#### **REVIEW**



# **Antimicrobial resistance dynamics and the one‑health strategy: a review**

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Received: 10 March 2021 / Accepted: 5 April 2021 / Published online: 15 April 2021 © The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

### **Abstract**

Antimicrobial resistance is a global threat that kills at least 75,000 people every year worldwide and causes extended hospital stays. In the coming 10 years, antimicrobial resistance is projected to have huge health and economic burden on countries, and the scarcity of available antibiotics further worsens the situation. Antimicrobial resistance results mainly from indiscriminate antibiotic usage in humans, animals and agriculture, and from the rapid emergence and dissemination of resistant pathogens. This issue is challenging for antibiotic stewardship, strict regulations on antibiotics usage, large-scale surveillance and responsible public behavior. This demands international cooperation and integrated eforts under the 'one-health' strategy. Here, we review antimicrobial resistance and the one-health strategy. We discuss the historical issue of using antibiotics. We highlight the efectiveness of hygiene in livestock rearing, careful antibiotic usage and large-scale surveillance of animals, humans and environment domains. We present strategies for mitigation of antimicrobial resistance, exemplifed by the successful ban of triclosan which induced a signifcant decline of resistant pathogens. We emphasize the benefts of the global antibiotic resistance partnership and of the one-health participation of stakeholders from public, healthcare professionals and government to mitigate antimicrobial resistance.

**Keywords** Antimicrobial resistance · One-health · Antibiotics and agriculture · Animal health · Horizontal gene transfer · Environmental surveillance

#### **Abbreviations**



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## **Introduction**

There have been many stages of discovery and development in the knowledge and status of antibiotics usage and menace of antimicrobial resistance. After the initial discovery of

Penicillin in 1928, many new antibiotics were found through intense industrial research during 1950–1970. However, regulatory constraints, long development phase, accruing fnancial inputs dampened the pharmaceutical interest into new antibiotics and fewer antibiotics were developed after the mid-1980s (Spellberg and Gilbert [2014\)](#page-11-0).

On the other hand, indiscriminate antibiotics usage led to pollution by antibiotics and/or their residues across all niches such as food, soil, aquatic water bodies and sewage (Shi et al. [2020\)](#page-11-1). These antibiotics originate from hot spots such as hospitals, pharmaceutical units, agricultural setups and livestock farms. High antibiotic concentration is linked to antimicrobial resistance often through horizontal gene transfer of antibiotic resistance determinants such as genes, transposons, integrons in deadly pathogens such as *Acinetobacter baumannii* (Gillings [2013](#page-9-0)), (Rodgers et al. [2019](#page-11-2)). Many such microbes pass through overlapping ecological niches between humans, animals and environment, thereby gaining resistance at even faster rate. Rapid emergence of resistant clinical pathogens is an imminent threat to public health, and 2.4 million people are predicted to die due to antimicrobial resistance in Europe, North America and Australia over next 30 years (Bianco et al. [2020\)](#page-8-0).

Improved international mobility, immigration and economic exchange further lead to intercontinental distribution of resistant microbial phenotypes (Church [2004](#page-9-1)). Since 1950, many studies have reported increasing incidences of antimicrobial resistance (Davies and Davies [2010](#page-9-2)) and WHO has advocated research and antibiotics usage as per guidelines (WHO [2016\)](#page-12-0). Subsequent research and attention to pharmaceutical neglect toward new antibiotics has been recognized, and efforts are directed toward aiding new antibiotic development and understanding the antimicrobial resistance landscape between animals, humans and the environment.

The interconnections and microbial exchange between ecological niches involving humans, animals and poultry, agriculture, environment and wastewater (sewage) cannot be neglected. This is frequently underscored by recognition of animal carriers and zoonotic sources during disease outbreaks (He et al. [2021](#page-10-0)). This scenario calls for adoption of a powerful emerging concept called 'one health.' It refers to a collaborative, inter-disciplinary and multi-domain approach of actions applied at local, national and inter-national scale. This collaborative approach needs to be implemented for antimicrobial resistance to tackle the omnipresence of drugresistant microbes (Fisman and Laupland [2010](#page-9-3)). The underlying principles of 'one health' help to achieve long-term sustainability by strengthening human-animal bonds, providing safe food and water, sustainable agricultural practices, surveillance of disease, antimicrobial resistance and environment, alignment of government policies and regulations for efficient communication and global outreach. Direct outcomes of this 'one-health' approach would be interdisciplinary solutions emanating from strategic education, training, prevention, diagnosis and novel therapies against antimicrobial resistance (Fig. [1](#page-1-0)). The issue of antimicrobial resistance can only be dealt through knowledge and multi-thronged mitigation approaches, and thus, we discuss the overlapping landscape of antimicrobial resistance and stakeholders in

<span id="page-1-0"></span>**Fig. 1** One-health paradigm and stakeholders. 'One-health' refers to integrative strategies in design and implementation of legislation, policies and research, which facilitates communication between multiple sectors to achieve better solutions for public health. This holistic approach can aid in integration of information from diferent stakeholders such as regulatory agencies, economists, consumers and other contributors to provide sustainable solutions against antimicrobial resistance



this review. This article is an abridged version of the book chapter on antimicrobial resistance and One-health by Singh et al. [\(2020\)](#page-11-3).

#### **Antibiotics and antimicrobial resistance**

#### **Antibiotics in humans, veterinary and agriculture**

Various antibiotics are used for the treatment of infections in humans and animals. Some of these antibiotics could be used in both animals and humans, whereas due to toxicity and/or diferent incidences of infection in humans, some of them are restricted only for use in animals (Church [2004](#page-9-1)), (Davies and Davies [2010](#page-9-2)), (Van Boeckel et al. [2015](#page-11-4)). Nontherapeutic usage of antibiotics has been found disproportionately more in animals, especially farm animals, as compared to humans because the antibiotics are added in their water and/or feed to prevent them from various infections (Cully [2014\)](#page-9-4), (Van Boeckel et al. [2015](#page-11-4)). In farm animals, antibiotics are also used at sub-therapeutic levels as a growth enhancer (Landers et al. [2012](#page-10-1)), (Chattopadhyay [2014](#page-8-1)), a procedure called 'metaphylaxis' (Butaye et al. [2003](#page-8-2)). The period of metaphylaxis may range from 2 weeks to throughout the life of animals and increases their growth rate by up to 10% (WHO [2003](#page-12-1)). All such activities lead to the direct or indirect leakage of antibiotics in the environment and possess extreme threats to the human population.

What makes it more adverse is that certain antibiotics used on farm animals are reserved for human usage to treat terminal infection or infections caused by multi-drug-resistant pathogens (MDR) (WHO [2016\)](#page-12-0). MDR pathogens have intrinsic resistance against many antibiotics, and they extensive usage of antibiotics increases their number in the farm waste, thereby creating more number of microbes showing resistance against more than one antibiotic (Tacconelli et al. [2018](#page-11-5)). According to a report published in the USA, it has been estimated that a large group of people may get afected by fluoroquinolone-resistant *Campylobacter* infections via chicken (Food and Drug Administration [2000](#page-9-5)). It has been reported from Canada and Europe, that poor hygiene and sanitation conditions lead to excessive use of antibiotics (Health Canada [2014;](#page-10-2) European Union [2015](#page-9-6)). Without compromising on economic front, antibiotic usage can be reduced largely by ensuring hygiene and routine vaccination (World Health Organization [2003](#page-12-1); Tang et al. [2019\)](#page-11-6). Under the guidelines of WHO, many countries have started good practices in their farm animals and have prohibited the use of certain antibiotics in their countries (Food and Drug Administration [2013;](#page-9-7) Health Canada [2014;](#page-10-2) European Union, [2015](#page-9-6)).

The risks associated with metaphylaxis of common classes of antibiotics used between humans and animals is clearly evident for third-generation antibiotics such as cephalosporins (WHO [2016\)](#page-12-0). Upon widespread metaphylaxis of cephalosporins in animals, resistance across all cephalosporin antibiotics developed and spread to human pathogens. Now, it has been reported that the cephalosporin dosage had to be increased in humans as compared to animals (Food and Drug Administration 2012) and two genes, extended-spectrum beta-lactamases (ESBLs) and AmpC, which both confer resistance against these antibiotics, have been found on the bacterial plasmids in a highly mobile genomic region (European Medicines Agency [2009](#page-9-8)). ESBLs are a group of highly mobile antibiotic resistance genes providing resistance against beta-lactam group of antibiotics, and it poses major health risk to humans (Rawat and Nair [2010](#page-11-7)). Resistance in one class of antibiotics can also result in emergence of resistance against other antibiotics due to co-transfer of resistance genes, owing to their proximity in the genome, through plasmids. One such example is the carbapenems, against which common human pathogens like *E. coli, Salmonella* spp. and *K. pneumoniae* have gained co-resistance after overuse of cephalosporins (Nordmann [2014\)](#page-10-3). Many studies have found similar ESBL-containing *E. coli* strains between animal, food and humans (Tadesse et al. [2017](#page-11-8)), (Odsbu et al. [2018](#page-10-4)), (Rizzo et al. [2019](#page-11-9)). The mode of dissemination of resistance genes between animals and humans has also been found to be through plasmids (de Been et al. [2014](#page-9-9)). Thus, excessive use of these antibiotics as metaphylaxis further aggravates the problem of resistant microbes in the environment (Canadian Integrated Program for Antimicrobial Resistance 2008) and urgently needs to be regulated. Many studies done in Japan, Denmark and Canada have shown the positive impact on the reduction of cephalosporin resistant microbes in food animals if it is not being used (Hiki et al. [2015](#page-10-5)), and a similar pattern was observed with the usage of fuoroquinolones in Australia (Nelson et al. [2007](#page-10-6)), (Rusu et al. [2015](#page-11-10)).

With the emergence of many multi-drug-resistant pathogens, the efectiveness of many general use antibiotics has largely reduced. Thus, certain antibiotics with high side efects in humans, such as colistin, are now being approved as last resort antibiotics against resistant pathogens. Colistin was only limited for metaphylaxis in food animals, and thus, high levels were being used. Its overuse in food animals led to many colistin-resistant pathogens and reports of horizontal transfer of colistin resistance gene (mcr-1) via plasmid (Liu et al. [2016](#page-10-7)) prompted the regulatory agencies of many countries to reduce colistin administration in food animals. Likewise, the extensive use of avoparcin, a glycopeptide antimicrobial, in food animals (Aarestrup et al. [1996](#page-8-3)) has created the problem for other candidates of the glycopeptide class of antibiotics. One such case is vancomycin (also a glycopeptide antibiotic), which is being used in case of multi-drug resistance and is now facing the problem of losing its efectiveness on humans as well as animals (Tang

et al. [2014](#page-11-11)). Altogether, careful regulations need to be put in place for the use of veterinary antibiotics especially when they are highly potent and widespread surveillance of antibiotic seepage into the environment needs to be done.

## **Antimicrobial resistance, bacteriophages, animals and public health**

The indiscriminate use of antibiotics has led to the increased prevalence of antibiotic resistance genes among pathogens (Fig. [2](#page-3-0)). In livestock, antibiotics are said to stimulate overall body development by improving digestion or preventing bacterial infections. This is postulated to be caused by suppression of unwanted intestinal bacteria which otherwise will cause infections leading to release of certain cytokines which reduces muscle mass (Liao and Nyachoti [2017](#page-10-8)), (Gonzalez Ronquillo and Angeles Hernandez [2017\)](#page-9-10). In pork industry, China's use of the antibiotics for growth is four times higher than the USA (Cully [2014](#page-9-4)). Since antibiotics are often used for metaphylaxis in high levels, they are poorly absorbed in the animal body. Most of the antibiotics are excreted in feces which leads to emergence of resistant bacteria in the animal manure. In accordance, Denmark and the European Union have forbidden the use of 11 antibiotics since 1995, causing a decrease of up to 50% in antibiotic resistance (Hollis and Ahmed [2013](#page-10-9)).

It has been observed that manure enhances antibiotic genes transfer to bacteria infecting human populations and even exposes soil to it, causing a build-up of antibioticresistant genes in the environment (Zhu et al. [2013](#page-12-2)), (Pruden et al. [2012\)](#page-11-12). These antibiotics along with metals used in livestock feed such as Cu, Zn, As, Cr, Cd, Pb and further generate a co-selection pressure on bacterial populations by hindering their growth and are positively correlated with antibiotic resistance genes (*sul*A and *sul*III) (Baker-Austin et al. [2006](#page-8-4)), (Zhang et al. [2012](#page-12-3)), (Seiler and Berendonk [2012\)](#page-11-13). Even the unculturable portion of the microbiome of niches potentially houses antibiotic-resistant genes (Enne et al. [2008](#page-9-11)), (Zhu et al. [2013\)](#page-12-2) and is speculated to act as sink for them. Further, it has been noted that human waste processed in sewage treatment plants results in less frequent antimicrobial resistance than untreated animal waste (Di Cesare et al. [2016](#page-9-12)).

Bacteriophages (bacteria-killing viruses) are an often ignored but important player in the antimicrobial resistance landscape. They carry many antibiotic resistance genes, and they can mobilize these genes similar to plasmids and integrons through horizontal gene transfer (Brabban et al. [2005](#page-8-5)). Phage-mediated transfer of resistance genes has been found responsible for conferring resistance against ampicillin in *Escherichia coli* (Colomer-Lluch et al. [2011\)](#page-9-13), multiple drugs in *Salmonella enterica* (Schmieger and Schicklmaier [1999](#page-11-14)) and *Pseudomonas aeruginosa* (Blahová et al. [2000\)](#page-8-6). Their attribute as vectors for gene transfer has been reported from many niches including sewage, which also has high numbers of resistant microbes and genes ( $qnrs$ ,  $bla_{\text{TEM}}$ ,  $bla_{\text{CTX-M}}$ ,  $bla_{\text{SHV}}$ , *mec*A, *sul*1) conferring multi-drug resistance (Colomer-Lluch et al. [2011\)](#page-9-13), (Marti et al. [2014\)](#page-10-10), (Calero-Cáceres and Muniesa [2016\)](#page-8-7). More research needs to be done on the extent of involvement of phages, but their omnipresence and high capacity for gene mobilization between microbes

<span id="page-3-0"></span>**Fig. 2** Inter-connectedness and transferability between the diferent niches. Many routes of transmission exist for exchange of antimicrobial resistance determinants such as antibiotic resistance genes (ARGs) between animals, humans and the environment. Anthropogenic activities remain at the center of all major activities related to antimicrobial resistance



make them important in animal, human and environmental niches.

Another interesting study argues that street food indirectly exposes humans to antibiotics (Campos et al. [2015](#page-8-8)), owing to compromising practices in hygiene. This also relates to storage and purchase of raw materials from farms using cheaper farm practices, unregulated antibiotic use and diseased animals with high antibiotic levels (Pham Kim et al. [2013](#page-10-11)). One striking example of feed is the use of poultry litter as protein supplement for farm animals in South Africa, which is often contaminated with residues of antibiotics to which the poultry was exposed (Soto [2013\)](#page-11-15). Food from animal origin can carry major proportions of antibiotic-resistant bacteria with about 47% of resistant *Salmonella* being isolated from meat and milk in Ethiopia (Ejo et al. [2016](#page-9-14)) and 14% of resistant *E. coli* isolated from chicken, milk and eggs in India (Laxminarayan and Chaudhury [2016](#page-10-12)). Unregulated antibiotic use can cause emergence of resistant pathogens like *Salmonella* spp., *Enterococcus* spp., *Yersinia enterocolitica, Listeria monocytogenes, Staphylococcus* spp*.* and many more (Phillips et al. [2004\)](#page-10-13) which generally have higher recombination frequency and gene transfer ability, thus indicating fecal, soil and water contamination. Also the lack of quick and efective antibiotic resistance detection techniques, complex patterns of exchange of antibiotic resistance genes, unreliable prediction tools to understand the consequences of human mortality and morbidity, and fuctuating costs of care are associated with infections induced by such pathogens (Wegener [2012](#page-12-4)).

Residues of antibiotics such as tetracycline and chloramphenicol (Cameroon, Iran, Egypt) above their maximum residue limit (European Union standards) have been detected in muscle, heart and kidney of farm chicken (Daghrir and Drogui [2013\)](#page-9-15), (Tavakoli [2015](#page-11-16)), (Guetiya Wadoum et al. [2016\)](#page-9-16). Similarly, ciprofoxacin has been detected in eggs of terminally ill birds receiving antibiotic treatment (Billah et al. [2015](#page-8-9)), quinolones inside chicken and beef in Turkey (Er et al. [2013\)](#page-9-17), amoxicillin in milk and eggs in Bangladesh (Chowdhury et al. [2015](#page-9-18)), sulfonamides and quinolones in milk in China, Malaysia and India (Cheong et al. [2010](#page-8-10)), (Zheng et al. [2013](#page-12-5)), (Kumari Anjana and Jayachandran [2017\)](#page-10-14). This exposure of antibiotics to humans can lead to neuropathological problems, drug hypersensitivity, aplastic anemia, mutagenesis, disturbance of normal gut fora and many more such complications (Lee et al. [2001](#page-10-15)), (Nisha [2008](#page-10-16)), (Beyene [2015\)](#page-8-11). European Union and other authorities have recommended a maximum residue limit for antibiotics in animal-derived food, but it is quite difficult to ascertain their safe level, thus monitoring antibiotic use and their clearance period from animal becomes crucial (Codex Alimentarius [2012\)](#page-9-19). Many such guidelines on the use of medically important antibiotics in food-producing animals have been recommended by WHO in 20[1](#page-4-0)6 (Table 1) (Aidara-Kane et al. [2018\)](#page-8-12). The historical linkage of antibiotic and antimicrobial resistance in the modern age could not be overstated, and direct effects of antibiotics and interconnected microbial entities on human health must be considered when probing the emergence and spread of antimicrobial resistance.

#### **Antimicrobial resistance in soil and water bodies**

Soil is a complex ecological niche and acts as source and sink for physical exchange between the living and nonliving components of the earth. Naturally, it also serves as a reservoir for antibiotic resistance genes and determinants coming from various sources (Fig. [1,](#page-1-0) [2](#page-3-0)) (Monier et al. [2011\)](#page-10-17). Metagenomic studies of soil have shown that many antibiotic resistance genes exist which are yet to be fully characterized (Riesenfeld et al. [2004](#page-11-17)). Agriculture land may receive antibiotics washed off from other sources, but manure-supplemented soil especially with inputs from

<span id="page-4-0"></span>**Table 1** WHO recommendations on the use of medically important antimicrobials in food-producing animals

1 The Guideline Development Group (GDG) recommends an overall reduction in use of all classes of medically important antimicrobials in food-producing animals

2 The GDG recommends complete restriction of use of all classes of medically important antimicrobials in food-producing animals for growth promotion

- <sup>3</sup> \*The GDG recommends complete restriction of use of all classes of medically important antimicrobials in food-producing animals for prevention of infectious diseases that have not yet been clinically diagnosed
- 4a **†**The GDG suggests that antimicrobials classifed as critically important for human medicine should not be used for control of the dissemination of a clinically diagnosed infectious disease identifed within a group of food-producing animals
- 4b **†**The GDG suggests that antimicrobials classifed as critically important for human medicine should not be used for treatment of food-producing animals with a clinically diagnosed infectious disease

**<sup>\*</sup>** Specifc considerations: when a veterinary professional judges that there is a high risk of spread of a particular infectious disease, use of antimicrobials for disease prevention is justifed, if such a judgment is made on the basis of recent culture and sensitivity testing results

**<sup>†</sup>**To prevent harm to animal health and welfare, exceptions to recommendations 4a and 4b can be made when, in the judgment of veterinary professionals, bacterial culture and sensitivity results demonstrate that the selected drug is the only treatment option

animal farms often contains high concentration of antibiotics (Marti et al. [2013](#page-10-18)), such as tetracycline residues and resistant microbes (Zhu et al. [2013](#page-12-2)). Antimicrobial resistance in soil is further aggravated if improperly treated wastewater which is a nutrient-rich niche for microbes is used for irrigation (Chee-Sanford et al. [2009](#page-8-13); Oluyege and C [2015\)](#page-10-19). Many antibiotics and resistance genes such as the  $bla_{CTX-M}$  gene (confers resistance against cefotaxime) have been reported from soil near pig farms and even rice felds (Xiao et al. [2016\)](#page-12-6).

The waste generated from farms, manures, irrigation water, drug manufacturing plants, hospitals, humans and antibiotic-contaminated ground surfaces often ends up in agricultural land which increases the level of antibiotics (Rodgers et al. [2019\)](#page-11-2). Depending upon class of antibiotics, they degrade at diferent rates and thereby aggravate resistance against antibiotics (Du and Liu [2012\)](#page-9-20) such as ampicillin, tetracycline, sulfamethoxazole, nalidixic acid and chloramphenicol (Carballo et al. [2013\)](#page-8-14). The vegetables and fruits from these farming lands often pick up the antibiotics and resistant microbes present in the soil and pose risk of transmission of antimicrobial resistance to humans during transport and storage (Beuchat [2002](#page-8-15)), (Kumar et al. [2005](#page-10-20)). Although data for direct use of antibiotics in agriculture is limited, inputs from aforementioned antimicrobial resistance sources are often ignored and there is hardly any surveillance for resistance in agricultural land.

Water bodies such as freshwater lakes, rivers, artifcial water systems and sea also act as dynamic reservoirs for antibiotic-resistant bacteria, antibiotic resistance genes and antibiotics owing to multiple edaphic and biotic factors (Ding and He [2010](#page-9-21)), (Cizmas et al. [2015](#page-9-22)) (Khan et al. [2019](#page-10-21)). Many resistance determinants such as ESBLs (*bla*<sub>CTX-M-15</sub>,  $bla$   $_{\text{CTX-M-15}}$   $bla_{\text{OXA-58}}$ ) were found from rivers and drinking water sources, which exposes a large population to resistant microbes (Cacace et al. [2019\)](#page-8-16). This is particularly worsened if animal farms, pharmaceutical plants, manure storages, wastewater treatment plants are in close vicinity of these water bodies (Kümmerer [2009](#page-10-22)). Very diverse and high numbers of MDR strains have often been reported from ground water and rivers across the world (Mulamattathil et al. [2014](#page-10-23); (Wahome [2013\)](#page-12-7). Certain mass gathering events such as religious congregations, sports, water parks, beach sports near water bodies witness a huge infux of resistant microbes such as *Enterobacter, Serratia, Klebsiella, Citrobacter* and *Pantoea* and inter-mixing of large populations in such hot spots also facilitates the spread of antimicrobial resistance (Khan, Memish, et al. [2010;](#page-10-24) Jani et al. [2018](#page-10-25)). These reports indicate toward the enormous efects of the antimicrobial resistance in the soil and water bodies and the role of environment as a source–sink pair for antimicrobial resistance. This demands for careful surveillance, strategic management of public events for efective control over spread of antimicrobial resistance.

#### **Antimicrobial resistance and humans**

Antibiotic consumption varies across the world owing to diferences in incidences of infectious diseases, living standards, community hygiene and often broad-spectrum antibiotics are unnecessarily prescribed. Although new treatments alternatives such as use of probiotics (Hatakka and Saxelin [2008](#page-10-26)), probiotics surface proteins such as mucus binding protein (Singh et al. [2018](#page-11-18)) (Singh et al., [2017](#page-11-19)), fbronectin binding protein (Bisht et al. [2018\)](#page-8-17) or oligosaccharides (Anand et al. [2018\)](#page-8-18) have been proposed, they are still far from being commercial therapeutic success. This leaves fewer options to treat MDR strains, and extensive antibiotic stewardship of health professionals is needed. Antibiotic stewardship approach can be further augmented by increasing awareness in the general public about responsible usage of antibiotics. Humans can potentially act as most aggressive vectors for transmission of antimicrobial resistance, given their omnipresence, long distance travel, socializing and work. Human activities around antimicrobial resistance hot spots such as farms, drug manufacturers, hospitals, further load them with resistant microbes. Resistant pathogens can afect humans directly (Scallan et al. [2011\)](#page-11-20), use them as transmission vectors (Spoor et al. [2013\)](#page-11-21) and also utilize humans as niche for example 'human gut' for the exchange of antibiotic resistance genes among microbes (Willems et al. [2011](#page-12-8)). It is correctly reasoned that antimicrobial resistance in humans is linked to high exposure of antibiotics either by drugs or contaminated food or water. The antibiotic prescription pattern, market dynamics and disease state of patients decide antimicrobial resistance in humans, which directly affects patient's amicability for vital invasive procedures in cancer and transplantation (Friedman et al. [2016\)](#page-9-23).

Many nations have taken steps to curb indiscriminate antibiotic usage at both pre-prescription and post-prescription levels, by strengthening public health and structured antibiotic stewardship (Garnacho-Montero et al. [2015\)](#page-9-24) (Dyar et al. [2017](#page-9-25)). This has been further augmented by strict laws and regulations on the sale without prescription and overthe-counter antibiotics in many countries including Thailand, Mexico and Brazil (Santa-Ana-Tellez et al. [2013](#page-11-22)), (Holloway et al. [2017](#page-10-27)). Similar strides have been initiated in India (Laxminarayan and Chaudhury [2016\)](#page-10-12) and South Africa (Gelband and Laxminarayan [2015](#page-9-26)) for strengthening of antibiotic stewardship program, tight regulatory control over antibiotics usage and data register of antibiotics sale (Masterton [2011\)](#page-10-28), (Valsamatzi-Panagiotou et al. [2021\)](#page-11-23). The direct effects of anthropogenic activities and fast transfer of antimicrobial resistance through food and drinking water to humans are easily perceptible. Apart from strategic antibiotic stewardship in hospitals, there is a need to adopt safe handling practices of antibiotic-polluted environmental samples originating from soil, water, etc.

## **Prevention and regulation of antimicrobial resistance**

The issue of antimicrobial resistance needs to be tackled both at prevention and mitigation levels, which needs awareness in general public, tight regulatory measures and responsible antibiotics usage. Sensitization of the public toward antimicrobials could be done through radio, television and awareness campaigns by non-governmental organizations and can result in success stories such as of 'triclosan.' Earlier, triclosan was a widely used antimicrobial in many countries, but it was banned for use when its role in antimicrobial resistance became public and public pressure mounted on the government (McGinley [2016](#page-10-29)). Healthcare systems could beneft from antibiotic stewardships, identifcation of critical control points and strict adherence to regulatory measures (Uchil et al. [2014](#page-11-24)). A major preventive measure to mitigate antimicrobial resistance is surveillance, further augmented by use of high-throughput technologies such as next-generation DNA sequencing and inclusive classifcation systems such as Rescon (resistance readiness condition), which links severity and spread potential of antimicrobial resistance (Martínez et al. [2015\)](#page-10-30). This applies directly to resistance determinants and mobilizing elements like plasmids, transposons and integrons and also to other environmental stress factors like heavy metals, temperature (Vorholt [2012](#page-12-9)). Other omics technologies further resolve information about antibiotics levels, mobile genetic elements, resistance genes and their expression patterns coupled with actual antibiotic residues present in the niche (Spicknall et al. [2013\)](#page-11-25).

These approaches are important to identify patterns and overlapping ecological factors involved in emergence and spread of antimicrobial resistance including those due to bioflms, phyllosphere and water bodies (Calero-Cáceres et al. [2014\)](#page-8-19). Overall, this helps in precise statistical prediction of abundance and mobilization of resistant genes and accessory genetic elements, spread between humans, animals and environment and predictive model to aid in the design of mitigation strategies and critical regulatory policies against antimicrobial resistance. These surveillance measures are not efective in standalone countries, and thus, inter-continental partnerships between regulatory agencies of diferent countries are being strengthened (Rogers Van Katwyk et al. [2020](#page-11-26)). Some examples are GARP (Global Antibiotic Resistance Partnership)—(Ganguly et al. [2011\)](#page-9-27), (Leung et al. [2011\)](#page-10-31), INSAR-India, NARMS-USA,

ESAC-EU, SASCM-South Africa, PAHO (Pan American Health Organization) and EARS-Net (European Resistance Surveillance Network), GLASS – WHO. All of these networks focus on information exchange and partnership in research and action for efficient tackling of antimicrobial resistance (Berendonk et al. [2015\)](#page-8-20). These preventive measures need a boost from another front, which is the development of novel and potent antibiotics augmented with improved drug delivery strategies to bypass bioflms (Singh et al. [2021](#page-11-27)). Reducing the mass usage of antibiotics by imposing extra charges on non-human antibiotic usage and strict regulations to encourage raising animals in clean, hygienic conditions would also help the overall effort (Hollis and Ahmed  $2013$ ). However, all such efforts need to be integrated under 'One-Health' approach across the world with further consideration of other ecological factors and niche diversity to customize the prevention and mitigation plan against antimicrobial resistance. The efectiveness of highthroughput surveillance and public awareness is paramount, and we need to consider the interplay of ecological factors in the antimicrobial resistance landscape. The discernible need for worldwide networks to fght against antimicrobial resistance is also a pertinent concern, and recent efforts by many countries need to be continued and strengthened.

## **Antimicrobial resistance and the one‑health approach**

The 'one-health' concept offers multiple benefits in systematic dealing with the menace of antimicrobial resistance. A big concern in the antimicrobial resistance issue is the acquisition of resistance by human pathogens from their non-pathogenic relatives in the environment (Forsberg et al. [2012](#page-9-28)). There are many direct evidences for human–animal–environment interrelatedness, and inclusive strategies such as 'One health' have to be adopted for effective control and mitigation of antimicrobial resistance. The 'one-health' approach is directly benefcial for humans when applied to surveillance network and public awareness programs where it facilitates precise understanding of the pattern and active dynamics of antimicrobial resistance. For example, documentation of resistant microbes from human origin in an area provides ideas about endemic patterns of such resistant microbes (Critchley and Karlowsky [2004\)](#page-9-29). This information could be utilized for prioritizing antibiotic susceptibility screens for infection in incoming patients and would promote evidencebased antibiotic prescription by physicians. Furthermore, the sensitized physicians themselves can encourage patients to adopt good hygiene practices, safe disposal of animal waste and general behavior during zoonotic disease outbreaks, thereby leading to low infections and robust addressal of public health issues.

Although not considered as measure of antimicrobial resistance, the antibiotics and their degraded products largely drive resistance in all niches (Subbiah et al. [2016\)](#page-11-28) and are sometimes responsible for activating bioflm formation, motility and stress response in microbes (Romero et al. [2011\)](#page-11-29). There are myriad types of applicable regulations and stakeholders in areas related to human, animals and environmental health, which handle the issue of antimicrobial resistance with diferent levels of seriousness. The regulation on use of last resort antibiotics (for humans) such as carbapenems and colistin in agriculture settings is very relaxed and allows mass level usage (Pereko et al. [2016](#page-10-32)). Less regulatory oversight into agricultural systems and their neglect toward recognizing it as reservoir of antimicrobial resistance further worsens the situation. The risks estimated due to pathogens arising from these domains directly afect humans involved in handling agriculture products during production, transport and even the disposal of agricultural wastes. Careless antibiotic prescription to animals without giving any consideration for resistance against carbapenems and colistin in case of veterinary pathogens still ends up being part of the animal–human–environment interplay and aggravates the development of resistance. The antibiotic administration pattern at individual animal and herd levels also difers, and integrated approaches to understand longterm effects in the context of antimicrobial resistance must be considered.

Across many developing nations, hunting of wild animals for food also referred as 'bush meat' has been linked with infections including, e.g., Ebola virus (Peterson [2003](#page-10-33)). Low vaccination, frequent mixing of animal–human habitats and poor hygiene in such countries further drive infections and concomitantly complicate the issue of antimicrobial resistance. Such situations have prompted many agencies such as WHO and UN-FAO (United Nations Food and Agricultural Organization) to join hands and design strategies using the 'one-health' approach to deal with endemic diseases such as echinococcosis, bovine tuberculosis, rabies and brucellosis in Africa while considering animal interactions (Aidara-Kane et al. 2018). Similar strides have been initiated by UN-FAO and WHO to tackle the risk of avian infuenza (infuenza virus A) in humans by collaborative eforts of veterinarians and physicians (OFFLU-OIE/FAO 2019).

Given the multidimensional issue of antimicrobial resistance and overlapping spheres of its sources, treatment regimen and control of regulations demand inclusive mitigation under the 'one-health' framework. This framework collates information from all stakeholders and flls knowledge gaps (Fisman and Laupland [2010\)](#page-9-3) to compare regulatory standards for resistant strains isolated from human, animal and environmental sources. This facilitates clarity in mitigation strategy and design of logicbased risk mitigation policies for precise administrative and structural support in dealing with antimicrobial resistance (McLain et al. [2016\)](#page-10-29). The one-health model can efectively mitigate the issue of antimicrobial resistance and is equally applicable to developed, developing and under-developed countries (Gebreyes et al. [2014](#page-9-30)) which is necessary for a sustainable control of mankind over antimicrobial resistance. The overreaching arc of 'one health' to provide efective control over the confounding factors of antimicrobial resistance is widely recognized. It is prudent to work within the integrated framework of 'one health' and mitigate antimicrobial resistance in a sustainable and long-term fashion.

## **Conclusion**

Numerous studies have established overlapping interactions between humans, animals and the environment, especially in the context of emergence and spread of antimicrobial resistance. Frequent outbreaks of zoonotic diseases and their clear association with both environmental and human domains underscore their interwoven relationship. Clinicians, veterinarians and regulatory agencies have achieved wonderful milestones in infectious diseases, but quantifable and integrated measures against antimicrobial resistance between human, animals and environment are largely missing. Due to this, even though the link between these ecological domains is obvious, only few predictive models exist which could inform about their source–sink–reservoir relationships during most disease outbreaks. A big reason for this knowledge gap is due to focus on human research and complete ignorance of other stakeholders spanning the animal, plant and environmental niches. This ignorance has been rampant at almost all levels including relaxed regulations, indiscriminate antibiotic usage, surveillance, control and research.

Recent studies on the rapid rate of exchange of antimicrobial resistance determinants between these spheres of life further underscore futility of all domain-limited eforts and demands inter-sectoral focus for tackling antimicrobial resistance. Recent research has shown that strict regulatory control, antibiotic stewardship, novel antibiotic development and widespread surveillance have to be adopted. The multi-domain nature of the issue of antimicrobial resistance demands its handling through an inclusive approach called 'one health,' which provides integrated solutions with multisectoral representation at the level of surveillance, detection and targeted mitigation. This deals comprehensively with the biotic and abiotic confounding factors involved in antimicrobial resistance and provides sustainable solutions. One-health approaches are useful for all countries at diferent economic levels and are particularly suitable for efective implementation in low- and middle-income countries that often tend to have high disease burden and worse situations of antimicrobial resistance.

**Authors' contributions** *KSS, SA and SD* wrote the manuscript. *JKS* made the fgures. *WB* and *YS* edited the manuscript with comments and changes as and when required.

**Funding** No funding from any source.

**Availability of data material** All data have been assessed from open/ public domain, and no data/material needs to be made available from our side.

**Code availability** Not applicable.

#### **Declarations**

**Conflict of interest** All authors declare no confict/competing interest.

**Ethical approval** Not applicable.

**Consent participate** All authors have given consent for participation, if applicable.

**Consent publication** All authors have given consent for publication.

#### **References**

- <span id="page-8-3"></span>Aarestrup FM, Ahrens P, Madsen M, Pallesen LV, Poulsen RL, Westh H (1996) Glycopeptide susceptibility among Danish Enterococcus faecium and Enterococcus faecalis isolates of animal and human origin and PCR identifcation of genes within the VanA cluster. Antimicrob Agents Chemother 40(8):1938–1940.<https://doi.org/10.1128/AAC.40.8.1938>
- <span id="page-8-12"></span>Aidara-Kane A, Angulo FJ, Conly JM, Minato Y, Silbergeld EK, McEwen SA, Collignon PJ, Balkhy H, Collignon P, Conly J, Friedman C, Hollis A, Kariuki S, Kwak HS, McEwen S, Moulin G, Ngandjio A, Rollin B, Rossi F, … for the WHO Guideline Development Group (2018) World Health Organization (WHO) guidelines on use of medically important antimicrobials in food-producing animals. Antimicro Resist Infect Control *7*(1), 7[.https://doi.org/10.1186/s13756-017-0294-9](https://doi.org/10.1186/s13756-017-0294-9)
- <span id="page-8-18"></span>Anand S, Mandal S, Singh KS, Patil P, Tomar SK (2018) Synbiotic combination of Lactobacillus rhamnosus NCDC 298 and short chain fructooligosaccharides prevents enterotoxigenic Escherichia coli infection. LWT 98:329–334. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.lwt.2018.08.061) [lwt.2018.08.061](https://doi.org/10.1016/j.lwt.2018.08.061)
- <span id="page-8-4"></span>Baker-Austin C, Wright MS, Stepanauskas R, McArthur JV (2006) Co-selection of antibiotic and metal resistance. Trends Microbiol 14(4):176–182. <https://doi.org/10.1016/j.tim.2006.02.006>
- <span id="page-8-20"></span>Berendonk TU, Manaia CM, Merlin C, Fatta-Kassinos D, Cytryn E, Walsh F, Bürgmann H, Sørum H, Norström M, Pons M-N, Kreuzinger N, Huovinen P, Stefani S, Schwartz T, Kisand V, Baquero F, Martinez JL (2015) Tackling antibiotic resistance: The environmental framework. Nat Rev Microbiol 13(5):310– 317.<https://doi.org/10.1038/nrmicro3439>
- <span id="page-8-15"></span>Beuchat LR (2002) Ecological factors infuencing survival and growth of human pathogens on raw fruits and vegetables. Microbes

Infect 4(4):413–423. [https://doi.org/10.1016/S1286-4579\(02\)](https://doi.org/10.1016/S1286-4579(02)01555-1) [01555-1](https://doi.org/10.1016/S1286-4579(02)01555-1)

- <span id="page-8-11"></span>Beyene T (2015) Veterinary drug residues in food-animal products: its risk factors and potential efects on public health. J Vet Sci Tech 07(01). <https://doi.org/10.4172/2157-7579.1000285>
- <span id="page-8-0"></span>Bianco A, Licata F, Zucco R, Papadopoli R, Pavia M (2020) Knowledge and practices regarding antibiotics use. Evol Med Public Health 2020(1):129–138. <https://doi.org/10.1093/emph/eoaa028>
- <span id="page-8-9"></span>Billah MdM, Rana SMM, Hossain MS, Ahamed SK, Banik S, Hasan M (2015) Ciprofoxacin residue and their impact on biomolecules in eggs of laying hens following oral administration. Int J Food Contam 2(1):13. <https://doi.org/10.1186/s40550-015-0019-x>
- <span id="page-8-17"></span>Bisht S, Singh KS, Choudhary R, Kumar S, Grover S, Mohanty AK, Pande V, Kaushik JK (2018) Expression of fbronectin-binding protein of L. acidophilus NCFM and in vitro refolding to adhesion capable native-like protein from inclusion bodies. Protein Expr Purif 145:7–13.<https://doi.org/10.1016/j.pep.2017.11.007>
- <span id="page-8-6"></span>Blahová J, Králiková K, Krcméry V, Jezek P (2000) Low-Frequency transduction of imipenem resistance and high-frequency transduction of ceftazidime and aztreonam resistance by the bacteriophage AP-151 isolated from a Pseudomonas aeruginosa strain. J Chemother (Florence, Italy) 12(6):482–486. [https://doi.org/10.](https://doi.org/10.1179/joc.2000.12.6.482) [1179/joc.2000.12.6.482](https://doi.org/10.1179/joc.2000.12.6.482)
- <span id="page-8-5"></span>Brabban Ad, Hite E, Callaway Tr (2005) Evolution of foodborne pathogens via temperate bacteriophage-mediated gene transfer. Foodborne Pathog Dis 2(4):287–303. [https://doi.org/10.1089/](https://doi.org/10.1089/fpd.2005.2.287) [fpd.2005.2.287](https://doi.org/10.1089/fpd.2005.2.287)
- <span id="page-8-2"></span>Butaye P, Devriese LA, Haesebrouck F (2003) Antimicrobial Growth Promoters Used in Animal Feed: Effects of Less Well Known Antibiotics on Gram-Positive Bacteria. Clin Microbiol Rev 16(2):175–188. <https://doi.org/10.1128/CMR.16.2.175-188.2003>
- <span id="page-8-16"></span>Cacace D, Fatta-Kassinos D, Manaia CM, Cytryn E, Kreuzinger N, Rizzo L, Karaolia P, Schwartz T, Alexander J, Merlin C, Garelick H, Schmitt H, de Vries D, Schwermer CU, Meric S, Ozkal CB, Pons M-N, Kneis D, Berendonk TU (2019) Antibiotic resistance genes in treated wastewater and in the receiving water bodies: A pan-European survey of urban settings. Water Res 162:320–330. <https://doi.org/10.1016/j.watres.2019.06.039>
- <span id="page-8-19"></span>Calero-Cáceres W, Melgarejo A, Colomer-Lluch M, Stoll C, Lucena F, Jofre J, Muniesa M (2014) Sludge As a Potential Important Source of Antibiotic Resistance Genes in Both the Bacterial and Bacteriophage Fractions. Environ Sci Technol 48(13):7602– 7611.<https://doi.org/10.1021/es501851s>
- <span id="page-8-7"></span>Calero-Cáceres W, Muniesa M (2016) Persistence of naturally occurring antibiotic resistance genes in the bacteria and bacteriophage fractions of wastewater. Water Res 95:11–18. [https://doi.org/10.](https://doi.org/10.1016/j.watres.2016.03.006) [1016/j.watres.2016.03.006](https://doi.org/10.1016/j.watres.2016.03.006)
- <span id="page-8-8"></span>Campos J, Gil J, Mourão J, Peixe L, Antunes P (2015) Ready-to-eat street-vended food as a potential vehicle of bacterial pathogens and antimicrobial resistance: an exploratory study in Porto region, Portugal. Int J Food Microbiol 206:1–6. [https://doi.org/](https://doi.org/10.1016/j.ijfoodmicro.2015.04.016) [10.1016/j.ijfoodmicro.2015.04.016](https://doi.org/10.1016/j.ijfoodmicro.2015.04.016)
- <span id="page-8-14"></span>Carballo M, Esperón F, Sacristán C, González M, Vázquez B, Aguayo S, Torre A de la (2013) Occurrence of tetracycline residues and antimicrobial resistance in gram negative bacteria isolates from cattle farms in Spain. <https://doi.org/10.4236/abb.2013.42A040>
- <span id="page-8-1"></span>Chattopadhyay MK (2014) Use of antibiotics as feed additives: a burning question. Front Microbio 5[.https://doi.org/10.3389/fmicb.](https://doi.org/10.3389/fmicb.2014.00334) [2014.00334](https://doi.org/10.3389/fmicb.2014.00334)
- <span id="page-8-13"></span>Chee-Sanford JC, Mackie RI, Koike S, Krapac IG, Lin Y-F, Yannarell AC, Maxwell S, Aminov RI (2009) Fate and transport of antibiotic residues and antibiotic resistance genes following land application of manure waste. J Environ Qual 38(3):1086–1108. <https://doi.org/10.2134/jeq2008.0128>
- <span id="page-8-10"></span>Cheong CK, Parvaneh H, Selamat J, Rashedi IFM (2010) Sulfonamides determination in chicken meat products from Malaysia. [https://](https://www.semanticscholar.org/paper/Sulfonamides-determination-in-chicken-meat-products-Cheong-Parvaneh/ba72a9f483d18e6fd9e9563149cc15dd25c9dbe5)

[www.semanticscholar.org/paper/Sulfonamides-determination](https://www.semanticscholar.org/paper/Sulfonamides-determination-in-chicken-meat-products-Cheong-Parvaneh/ba72a9f483d18e6fd9e9563149cc15dd25c9dbe5)[in-chicken-meat-products-Cheong-Parvaneh/ba72a9f483d18e6](https://www.semanticscholar.org/paper/Sulfonamides-determination-in-chicken-meat-products-Cheong-Parvaneh/ba72a9f483d18e6fd9e9563149cc15dd25c9dbe5) [fd9e9563149cc15dd25c9dbe5](https://www.semanticscholar.org/paper/Sulfonamides-determination-in-chicken-meat-products-Cheong-Parvaneh/ba72a9f483d18e6fd9e9563149cc15dd25c9dbe5)

- <span id="page-9-18"></span>Chowdhury S, Hassan MM, Alam M, Sattar S, Bari Md. S, Saifuddin AKM, Hoque Md A (2015) Antibiotic residues in milk and eggs of commercial and local farms at Chittagong, Bangladesh. Veterinary World, 8(4), 467–471. [https://doi.org/10.14202/vetwo](https://doi.org/10.14202/vetworld.2015.467-471) [rld.2015.467-471](https://doi.org/10.14202/vetworld.2015.467-471)
- <span id="page-9-1"></span>Church DL (2004) Major factors afecting the emergence and reemergence of infectious diseases. Clin Lab Med 24(3):559–586. <https://doi.org/10.1016/j.cll.2004.05.008>
- <span id="page-9-22"></span>Cizmas L, Sharma VK, Gray CM, McDonald TJ (2015) Pharmaceuticals and personal care products in waters: Occurrence, toxicity, and risk. Environ Chem Lett 13(4):381–394. [https://doi.org/10.](https://doi.org/10.1007/s10311-015-0524-4) [1007/s10311-015-0524-4](https://doi.org/10.1007/s10311-015-0524-4)
- <span id="page-9-19"></span>Codex Alimentarius Commission (2012) Maximum Residue Limits for Veterinary Drugs in Foods; Updated as at the 35th Session of the Codex Alimentarius Commission, CAC/MRL, vol 2. Codex Alimentarius Commission, Fribourg, Switzerland, pp 1–40
- <span id="page-9-13"></span>Colomer-Lluch M, Jofre J, Muniesa M (2011) Antibiotic resistance genes in the bacteriophage DNA fraction of environmental samples. PLoS ONE, 6(3). [https://doi.org/10.1371/journal.pone.](https://doi.org/10.1371/journal.pone.0017549) [0017549](https://doi.org/10.1371/journal.pone.0017549)
- <span id="page-9-29"></span>Critchley IA, Karlowsky JA (2004) Optimal use of antibiotic resistance surveillance systems. Clin Microbiol Infect 10(6):502–511. <https://doi.org/10.1111/j.1469-0691.2004.00911.x>
- <span id="page-9-4"></span>Cully M (2014) Public health: the politics of antibiotics. Nature 509(7498):S16–S17.<https://doi.org/10.1038/509S16a>
- <span id="page-9-15"></span>Daghrir R, Drogui P (2013) Tetracycline antibiotics in the environment: a review. Environ Chem Lett 11(3):209–227. [https://doi.org/10.](https://doi.org/10.1007/s10311-013-0404-8) [1007/s10311-013-0404-8](https://doi.org/10.1007/s10311-013-0404-8)
- <span id="page-9-2"></span>Davies J, Davies D (2010) Origins and evolution of antibiotic resistance. Microbiol Mol Biol Rev 74(3):417–433. [https://doi.org/10.](https://doi.org/10.1128/MMBR.00016-10) [1128/MMBR.00016-10](https://doi.org/10.1128/MMBR.00016-10)
- <span id="page-9-9"></span>de Been M, Lanza VF, de Toro M, Scharringa J, Dohmen W, Du Y, Hu J, Lei Y, Li N, Tooming-Klunderud A, Heederik DJJ, Fluit AC, Bonten MJM, Willems RJL, de la Cruz F, van Schaik W (2014) Dissemination of cephalosporin resistance genes between escherichia coli strains from farm animals and humans by specifc plasmid lineages. PLoS Genetics, 10(12). [https://doi.org/10.](https://doi.org/10.1371/journal.pgen.1004776) [1371/journal.pgen.1004776](https://doi.org/10.1371/journal.pgen.1004776)
- <span id="page-9-12"></span>Di Cesare A, Eckert EM, D'Urso S, Bertoni R, Gillan DC, Wattiez R, Corno G (2016) Co-occurrence of integrase 1, antibiotic and heavy metal resistance genes in municipal wastewater treatment plants. Water Res 94:208–214. [https://doi.org/10.1016/j.watres.](https://doi.org/10.1016/j.watres.2016.02.049) [2016.02.049](https://doi.org/10.1016/j.watres.2016.02.049)
- <span id="page-9-21"></span>Ding C, He J (2010) Effect of antibiotics in the environment on microbial populations. Appl Microbiol Biotechnol 87(3):925–941. <https://doi.org/10.1007/s00253-010-2649-5>
- <span id="page-9-20"></span>Du L, Liu W (2012) Occurrence, fate, and ecotoxicity of antibiotics in agro-ecosystems: a review. Agron Sustain Develop 32(2):309– 327. <https://doi.org/10.1007/s13593-011-0062-9>
- <span id="page-9-25"></span>Dyar OJ, Huttner B, Schouten J, Pulcini C (2017) What is antimicrobial stewardship? Clin Microbiol Infect 23(11):793–798. [https://doi.](https://doi.org/10.1016/j.cmi.2017.08.026) [org/10.1016/j.cmi.2017.08.026](https://doi.org/10.1016/j.cmi.2017.08.026)
- <span id="page-9-14"></span>Ejo M, Garedew L, Alebachew Z, Worku W (2016) Prevalence and antimicrobial resistance of salmonella isolated from animal-origin food items in gondar. Ethiopia BioMed Res Int 2016:4290506.<https://doi.org/10.1155/2016/4290506>
- <span id="page-9-11"></span>Enne VI, Cassar C, Sprigings K, Woodward MJ, Bennett PM (2008) A high prevalence of antimicrobial resistant Escherichia coli isolated from pigs and a low prevalence of antimicrobial resistant E. coli from cattle and sheep in Great Britain at slaughter. FEMS Microbiol Lett 278(2), 193–199. [https://doi.org/10.1111/j.1574-](https://doi.org/10.1111/j.1574-6968.2007.00991.x) [6968.2007.00991.x](https://doi.org/10.1111/j.1574-6968.2007.00991.x)
- <span id="page-9-17"></span>Er B, Onurdağ FK, Demirhan B, Özgacar SÖ, Öktem AB, Abbasoğlu U (2013) Screening of quinolone antibiotic residues in chicken meat and beef sold in the markets of Ankara Turkey. Poultry Sci 92(8):2212–2215. <https://doi.org/10.3382/ps.2013-03072>
- <span id="page-9-8"></span>European Medicines Agency (EMA) (2009) Revised refection paper on the use of 3rd and 4th generation cephalosporins in food producing animals in the European Union: development of resistance and impact on human and animal health. EMA, London, United Kingdom. EMEA/CVMP/SAGAM/81730/2006-Rev.1
- <span id="page-9-6"></span>European Union (EU) (2015) Guidelines for the prudent use of antimicrobials in veterinary medicine (2015/C 299/04). Official Journal of the European Union 11.9.2015. C 299/7-26
- <span id="page-9-3"></span>Fisman DN, Laupland KB (2010) The 'One Health' paradigm: time for infectious diseases clinicians to take note? Canadian J Infect Dis Med Microbiol 21(3):111–114
- <span id="page-9-5"></span>Food and Drug Administration. Center for Veterinary Medicine (2000) The Human Health Impact of Fluoroquinolone-Resistant Campylobacter Attributed to the Consumption of Chicken. Food Drug Admin, Washington, DC
- <span id="page-9-7"></span>Food and Drug Administration (FDA) (2013) New animal drugs and new animal drug combination products administered in or on medicated feed ordrinking water of food-producing animals: recommendations for drug sponsors for voluntarily aligning product use conditions with GFI #209. U.S. Department of Health and Human Services, Washington, DC
- <span id="page-9-28"></span>Forsberg KJ, Reyes A, Wang B, Selleck EM, Sommer MOA, Dantas G (2012) The Shared Antibiotic Resistome of Soil Bacteria andHuman Pathogens. Science 337(6098):1107–1111. [https://doi.org/](https://doi.org/10.1126/science.1220761) [10.1126/science.1220761](https://doi.org/10.1126/science.1220761)
- <span id="page-9-23"></span>Friedman ND, Temkin E, Carmeli Y (2016) The negative impact of antibiotic resistance. Clin Microbiol Infect 22(5):416–422. <https://doi.org/10.1016/j.cmi.2015.12.002>
- <span id="page-9-27"></span>Ganguly NK, Arora NK, Chandy SJ, Fairoze MN, Gill JPS, Gupta U, Hossain S, Joglekar S, Joshi PC, Kakkar M, Kotwani A, Rattan A, Sudarshan H, Thomas K, Wattal C, Easton A, Laxminarayan R, Global Antibiotic Resistance Partnership (GARP)—India Working Group (2011) Rationalizing antibiotic use to limit antibiotic resistance in India. Ind J Med Res 134, 281–294
- <span id="page-9-24"></span>Garnacho-Montero J, Escoresca-Ortega A, Fernández-Delgado E (2015) Antibiotic de-escalation in the ICU: How is it best done? Curr Opin Infect Dis 28(2):193–198. [https://doi.org/10.1097/](https://doi.org/10.1097/QCO.0000000000000141) [QCO.0000000000000141](https://doi.org/10.1097/QCO.0000000000000141)
- <span id="page-9-30"></span>Gebreyes WA, Dupouy-Camet J, Newport MJ, Oliveira CJB, Schlesinger LS, Saif YM, Kariuki S, Saif LJ, Saville W, Wittum T, Hoet A, Quessy S, Kazwala R, Tekola B, Shryock T, Bisesi M, Patchanee P, Boonmar S, King LJ (2014) The global one health paradigm: challenges and opportunities for tackling infectious diseases at the human, animal, and environment interface in lowresource settings. PLoS Neglect Trop Dis 8(11). [https://doi.org/](https://doi.org/10.1371/journal.pntd.0003257) [10.1371/journal.pntd.0003257](https://doi.org/10.1371/journal.pntd.0003257)
- <span id="page-9-26"></span>Gelband H, Laxminarayan R (2015) Tackling antimicrobial resistance at global and local scales. Trends Microbiol 23(9):524–526. <https://doi.org/10.1016/j.tim.2015.06.005>
- <span id="page-9-0"></span>Gillings MR (2013) Evolutionary consequences of antibiotic use for the resistome, mobilome and microbial pangenome. Front Microbio 4.<https://doi.org/10.3389/fmicb.2013.00004>
- <span id="page-9-10"></span>Gonzalez Ronquillo M, Angeles Hernandez JC (2017) Antibiotic and synthetic growth promoters in animal diets: review of impact and analytical methods. Food Control 72:255–267. [https://doi.org/10.](https://doi.org/10.1016/j.foodcont.2016.03.001) [1016/j.foodcont.2016.03.001](https://doi.org/10.1016/j.foodcont.2016.03.001)
- <span id="page-9-16"></span>Guetiya Wadoum RE, Zambou NF, Anyangwe FF, Njimou JR, Coman MM, Verdenelli MC, Cecchini C, Silvi S, Orpianesi C, Cresci A, Colizzi V (2016) Abusive use of antibiotics in poultry farming in Cameroon and the public health implications. Br Poult Sci 57(4):483–493.<https://doi.org/10.1080/00071668.2016.1180668>
- <span id="page-10-26"></span>Hatakka K, Saxelin M (2008) Probiotics in intestinal and non-intestinal infectious diseases—Clinical evidence. Curr Pharm Des 14(14):1351–1367. [https://doi.org/10.2174/138161208784480](https://doi.org/10.2174/138161208784480162) [162](https://doi.org/10.2174/138161208784480162)
- <span id="page-10-0"></span>He S, Han J, Lichtfouse E (2021) Backward transmission of COVID-19 from humans to animals may propagate reinfections and induce vaccine failure. Environ Chem Lett. [https://doi.org/10.1007/](https://doi.org/10.1007/s10311-020-01140-4) [s10311-020-01140-4](https://doi.org/10.1007/s10311-020-01140-4)
- <span id="page-10-2"></span>Health Canada (2014) Notice to stakeholders: collaborative efforts to promote the judicious use of medically-important antimicrobial drugs in food animal production. [http://www.hc-sc.gc.ca/dhp](http://www.hc-sc.gc.ca/dhp-mps/vet/antimicrob/amr-notice-ram-avis-20140410-eng.php)[mps/vet/antimicrob/amr-notice-ram-avis-20140410-eng.php](http://www.hc-sc.gc.ca/dhp-mps/vet/antimicrob/amr-notice-ram-avis-20140410-eng.php). Accessed 3 Jan 2017
- <span id="page-10-5"></span>Hiki M, Kawanishi M, Abo H, Kojima A, Koike R, Hamamoto S, Asai T (2015) Decreased resistance to broad-spectrum cephalosporin in escherichia coli from healthy broilers at farms in Japan after voluntary withdrawal of Ceftiofur. Foodborne Pathog Dis 12(7):639–643. <https://doi.org/10.1089/fpd.2015.1960>
- <span id="page-10-9"></span>Hollis A, Ahmed Z (2013) Preserving antibiotics, rationally. N Engl J Med 369(26):2474–2476. [https://doi.org/10.1056/NEJMp13114](https://doi.org/10.1056/NEJMp1311479) [79](https://doi.org/10.1056/NEJMp1311479)
- <span id="page-10-27"></span>Holloway KA, Kotwani A, Batmanabane G, Puri M, Tisocki K (2017) Antibiotic use in South East Asia and policies to promote appropriate use: reports from country situational analyses. BMJ 35[8https://doi.org/10.1136/bmj.j2291](https://doi.org/10.1136/bmj.j2291)
- <span id="page-10-25"></span>Jani K, Dhotre D, Bandal J, Shouche Y, Suryavanshi M, Rale V, Sharma A (2018) World's largest mass bathing event infuences the bacterial communities of godavari, a holy river of India. Microb Ecol 76(3):706–718. [https://doi.org/10.1007/](https://doi.org/10.1007/s00248-018-1169-1) [s00248-018-1169-1](https://doi.org/10.1007/s00248-018-1169-1)
- <span id="page-10-24"></span>Khan K, Memish ZA, Chabbra A, Liauw J, Hu W, Janes DA, Sears J, Arino J, Macdonald M, Calderon F, Raposo P, Heidebrecht C, Wang J, Chan A, Brownstein J, Gardam M (2010) Global public health implications of a mass gathering in mecca, saudi arabia during the midst of an infuenza pandemic. J Travel Med 17(2):75–81.<https://doi.org/10.1111/j.1708-8305.2010.00397.x>
- <span id="page-10-21"></span>Khan S, Beattie TK, Knapp CW (2019) Rapid selection of antimicrobial-resistant bacteria in complex water systems by chlorine and pipe materials. Environ Chem Lett 17(3):1367–1373. [https://doi.](https://doi.org/10.1007/s10311-019-00867-z) [org/10.1007/s10311-019-00867-z](https://doi.org/10.1007/s10311-019-00867-z)
- <span id="page-10-20"></span>Kumar KC, Gupta S, Chander Y, Singh AK (2005) Antibiotic use in agriculture and its impact on the terrestrial environment. In Adv Agron (vol. 87, pp. 1–54). Academic Press. [https://doi.org/10.](https://doi.org/10.1016/S0065-2113(05)87001-4) [1016/S0065-2113\(05\)87001-4](https://doi.org/10.1016/S0065-2113(05)87001-4)
- <span id="page-10-14"></span>Kumari Anjana RKN, Jayachandran KGM (2017) Persistence of antibiotic residue in milk under Region of Bihar, India. Int J Current Microbio Appl Sci 6(3), 2296–2299. [https://doi.org/10.20546/](https://doi.org/10.20546/ijcmas.2017.603.262) [ijcmas.2017.603.262](https://doi.org/10.20546/ijcmas.2017.603.262)
- <span id="page-10-22"></span>Kümmerer K (2009) Antibiotics in the aquatic environment—a review—Part I. Chemosphere 75(4):417–434. [https://doi.org/](https://doi.org/10.1016/j.chemosphere.2008.11.086) [10.1016/j.chemosphere.2008.11.086](https://doi.org/10.1016/j.chemosphere.2008.11.086)
- <span id="page-10-1"></span>Landers TF, Cohen B, Wittum TE, Larson EL (2012) A review of antibiotic use in food animals: perspective, policy, and potential. Public Health Rep 127(1):4–22
- <span id="page-10-12"></span>Laxminarayan R, Chaudhury RR (2016) Antibiotic resistance in India: drivers and opportunities for action. PLoS Medicine 13(3). <https://doi.org/10.1371/journal.pmed.1001974>
- <span id="page-10-15"></span>Lee MH, Lee HJ, Ryu PD (2001) public health risks: chemical and antibiotic residues—review. Asian-Austr J Anim Sci 14(3), 402–413
- <span id="page-10-31"></span>Leung E, Weil DE, Raviglione M, Nakatani H (2011) The WHO policy package to combat antimicrobial resistance. Bull World Health Organ 89:390–392.<https://doi.org/10.2471/BLT.11.088435>
- <span id="page-10-8"></span>Liao SF, Nyachoti M (2017) Using probiotics to improve swine gut health and nutrient utilization. Anim Nutr 3(4):331–343. [https://](https://doi.org/10.1016/j.aninu.2017.06.007) [doi.org/10.1016/j.aninu.2017.06.007](https://doi.org/10.1016/j.aninu.2017.06.007)
- <span id="page-10-7"></span>Liu Y-Y, Wang Y, Walsh TR, Yi L-X, Zhang R, Spencer J, Doi Y, Tian G, Dong B, Huang X, Yu L-F, Gu D, Ren H, Chen X, Lv L, He D, Zhou H, Liang Z, Liu J-H, Shen J (2016) Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. Lancet Infect Dis 16(2):161–168. [https://doi.](https://doi.org/10.1016/S1473-3099(15)00424-7) [org/10.1016/S1473-3099\(15\)00424-7](https://doi.org/10.1016/S1473-3099(15)00424-7)
- <span id="page-10-18"></span>Marti R, Scott A, Tien Y-C, Murray R, Sabourin L, Zhang Y, Topp E (2013) Impact of Manure Fertilization on the Abundance of Antibiotic-Resistant Bacteria and Frequency of Detection of Antibiotic Resistance Genes in Soil and on Vegetables at Harvest. Appl Environ Microbiol 79(18):5701–5709. [https://doi.org/](https://doi.org/10.1128/AEM.01682-13) [10.1128/AEM.01682-13](https://doi.org/10.1128/AEM.01682-13)
- <span id="page-10-10"></span>Marti E, Variatza E, Balcazar JL (2014) The role of aquatic ecosystems as reservoirs of antibiotic resistance. Trends Microbiol 22(1):36– 41.<https://doi.org/10.1016/j.tim.2013.11.001>
- <span id="page-10-30"></span>Martínez JL, Coque TM, Baquero F (2015) Prioritizing risks of antibiotic resistance genes in all metagenomes. Nat Rev Microbiol 13(6):396–396.<https://doi.org/10.1038/nrmicro3399-c2>
- <span id="page-10-28"></span>Masterton RG (2011) Antibiotic de-escalation. Crit Care Clin 27(1):149–162.<https://doi.org/10.1016/j.ccc.2010.09.009>
- <span id="page-10-29"></span>McLain JE, Cytryn E, Durso LM, Young S (2016) Culture-based methods for detection of antibiotic resistance in agroecosystems: advantages, challenges, and gaps in knowledge. J Environ Qual 45(2):432–440.<https://doi.org/10.2134/jeq2015.06.0317>
- <span id="page-10-17"></span>Monier J-M, Demanèche S, Delmont TO, Mathieu A, Vogel TM, Simonet P (2011) Metagenomic exploration of antibiotic resistance in soil. Curr Opin Microbiol 14(3):229–235. [https://doi.org/](https://doi.org/10.1016/j.mib.2011.04.010) [10.1016/j.mib.2011.04.010](https://doi.org/10.1016/j.mib.2011.04.010)
- <span id="page-10-23"></span>Mulamattathil SG, Bezuidenhout C, Mbewe M, Ateba CN (2014) Isolation of environmental bacteria from surface and drinking water in mafkeng, south africa, and characterization using their antibiotic resistance profles [Research Article]. J Pathog; Hindawi. [https://](https://doi.org/10.1155/2014/371208) [doi.org/10.1155/2014/371208](https://doi.org/10.1155/2014/371208)
- <span id="page-10-6"></span>Nelson JM, Chiller TM, Powers JH, Angulo FJ (2007) Fluoroquinolone-resistant campylobacter species and the withdrawal of fuoroquinolones from use in poultry: a public health success story. Clin Infect Dis 44(7):977–980. [https://doi.org/10.1086/](https://doi.org/10.1086/512369) [512369](https://doi.org/10.1086/512369)
- <span id="page-10-16"></span>Nisha A (2008) Antibiotic residues—a global health hazard. Veterinary World 2(2):375.<https://doi.org/10.5455/vetworld.2008.375-377>
- <span id="page-10-3"></span>Nordmann P (2014) Carbapenemase-producing Enterobacteriaceae: Overview of a major public health challenge. Med Mal Infect 44(2):51–56.<https://doi.org/10.1016/j.medmal.2013.11.007>
- <span id="page-10-4"></span>Odsbu I, Khedkar S, Lind F, Khedkar U, Nerkar SS, Orsini N, Tamhankar AJ, Stålsby Lundborg C (2018) Trends in resistance to extended-spectrum cephalosporins and carbapenems among escherichia coli and klebsiella spp. Isolates in a District in Western India during 2004–2014. Int J Environ Res Pub Health 15(1), 155. <https://doi.org/10.3390/ijerph15010155>
- <span id="page-10-19"></span>Oluyege OJ, T OTIO (2015) Composition of antibiotic resistant bacteria from irrigated vegetable farmland. J Microbio Res 5(5):161–168
- <span id="page-10-32"></span>Pereko DD, Lubbe MS, Essack SY (2016) Surveillance of antibiotic use in the private sector in Namibia using sales and claims data. J Infect Develop Ctries 10(11):1243–1249. [https://doi.org/10.](https://doi.org/10.3855/jidc.7329) [3855/jidc.7329](https://doi.org/10.3855/jidc.7329)
- <span id="page-10-33"></span>Peterson D (2003) Eating Apes. University of California Press
- <span id="page-10-11"></span>Pham Kim D, Saegerman C, Douny C, Vu Dinh T, Ha Xuan B, Dang Vu B, Pham Hong N, Scippo ML (2013) First survey on the use of antibiotics in pig and poultry production in the red river delta region of Vietnam. Food Public Health 3(5). [https://doi.org/10.](https://doi.org/10.5923/j.fph.20130305.03) [5923/j.fph.20130305.03](https://doi.org/10.5923/j.fph.20130305.03)
- <span id="page-10-13"></span>Phillips I, Casewell M, Cox T, De Groot B, Friis C, Jones R, Nightingale C, Preston R, Waddell J (2004) Does the use of antibiotics in food animals pose a risk to human health? a critical review of

published data. J Antimicrob Chemother 53(1):28–52. [https://](https://doi.org/10.1093/jac/dkg483) [doi.org/10.1093/jac/dkg483](https://doi.org/10.1093/jac/dkg483)

- <span id="page-11-12"></span>Pruden A, Arabi M, Storteboom HN (2012) Correlation between upstream human activities and riverine antibiotic resistance genes. Environ Sci Technol 46(21):11541–11549. [https://doi.](https://doi.org/10.1021/es302657r) [org/10.1021/es302657r](https://doi.org/10.1021/es302657r)
- <span id="page-11-7"></span>Rawat D, Nair D (2010) Extended-spectrum β-lactamases in gram negative bacteria. J Glob Infect Dis 2(3):263–274. [https://doi.](https://doi.org/10.4103/0974-777X.68531) [org/10.4103/0974-777X.68531](https://doi.org/10.4103/0974-777X.68531)
- <span id="page-11-17"></span>Riesenfeld CS, Goodman RM, Handelsman J (2004) Uncultured soil bacteria are a reservoir of new antibiotic resistance genes. Environ Microbiol 6(9):981–989. [https://doi.org/10.1111/j.1462-](https://doi.org/10.1111/j.1462-2920.2004.00664.x) [2920.2004.00664.x](https://doi.org/10.1111/j.1462-2920.2004.00664.x)
- <span id="page-11-9"></span>Rizzo K, Horwich-Scholefeld S, Epson E (2019) Carbapenem and cephalosporin resistance among enterobacteriaceae in healthcare-associated infections, California, USA1. Emerg Infect Dis 25(7):1389–1393.<https://doi.org/10.3201/eid2507.181938>
- <span id="page-11-2"></span>Rodgers K, McLellan I, Peshkur T, Williams R, Tonner R, Hursthouse AS, Knapp CW, Henriquez FL (2019) Can the legacy of industrial pollution infuence antimicrobial resistance in estuarine sediments? Environ Chem Lett 17(2):595-607. [https://doi.org/](https://doi.org/10.1007/s10311-018-0791-y) [10.1007/s10311-018-0791-y](https://doi.org/10.1007/s10311-018-0791-y)
- <span id="page-11-26"></span>Rogers Van Katwyk S, Hofman SJ, Mendelson M, Taljaard M, Grimshaw JM (2020) Strengthening the science of addressing antimicrobial resistance: a framework for planning, conducting and disseminating antimicrobial resistance intervention research. Health Res Policy Syst 18(1):60. [https://doi.org/10.1186/](https://doi.org/10.1186/s12961-020-00549-1) [s12961-020-00549-1](https://doi.org/10.1186/s12961-020-00549-1)
- <span id="page-11-29"></span>Romero D, Traxler MF, López D, Kolter R (2011) Antibiotics as signal molecules. Chem Rev 111(9):5492–5505. [https://doi.org/10.](https://doi.org/10.1021/cr2000509) [1021/cr2000509](https://doi.org/10.1021/cr2000509)
- <span id="page-11-10"></span>Rusu A, Hancu G, Uivaroşi V (2015) Fluoroquinolone pollution of food, water and soil, and bacterial resistance. Environ Chem Lett 13(1):21–36.<https://doi.org/10.1007/s10311-014-0481-3>
- <span id="page-11-22"></span>Santa-Ana-Tellez Y, Mantel-Teeuwisse AK, Dreser A, Leufkens HGM, Wirtz VJ (2013) Impact of over-the-counter restrictions on antibiotic consumption in Brazil and Mexico. PLoS ONE 8(10):e75550.<https://doi.org/10.1371/journal.pone.0075550>
- <span id="page-11-20"></span>Scallan E, Hoekstra RM, Angulo FJ, Tauxe RV, Widdowson M-A, Roy SL, Jones JL, Grifn PM (2011) Foodborne Illness Acquired in the United States—Major Pathogens. Emerg Infect Dis 17(1):7– 15.<https://doi.org/10.3201/eid1701.P11101>
- <span id="page-11-14"></span>Schmieger H, Schicklmaier P (1999) Transduction of multiple drug resistance of Salmonella enterica serovar typhimurium DT104. FEMS Microbiol Lett 170(1):251–256. [https://doi.org/10.1111/j.](https://doi.org/10.1111/j.1574-6968.1999.tb13381.x) [1574-6968.1999.tb13381.x](https://doi.org/10.1111/j.1574-6968.1999.tb13381.x)
- <span id="page-11-13"></span>Seiler C, Berendonk TU (2012) Heavy metal driven co-selection of antibiotic resistance in soil and water bodies impacted by agriculture and aquaculture. Front Microbio *3.* [https://doi.org/10.3389/](https://doi.org/10.3389/fmicb.2012.00399) [fmicb.2012.00399](https://doi.org/10.3389/fmicb.2012.00399)
- <span id="page-11-1"></span>Shi H, Ni J, Zheng T, Wang X, Wu C, Wang Q (2020) Remediation of wastewater contaminated by antibiotics: a review. Environ Chem Lett 18(2):345–360. <https://doi.org/10.1007/s10311-019-00945-2>
- <span id="page-11-27"></span>Singh BP, Ghosh S, Chauhan A (2021) Development, dynamics and control of antimicrobial-resistant bacterial bioflms: a review. Environ Chem Lett. <https://doi.org/10.1007/s10311-020-01169-5>
- <span id="page-11-3"></span>Singh KS, Anand S, Dholpuria S, Sharma JK, Shouche Y (2020) Antimicrobial Resistance Paradigm and One-Health Approach. In: Panwar H, Sharma C, Lichtfouse E (Eds.), Sustainable agriculture reviews 46: mitigation of antimicrobial resistance Vol 1 tools and targets (pp. 1–32). Springer International Publishing, New York. [https://doi.org/10.1007/978-3-030-53024-2\\_1](https://doi.org/10.1007/978-3-030-53024-2_1)
- <span id="page-11-19"></span>Singh KS, Choudhary R, Bisht S, Grover S, Kumar S, Mohanty AK, Kaushik JK (2017) Expression of recombinant truncated domains of mucus-binding (Mub) protein of Lactobacillus plantarum in

soluble and biologically active form. Protein Expr Purif 135:54– 60.<https://doi.org/10.1016/j.pep.2017.04.015>

- <span id="page-11-18"></span>Singh KS, Kumar S, Mohanty AK, Grover S, Kaushik JK (2018) Mechanistic insights into the host-microbe interaction and pathogen exclusion mediated by the Mucus-binding protein of Lactobacillus plantarum. Sci Rep 8(1):14198. [https://doi.org/10.1038/](https://doi.org/10.1038/s41598-018-32417-y) [s41598-018-32417-y](https://doi.org/10.1038/s41598-018-32417-y)
- <span id="page-11-15"></span>Soto SM (2013) Role of efflux pumps in the antibiotic resistance of bacteria embedded in a bioflm. Virulence 4(3):223–229. [https://](https://doi.org/10.4161/viru.23724) [doi.org/10.4161/viru.23724](https://doi.org/10.4161/viru.23724)
- <span id="page-11-0"></span>Spellberg B, Gilbert DN (2014) The future of antibiotics and resistance: a tribute to a career of leadership by John Bartlett. Clinical Infect Dis 59(suppl\_2), S71–S75. [https://doi.org/10.1093/cid/](https://doi.org/10.1093/cid/ciu392) [ciu392](https://doi.org/10.1093/cid/ciu392)
- <span id="page-11-25"></span>Spicknall IH, Foxman B, Marrs CF, Eisenberg JNS (2013) A modeling framework for the evolution and spread of antibiotic resistance: literature review and model categorization. Am J Epidemiol 178(4):508–520.<https://doi.org/10.1093/aje/kwt017>
- <span id="page-11-21"></span>Spoor LE, McAdam PR, Weinert LA, Rambaut A, Hasman H, Aarestrup FM, Kearns AM, Larsen AR, Skov RL, Fitzgerald JR (2013) Livestock origin for a human pandemic clone of community-associated methicillin-resistant staphylococcus aureus. MBio 4(4).<https://doi.org/10.1128/mBio.00356-13>
- <span id="page-11-28"></span>Subbiah M, Mitchell SM, Call DR (2016) Not all antibiotic use practices in food-animal agriculture aford the same risk. J Environ Qual 45(2):618–629. <https://doi.org/10.2134/jeq2015.06.0297>
- <span id="page-11-5"></span>Tacconelli E, Carrara E, Savoldi A, Harbarth S, Mendelson M, Monnet DL, Pulcini C, Kahlmeter G, Kluytmans J, Carmeli Y, Ouellette M, Outterson K, Patel J, Cavaleri M, Cox EM, Houchens CR, Grayson ML, Hansen P, Singh N … Zorzet A (2018) Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. Lancet Infect Dis 18(3), 318–327.[https://doi.org/10.1016/S1473-3099\(17\)](https://doi.org/10.1016/S1473-3099(17)30753-3) [30753-3](https://doi.org/10.1016/S1473-3099(17)30753-3)
- <span id="page-11-8"></span>Tadesse BT, Ashley EA, Ongarello S, Havumaki J, Wijegoonewardena M, González IJ, Dittrich S (2017) Antimicrobial resistance in Africa: a systematic review. BMC Infect Dis 17(1):616. [https://](https://doi.org/10.1186/s12879-017-2713-1) [doi.org/10.1186/s12879-017-2713-1](https://doi.org/10.1186/s12879-017-2713-1)
- <span id="page-11-11"></span>Tang SS, Apisarnthanarak A, Hsu LY (2014) Mechanisms of β-lactam antimicrobial resistance and epidemiology of major communityand healthcare-associated multidrug-resistant bacteria. Adv Drug Deliv Rev 78:3–13. <https://doi.org/10.1016/j.addr.2014.08.003>
- <span id="page-11-6"></span>Tang KL, Cafrey NP, Nóbrega DB, Cork SC, Ronksley PE, Barkema HW, Polachek AJ, Ganshorn H, Sharma N, Kellner JD, Checkley SL, Ghali WA (2019) Comparison of diferent approaches to antibiotic restriction in food-producing animals: Stratifed results from a systematic review and meta-analysis. BMJ Glob Health 4(4):e001710.<https://doi.org/10.1136/bmjgh-2019-001710>
- <span id="page-11-16"></span>Tavakoli H (2015) Detecting antibiotic residues by HPLC method in chicken and calves meat in diet of a military center in Tehran | Request PDF. ResearchGate. [https://www.researchgate.net/publi](https://www.researchgate.net/publication/291835452_Detecting_antibiotic_residues_by_HPLC_method_in_chicken_and_calves_meat_in_diet_of_a_military_center_in_Tehran) [cation/291835452\\_Detecting\\_antibiotic\\_residues\\_by\\_HPLC\\_](https://www.researchgate.net/publication/291835452_Detecting_antibiotic_residues_by_HPLC_method_in_chicken_and_calves_meat_in_diet_of_a_military_center_in_Tehran) [method\\_in\\_chicken\\_and\\_calves\\_meat\\_in\\_diet\\_of\\_a\\_military\\_](https://www.researchgate.net/publication/291835452_Detecting_antibiotic_residues_by_HPLC_method_in_chicken_and_calves_meat_in_diet_of_a_military_center_in_Tehran) [center\\_in\\_Tehran](https://www.researchgate.net/publication/291835452_Detecting_antibiotic_residues_by_HPLC_method_in_chicken_and_calves_meat_in_diet_of_a_military_center_in_Tehran)
- <span id="page-11-24"></span>Uchil RR, Kohli GS, Katekhaye VM, Swami OC (2014) Strategies to combat antimicrobial resistance. J Clinic Diagn Res: JCDR, 8(7), ME01–ME04.<https://doi.org/10.7860/JCDR/2014/8925.4529>
- <span id="page-11-23"></span>Valsamatzi-Panagiotou A, Popova KB, Penchovsky R (2021) Methods for prevention and constraint of antimicrobial resistance: A review. Environ Chem Lett. [https://doi.org/10.1007/](https://doi.org/10.1007/s10311-021-01206-x) [s10311-021-01206-x](https://doi.org/10.1007/s10311-021-01206-x)
- <span id="page-11-4"></span>Van Boeckel TP, Brower C, Gilbert M, Grenfell BT, Levin SA, Robinson TP, Teillant A, Laxminarayan R (2015) Global trends in antimicrobial use in food animals. Proc Natl Acad Sci 112(18):5649. <https://doi.org/10.1073/pnas.1503141112>
- <span id="page-12-9"></span>Vorholt JA (2012) Microbial life in the phyllosphere. Nat Rev Microbiol 10(12):828–840. <https://doi.org/10.1038/nrmicro2910>
- <span id="page-12-7"></span>Wahome CN (2013) Contamination levels of groundwater, antimicrobial resistance patterns, plasmid profiles and chlorination efficacy in ongata rongai, kajiado north county, kenya. 116
- <span id="page-12-4"></span>Wegener HC (2012) Antibiotic resistance—linking human and animal health. In Improving food safety through a one health approach: workshop summary. National Academies Press (US). [https://](https://www.ncbi.nlm.nih.gov/books/NBK114485/) [www.ncbi.nlm.nih.gov/books/NBK114485/](https://www.ncbi.nlm.nih.gov/books/NBK114485/)
- <span id="page-12-1"></span>World Health Organization (WHO) (2003) Impacts of antimicrobial growth promoter termination in Denmark. The WHO international review panel's evaluation of the termination of the use of antimicrobial growth promoters in Denmark. WHO, Geneva, Switzerland
- <span id="page-12-0"></span>WHO Advisory Group on Integrated Surveillance of Antimicrobial Resistance (AGISAR) (2016) Critically Important Antimicrobials for Human Medicine, 4th revision. WHO, Geneva, Switzerland
- <span id="page-12-8"></span>Willems RJL, Hanage WP, Bessen DE, Feil EJ (2011) Population biology of Gram-positive pathogens: High-risk clones for dissemination of antibiotic resistance. FEMS Microbiol Rev 35(5):872– 900. <https://doi.org/10.1111/j.1574-6976.2011.00284.x>
- <span id="page-12-6"></span>Xiao KQ, Li B, Ma L, Bao P, Zhou X, Zhang T, Zhu YG (2016) Metagenomic profles of antibiotic resistance genes in paddy soils from South China. FEMS Microbio Ecol 92(3). [https://doi.](https://doi.org/10.1093/femsec/fiw023) [org/10.1093/femsec/fw023](https://doi.org/10.1093/femsec/fiw023)
- <span id="page-12-3"></span>Zhang F, Li Y, Yang M, Li W (2012) Content of Heavy metals in animal feeds and manures from farms of diferent scales in Northeast China. Int J Environ Res Public Health 9(8):2658–2668. <https://doi.org/10.3390/ijerph9082658>
- <span id="page-12-5"></span>Zheng N, Wang J, Han R, Xu X, Zhen Y, Qu X, Sun P, Li S, Yu Z (2013) Occurrence of several main antibiotic residues in raw milk in 10 provinces of China. Food Addit Contam: Part B 6(2):84–89.<https://doi.org/10.1080/19393210.2012.727189>
- <span id="page-12-2"></span>Zhu Y-G, Johnson TA, Su J-Q, Qiao M, Guo G-X, Stedtfeld RD, Hashsham SA, Tiedje JM (2013) Diverse and abundant antibiotic resistance genes in Chinese swine farms. Proc Natl Acad Sci 110(9):3435.<https://doi.org/10.1073/pnas.1222743110>

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