785-9>
- Cui J, Purton S, Baganz F (2022) Characterization of a simple 'hanging bag' photobioreactor for low-cost cultivation of microalgae. J Chem Technol Biotechnol 97(3):608–619. [https://doi.org/10.](https://doi.org/10.1002/jctb.6985) [1002/jctb.6985](https://doi.org/10.1002/jctb.6985)
- de Alva MS, Pabello VML, Ledesma MTO, Gómez MJC (2018) Carbon, nitrogen, and phosphorus removal, and lipid production by three saline microalgae grown in synthetic wastewater irradiated with diferent photon fuxes. Algal Res 34:97–103. [https://doi.](https://doi.org/10.1016/j.algal.2018.07.006) [org/10.1016/j.algal.2018.07.006](https://doi.org/10.1016/j.algal.2018.07.006)
- de Wilt A, Butkovskyi A, Tuantet K, Leal LH, Fernandes TV, Langenhoff A, Zeeman G (2016) Micropollutant removal in an algal treatment system fed with source separated wastewater streams. J Hazard Mater 304:84–92. [https://doi.org/10.1016/j.jhazmat.](https://doi.org/10.1016/j.jhazmat.2015.10.033) [2015.10.033](https://doi.org/10.1016/j.jhazmat.2015.10.033)
- Ding N, Li C, Wang T, Guo M, Mohsin A, Zhang S (2021) Evaluation of an enclosed air-lift photobioreactor (ALPBR) for biomass and lipid biosynthesis of microalgal cells grown under fuid-induced shear stress. Biotechnol Biotechnol Equip 35(1):139–149. [https://](https://doi.org/10.1080/13102818.2020.1856717) doi.org/10.1080/13102818.2020.1856717
- Dong X, Zhang X, Zeng S (2017) Measuring and explaining eco-efficiencies of wastewater treatment plants in China: an uncertainty analysis perspective. Water Res 112:195–207. [https://doi.org/10.](https://doi.org/10.1016/j.watres.2017.01.026) [1016/j.watres.2017.01.026](https://doi.org/10.1016/j.watres.2017.01.026)
- Dvir I, vam Moppes D, Arad SM (2021) Foodomics: To discover the health potential of microalgae. In: (Ed.) Comprehensive foodomics A Cifuentes, Elsevier, pp 658–671 [https://doi.org/10.1016/](https://doi.org/10.1016/b978-0-08-100596-5.22888-6) [b978-0-08-100596-5.22888-6.](https://doi.org/10.1016/b978-0-08-100596-5.22888-6)
- DWA (2016) New Alternative Sanitation Systems - NASS: Terminology, Material Flows Treatment of Partial Flows, Utilization. VDG Bauhaus-Universit¨ atsverlag, Weimar, Germany
- EFSA, NDA Panel (Panel on Nutrition, Novel Foods and Food Allergens) Turck D, Castenmiller J, De Henauw SH, Ferreira L, Knutsen HKEFSA (2020) Scientifc opinion on the safety of Schizochytrium sp. oil as a novel food pursuant to regulation (EU) 2015/2283. EFSA J 8(10): 6242 doi[:https://doi.org/10.2903/j.](https://doi.org/10.2903/j.efsa.2020.6242) [efsa.2020.6242,](https://doi.org/10.2903/j.efsa.2020.6242) 24
- Ejike CECC, Collins SA, Balasuriya N, Swanson AK, Mason B, Udenigwe CC (2016) Prospects of microalgae proteins in producing peptide-based functional foods for promoting cardiovascular health. Trends Food Sci Tech 59:30-36. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.tifs.2016.10.026) [tifs.2016.10.026](https://doi.org/10.1016/j.tifs.2016.10.026)
- Elaya Perumal U, Sundararaj R (2020) Algae: a potential source to prevent and cure the novel coronavirus – a review. Int J Emerg Technol 11(2):479–483
- Emparan Q, Harun R, Danquah MK (2019) Role of phycoremediation for nutrient removal from wastewaters: a review. Appl Ecol Environ Res 17(1):889–915. [https://doi.org/10.15666/aeer/1701_](https://doi.org/10.15666/aeer/1701_889915) [889915](https://doi.org/10.15666/aeer/1701_889915)
- Emparan Q, Harun R, Sing Jye Y (2021) Efficiency of pollutants removal in treated palm oil mill effluent (TPOME) using different concentrations of sodium alginate-immobilized Nannochloropsis sp. cells. Int J Phytoremediation 23(5):454–461. [https://doi.org/](https://doi.org/10.1080/15226514.2020.1825327) [10.1080/15226514.2020.1825327](https://doi.org/10.1080/15226514.2020.1825327)
- Enzing C, Ploeg M, Barbosa M. Sijtsma L (2014) Microalgae-Based Products for the Food and Feed Sector: In: (Eds) An Outlook for Europe; Vigani M, Parisi C, Cerezo ER, Publications Office of the European Union: Luxembourg
- Essner JB, Laber CH, Ravula S, Polo-Parada L, Baker GA (2016) Pee-dots: biocompatible fuorescent carbon dots derived from the upcycling of urine. Green Chem 18(1):243–250. [https://](https://doi.org/10.1039/c5gc02032h) doi.org/10.1039/c5gc02032h
- EU (2015) Regulation (EU) 2015/2283 of the European Parliament and the Council of 25 November 2015 on novel foods, amending Regulation (EU) No 1169/2011 of the European Parliament and the Council and repealing Regulation (EC) No 258/97 of the European Parliament and the Council and Commission Regulation (EC) No 1852/2001. [http://data.europa.eu/eli/reg/](http://data.europa.eu/eli/reg/2015/2283/oj) [2015/2283/oj](http://data.europa.eu/eli/reg/2015/2283/oj)
- EU (2018) Regulation (EU) 2018/1032 of 20 July 2018. Authorizing the extension of use of oil from the microalgae Schizochytrium sp. as a novel food under Regulation (EU) 2015/2283 of the European Parliament and of the Council and amending Commission Implementing Regulation (EU) 2017/2470. [http://data.](http://data.europa.eu/eli/reg_impl/2018/1032/oj) [europa.eu/eli/reg_impl/2018/1032/oj](http://data.europa.eu/eli/reg_impl/2018/1032/oj)
- Ferguson DT (2014) Night soil and the 'Great Divergence': human waste, the urban economy, and economic productivity, 1500– 1900. J Glob Hist 9(3):379–402. [https://doi.org/10.1017/S1740](https://doi.org/10.1017/S1740022814000175) [022814000175](https://doi.org/10.1017/S1740022814000175)
- Fernandez I, Acién FG, Berenguel M, Guzmán JL (2014) First principles model of a tubular photobioreactor for microalgal production. Ind Eng Chem Res 53(27):11121–11136. [https://doi.](https://doi.org/10.1021/ie501438r) [org/10.1021/ie501438r](https://doi.org/10.1021/ie501438r)
- Ferro L, Gorzsas A, Gentili FG, Funk C (2018) Subarc microalgal strains treat wastewater and produce biomass at low temperature and short photoperiod. Algal Res 35:160–167. [https://doi.](https://doi.org/10.1016/j.algal.2018.08.031) [org/10.1016/j.algal.2018.08.031](https://doi.org/10.1016/j.algal.2018.08.031)
- Ferro L, Hu YO, Gentili FG, Andersson AF, Funk C (2020) DNA metabarcoding reveals microbial community dynamics in a microalgae-based municipal wastewater treatment open

 $\circled{2}$ Springer

photobioreactor. Algal Res 51:102043. [https://doi.org/10.](https://doi.org/10.1016/j.algal.2020.102043) [1016/j.algal.2020.102043](https://doi.org/10.1016/j.algal.2020.102043)

- Fu J, Huang Y, Liao Q, Xia A, Fu Q, Zhu X (2019) Photo-bioreactor design for microalgae: a review from the aspect of CO2 transfer and conversion. Bioresour technol 292:121947. [https://doi.](https://doi.org/10.1016/j.biortech.2019.121947) [org/10.1016/j.biortech.2019.121947](https://doi.org/10.1016/j.biortech.2019.121947)
- Fu Y, Chen T, Chen SHY, Liu B, Sun P, Sun H, Chen F (2021) The potentials and challenges of using microalgae as an ingredient to produce meat analogues. Trends Food Sci Tech 112:188– 200.<https://doi.org/10.1016/j.tifs.2021.03.050>
- Gajda I, Stinchcombe A, Greenman J, Melhuish C, Ieropoulos I (2015) Ceramic MFCs with internal cathode producing suffcient power for practical applications. Int J Hydrog Energy 40(42):14627–14631. [https://doi.org/10.1016/j.ijhydene.2015.](https://doi.org/10.1016/j.ijhydene.2015.06.039) [06.039](https://doi.org/10.1016/j.ijhydene.2015.06.039)
- Gandiglio M, Lanzini A, Soto A, Leone P, Santarelli M (2017) Enhancing the energy efficiency of wastewater treatment plants through co-digestion and fuel cell systems. Front Environ Sci 5:70. <https://doi.org/10.3389/fenvs.2017.00070>
- Ganesapillai M, Simha P, Beknalkar SS, Sekhar DMR (2016) Lowgrade rock phosphate enriched human urine as novel fertilizer for sustaining and improving agricultural productivity of Cicer arietinum. Sustain Prod Consum 6:62–66. [https://doi.org/10.](https://doi.org/10.1016/j.spc.2016.01.005) [1016/j.spc.2016.01.005](https://doi.org/10.1016/j.spc.2016.01.005)
- Garbowski T, Charazinska S, Pulikowski K, Wiercik P (2020) Application of microalgae cultivated on pine bark for the treatment of municipal wastewater in cylindrical photobioreactors. Water Environ J 34:949–959. <https://doi.org/10.1111/wej.12606>
- García JL, de Vicente M, Galán B (2017) Microalgae, old sustainable food and fashion nutraceuticals. Microb Biotechnol 10:1017–1024
- García-Segovia P, Pagán-Moreno MJ, Lara IF, Martínez-Monzó J (2017) Efect of microalgae incorporation on physicochemical and textural properties in wheat bread formulation. Food Sci Technol Int 23(5):437–447. [https://doi.org/10.1177/10820](https://doi.org/10.1177/1082013217700259) [13217700259](https://doi.org/10.1177/1082013217700259)
- Gentili FG, Fick J (2017) Algal cultivation in urban wastewater: an efficient way to reduce pharmaceutical pollutants. J Appl Phycol 29(1):255–262.<https://doi.org/10.1007/s10811-016-0950-0>
- Gerdes CR, Suess TN, Anderson GA, Gent SP (2014) Investigation of hydrodynamics and heat transfer efects due to light guides in a column photobioreactor. J Fuel Cell Sci Technol 10(1115/1):4026623
- Gerotto C, Norici A, Giordano M (2020) Toward enhanced fxation of CO2 in aquatic biomass: Focus on microalgae. Front Energy Res 8:213.<https://doi.org/10.3389/fenrg.2020.00213>
- Ghayal MS, Pandya MT (2013) Microalgae biomass: a renewable source of energy. Energy Procedia 32:242–250. [https://doi.org/](https://doi.org/10.1016/j.egypro.2013.05.031) [10.1016/j.egypro.2013.05.031](https://doi.org/10.1016/j.egypro.2013.05.031)
- Giannakis S, Androulaki B, Comninellis C, Pulgarin C (2018) Wastewater and urine treatment by UVC-based advanced oxidation processes: Implications from the interactions of bacteria, viruses, and chemical contaminants. Chem Eng J 343:270–282. [https://](https://doi.org/10.1016/j.cej.2018.03.019) doi.org/10.1016/j.cej.2018.03.019
- Giannelli L, Torzillo G (2012) Hydrogen production with the microalga Chlamydomonas reinhardtii grown in a compact tubular photobioreactor immersed in a scattering light nanoparticle suspension. Int J Hydrog Energy 37(22):16951–16961. [https://doi.org/](https://doi.org/10.1016/j.ijhydene.2012.08.103) [10.1016/j.ijhydene.2012.08.103](https://doi.org/10.1016/j.ijhydene.2012.08.103)
- Gouveia L, Graça S, Sousa C, Ambrosano L, Ribeiro B, Botrel EP, Neto PC, Ferreira AF, Silva CM (2016) Microalgae biomass production using wastewater: treatment and costs: scale-up considerations. Algal Res 16:167–176. [https://doi.org/10.1016/j.algal.](https://doi.org/10.1016/j.algal.2016.03.010) [2016.03.010](https://doi.org/10.1016/j.algal.2016.03.010)
- Government of Canada (2021) About Novel and Genetically-Modifed (GM) Foods. Available online: [https://www.canada.ca/en/](https://www.canada.ca/en/health-canada/services/food-nutrition/genetically-modified-foods-other-novel-foods.html)

[health-canada/services/food-nutrition/genetically-modified](https://www.canada.ca/en/health-canada/services/food-nutrition/genetically-modified-foods-other-novel-foods.html)[foods-other-novel-foods.html](https://www.canada.ca/en/health-canada/services/food-nutrition/genetically-modified-foods-other-novel-foods.html) (accessed on 20 April 2021).

- Grahl S, Strack M, Weinrich R, Morlein D (2018) Consumer-oriented product development: the conceptualization of novel food products based on Spirulina (Arthrospira platensis) and resulting consumer expectations. J Food Qual. [https://doi.org/10.1155/](https://doi.org/10.1155/2018/1919482) [2018/1919482](https://doi.org/10.1155/2018/1919482)
- Grossmann L, Hinrichs J, Weiss J (2019) Cultivation and downstream processing of microalgae and cyanobacteria to generate proteinbased techno functional food ingredients. Crit Rev Food Sci Nutr 60(17):2961–2989. [https://doi.org/10.1080/10408398.2019.](https://doi.org/10.1080/10408398.2019.1672137) [1672137](https://doi.org/10.1080/10408398.2019.1672137)
- Guo X, Yao L, Huang Q (2015) Aeration and mass transfer optimization in a rectangular airlift loop photobioreactor for the production of microalgae. Bioresour Technol 190:189–195. [https://doi.](https://doi.org/10.1016/j.biortech.2015.04.077) [org/10.1016/j.biortech.2015.04.077](https://doi.org/10.1016/j.biortech.2015.04.077)
- Gupta SK, Ansari FA, Bauddh K, Singh B, Nema AK, Pant KK (2017a) Harvesting of microalgae for biofuels: comprehensive performance evaluation of natural, inorganic, and synthetic focculants. In: (Ed.) Green Technologies and Environmental Sustainability, Cham. Springer, pp 131–156 DOI: [https://doi.org/10.](https://doi.org/10.1007/978-3-319-50654-8_6) [1007/978-3-319-50654-8_6](https://doi.org/10.1007/978-3-319-50654-8_6)
- Gupta SK, Ansari FA, Shriwastav A, Sahoo NK, Rawat I, Bux F (2016) Dual role of Chlorella sorokiniana and Scenedesmus obliquus for comprehensive wastewater treatment and biomass production for bio-fuels. J Clean Prod 115:255–264. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jclepro.2015.12.040) [jclepro.2015.12.040](https://doi.org/10.1016/j.jclepro.2015.12.040)
- Gupta SK, Shriwastav A, Kumari S, Ansari FA, Malik A, Bux F (2015) Phycoremediation of emerging contaminants. In: (Ed.), Algae and environmental sustainability. Springer, New Delhi, pp 129– 146 DOI <https://doi.org/10.1007/978-81-322-2641-3>
- Gupta SK, Sriwastav A, Ansari FA, Nasr M, Nema AK (2017b) Phycoremediation: an eco-friendly algal technology for bioremediation and bioenergy production. In: (Ed.) Phytoremeation potential of bioenergy plants. Springer, Singapore, pp 431–456 DOI: https://doi.org/10.1007/978-981-10-3084-0_18
- Ha PT, Lee TK, Rittmann BE, Park J, Chang IS (2012) Treatment of alcohol distillery Sewage using a Bacteroidetes-dominant thermophilic microbial fuel cell. J Environ Sci Technol 46(5):3022– 3030. <https://doi.org/10.1021/es203861v>
- Hachicha R, Elleuch F, benHlimaDubessay HP, de Baynast H, Fendri I (2022) Biomolecules from microalgae and cyanobacteria: applications and market survey. Appl Sci 12(4):1924. [https://doi.org/](https://doi.org/10.3390/app12041924) [10.3390/app12041924](https://doi.org/10.3390/app12041924)
- Health Canada (2021) Natural Health Products Ingredients Database. Available online: <http://webprod.hc-sc.gc.ca/nhpid-bdipsn/> search-rechercheReq.do?lang=eng (accessed on 20 April 2021).
- Heinsoo D (2014) Cultivation of Spirulina on Conventional and Urine Based Medium in a Household Scale System. Master of Science Thesis KTH School of Biotechnology, SE-100: 44
- Hu B, Min M, Zhou W, Du Z, Mohr M, Chen P, Ruan R (2012) Enhanced mixotrophic growth of microalga Chlorella sp. on pretreated swine manure for simultaneous biofuel feedstock production and nutrient removal. Bioresour Technol 126:71–79. [https://](https://doi.org/10.1016/j.biortech.2012.09.031) doi.org/10.1016/j.biortech.2012.09.031
- Hu B, Zhou W, Min M, Du Z, Chen P, Ma X, Liu Y, Lei H, Shi J, Ruan R (2013) Development of an efective acidogenically digested swine manure-based algal system for improved wastewater treatment and biofuel and feed production. Appl Energy 107:255– 263. <https://doi.org/10.1016/j.apenergy.2013.02.033>
- Huang J, Feng F, Wan M, Ying J, Li Y, Qu X, Pan R, Shen G, Li W (2015) Improving performance of fat-plate photobioreactors by installation of novel internal mixers optimized with computational fuid dynamics. Bioresour Technol 182:151–159. [https://](https://doi.org/10.1016/j.biortech.2015.01.067) doi.org/10.1016/j.biortech.2015.01.067
- Huang Q, Jiang F, Wang L, Yang C (2017) Design of photobioreactors for mass cultivation of photosynthetic organisms. Engineering 3(3):318–329. <https://doi.org/10.1016/J.ENG.2017.03.020>
- Hulatt CJ, Lakaniemi AM, Puhakka JA, Thomas DN (2012) Energy demands of nitrogen supply in mass cultivation of two commercially important microalgal species, Chlorella vulgaris and Dunaliella tertiolecta. Bio Energy Res 5(3):669–684. [https://doi.](https://doi.org/10.1007/s12155-011-9175-x) [org/10.1007/s12155-011-9175-x](https://doi.org/10.1007/s12155-011-9175-x)
- Hussein HJ, Naji SS, Al-Khafaji NMS (2018) Antibacterial properties of the Chlorella vulgaris isolated from polluted water in Iraq. J Pharm Sci Res 10(10):2457–2460
- Hwang JH, Church J, Lee SJ, Park J, Lee WH (2016) Use of microalgae for advanced wastewater treatment and sustainable bioenergy generation. Environ Eng Sci 33(11):882–897. [https://doi.org/10.](https://doi.org/10.1089/ees.2016.0132) [1089/ees.2016.0132](https://doi.org/10.1089/ees.2016.0132)
- Ieropoulos I, Greenman J, Melhuish C (2012) Urine utilization by microbial fuel cells; energy fuel for the future. Phys Chem Chem Phys 14(1):94–98. <https://doi.org/10.1039/C1CP23213D>
- Ieropoulos IA, Ledezma P, Stinchcombe A, Papaharalabos G, Melhuish C, Greenman J (2013) Waste to real energy: the frst MFC powered mobile phone. Phys Chem Chem Phys 15(37):15312–15316. <https://doi.org/10.1039/c3cp52889h>
- Ieropoulos IA, Stinchcombe A, Gajda I, Forbes S, Merino-Jimenez I, Pasternak G, Sanchez-Herranz D, Greenman J (2016) Pee power urinal–microbial fuel cell technology feld trials in the context of sanitation. Environ Sci Water Res Technol 2(2):336–343. [https://](https://doi.org/10.1039/C5EW00270B) doi.org/10.1039/C5EW00270B
- Igos E, Besson M, Gutierrez TN, de Faria ABB, Benetto E, Barna L, Ahmadi A, Spérandio M (2017) Assessment of environmental impacts and operational costs of the implementation of an innovative source-separated urine treatment. Water Res 126:50–59. <https://doi.org/10.1016/j.watres.2017.09.016>
- Ishii SKL, Boyer TH (2015) Life cycle comparison of centralized wastewater treatment and urine source separation with struvite precipitation: Focus on urine nutrient management. Water Res 79:88–103. <https://doi.org/10.1016/j.watres.2015.04.010>
- Ismailkhodjaev BS, Khalmurzayeva BA, Satayev MI, Alibekov RS (2019) Study of the vitamins content of microalgae. Series Chem and Techno 4:19–24. [https://doi.org/10.32014/2019.2518-1491.](https://doi.org/10.32014/2019.2518-1491.37) [37](https://doi.org/10.32014/2019.2518-1491.37)
- Jaatinen S, Lakaniemi AM, Rintala J (2016) Use of diluted urine for cultivation of Chlorella vulgaris. Environ Technol 37(9):1159– 1170.<https://doi.org/10.1080/09593330.2015.1105300>
- Jacobi A, Steinweg C, Sastre RR, Posten C (2012) Advanced photobioreactor LED illumination system: scale-down approach to study microalgal growth kinetics. Eng Life Sci 12(6):621–630. [https://](https://doi.org/10.1002/elsc.201200004) doi.org/10.1002/elsc.201200004
- Jadhav DA, Jain SC, Ghangrekar MM (2016) Cow's urine as a yellow gold for bioelectricity generation in low cost clayware microbial fuel cell. Energy 113:76–84. [https://doi.org/10.1016/j.energy.](https://doi.org/10.1016/j.energy.2016.07.025) [2016.07.025](https://doi.org/10.1016/j.energy.2016.07.025)
- Jämsä M, Lynch F, Santana-Sánchez A, Laaksonen P, Zaitsev G, Solovchenko A, Allahverdiyeva Y (2017) Nutrient removal and biodiesel feedstock potential of green alga UHCC00027 grown in municipal wastewater under Nordic conditions. Algal Res 26:65–73.<https://doi.org/10.1016/j.algal.2017.06.019>
- Jana BB, Rana S, Bag SK (2012) Use of human urine in phytoplankton production as a tool for ecological sanitation. Water Sci Technol 65(8):1350–1356. <https://doi.org/10.2166/wst.2012.044>
- Jimenez J, Bott C, Love N, Bratby J (2015) Source separation of urine as an alternative solution to nutrient management in biological nutrient removal treatment plants. Water Environ Res 87(12):2120–2129. [https://doi.org/10.2175/106143015X14212](https://doi.org/10.2175/106143015X14212658613884) [658613884](https://doi.org/10.2175/106143015X14212658613884)
- Johnson TJ, Katuwal S, Anderson GA, Gu L, Zhou R, Gibbons WR (2018) Photobioreactor cultivation strategies for microalgae and cyanobacteria. Biotechnol Prog 34(4):811–827. [https://doi.org/](https://doi.org/10.1002/btpr.2628) [10.1002/btpr.2628](https://doi.org/10.1002/btpr.2628)
- Kadir WNA, Lam MK, Uemura Y, Lim JW, Lee KT (2018) Harvesting and pre-treatment of microalgae cultivated in wastewater for biodiesel production: a review. Energy Convers Manag 171:1416– 1429. <https://doi.org/10.1016/j.enconman.2018.06.074>
- Kainer D (2018) Microalgae protocols. In: K. Whitfeld, S. Burks, A. B. Cortez, & C Kainer (Eds.) International space station (ISS) mission: Microalgae. National Aeronautics and Space Administration (NASA). Retrieved from <https://www.nasa>. gov/sites/ default/fes/atoms/fes/microalgaeprotocols_508.pdf . (Accessed 20June 2021).
- Kajjumba GW, Fischer D, Risso L, Koury D, Marti EJ (2021) Application of cerium and lanthanum coagulants in wastewater treatment—A comparative assessment to magnesium, aluminum, and iron coagulants. Chem Eng J 426:131268. [https://doi.org/](https://doi.org/10.1016/j.cej.2021.131268) [10.1016/j.cej.2021.131268](https://doi.org/10.1016/j.cej.2021.131268)
- Kalra R, Gaur S, Goel M (2021) Microalgae bioremediation: A perspective towards wastewater treatment along with industrial carotenoids production. J Water Process Eng 40:101794. [https://](https://doi.org/10.1016/j.jwpe.2020.101794) doi.org/10.1016/j.jwpe.2020.101794
- Kaur R (2021) Proftability analysis for microalgae cultivation in fnland-case company: Redono Oy
- Kc S, Shinjo H (2020) Efects of human urine and ecosan manure on plant growth and soil properties in Central Nepal. Sanitation Value Chain 4(2):19–37.<https://doi.org/10.34416/svc.00021>
- Khaldi H, Maatoug M, Dube CS, Ncube M, Tandlich R, Heilmeier H, Dellal A (2017) Efficiency of wastewater treatment by a mixture of sludge and microalgae. J Fundam Appl Sci 9(3):1454–1472. <https://doi.org/10.4314/jfas.v9i3.13>
- Khanra S, Mondal M, Halder G, Tiwari ON, Gayen K, Bhowmick TK (2018) Downstream processing of microalgae for pigments, protein and carbohydrate in industrial application: a review. Food Bioprod Process 110:60–84. [https://doi.org/10.1016/j.fbp.2018.](https://doi.org/10.1016/j.fbp.2018.02.002) [02.002](https://doi.org/10.1016/j.fbp.2018.02.002)
- Khan AH, Khan NA, Ahmed S, Dhingra A, Singh CP, Khan SU, Mohammadi AA, Changani F, Yousef M, Alam S (2020) Application of advanced oxidation processes followed by diferent treatment technologies for hospital wastewater treatment. J Clean Prod 269:122411.<https://doi.org/10.1016/j.jclepro.2020.122411>
- Kim J, Kim B, Kim H, Yun Z (2014) Efects of ammonium ions from the analyte within bio-cathode microbial fuel cells on nitrate reduction and current density. Int Biodeterior Biodegrad 95:122– 126. <https://doi.org/10.1016/j.ibiod.2014.04.015>
- Kishor N, Kale S, Agrawal PS (2020) Use of fertilizers derived from urine as a plant growth regulator. Mater Today: Proceed 32:504– 509. <https://doi.org/10.1016/j.matpr.2020.02.761>
- Korving L, Loosdrecht MV, Wilfert P (2019) Effect of iron on phosphate recovery from sewage sludge. In: (Ed.) Phosphorus recovery and recycling. Springer, Singapore, pp 303–326 DOI: [https://](https://doi.org/10.1007/978-981-10-8031-9_21) doi.org/10.1007/978-981-10-8031-9_21
- Koutra E, Papavasileiou P, Andriopoulos V, Mastropetros SG, Kornaros M (2022) Bioactive compounds from microalgae cultivated in wastewaters. In: (Ed.) An Integration of Phycoremediation Processes in Wastewater Treatment. Elsevier, pp 177–202 [https://](https://doi.org/10.1016/B978-0-12-823499-0.00009-2) doi.org/10.1016/B978-0-12-823499-0.00009-2
- Koutsoumanis K, Allende A, Alvarez-Ord A, Romero P (2020) Peer review of the pesticide risk assessment of the active substance Reynoutria sachalinensis extract. EFSA J 18(2):5966. [https://doi.](https://doi.org/10.2903/j.efsa.2020.5966) [org/10.2903/j.efsa.2020.5966](https://doi.org/10.2903/j.efsa.2020.5966)
- Koyande AK, Chew KW, Rambabu K, Tao Y, Chu D, Show P (2019) Microalgae: A potential alternative to health supplementation for humans. Food Sci Hum Wellness 8(1):16–24. [https://doi.org/10.](https://doi.org/10.1016/j.fshw.2019.03.001) [1016/j.fshw.2019.03.001](https://doi.org/10.1016/j.fshw.2019.03.001)
- Krahenbühl M, Etter B, Udert KM (2016) Pretreated magnesite as a source of low-cost magnesium for producing struvite from urine in Nepal. Sci Total Environ 542:1155–1161. [https://doi.org/10.](https://doi.org/10.1016/j.scitotenv.2015.08.060) [1016/j.scitotenv.2015.08.060](https://doi.org/10.1016/j.scitotenv.2015.08.060)
- Kube M, Fan L, Roddick F (2021) Alginate-immobilized algal wastewater treatment enhanced by species selection. Algal Res 54:102219. <https://doi.org/10.1016/j.algal.2021.102219>
- Kube M, Mohseni A, Fan L, Roddic F (2019) Impact of alginate selection for wastewater treatment by immobilized Chlorella vulgaris. Chem Eng J 358:1601–1609. [https://doi.org/10.1016/j.cej.2018.](https://doi.org/10.1016/j.cej.2018.10.065) [10.065](https://doi.org/10.1016/j.cej.2018.10.065)
- Kumar A, Singh AK, Chandra R (2021) Recent advances in physicochemical and biological approaches for degradation and detoxifcation of industrial wastewater. Emerg Treat Technolo Waste Manage. https://doi.org/10.1007/978-981-16-2015-7_1
- Kumar V, Kumar A, Nanda M (2018) Pretreated animal and human waste as a substantial nutrient source for cultivation of microalgae for biodiesel production. Environ Sci Pollut Res 25(22):22052– 22059.<https://doi.org/10.1007/s11356-018-2339-x>
- Kuntke P, Smiech K, Bruning H, Zeeman G, Saakes M, Sleutels THJA, Hamelers HVM, Buisman CJN (2012) Ammonium recovery and energy production from urine by a microbial fuel cell. Water Res 46(8):2627–2636. <https://doi.org/10.1016/j.watres.2012.02.025>
- Lafarga T, Rodríguez-Bermúdez R, Morillas-España A, Villaró S, García-Vaquero M, Morán L, Sánchez-Zurano A, González-López CV, Acién-Fernández FG (2021) Consumer knowledge and attitudes towards microalgae as food: the case of Spain. Algal Res 54:102174
- Lam MK, Lee KT (2012) Microalgae biofuels: a critical review of issues, problems and the way forward. Biotechnol Adv 30(3):673–690. <https://doi.org/10.1016/j.biotechadv.2011.11.008>
- Lamichhane KM, Babcock RW Jr (2013) Survey of attitudes and perceptions of urine-diverting toilets and human waste recycling in Hawaii. Sci Total Environ 443:749–756. [https://doi.org/10.](https://doi.org/10.1016/j.scitotenv.2012.11.039) [1016/j.scitotenv.2012.11.039](https://doi.org/10.1016/j.scitotenv.2012.11.039)
- Lamminen M, Halmemies-Beauchet-Filleau A, Kokkonen T, Vanhatalo A, Jaakkola S (2019) The efect of partial substitution of rapeseed meal and faba beans by Spirulina platensis microalgae on milk production, nitrogen utilization, and amino acid metabolism of lactating dairy cows. J Dairy Sci 102(8):7102– 7117. <https://doi.org/10.3168/jds.2018-16213>
- Larsen TA, Hoffmann S, Lüthi C, Truffer B, Maurer M (2016) Emerging solutions to the water challenges of an urbanizing world. Science 352(6288):928–933. [https://doi.org/10.1126/](https://doi.org/10.1126/science.aad8641) [science.aad8641](https://doi.org/10.1126/science.aad8641)
- Larsen TA, Riechmann ME, Udert KM (2021) State of the art of urine treatment technologies: a critical review. Water Res X 13:100114. <https://doi.org/10.1016/j.wroa.2021.100114>
- Ledezma P, Greenman J, Ieropoulos I (2013) MFC-cascade stacks maximize COD reduction and avoid voltage reversal under adverse conditions. Bioresour Technol 134:158–165. [https://](https://doi.org/10.1016/j.biortech.2013.01.119) doi.org/10.1016/j.biortech.2013.01.119
- Lee H, Jeong D, Im S, Jang A (2020) Optimization of alginate bead size immobilized with Chlorella vulgaris and Chlamydomonas reinhardtii for nutrient removal. Bioresour Technol 302:122891. [https://doi.org/10.1016/j.biortech.2020.](https://doi.org/10.1016/j.biortech.2020.122891) [122891](https://doi.org/10.1016/j.biortech.2020.122891)
- Leonard C, Ferrasse JH, Boutin O, Lefevre S, Viand A (2015) Bubble column reactors for high pressures and high temperatures operation. Chem Eng Res Des 100:391–421. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.cherd.2015.05.013) [cherd.2015.05.013](https://doi.org/10.1016/j.cherd.2015.05.013)
- Levasseur W, Perr´e P, Pozzobon V (2020) A review of high valueadded molecules production by microalgae in light of the classifcation. Biotechnol Adv. [https://doi.org/10.1016/j.biotechadv.](https://doi.org/10.1016/j.biotechadv.2020.107545) [2020.107545](https://doi.org/10.1016/j.biotechadv.2020.107545)
- Levasseur W, Perre P, Pozzobon V (2020) A review of high valueadded molecules production by microalgae in light of the classifcation. Biotechnol Adv. [https://doi.org/10.1016/j.biotechadv.](https://doi.org/10.1016/j.biotechadv.2020.107545) [2020.107545](https://doi.org/10.1016/j.biotechadv.2020.107545)
- Li J, Stamato M, Velliou E, Jefryes C, Agathos SN (2015) Design and characterization of a scalable airlift fat panel photobioreactor for microalgae cultivation. J Appl Phycol 27(1):75–86. [https://doi.](https://doi.org/10.1007/s10811-014-0335-1) [org/10.1007/s10811-014-0335-1](https://doi.org/10.1007/s10811-014-0335-1)
- Li K, Liu Q, Fang F, Luo R, Lu Q, Zhou W, Huo S, Cheng P, Liu J, Addy M, Chen P, Chem D, Ruan R (2019) Microalgae-based wastewater treatment for nutrients recovery: a review. Bioresour Technol 291:121934. [https://doi.org/10.1016/j.biortech.2019.](https://doi.org/10.1016/j.biortech.2019.121934) [121934](https://doi.org/10.1016/j.biortech.2019.121934)
- Li L, Wang X, Miao J, Abulimiti A, Jing X, Ren N (2022) Carbon neutrality of wastewater treatment-A systematic concept beyond the plant boundary. Environ Sci Ecotechnol 11:100180. [https://](https://doi.org/10.1016/j.ese.2022.100180) doi.org/10.1016/j.ese.2022.100180
- Lienert J, Haller M, Berner A, Staufacher M, Larsen TA (2003) How farmers in Switzerland perceive fertilizers from recycled anthropogenic nutrients (urine). Water Sci Technol 48(1):47–56 (**PMID: 12926620**)
- Lindberg RH, Namazkar S, Lage S, Ostman M, Gojkovic Z, Funk C, Tysklind M (2021) Fate of active pharmaceutical ingredients in a northern high-rate algal pond fed with municipal waste. Chemosphere 271:129763. [https://doi.org/10.1016/j.chemosphere.](https://doi.org/10.1016/j.chemosphere.2021.129763) [2021.129763](https://doi.org/10.1016/j.chemosphere.2021.129763)
- Liyanaarachchi VC, Premaratne M, Ariyadasa TU, Nimarshana PHV, Malik A (2021) Two-stage cultivation of microalgae for production of high-value compounds and biofuels: a review. Algal Res 57:102353.<https://doi.org/10.1016/j.algal.2021.102353>
- Ljubic A, Jacobsen C, Holdt SL, Jakobsen J (2020) Microalgae Nannochloropsis oceanica as a future new natural source of vitamin D3. Food Chem. <https://doi.org/10.1016/j.foodchem.2020.126627>
- López-Rosales L, García-Camacho F, Sánchez-Mirón A, Beato EM, Chisti Y, Grima EM (2016) Pilot-scale bubble column photobioreactor culture of a marine dinofagellate microalga illuminated with light emission diodes. Bioresour Technol 216:845–855. <https://doi.org/10.1016/j.biortech.2016.06.027>
- Lu J, Bo X, Wang H, Guo L (2013) Nitrogen-doped ordered mesoporous carbons synthesized from honey as metal-free catalyst for oxygen reaction reduction. Electrochim Acta 108:10–16. <https://doi.org/10.1016/j.electacta.2013.06.066>
- Lucas S, Gouin S, Lesueur M (2019) Seaweed consumption and label preferences in France. Mar Resour Econ 34:143–162
- Luo Y, Le-Clech P, Henderson RK (2017) Simultaneous microalgae cultivation and wastewater treatment in submerged membrane photobioreactors: a review. Algal Res 24:425–437. [https://doi.](https://doi.org/10.1016/j.algal.2016.10.026) [org/10.1016/j.algal.2016.10.026](https://doi.org/10.1016/j.algal.2016.10.026)
- Lynch F, Santana-Sánchez A, Jämsä M, Sivonen K, Aro EM, Allahverdiyeva Y (2015) Screening native isolates of cyanobacteria and a green alga for integrated wastewater treatment, biomass accumulation and neutral lipid production. Algal Res 11:411–420. <https://doi.org/10.1016/j.algal.2015.05.015>
- Martin TM, Esculier F, Levavasseur F, Houot S (2022) Human urine-based fertilizers: a review. Crit Rev Environ Sci Technol 52(6):890–936. <https://doi.org/10.1080/10643389.2020.1838214>
- Mastropetros SG, Pispas K, Zagklis D, Ali SS, Kornaros M (2022) Biopolymers production from microalgae and cyanobacteria cultivated in wastewater: recent advances. Biotechnol Adv. [https://](https://doi.org/10.1016/j.biotechadv.2022.107999) doi.org/10.1016/j.biotechadv.2022.107999
- Mbir BI, Mensah AJK (2017) The wastewater nutrient removal efficiences of Chlorella sorokiniana and Scenedesmus obtusiusculus. Bioprocess Eng 1(3): 69–76. [https://doi.org/10.11648/j.be.20170](https://doi.org/10.11648/j.be.20170103.12) [103.12](https://doi.org/10.11648/j.be.20170103.12)
- Mehariya S, Goswami RK, Karthikeysan OP, Verma P (2021) Microalgae for high-value products: A way towards green nutraceutical

and pharmaceutical compounds. Chemosphere. [https://doi.org/](https://doi.org/10.1016/j.chemosphere.2021.130553) [10.1016/j.chemosphere.2021.130553](https://doi.org/10.1016/j.chemosphere.2021.130553)

- Mehta CM, Khunjar WO, Nguyen V, Tait S, Batstone DJ (2015) Technologies to recover nutrients from waste streams: a critical review. Crit Rev Environ Sci Technol 45(4):385–427. [https://doi.](https://doi.org/10.1080/10643389.2013.866621) [org/10.1080/10643389.2013.866621](https://doi.org/10.1080/10643389.2013.866621)
- Min M, Hu B, Mohr MJ, Shi A, Ding J, Sun Y, Chen P, Zhou WG, Ruan R (2014) Swine manure-based pilot-scale algal biomass production system for fuel production and wastewater treatment—a case study. Appl Biochem Biotechnol 172(3):1390–1406. [https://doi.](https://doi.org/10.1007/s12010-013-0603-6) [org/10.1007/s12010-013-0603-6](https://doi.org/10.1007/s12010-013-0603-6)
- Miranda AF, Taha M, Wrede D, Morrison P, Ball AS, Stevenson T, Mouradov A (2015) Lipid production in association of flamentous fungi with genetically modifed cyanobacterial cells. Biotechnol Biofuels 8(1):1–18. [https://doi.org/10.1016/j.algal.2016.](https://doi.org/10.1016/j.algal.2016.10.026) [10.026](https://doi.org/10.1016/j.algal.2016.10.026)
- Mishra S, Pang S, Zhang W, Lin Z, Bhatt P, Chen S (2021) Insights into the microbial degradation and biochemical mechanisms of carbamates. Chemosphere 279:130500. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.chemosphere.2021.130500) [chemosphere.2021.130500](https://doi.org/10.1016/j.chemosphere.2021.130500)
- Mohsenpour SF, Hennige S, Willoughby N, Adeloye A, Gutierrez T (2021) Integrating micro-algae into wastewater treatment: a review. Sci Total Environ 752:142168. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.scitotenv.2020.142168) [scitotenv.2020.142168](https://doi.org/10.1016/j.scitotenv.2020.142168)
- Moons I, Barbarossa C, De Pelsmacker P (2018) The determinants of the adoption intention of eco-friendly functional food in diferent market segments. Ecol Econ 151:151–161
- Moraes L, Rosa GM, Cara IM, Santos LO, Morais MG, Grima EM, Costa JAV, Fernández FG (2020) Bioprocess strategies for enhancing the outdoor production of Nannochloropsis gaditana: an evaluation of the efects of pH on culture performance in tubular photobioreactors. Bioprocess Biosyst Eng 43(10):1823–1832. <https://doi.org/10.1007/s00449-020-02373-x>
- Mubarak M, Shaija A (2019) Prashanth P (2019) Bubble column photobioreactor for Chlorella pyrenoidosa cultivation and validating gas hold up and volumetric mass transfer coefficient energy sources part a recovery util. Environ Eff 10(1080/15567036):1680769
- Mugivhisa LL, Olowoyo JO (2015) An assessment of university students and staff perceptions regarding the use of human urine as a valuable soil nutrient in South Africa. Afr Health Sci 15(3):999– 1010.<https://doi.org/10.4314/ahs.v15i3.39>
- Muluye A, Sali B, Bahta B, Melese B, Girma B, Kebede M, Suresh A (2021) Potential of Human Urine as a Nutrient Medium for the Biomass Production of Microalga Scenedesmus sp. East Afr J Agric Biotechnol 3(1):35–41. [https://doi.org/10.37284/eajab.3.](https://doi.org/10.37284/eajab.3.1.409) [1.409](https://doi.org/10.37284/eajab.3.1.409)
- Naidoo S, Olaniran AO (2014) Treated wastewater effluent as a source of microbial pollution of surface water resources. Int J Environ Res Public Health 11(1):249–270. [https://doi.org/10.3390/ijerp](https://doi.org/10.3390/ijerph110100249) [h110100249](https://doi.org/10.3390/ijerph110100249)
- Nassar MY, Ali EI, Zakaria ES (2017) Tunable auto-combustion preparation of TiO2 nanostructures as efficient adsorbents for the removal of an anionic textile dye. RSC Adv 7:8034–8050. [https://](https://doi.org/10.1039/C6RA27924D) doi.org/10.1039/C6RA27924D
- Nazih H, Bard JM (2018) Microalgae in human health: Interest as a functional food. In: I. A. Levine, & J. Fleurence (Eds.), Microalgae in health and disease prevention Elsevier, Inc. pp 211–226 <https://doi.org/10.1016/B978-0-12-811405-6.00010-4>
- Nguyen TT, Bui XT, Ngo HH, Nguyen KQ, Nguyen HH, Némery J, Fujioka T, Duong CH, Dang BT, Varjani S (2021) Nutrient recovery and microalgae biomass production from urine by membrane photobioreactor at low biomass retention times. Sci Total Environ 785:147423. [https://doi.org/10.1016/j.scitotenv.](https://doi.org/10.1016/j.scitotenv.2021.147423) [2021.147423](https://doi.org/10.1016/j.scitotenv.2021.147423)
- Niccolai A, Chini-Zittelli G, Rodolf L, Biondi N, Tredici MR (2019) Microalgae of interest as food source: biochemical composition and digestibility. Algal Res 42:101617. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.ALGAL.2019.101617) [ALGAL.2019.101617](https://doi.org/10.1016/J.ALGAL.2019.101617)
- Nie X, Mubashar M, Zhang S, Qin Y, Zhang X (2020) Current progress, challenges and perspectives in microalgae-based nutrient removal for aquaculture waste: A comprehensive review. J Clean Prod 277:124209.<https://doi.org/10.1016/j.jclepro.2020.124209>
- Nirmalakhandan N, Selvaratnam T, Henkanatte-Gedera SM, Tchinda D, Abeysiriwardana-Arachchige ISA, Delanka-Pedige HMK, Munasinghe Arachchige SP, Zhang Y, Holguin FO, Lammers PJ (2019) Algal wastewater treatment: photoautotrophic vs mixotrophic processes. Algal Res 41:101569. [https://doi.org/](https://doi.org/10.1016/j.algal.2019.101569) [10.1016/j.algal.2019.101569](https://doi.org/10.1016/j.algal.2019.101569)
- Nova P, Pimenta-Martins A, Silva JL, Silva AM, Gomes AM, Freita AC (2020) Health benefts and bioavailability of marine resources components thatcontribute to health – what's new? Crit Rev Food Sci Nutr 60(21):3680–3692. [https://doi.org/10.](https://doi.org/10.1080/10408398.2019.1704681) [1080/10408398.2019.1704681](https://doi.org/10.1080/10408398.2019.1704681)
- Nutrient recovery from wastewater. Evoqua Water Technologies. (n.d.). Retrieved October 30, 2022, from [https://www.evoqua.com/en/](https://www.evoqua.com/en/markets/applications/nutrient-recovery/#:~:text=Nutrient%20recovery%20as%20a%20treatment,transformed%20into%20high%2Dperforming%20fertilizers) [markets/applications/nutrient-recovery/#:~:text=Nutrient%20rec](https://www.evoqua.com/en/markets/applications/nutrient-recovery/#:~:text=Nutrient%20recovery%20as%20a%20treatment,transformed%20into%20high%2Dperforming%20fertilizers) [overy%20as%20a%20treatment,transformed%20into%20high%](https://www.evoqua.com/en/markets/applications/nutrient-recovery/#:~:text=Nutrient%20recovery%20as%20a%20treatment,transformed%20into%20high%2Dperforming%20fertilizers) [2Dperforming%20fertilizers.](https://www.evoqua.com/en/markets/applications/nutrient-recovery/#:~:text=Nutrient%20recovery%20as%20a%20treatment,transformed%20into%20high%2Dperforming%20fertilizers)
- Odoemena KI, Rowshon MD, KHasfalinaCMBinti (2020) Advances in utilization of wastewater in agricultural practice: a technical note. Irrig Drain Syst 69(1):149–163.<https://doi.org/10.1002/ird.2384>
- Ojanen S, Tyystjärvi E, Holmberg H, Ahtila P (2015) Porous membrane as a means of gas and nutrient exchange in a tubular photobioreactor. J Appl Phycol 27(3):1169–1175. [https://doi.org/10.](https://doi.org/10.1007/s10811-014-0397-0) [1007/s10811-014-0397-0](https://doi.org/10.1007/s10811-014-0397-0)
- Pandorf M, Hochmuth G, Boyer TH (2018) Human urine as a fertilizer in the cultivation of snap beans (Phaseolus vulgaris) and turnips (Brassica rapa). J Agric Food Chem 67(1):50–62. [https://doi.org/](https://doi.org/10.1021/acs.jafc.8b06011) [10.1021/acs.jafc.8b06011](https://doi.org/10.1021/acs.jafc.8b06011)
- Parwada C, Chigiya V, Ngezimana W, Chipomho J, Bandason W, Nyamushamba GB (2021) Use and management of animal manure by the communal farmers, Seke District, Mashonaland East Province, Zimbabwe. Res World Agric Econ 2:6–15
- Patel A, Mungray AA, Mungray AK (2020) Technologies for the recovery of nutrients, water and energy from human urine: a review. Chemosphere 259:127372. [https://doi.org/10.1016/j.chemo](https://doi.org/10.1016/j.chemosphere.2020.127372) [sphere.2020.127372](https://doi.org/10.1016/j.chemosphere.2020.127372)
- Pathy A, Ray J, Paramasivan B (2021) Challenges and opportunities of nutrient recovery from human urine using biochar for fertilizer applications. J Clean Prod 304:127019. [https://doi.](https://doi.org/10.1016/j.jclepro.2021.127019) [org/10.1016/j.jclepro.2021.127019](https://doi.org/10.1016/j.jclepro.2021.127019)
- Peter AP, Koyande AK, Chew KW, Ho SH, Chen WH, Chang JS, Krishnamoorthy R, Banat F, Show PL (2022) Continuous cultivation of microalgae in photobioreactors as a source of renewable energy: current status and future challenges. Renew Sustain Energy Rev 154:111852. [https://doi.org/10.1016/j.rser.](https://doi.org/10.1016/j.rser.2021.111852) [2021.111852](https://doi.org/10.1016/j.rser.2021.111852)
- Pfafnger CE, Severin TS, Apel AC, Göbel J, Sauter J, Weuster-Botz D (2019) Light-dependent growth kinetics enable scale-up of well-mixed phototrophic bioprocesses in diferent types of photobioreactors. J Biotechnol 297:41–48. [https://doi.org/10.](https://doi.org/10.1016/j.jbiotec.2019.03.003) [1016/j.jbiotec.2019.03.003](https://doi.org/10.1016/j.jbiotec.2019.03.003)
- Piltz B, Melkonian M (2018) Immobilized microalgae for nutrient recovery from source-separated human urine. J Appl Phycol 30(1):421–429. <https://doi.org/10.1007/s10811-017-1266-4>
- Pittman JK, Dean AP, Osundeko O (2011) The potential of sustainable algal biofuel production using wastewater resources. Bioresour Technol 102(1):17–25. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.biortech.2010.06.035) [biortech.2010.06.035](https://doi.org/10.1016/j.biortech.2010.06.035)
- Pomport PH, Warren HE, Taylor-Pickard J (2021) Effect of total replacement of inorganic with organic sources of key trace minerals on performance and health of high producing dairy cows. J Appl Anim Nutr 9(1):23–30. [https://doi.org/10.3920/](https://doi.org/10.3920/JAAN2020.0018) [JAAN2020.0018](https://doi.org/10.3920/JAAN2020.0018)
- Posadas E, Alcántara C, García-Encina PA, Gouveia L, Guieysse B, Norvill Z, Acien FG, Markou G, Congestri R, Koreiviene J, Muñoz R (2017) Microalgae cultivation in wastewater. In: Microalgae-based biofuels and bioproducts. Woodhead Publishing, pp 67–91 [https://doi.org/10.1016/B978-0-08-101023-](https://doi.org/10.1016/B978-0-08-101023-5.00003-0) [5.00003-0](https://doi.org/10.1016/B978-0-08-101023-5.00003-0)
- Praveen P, Heng JYP, Loh KC (2016) Tertiary wastewater treatment in membrane photobioreactor using microalgae: Comparison of forward osmosis & microfltration. Bioresour Technol 222:448–457. <https://doi.org/10.1016/j.biortech.2016.09.124>
- Raheem A, Azlina WW, Yap YT, Danquah MK, Harun R (2015) Thermochemical conversion of microalgal biomass for biofuel production. Renew Sustain Energy Rev 49:990–999. [https://doi.org/](https://doi.org/10.1016/j.rser.2015.04.186) [10.1016/j.rser.2015.04.186](https://doi.org/10.1016/j.rser.2015.04.186)
- Ranasinghe ESS, Karunarathne CLSM, Beneragama CK, Wijesooriya BGG (2016) Human urine as a low cost and effective nitrogen fertilizer for bean production. Procedia Food Sci 6:279–282. <https://doi.org/10.1016/j.profoo.2016.02.055>
- Rao PH, Kumar RR, Mohan N (2019) Phycoremediation: role of algae in waste management. In: (Ed.) Environmental Contaminants: Ecological Implications and Management. Springer, Singapore, pp 49–82 DOI: https://doi.org/10.1007/978-981-13-7904-8_3
- Rawat I, Bhola V, Kumar RR, Bux F (2013) Improving the feasibility of producing biofuels from microalgae using wastewater. Environ Technolo 34(13–14):1765–1775. [https://doi.org/10.1080/09593](https://doi.org/10.1080/09593330.2013.826287) [330.2013.826287](https://doi.org/10.1080/09593330.2013.826287)
- Razzak SA, Ali SAM, Hossain MM, deLasa H (2017) Biological CO2 fxation with production of microalgae in wastewater–a review. Renew Sustain Energy Rev 76:379–390. [https://doi.org/10.](https://doi.org/10.1016/j.rser.2017.02.038) [1016/j.rser.2017.02.038](https://doi.org/10.1016/j.rser.2017.02.038)
- Rengasamy K, Berchmans S (2012) Simultaneous degradation of bad wine and electricity generation with the aid of the coexisting biocatalysts Acetobacter aceti and Gluconobacter roseus. Bioresour Technolo 104:388–393. [https://doi.org/10.1016/j.biortech.](https://doi.org/10.1016/j.biortech.2011.10.092) [2011.10.092](https://doi.org/10.1016/j.biortech.2011.10.092)
- Renuka N, Sood A, Prasanna R, Ahluwalia AS (2015) Phycoremediation of Waters: a synergistic approach using microalgae for bioremediation and biomass generation. Int J Environ Sci Tech 12(4):1443–1460. <https://doi.org/10.1007/s13762-014-0700-2>
- Rezvani S, Saadaoui I, Al Jabri H, Moheimani NR (2022) Technoeconomic modeling of high-value metabolites and secondary products from microalgae cultivated in closed photobioreactors with supplementary lighting. Algal Res 65:102733. [https://doi.](https://doi.org/10.1016/j.algal.2022.102733) [org/10.1016/j.algal.2022.102733](https://doi.org/10.1016/j.algal.2022.102733)
- Ricci EC, Banterle A, Stranieri S (2018) Trust to go green: an exploration of consumer intentions for eco-friendly convenience food. Ecol Econ 148:54–65
- Rodriguez-Rangel H, Arias DM, Morales-Rosales LA, Gonzalez-Huitron V, Partida MV, Garcia J (2022) Machine learning methods modeling carbohydrate enriched cyanobacteria biomass production in wastewater treatment systems. Energies 15(7):2500. <https://doi.org/10.3390/en15072500>
- Ronteltap M, Maurer M, Gujer W (2007) The behaviour of pharmaceuticals and heavy metals during struvite precipitation in urine. Water Res 41(9):1859–1868. [https://doi.org/10.1016/j.watres.](https://doi.org/10.1016/j.watres.2007.01.026) [2007.01.026](https://doi.org/10.1016/j.watres.2007.01.026)
- Rose C, Parker A, Jeferson B, Cartmell E (2015) The characterization of feces and urine: a review of the literature to inform advanced treatment technology. Crit Rev Environ Sci Technol 45(17):1827–1879. [https://doi.org/10.1080/10643389.2014.](https://doi.org/10.1080/10643389.2014.1000761) [1000761](https://doi.org/10.1080/10643389.2014.1000761)
- Roßmann M, Rösch C (2020) Key-Narratives of microalgae nutrition: exploring futures through a public delphi survey in Germany. Sci Public Policy 47(1):137–147
- Ruiz J, Olivieri G, De Vree J, Bosma R, Willems P, Reith JH, Eppink MHM, Kleinegris DMM, Wijffels RH, Barbosa MJ (2016) Towards industrial products from microalgae. Energy Environ Sci 9(10):3036–3043.<https://doi.org/10.1039/C6EE01493C>
- Rumin J, Nicolau E, de Oliveira G, Junior R, Fuentes-Grünewald C, Picot L (2020) Analysis of scientifc research driving microalgae market opportunities in Europe. Mar Drugs 18:264
- Rushan NH, Yasin NHM, Sepian NRA, Said FM, Shafei NI (2019) Efect of immobilization method on the growth of Chlorella vulgaris and fatty acid profle for biodiesel production. Indones J Chem 19(3):767–776.<https://doi.org/10.22146/ijc.39800>
- Sahin OI (2020) Functional and sensorial properties of cookies enriched with Spirulina and Dunaliella biomass. J Food Sci Technol 57(10):3639–3646. [https://doi.org/10.1007/](https://doi.org/10.1007/s13197-020-04396-4) [s13197-020-04396-4](https://doi.org/10.1007/s13197-020-04396-4)
- Sahin OI, Ozturk B (2021) Microalgal biomass — a bio-based additive: Evaluation of green smoothies during storage. Int Food Res J 28(2):309–316
- Sahu O (2019) Sustainable and clean treatment of industrial wastewater with microbial fuel cell. Results Eng 4:100053. [https://doi.org/](https://doi.org/10.1016/j.rineng.2019.100053) [10.1016/j.rineng.2019.100053](https://doi.org/10.1016/j.rineng.2019.100053)
- Sakarika M, Koutra E, Mastropetros SG, Giannakopoulos S, Kornaros M (2022) Wastewater treatment coupled to algal biomass production. In: (Ed.) An Integration of Phycoremediation Processes in Wastewater Treatment. Elsevier, pp 203–230 [https://doi.org/10.](https://doi.org/10.1016/B978-0-12-823499-0.00020-1) [1016/B978-0-12-823499-0.00020-1](https://doi.org/10.1016/B978-0-12-823499-0.00020-1)
- Saldarriaga LF, Almenglo F, Ramírez M, Cantero D (2020) Kinetic characterization and modeling of a microalgae consortium isolated from landfll leachate under a high CO2 concentration in a bubble column photobioreactor. Electron J Biotechnol 44:47–57. <https://doi.org/10.1016/j.ejbt.2020.01.006>
- Salehi AS, Yang SO, Earl CC, Tang MJS, Hunt JP, Smith MT, Wood DW, Bundy BC (2018) Biosensing estrogenic endocrine disruptors in human blood and urine: a RAPID cell-free protein synthesis approach. Toxicol Appl Pharmacol 345:19–25. [https://doi.](https://doi.org/10.1016/j.taap.2018.02.016) [org/10.1016/j.taap.2018.02.016](https://doi.org/10.1016/j.taap.2018.02.016)
- Salgot M, Folch M (2018) Wastewater treatment and water reuse. Curr Opin Environ Sci Health 2:64–74. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.coesh.2018.03.005) [coesh.2018.03.005](https://doi.org/10.1016/j.coesh.2018.03.005)
- Samsudeen N, Radhakrishnan TK, Matheswaran M (2015) Bioelectricity production from microbial fuel cell using mixed bacterial culture isolated from distillery wastewater. Bioresour Technol 195:242–247.<https://doi.org/10.1016/j.biortech.2015.07.023>
- Santiago AF, Calijuri ML, Assemany PP, Calijuri MDC, Reis AJDD (2013) Algal biomass production and wastewater treatment in high rate algal ponds receiving disinfected effluent. Environ Technol 34(13–14):1877–1885. [https://doi.org/10.1080/09593](https://doi.org/10.1080/09593330.2013.812670) [330.2013.812670](https://doi.org/10.1080/09593330.2013.812670)
- Saravanan A, Kumar PS, Varjani S, Jeevanantham S, Yaashikaa PR, Thamarai P, Abirami B, George CS (2021) A review on algalbacterial symbiotic system for efective treatment of wastewater. Chemosphere 271:129540. [https://doi.org/10.1016/j.chemo](https://doi.org/10.1016/j.chemosphere.2021.129540) [sphere.2021.129540](https://doi.org/10.1016/j.chemosphere.2021.129540)
- Sarbatly RH, Suali E (2013) Membrane Photobioreactor as a Device to Increase CO₂ Mitigation by Microalgae. In: Advances in Biofuels. Springer, Boston, MA. pp 241–258 [https://doi.org/10.1007/](https://doi.org/10.1007/978-1-4614-6249-1_14) [978-1-4614-6249-1_14](https://doi.org/10.1007/978-1-4614-6249-1_14)
- Schagerl M, Ludwig I, El-Sheekh M, Kornaros M, Ali SS (2022) The efficiency of microalgae-based remediation as a green process for industrial wastewater treatment. Algal Res 66:102775. [https://](https://doi.org/10.1016/j.algal.2022.102775) doi.org/10.1016/j.algal.2022.102775
- Sharma R, Kumari R, Pant D, Malaviya P (2021) Bioelectricity generation from human urine and simultaneous nutrient recovery: role of microbial fuel cells. Chemosphere. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.chemosphere.2021.133437) [chemosphere.2021.133437](https://doi.org/10.1016/j.chemosphere.2021.133437)
- Shin YH Schideman L (2015) Characterizing the fate and transport of Chemicals of Emerging Concerns (CECs) from integrated bioenergy and manure management system. In: 2015 ASABE Annual International Meeting, American Society of Agricultural and Biological Engineers, p 1
- Simha P, Lalander C, Vinnerås B, Ganesapillai M (2017) Farmer attitudes and perceptions to the re–use of fertiliser products from resource–oriented sanitation systems–the case of Vellore. South India Sci Total Environ 581:885–896. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.scitotenv.2017.01.044) [scitotenv.2017.01.044](https://doi.org/10.1016/j.scitotenv.2017.01.044)
- Singh S, Cheema PPS (2016) Use of Eco-sanitation process in rural areas of Punjab India. J Environ Sci toxicol Food Technol 10:29– 33.<https://doi.org/10.9790/2402-1011032933>
- Soares A (2020) Wastewater treatment in 2050: challenges ahead and future vision in a European context. Environ Sci EcoTechno l 2:100030.<https://doi.org/10.1016/j.ese.2020.100030>
- Solimeno A, Acíen FG, García J (2017) Mechanistic model for design, analysis, operation and control of microalgae cultures: calibration and application to tubular photobioreactors. Algal Res 21:236– 246. <https://doi.org/10.1016/j.algal.2016.11.023>
- Slatni I, Elberrichi FZ, Duplay J, Fardjaoui NEH, Guendouzi A, Guendouzi O, Gasmi B, Akbal F, Rekkab I (2020) Mesoporous silica synthesized from natural local kaolin as an efective adsorbent for removing of Acid Red 337 and its application in the treatment of real industrial textile effluent. Environ Sci Pollut Res Int 27:38422–38433.<https://doi.org/10.1007/s11356-020-08615-5>
- Sukla LB, Pradhan D, Subbaiah T (2019) Future prospects of microalgae in wastewater treatment. In: (Ed) The Role of Microalgae in Wastewater Treatment. Singapore: Springer pp 129–135 Doi: https://doi.org/10.1007/978-981-13-1586-2_10
- Sun H, Zhao W, Mao X, Li Y, Wu T, Chen F (2018) High-value biomass from microalgae production platforms: strategies and progress based on carbon metabolism and energy conversion. Biotechnol Biofuels 11(1):1–23. [https://doi.org/10.1186/](https://doi.org/10.1186/s13068-018-1225-6) [s13068-018-1225-6](https://doi.org/10.1186/s13068-018-1225-6)
- Sun Y, Huang Y, Liao Q, Fu Q, Zhu X (2016) Enhancement of microalgae production by embedding hollow light guides to a fat-plate photobioreactor. Bioresour Technol 207:31–38. [https://doi.org/](https://doi.org/10.1186/s13068-018-1225-6) [10.1186/s13068-018-1225-6](https://doi.org/10.1186/s13068-018-1225-6)
- Tan JS, Lee SY, Chew KW, Lam MK, Lim JW, Ho SH, Show PL (2020) A review on microalgae cultivation and harvesting, and their biomass extraction processing using ionic liquids. Bioengineered 11(1):116–129. [https://doi.org/10.1080/21655979.2020.](https://doi.org/10.1080/21655979.2020.1711626) [1711626](https://doi.org/10.1080/21655979.2020.1711626)
- Tang C, Sun B, Yu K, Shi J, Liu M, Jiang T, Huo Y, He P (2018) Environmental triggers of a Microcystis (Cyanophyceae) bloom in an artifcial lagoon of Hangzhou Bay, China. Mar Pollut Bull 135:776–782.<https://doi.org/10.1016/j.marpolbul.2018.08.005>
- Tao Y, Liu Z, Zheng J, Zhou J, He D, Ma J (2022) Microalgae production in human urine: fundamentals, opportunities, and perspectives. Front Microbiol. [https://doi.org/10.3389/fmicb.2022.](https://doi.org/10.3389/fmicb.2022.1067782) [1067782](https://doi.org/10.3389/fmicb.2022.1067782)
- Tarpeh WA, Udert KM, Nelson KL (2017) Comparing ion exchange adsorbents for nitrogen recovery from source-separated urine. Environ Sci Technol 51(4):2373–2381. [https://doi.org/10.1021/](https://doi.org/10.1021/acs.est.6b05816) [acs.est.6b05816](https://doi.org/10.1021/acs.est.6b05816)
- Thomas C, Symoneaux R, Pantin-Sohier G, Picouet P, Maître I (2020) Perceptions of Spirulina from French Consumers of Organic Products; hal-02615769; HAL Open Science: Lyon, France, 2020.
- Tilley E (2016) Cost-efectiveness and community impacts of two urine-collection programs in rural South Africa. Environ Sci: Water Res Technol 2(2):320–335. [https://doi.org/10.1039/](https://doi.org/10.1039/C5EW00237K) [C5EW00237K](https://doi.org/10.1039/C5EW00237K)
- Ting H, Haifeng L, Shanshan M, Zhang Y, Zhidan L, Na D (2017) Progress in microalgae cultivation photobioreactors and applications in wastewater treatment: a review. Int J Agric Biol 10(1):1–29. <https://doi.org/10.3965/j.ijabe.20171001.2705>
- Torres LG, Martinez M, García D, Fernández LC (2014) Three microalgae strains culture using human urine and light. J Chem Biol Phys Sci 4(5):74–80
- Tuantet K, Janssen M, Temmink H, Zeeman G, Wijfels RH, Buisman CJN (2014a) Microalgae growth on concentrated human urine. J Appl Phycol 26(1):287–297. [https://doi.org/10.1007/](https://doi.org/10.1007/s10811-013-0108-2) [s10811-013-0108-2](https://doi.org/10.1007/s10811-013-0108-2)
- Tuantet K, Temmink H, Zeeman G, Janssen M, Wijfels RH, Buisman CJ (2014b) Nutrient removal and microalgal biomass production on urine in a short light-path photobioreactor. Water Res 55:162–174. <https://doi.org/10.1016/j.watres.2014.02.027>
- Tuantet K (2015) Microalgae cultivation for nutrient recovery from human urine. PhD. Wageningen University, Wageningen
- Tuantet K, Temmink H, Zeeman G, Wijfels RH, Buisman CJ, Janssen M (2019) Optimization of algae production on urine. Algal Res 44:101667.<https://doi.org/10.1016/j.algal.2019.101667>
- Tunay D, Altinbas M, Ozkaya B (2022) Usage of source separated urine for the biodiesel production from algal biomass. Biochem Eng J. <https://doi.org/10.1016/j.bej.2022.108692>
- Udert KM, Buckley CA, Wächter M, McArdell CS, Kohn T, Strande L, Zöllig H, Fumasoli A, Oberson A, Etter B (2015) Technologies for the treatment of source-separated urine in the eThekwini municipality. Water SA 41(2):212–221. [https://doi.org/10.4314/](https://doi.org/10.4314/wsa.v41i2.06) [wsa.v41i2.06](https://doi.org/10.4314/wsa.v41i2.06)
- Udert KM, Wachter M (2012) Complete nutrient recovery from source-separated urine by nitrifcation and distillation. Water Res 46(2):453–464.<https://doi.org/10.1016/j.watres.2011.11.020>
- Viskari EL, Grobler G, Karimaki K, Gorbatova A, Vilpas R, Lehtoranta S (2018) Nitrogen recovery with source separation of human urine—preliminary results of its fertilizer potential and use in agriculture. Front Sustain Food Syst 2:32. [https://doi.org/10.](https://doi.org/10.3389/fsufs.2018.00032) [3389/fsufs.2018.00032](https://doi.org/10.3389/fsufs.2018.00032)
- Viskari EL, Lehtoranta S, Malila R (2021) Urine the nutrient recovery potential, value chain, and its sustainable management. Sanitation Value Chain 5(2):27–43.<https://doi.org/10.34416/svc.00065>
- Vogel V, Bergmann P (2018) Culture of Spirogyra sp. in a fat-panel airlift photobioreactor. Biotech 8(1):1–9. [https://doi.org/10.1007/](https://doi.org/10.1007/s13205-017-1026-9) [s13205-017-1026-9](https://doi.org/10.1007/s13205-017-1026-9)
- Volpin F, Chekli L, Phuntsho S, Cho J, Ghafour N, Vrouwenvelder JS, Shon HK (2018) Simultaneous phosphorous and nitrogen recovery from source-separated urine: a novel application for fertilizer drawn forward osmosis. Chemosphere 203:482–489. <https://doi.org/10.1016/j.chemosphere.2018.03.193>
- Volpin F, Yu H, Cho J, Lee C, Phuntsho S, Ghaffour N, Vrouwenvelder JS, Shon HK (2019) Human urine as a forward osmosis draw solution for the application of microalgae dewatering. J Hazard Mater 378:120724. [https://doi.org/10.1016/j.jhazmat.](https://doi.org/10.1016/j.jhazmat.2019.06.001) [2019.06.001](https://doi.org/10.1016/j.jhazmat.2019.06.001)
- Walter XA, Stinchcombe A, Greenman J, Ieropoulos I (2017) Urine transduction to usable energy: a modular MFC approach for smartphone and remote system charging. Appl Energy 192:575–581.<https://doi.org/10.1016/j.apenergy.2016.06.006>
- Wang B, Lan CQ, Horsman M (2012) Closed photobioreactors for production of microalgal biomasses. Biotechnol Adv 30(4):904–912. [https://doi.org/10.1016/j.biotechadv.2012.01.](https://doi.org/10.1016/j.biotechadv.2012.01.019) [019](https://doi.org/10.1016/j.biotechadv.2012.01.019)
- Wang J, Fu Z, Qiao H, Liu F (2019a) Assessment of eutrophication and water quality in the estuarine area of Lake Wuli, Lake

 $\circled{2}$ Springer

Taihu, China. Sci Total Environ 650:1392–1402. [https://doi.](https://doi.org/10.1016/j.scitotenv.2018.09.137) [org/10.1016/j.scitotenv.2018.09.137](https://doi.org/10.1016/j.scitotenv.2018.09.137)

- Wang J, Liu Q, Zhao X, Borthwick AG, Liu Y, Chen Q, Ni J (2019b) Molecular biogeography of planktonic and benthic diatoms in the Yangtze River. Microbiome 7(1):1–15. [https://doi.org/10.](https://doi.org/10.1186/s40168-019-0771-x) [1186/s40168-019-0771-x](https://doi.org/10.1186/s40168-019-0771-x)
- Wang S, Zhao S, Uzoejinwa BB, Zheng A, Wang Q, Huang J, Abomohra AEF (2020) A state-of-the-art review on dual purpose seaweeds utilization for wastewater treatment and crude bio-oil production. Energy Convers Manag 222:113253. [https://doi.](https://doi.org/10.1016/j.enconman.2020.113253) [org/10.1016/j.enconman.2020.113253](https://doi.org/10.1016/j.enconman.2020.113253)
- Wang Y, Ho SH, Cheng CL, Guo WQ, Nagarajan D, Ren NQ, Chang JS (2016) Perspectives on the feasibility of using microalgae for industrial wastewater treatment. Bioresour Technol 222:485–497. <https://doi.org/10.1016/j.biortech.2016.09.106>
- Whitton R, Ometto F, Pidou M, Jarvis P, Villa R, Jeferson B (2015) Microalgae for municipal wastewater nutrient remediation: mechanisms, reactors and outlook for tertiary treatment. Environ Technol Rev 4(1):133–148. [https://doi.org/10.1080/21622](https://doi.org/10.1080/21622515.2015.1105308) [515.2015.1105308](https://doi.org/10.1080/21622515.2015.1105308)
- Wiley P, Harris L, Reinsch S, Tozzi S, Embaye T, Clark K, Mckuin B, Kolber Z, Adams R, Kagawa H, Richardson TJ, Malinowski J, Beal C, Claxton MA, Geiger E, Rask J, Campbell JE, Trent J (2013) Microalgae cultivation using ofshore membrane enclosures for growing algae (OMEGA). J Sustain Bioenergy Syst Univ California Merced 3:18–32
- Winfeld J, Chambers LD, Rossiter J, Stinchcombe A, Walter XA, Greenman J, Ieropoulos I (2015) Fade to green: a biodegradable stack of microbial fuel cells. Chem Sus Chem 8(16):2705– 2712. <https://doi.org/10.1002/cssc.201500431>
- Xu M, Zhu S, Zhang Y, Wang H, Fan B (2019) Spatial-temporal economic analysis of modern sustainable sanitation in rural China: resource-oriented system. J Clean Prod 233:340–347. <https://doi.org/10.1016/j.jclepro.2019.06.103>
- Yadav G, Shanmugam S, Sivaramakrishnan R, Kumar D, Mathimani T, Brindhadevi K, Rajendran K (2021) Mechanism and challenges behind algae as a wastewater treatment choice for bioenergy production and beyond. Fuel 285:119093. [https://doi.](https://doi.org/10.1016/j.fuel.2020.119093) [org/10.1016/j.fuel.2020.119093](https://doi.org/10.1016/j.fuel.2020.119093)
- Yeiser M, Harris CL, Kirchoff AL, Patterson AC, Wampler JL, Zissman EN, Berseth CL (2016) Growth and tolerance of infants fed formula with a new algal source of docosahexaenoic acid: double-blind, randomized, controlled trial. Prostagland Leukot Essent Fat Acids 115:89–96. [https://doi.org/10.1016/j.plefa.](https://doi.org/10.1016/j.plefa.2016.09.001) [2016.09.001](https://doi.org/10.1016/j.plefa.2016.09.001)
- Yen HW, Hu IC, Chen CY, Nagarajan D, Chang JS (2019) Design of photobioreactors for algal cultivation. In: Biofuels from algae. Elsevier, pp 225–256 [https://doi.org/10.1016/B978-0-](https://doi.org/10.1016/B978-0-444-64192-2.00010-X) [444-64192-2.00010-X](https://doi.org/10.1016/B978-0-444-64192-2.00010-X)
- Zagklis D, Katrivesis FK, Sygouni V, Tsarouchi L, Tsigkou K, Kornaros M, Paraskeva CA (2021) Recovery of water from secondary effluent through pilot scale ultrafiltration membranes: implementation at Patras' wastewater treatment plant. Membranes 11(9):663. [https://doi.org/10.3390/membranes1](https://doi.org/10.3390/membranes11090663) [1090663](https://doi.org/10.3390/membranes11090663)
- Zamani N, Noshadi M, Amin S, Niazi A, Ghasemi Y (2012) Efect of alginate structure and microalgae immobilization method on orthophosphate removal from wastewater. J Appl Phycol 24(4):649–656. <https://doi.org/10.1007/s10811-011-9682-3>
- Zhang G, Zhang H, Zhang C, Zhang G, Yang F, Yuan G, Gao F (2013) Simultaneous nitrogen and carbon removal in a single chamber microbial fuel cell with a rotating biocathode. Process Biochem 48(5–6):893–900. [https://doi.org/10.1016/j.procbio.](https://doi.org/10.1016/j.procbio.2013.03.008) [2013.03.008](https://doi.org/10.1016/j.procbio.2013.03.008)
- Zhang S, Lim CY, Chen CL, Liu H, Wang JY (2014) Urban nutrient recovery from fresh human urine through cultivation of

Chlorella sorokiniana. J Environ Manage 145:129–136. [https://](https://doi.org/10.1016/j.jenvman.2014.06.013) doi.org/10.1016/j.jenvman.2014.06.013

- Zhao Y, Song X, Yu L, Han B, Li T, Yu X (2019) Infuence of cadmium stress on the lipid production and cadmium bioresorption by Monoraphidium sp. QLY-1. Energy Convers Manag 188:76–85. <https://doi.org/10.1016/j.enconman.2019.03.041>
- Zheng T, Ke CW (2017) Treatment of oilfeld wastewater using algal–bacterial fluidized bed reactor. Sep Sci Technol 52(13):2090–2097. [https://doi.org/10.1080/01496395.2017.](https://doi.org/10.1080/01496395.2017.1307853) [1307853](https://doi.org/10.1080/01496395.2017.1307853)
- Zhou W, Cheng Y, Li Y, Wan Y, Liu Y, Lin X, Ruan R (2012a) Novel fungal pelletization-assisted technology for algae harvesting and wastewater treatment. Appl Biochem Biotechnol 16(2):214–228. <https://doi.org/10.1007/s12010-012-9667-y>
- Zhou W, Li Y, Min M, Hu B, Zhang H, Ma X, Ruan R (2012b) Growing wastewater-born microalga Auxenochlorella protothecoides UMN280 on concentrated municipal wastewater for simultaneous nutrient removal and energy feedstock production. Appl Energy 98:433–440. [https://doi.org/10.1016/j.apene](https://doi.org/10.1016/j.apenergy.2012.04.005) [rgy.2012.04.005](https://doi.org/10.1016/j.apenergy.2012.04.005)
- Zhou W, Min M, Li Y, Hu B, Ma X, Cheng Y, Liu YH, Chen P, Ruan R (2012c) A hetero-photoautotrophic two-stage development process to improve wastewater nutrient removal and enhance algal lipid accumulation. Bioresour Technol 110:448–455. <https://doi.org/10.1016/j.biortech.2012.01.063>
- Zhou W, Wang J, Chen P, Ji C, Kang Q, Lu B, Li K, Liu J, Ruan R (2017) Bio-mitigation of carbon dioxide using microalgal systems: Advances and perspectives. Renew Sustain Energy Rev 76:1163–1175.<https://doi.org/10.1016/j.rser.2017.03.065>
- Zhou WG, Hu B, Li Y, Min M, Mohr M, Du Z, Chen P, Ruan R (2012d) Mass cultivation of microalgae on animal Wastewater: a sequential two-stage cultivation process for energy crop and omega-3-rich animal feed production. Appl Biochem Biotechnol 168(2):348–363. [https://doi.org/10.1007/](https://doi.org/10.1007/s12010-012-9779-4) [s12010-012-9779-4](https://doi.org/10.1007/s12010-012-9779-4)
- Zhou X, Li Z, Zheng T, Yan Y, Li P, Odey EA, Mang HP, Uddin SMN (2018) Review of global sanitation development. Environ Int 120:246–261.<https://doi.org/10.1016/j.envint.2018.07.047>
- Zhuang Q, Guo P, Zheng S, Lin Q, Lin Y, Wang Y, Ni Y (2018) Green synthesis of luminescent graphitic carbon nitride quantum dots from human urine and its bioimaging application. Talanta 188:35–40. [https://doi.org/10.1016/j.talanta.2018.05.](https://doi.org/10.1016/j.talanta.2018.05.060) [060](https://doi.org/10.1016/j.talanta.2018.05.060)
- Zou G, Liu Y, Zhang Q, Zhou T, Xiang S, Gu Z, Liu M (2020) Cultivation of Chlorella vulgaris in a light-receiving-plate (LRP)-enhanced raceway pond for ammonium and phosphorus removal from pretreated pig urine. Energies 13(7):1644. <https://doi.org/10.3390/en1307164>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.