#### **REVIEW**



# **Biosensors for toxic metals, polychlorinated biphenyls, biological oxygen demand, endocrine disruptors, hormones, dioxin, phenolic and organophosphorus compounds: a review**

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#### **Abstract**

The growing pollution by hazardous agents is a major concern due to pollutant transfer to water, air, soil and food. Since actual analytical methods are limited, there is a need for detectors that are more sensitive, more selective, faster and cheaper. For instance, advanced portable biosensors have better sensitivity compared to classical diagnostic devices. Here, we review ultrasensitive detection of pollutants by biosensors. In particular, nanobiosensors display remarkable nanomolar to picomolar detection of various pollutants including heavy metals, pesticides, endocrine disruptors, dioxin, biological oxygen demand and microbial pathogens.

**Keywords** Biosensor · Nanobiosensor · Pollutant · Environment · Detection · Selectivity · Sensitivity

# **Introduction**

Biosensor technology is one of the emerging technologies that have produced a major impact in the diverse sectors ranging from healthcare, food, pharmaceutical, agriculture and environmental industries (Campana et al. [2019;](#page-6-0) Verma [2017a](#page-9-0); Verma et al. [2010](#page-9-1)). According to a new market report published in 2020 by Allied Market Research "Biosensors market by product (wearable biosensors and non-wearable biosensors), technology (electrochemical biosensors, optical biosensors, piezoelectric biosensors, thermal biosensors and nanomechanical biosensors): Global opportunity analysis and industry forecast", the global biosensors market size was valued at \$17,500 million in 2018 and is expected to reach \$38,600 million by 2026, registering a compound annual growth rate of 10.4% from 2019 to 2026 ([https://www.allie](https://www.alliedmarketresearch.com/biosensors-market) [dmarketresearch.com/biosensors-market\)](https://www.alliedmarketresearch.com/biosensors-market).

Biosensors are remarkable portable tools employed for the detection of chemical and biological components of clinical, food and environmental monitoring (Kalyani et al. [2020](#page-7-0)). Biosensors are endowed with unique properties like higher specifcity, rapid response, compacted size, higher selectivity, higher stability, lower cost and user-friendly nature that make them the ideal sensing device. It combines a biologically derived recognition entity with a transducer for developing biochemical parameters quantitatively (Mishra et al. [2018;](#page-7-1) Verma [2017b\)](#page-9-2). Biological elements can be an antibody, enzyme, cell receptors, nucleic acids and microbes, while the sensing element can be an electric potential and electric current (Jain et al. [2010\)](#page-7-2). Diferent variants of the biosensors are based on the working mechanism of the transducer used. It can be resonant biosensors, electrochemical biosensors, thermal biosensors, ion-selective feld-efect transistor biosensors and optical biosensors (Prasad et al. [2017](#page-8-0); Verma [2017a](#page-9-0)).

Biosensors are being used in diferent felds such as food product quality control, environmental applications, medical applications, military and bioprocess control as depicted in Fig. [1.](#page-1-0) The role of biosensors in the food industry is to detect contaminants in the food products, content verifcation and freshness of the product as well as to monitor the conversion of raw material (Saravanan et al. [2020\)](#page-8-1). The detection of hazardous substances is also a vital application of the biosensors in defence services (Kuswandi [2018](#page-7-3); Verma [2017a](#page-9-0), [b](#page-9-2)).

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Environmental hazards can be monitored as well as controlled by using the specifc data about the contaminated site provided via biosensors. Biosensors can also measure the pollutants and determine the biological effects of pollutants like endocrinedisrupting efects and toxicity levels (Rodriguez-Mozaz et al. [2004\)](#page-8-2). Thus, the exceptional capabilities and performance of the biosensors provide the excellent services for the detection of chemical and biological components as well as monitoring the clinical, environmental and the food products (Malhotra et al. [2017;](#page-7-4) Amine et al. [2006](#page-6-1)).

This article reviews the role of biosensors in environmental monitoring like nitrogen compounds, heavy metals, phenolic compounds, biological oxygen demand, pesticides, insecticides, endocrine disruptors, hormones, organophosphorus compounds and polychlorinated biphenyls. This article is an abridged version of the chapter published by Rani and Verma [\(2020](#page-8-3)) in Environmental Chemistry for a Sustainable World [\(https://www.springer.com/series/11480\)](https://www.springer.com/series/11480).

## **Application of biosensors in environmental monitoring**

Pollutants are producing harmful efects globally. Thus, there is a pressing need to develop excellent portable tools which are user-friendly, cost-efective, fast to monitor and control these pollutants. Biosensors work like ultrasensitive tools for the environment impact assessment of ecological, biological and chemically monitoring of the inorganic as well as organic pollutants (Rogers [2006](#page-8-4); Sharpe [2003;](#page-8-5) Rogers and Gerlach [1996\)](#page-8-6).

Sulphur dioxide is a major contaminant of air, mainly produced by biological decay, oceans, forest fres and volcanoes. Anthropogenic sulphur dioxide pollutants are produced from the processes like sulphuric acid manufacturing, combustion of fossil fuels, coal burning and wood



<span id="page-1-0"></span>**Fig. 1** Applications of biosensors in diferent felds. Biosensors can be used for the monitoring of various toxicants, pollutants and contaminants in various felds such as the environment, health, food and agriculture

pulp industry (Tayanc [2000](#page-8-7)). An amperometric biosensor based on sulphite oxidase and cytochrome c developed for the measurement of sulphur dioxide concentration in the fowing gas stream. Biosensor produced linear response of pollutant detection in the range of 4–50 parts per million at the low voltage (Hart et al. [2002](#page-7-5)).

An electrochemical biosensor was developed for the 30 nanogram detection of formaldehyde, an air pollutant (Herschkovitz et al. [2000\)](#page-7-6). Smoke from exhaustion of automobiles, forest fres and tobacco contains formaldehyde compounds. Table [1](#page-2-0) shows the list of some biosensors used for determining the pollutants.

Monitoring of environmental and food contaminants has been reported via sensors based on electrochemical aptamers. In the process of development of nucleic acidbased sensors, aptamers are small, stable and inexpensive biorecognition components. Due to the extraordinary design properties like better elasticity and expediency, aptamers showed excellent sensitivity and selectivity (Mishra et al. [2018\)](#page-7-1). Pharmaceutical industry produces many contaminants whose concentration is ever-increasing and produces adverse efects on human health (Tijani et al. [2016](#page-9-3)). Although there is a lack of data about the actual concentrations of these pollutants, there is an urgent need to regulate and monitor the resources for sustainable development. The use of miniaturized enzyme-based biosensors is a promising way to tackle these pollutants of high concern (Campana et al. [2019](#page-6-0)). Monitoring of the environment is a key process to manage the pollution. The most promising approach to monitor the atmospheric pollutants is electronic noses which are made up of a sensor array. Electronic noses work on controlling the pollution as well as odour (Sayago et al. [2019](#page-8-8)). The role of biosensors in the detection of diverse environmental pollutants has been discussed as follows:

### **Detection of heavy metals**

Heavy metals refer to metals having atomic weights in the range of 63.5–200.6 g mol<sup>-1</sup> with a specific gravity greater than 5 g cm<sup>-3</sup> (Srivastava and Majumder [2008\)](#page-8-9). Heavy metals are polluting the environment at the concentration of more than 10 parts per million. It produces adverse efects on human health and environment (Abdu et al. [2017\)](#page-6-2). Heavy metals are ubiquitously present in our biosphere (Verma and Singh [2005;](#page-9-4) Sharpe [2003](#page-8-5)). The genesis of heavy metals is due to the anthropogenic and natural sources. Being highly toxic, heavy metals are producing harmful efects on the environment (Dubey et al. [2018\)](#page-7-7). Metal exposure to humans is damaging health including signifcant deaths. Commonly found metal environmental contaminants are mercury, lead, chromium, zinc, copper and cadmium (Barrocas et al. [2008](#page-6-3)). A range of heavy metals can be traced via nanotechnologybased electrochemical and optical sensors (Malik et al. [2019;](#page-7-8) Ion et al. [2010\)](#page-7-9). Bacterial biosensors use resistant bacterial genes for the analysis of heavy metals present in the environment. Metal-resistant genes from the bacterial strains have been isolated as biological receptors (Rathnayake et al. [2009\)](#page-8-10). Durrieu and Tran-Minh ([2002](#page-7-10)) developed an optical biosensor for micromolar detection of cadmium and lead. Enzyme alkaline phosphatase was inhibited to develop this optical biosensor, which was present on the surface of microalga, namely *Chlorella vulgaris*. A biosensor was developed by using microalgae *Tetraselmis chuii* for the volumetrically measurement of  $4.6 \times 10^{-10}$  M copper ions (Alpat et al. [2007\)](#page-6-4). Performance of an electrochemical

<span id="page-2-0"></span>**Table 1** Biosensors for the determination of pollutants in the real samples

Analyte	Matrix	Transducing and recognition element	References
Pesticides	River water	Optical and immunochemical	Kalyani et al. (2020), Rodriguez-Mozaz et al. $(2004)$ and Mallat et al. $(2001)$
<b>Phenols</b>	Wastewater	Electrochemical and enzymatic	Mazhari et al. (2017) and Nistor et al. (2002)
Heavy metals	Wastewater	Electrochemical/optical and bacteria	De Benedetto et al. $(2019)$ , Philp et al. $(2003)$ and Farre et al. $(2001)$
Biological oxygen demand	River water	Optical and bacteria	Jouanneau et al. (2014) and Chee et al. (2000)
Linear alkyl benzene sulphonate	River water	Electrochemical and bacteria	Borji et al. (2020) and Nomura et al. (1998)
Daunomycin, polychlorinated biphenyls, aflatoxin	River water	Electrochemical and deoxyribonucleic acid	Mishra et al. (2018), Ponomareva et al. $(2011)$ , Kara et al. $(2008)$ and Marrazza et al. (1999)
Oestrogens and xenoestrogens	Lake and sewage water plant samples	Optical and human oestrogen receptor	Salehi et al. (2018), Bahadir and Sezginturk $(2015)$ and Seifert et al. $(1999)$
Alkanes	Groundwater	Optical and bacteria	<i>Kuswandi</i> (2019) and Sticher et al. (1997)

The spectacular applications of diferent types of biosensors is increased multifold in situ environmental samples testing for nanomolar to picomolar detection of diferent pollutants such as phenols, biological oxygen demand, alkanes, oestrogens, microbial contaminants and pesticides

biosensor was optimized by using a chemometric tool (De Benedetto et al. [2019\)](#page-6-5). The performance of this amperometric biosensor for detecting metal ions was based on the robust design of the facile methodology. The model prepared was validated and employed for micromolar detection of different metal ions such as  $Bi^{3+}$ ,  $Al^{3+}$ ,  $Ag^{+}$  and  $Ni^{2+}$ ions. This biosensor was efficiently used as an amperometric detector in the ion chromatographic system (De Benedetto et al. [2019](#page-6-5)). Thus, biosensor has signifcantly improved the detection limit of heavy metals.

## **Detection of biological oxygen demand**

Biological oxygen demand is a vital parameter used for validating the water quality. More the value of this parameter indicates the less suitability of drinking water. The method for biological oxygen demand determination is a slow process which is not suitable for online monitoring (Farre et al. [2001](#page-7-13)). However, biosensor-based methods are the best one for the fast monitoring of biological oxygen demand (Rodriguez-Mozaz et al. [2004](#page-8-2); Chee et al. [2000](#page-6-6)). Biological oxygen demand-based sensors are the most commonly used biosensors for environmental monitoring. A Japanese company, "Nisshin Electric", had developed frst commercial biological oxygen demand sensor in 1983, and a number of biological oxygen demand biosensors based on the microbial cells are marked by Medingen GmbH, Dr. Lange GmbH in Germany and Autoteam GmbH; Kelma (Belgium); US Filter (USA) and Bioscience, Inc. (Rodriguez-Mozaz et al. [2004,](#page-8-2) [2005\)](#page-8-18). A bacterium named *Pseudomonas putida*, isolated from river water, was used for development of biological oxygen demand sensors (Chee et al. [1999\)](#page-6-9). *Vibrio fsheri* genes lux AE and *Escherichia coli* recombinant cell-based biosensors were used for measuring the biological oxygen demand (Nakamura and Karube [2003\)](#page-8-19). An optical biosensor was reported to measure the biological oxygen demand of multiple samples (Kwok et al. [2005\)](#page-7-18). Biological oxygen demand also provides information about the biodegradable organic load of water (Jouanneau et al. [2014\)](#page-7-14). Organic matter of water is being oxidized by the amount of dissolved oxygen in water. The value of biological oxygen demand of surface water ranges from 0.5 to 4 mg per cubic decimetre due to diferent values in diferent seasons. Thus, it is vital to monitor the water bodies at regular intervals. Biological oxygen demand determination via biosensors is a step towards the biotechnology advances (Ponomareva et al. [2011](#page-8-14)).

Kara et al. ([2008](#page-7-15)) reported a biological oxygen demand sensor based on an immobilized *Pseudomonas syringae* in a microcellular polymer. Biosensor showed the detection linearity over the range of 5 to 100 mg per litre at a flow rate of 0.6 millilitre per minute. *Pseudomonas syringae* containing microcellular disc of polymer made this biological oxygen demand biosensor long shelf life and highly stable. Thus, with the aid of the whole micro-organism or its bioactive sub-part used in the development of the biological oxygen demand sensors, they have provided the early detection of water quality in the various natural and storage water systems.

#### **Detection of nitrogen compounds**

Nitrites are the widely used compounds as fertilizers as well as food preservatives. Continuous consumption of these compounds is causing serious health problems in human beings. Nitrites compounds react adversely with the haemoglobin component of the red blood cells, thereby impairing oxygen carrying capacity to the human body (Moorcroft et al. [2001\)](#page-7-19). Increased levels of nitrites in water bodies are also afecting aquatic life (Rodriguez-Mozaz et al. [2004\)](#page-8-2). A biosensor has been employed for the amperometric determination of nitrites using cytochrome c nitrite reductase enzyme of *Desulfovibrio desulfuricans.* Biosensor demonstrated a response time of 5 s with a linear range between 0.015 and 2.35 micromolar (Chen et al. [2007](#page-6-10)). An enzymatic conductometric biosensor was developed for the micromolar determination of nitrites in water samples (Khadro et al. [2008\)](#page-7-20). Electrochemical biosensors have been reported to detect the degradation of DNA and purine metabolites in nanomolar concentrations. This electrochemical biosensor is also used to check the DNA damage and the DNA interaction with an anticancer drug, 6-mercaptopurine (Shpigun and Andryukhina [2019\)](#page-8-20). Thus, biosensors have improved considerably the detection limit of nitrites in the water bodies. It also provides the stability status of biological macromolecules of high importance.

## **Detection of polychlorinated biphenyls**

Polychlorinated biphenyls, a group of manmade persistent organic compounds, are the environmental pollutants (Borji et al. [2020\)](#page-6-7). Such toxic persistent organic compounds are present ubiquitously (Centi et al. [2006\)](#page-6-11). Due to the lipophilic nature, polychlorinated biphenyls get accumulated in the food products easily and afect human health (Gavlasova et al. [2008](#page-7-21); Centi et al. [2006](#page-6-11)). Gas chromatography coupled mass spectrometry is the generally employed method to detect polychlorinated biphenyls in micromolar concentrations (Gavlasova et al. [2008;](#page-7-21) Centi et al. [2006](#page-6-11)). Immunoassay-based sensory platforms, particularly enzymelinked immunosorbent assays, are commonly used for the determination of polychlorinated biphenyls in felds and laboratories in the range of 0.2–0.5 mg per litre (Gavlasova et al. [2008\)](#page-7-21). Pribyl et al. [\(2006](#page-8-21)) reported the determination of the polychlorinated biphenyls by a novel piezoelectric immunosensor. Extracts were directly employed without any purifcation steps for the determination of polychlorinated biphenyls. Lower-cost sensors had been constructed successfully for the monitoring of soil for polychlorinated biphenyls in the range of 1–100 parts per million (Pribyl et al. [2006](#page-8-21)). Genetically engineered rhizosphere bacterium, namely *Pseudomonas fuorescens* F113Rifpcb, has been employed to degrade the polychlorinated biphenyls (Power et al. [2011](#page-8-22)). Thus, whole-cell- and immunoassay-based biosensors have shown the efficient methodology for the detection of toxic and persistent organic polychlorinated biphenyls.

#### **Detection of phenolic compounds**

Phenolic compounds are the organic pollutants that are present in the polymers, plastic, industrial effluents, pesticides, dyes, resins, drugs, paper, detergents, oil refneries and disinfectants (Luong et al. [2008\)](#page-7-22). Chloro- and nitro-phenols are the highly toxic compounds for aquatic life as well as for human beings (Lin and Juang [2009](#page-7-23)). Phenolic compounds produce adverse efects in animals and plants because they can easily penetrate in the cell membrane of plants and skin of the animals. Processes like photosynthesis, respiration, biocatalysed reactions, mutagenicity and hepatotoxicity are afected by phenolic compounds (Rodriguez-Mozaz et al. [2006](#page-8-23)). The US Environmental Protection Agency and European Commission have put phenols in the list of hazardous compounds due to the highly toxic and persistent environment. Scognamiglio et al. ([2012\)](#page-8-24) developed a biosensing system for micromolar detection of catechol and bisphenol-A by using laccase enzyme from *Trametes versicolor* and tyrosinase enzyme from *Agaricus bisporus*, respectively.

Effluents of paper, wine and plastic industries had detected the presence of phenols by using a paper biosensor. A bioconjugate of tyrosinase sourced from *Streptomyces tuirus* DBZ39 and gold nanoparticles was used to develop the paper biosensor. Paper-based biosensor was very efficient in the micromolar detection of phenols (Mazhari et al. [2017\)](#page-7-12). Phenolic compound detection from the effluent of the textile industry has been reported from a polyphenol oxidase biosensor. Jenipapo (*Genipa americana* L.) fruit had been used for the development of polyphenol oxidase enzymebased biosensor, and this biosensor was found very efective for the micromolar detection of the phenolic compounds (Antunes et al. [2018](#page-6-12)). Thus, biosensors based on enzymes (laccase/tyrosinase) and antibodies are the prominent players employed for the detection of the phenolic pollutants.

#### **Detection of endocrine disruptors and hormones**

Glands secreting hormones make up the endocrine system that controls growth, development, regulation and maturation of humans by binding with specifc receptors. Hormone mimicking compounds bind the hormonal receptors, and thus, blocking the normal passage for hormones is called endocrine-disrupting chemicals (Tijani et al. [2016](#page-9-3)). Ovaries in the female reproductive system on maturation produce oestrogen hormone. An example of endocrine-disrupting chemicals is xenoestrogens which bind the receptor of hormone oestrogen and mimic its activity (Badihi-Mossberg et al. [2007\)](#page-6-13). A novel impedance biosensor had been developed to monitor the endocrine disruptors (Granek and Rishpon [2002](#page-7-24)). Normal homeostasis has been altered via endocrine disruptors (Moraes et al. [2008](#page-7-25)). The presence of endocrine disruptors in the environment is increasing the incidences of a variety of tumour formation such as breast, testicular and thyroid cancers. Waring and Harris [\(2005\)](#page-9-5) reported an immunosensor for the determination of estrone, testosterone, progesterone and some organic pollutants. Oestrogens are afecting the reproductive system of the aquatic animals from the last decade. Rapid response action of monoclonal and polyclonal antibodies has been used for nanomolar detection using a biomacromolecular interaction analyser-based chip immunosensor (Samsonova et al. [2004](#page-8-25)).

An electrochemical biosensor has been developed for the detection of progesterone in the cow milk (Xu et al. [2005](#page-9-6)). Salehi et al. [\(2018\)](#page-8-15) had developed a Rapid Adaptable Portable In vitro Detection (RAPID) biosensor platform to detect the chemicals interacting with the human oestrogen receptor. Increasing population and intensive farming has increased the harmful xenohormones in the environment. The use of such compounds has been prohibited by the European Council (Muller et al. [2008](#page-7-26)). The analysis of hormones exploiting the binding ability of natural receptors for oestrogen had been performed via several electrochemical biosensors (Bahadir and Sezginturk [2015\)](#page-6-8). Thus, immunoassay-based biosensors are predominantly employed for the detection of estrone, testosterone, progesterone and some organic pollutants.

#### **Detection of pesticides**

Pesticides are used as insecticides in agricultural practices (Mulchandani et al. [2001](#page-7-27)). These pesticides contain toxic substances and therefore produce ill efects on human health as well as the environment. Toxic substances present in the pesticides are polluting the water and accumulates in the fruits, vegetables, grains and soil (Simonian et al. [2004](#page-8-26); Aspelin [1994\)](#page-6-14). Pesticides contain organophosphates which interfere with the functioning of acetylcholinesterase enzymes causing the failure of the human central nervous system. So, it is crucial to monitor the presence of these compounds in diferent sources and the environment. The detection of organophosphate compounds had been performed using anhydrolase enzyme sourced from *Altermonas* bacterium (Cock et al. [2009;](#page-6-15) DeFrank et al. [1993\)](#page-6-16). Dhull et al. [\(2013\)](#page-6-17) reported enzymatic biosensors for analysing the organophosphorus compounds.

Graphene-based electrodes have been used to monitor organophosphates by using Inkjet Maskless Lithography technique. Biosensor had rapidly quantifed the paraoxon insecticide residues present in the soil samples in the nanomolar concentration (Hondred et al. [2018](#page-7-28)). Tunesi et al. [\(2018\)](#page-9-7) developed an indium tin oxide-based biosensor for micromolar detection of chlorpyrifos, fenthion and methyl parathion.

Electrochemical biosensors are the most preferred for the determination of diferent classes of pesticides (Kalyani et al. [2020](#page-7-0); Compagnone et al. [2010](#page-6-18)). A bi-enzyme biosensor has been developed to discriminate between the nonorganophosphorus and organophosphorus pesticides (Zhang et al. [2015\)](#page-9-8). Paraoxon has been detected in feld analysis using an amperometric biosensor in the micromolar concentration (Arduini et al. [2006](#page-6-19)). Czech Republic developed a biosensor for micromolar detection of the cholinesterase inhibitors (Matejovsky and Pitschmann [2018](#page-7-29)). Thus, the enzyme organophosphorus acid anhydrolase-based biosensor is predominately used for the detection of a range of organophosphorus compounds.

#### **Detection of herbicides**

Vital process of photosynthesis in plants is inhibited by the triazine- and phenylurea-based herbicides (Singh et al. [2018](#page-8-27)). The detection of these herbicides can be performed via biosensors which are designed with membrane receptors of thylakoids, chloroplasts, photosystem and reaction centres employed by optical and amperometric transducers (Velasco-García and Mottram [2003\)](#page-9-9). The determination of simazine, an herbicide, has been reported in nanomolar concentration by using an immunosensor based on ion-selective field-effect transistor (Starodub et al. [2000](#page-8-28)). Kim et al. [\(2018](#page-7-30)) had established a direct competitive immunoassay method which is based upon thermistor enzymes for the rapid detection of atrazine, an herbicide. Pollutants like heavy metals and herbicides can be detected by microalgae. These photosynthetic micro-organisms are able to detect the traces of pollutants and are highly sensitive towards the changes in the environment. Cyanobacterial and diatoms are important groups for the detection of pollutants. Sensitivity of diverse pollutants detection was enhanced when bacterial biosensors were introduced with bioluminescence genes (Vismara and Garavaglia [1997](#page-9-10)). The determination of mycotoxins and herbicides has been performed by electrochemical enzymatic biosensors based on enzyme peroxidase of plant origin. The preparation of modifed electrodes with reduced graphene oxide is an advanced option for developing thirdgeneration biosensors, and the limit of detection was 1400 fold better than the conventional immunoassay (Fernández et al. [2017\)](#page-7-31). Thus, biosensor has shown multifold higher sensitivity over the conventional immunoassay.

#### **Detection of dioxins**

Dioxins are organosoluble, teratogenic and carcinogenic compounds. It has produced adverse efects on the environment, contaminating food, water, soil and food chain. Dioxins can be transported to longer distances through air and water all over the globe (Yulaev et al. [2001](#page-9-11)). The use of biosensors is extremely helpful for the monitoring of dioxins. Centi et al. ([2007\)](#page-6-20) developed immunosensors for detecting polychlorinated biphenyls in the milk samples. Nanomolar detection of 2,3,7,8-tetrachlorodibenzo-p-dioxin in the ash samples has been reported by Kurosawa et al. [\(2005\)](#page-7-32). Biosensor immunosensor has shown a quick detection system for the dioxins.

## **Nanobiosensors**

Nanotechnology involves the synthesis and application of the nanoscale materials (Verma et al. [2020](#page-9-12)). Application of nanomaterials in the integrated part of biosensor covers a vast and diverse array of devices which are employed in diferent felds ranging from the food to agriculture sectors (Chamundeeswari et al. [2019](#page-6-21); Verma et al. [2019](#page-9-13); Srivastava et al. [2018;](#page-8-29) Verma [2017a](#page-9-0), [b\)](#page-9-2). Nanobiosensor is highly efficient in monitoring the processes of sustainable agriculture in terms of food safety and indiscriminate inputs of excessive herbicides. Due to unique properties of nanoscale materials, nanobiosensors enables the rapid detection of contaminants of soil and water (Kuswandi [2018](#page-7-3); Prasad et al. [2017](#page-8-0)). Nanobiosensors exhibit the excellent detection limit of environmental pollutants in the range of nanomolar to picomolar level (Verma [2017a\)](#page-9-0). Xu et al. [\(2005](#page-9-6)) developed electrochemical biosensors for the determination of hydrogen peroxide, xanthine and glucose, respectively. Plant/ microbial mediated synthesis of nanomaterials follows the principles of green chemistry that are safe as compared to toxic chemically synthesized route of nanomaterials (Prasad 2014). The use of renewable resources for the construction of nanomaterials is targeted for the monitoring of the environmental pollutants (Srivastava et al. [2018;](#page-8-29) Prasad et al. [2014](#page-8-30),[2016\)](#page-8-31). Nanobiosensors are routinely employed for the detection of pathogenic bacteria present in the environment (Jyoti and Tomar [2017](#page-7-33)). Gas biosensors are employed for monitoring and diagnosing a wider range of diseases in human beings. Highly sensitive and specific nature of gas biosensor provides rapid and accurate monitoring of diseases (Nasiri and Clarke [2019](#page-8-32)). Quantum dot-based bioluminescence resonance energy transfer sensor was developed for monitoring the target molecules changes and bioimaging in vivo (Hwang et al. [2019](#page-7-34)). Nanobiosensors are expected to be integrated into small devices for quick real-time monitoring of a plethora of environmental pollutants (Kuswandi [2019](#page-7-17)).

## **Conclusion**

Extensive agrochemical usages to enhance the agricultural yield not only resulted in pollution of the upper layer of soil, but also caused the pollution of underground water. To meet the needs of increasing population, agricultural produce should be enhanced in an eco-friendly and sustainable way. Safe ecosystems and healthy lifestyles can be maintained by monitoring the toxicants and pollutants present in the environment. Biosensors are the best tools for monitoring various sectors such as food, healthcare, pharmaceutical as well as agricultural industries. Nanotechnology is becoming an integral component of the biosensing system for monitoring various pollutants in the environment. Thus, the performance of nanobiosensors, a reinvigoration of conventional biosensors, has been improved considerably with the intervention of advanced functional nanomaterial.

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## **References**

- <span id="page-6-2"></span>Abdu N, Abdullahi AA, Abdulkadir A (2017) Heavy metals and soil microbes. Environ Chem Lett 15:65–84. [https://doi.org/10.1007/](https://doi.org/10.1007/s10311-016-0587-x) [s10311-016-0587-x](https://doi.org/10.1007/s10311-016-0587-x)
- <span id="page-6-4"></span>Alpat SK, Alpar S, Kutlu B, Ozbayrak B, Buyukisik HB (2007) Development of biosorption based algal biosensor for cu(II) using *Tetraselmis chuii*. Sensors Actuators B Chem 128:273–278. [https://](https://doi.org/10.1016/j.snb.2007.06.011) [doi.org/10.1016/j.snb.2007.06.011](https://doi.org/10.1016/j.snb.2007.06.011)
- <span id="page-6-1"></span>Amine A, Mohammadi H, Bourais I, Palleschi G (2006) Enzyme inhibition-based biosensors for food safety and environmental monitoring. Biosens Bioelectron 21:1405–1423. [https://doi.](https://doi.org/10.1016/j.bios.2005.07.012) [org/10.1016/j.bios.2005.07.012](https://doi.org/10.1016/j.bios.2005.07.012)
- <span id="page-6-12"></span>Antunes RS, Ferraz D, Garcia LF, Thomaz DV, Luque R, Lobon GS, Gil ES, Lopes FM (2018) Development of a polyphenol oxidase biosensor from Jenipapo fruit extract (*Genipa americana* L) and determination of phenolic compounds in textile industrial effluents. Biosensors 8(2):47. [https://doi.org/10.3390/](https://doi.org/10.3390/bios8020047) [bios8020047](https://doi.org/10.3390/bios8020047)
- <span id="page-6-19"></span>Arduini F, Ricci F, Tuta CS, Moscone D, Amine A, Palleschi G (2006) Detection of carbamic and organophosphorous pesticides in water samples using a cholinesterase biosensor based

on Prussian Blue-modifed screen-printed electrode. Anal Chim Acta 580:155–162.<https://doi.org/10.1016/j.aca.2006.07.052>

- <span id="page-6-14"></span>Aspelin L (1994) Pesticide industry sales and usage, 1992 and 1993 market estimates. U.S. Environmental Protection Agency, Washington, DC.<https://nepis.epa.gov/Exe/ZyNET.exe/>
- <span id="page-6-13"></span>Badihi-Mossberg M, Buchner V, Rishpon J (2007) Electrochemical biosensors for pollutants in the environment. Elecroanalysis 19:19–20.<https://doi.org/10.1002/elan.200703946>
- <span id="page-6-8"></span>Bahadır EB, Sezginturk MK (2015) Electrochemical biosensors for hormone analyses. Biosens Bioelectron 68:62-71. [https://doi.](https://doi.org/10.1016/j.bios.2014.12.054) [org/10.1016/j.bios.2014.12.054](https://doi.org/10.1016/j.bios.2014.12.054)
- <span id="page-6-3"></span>Barrocas PRG, Vasconcellos ACS, Duque SS, Santos LMG, Jacob SC, LauriaFilgueiras AL, Moreira JC (2008) Biossensores para o monitoramento da exposição a poluentesambientais. Cad Saúde Colet Rio de Janeiro 16:677–700
- <span id="page-6-7"></span>Borji H, Ayoub GM, Al-Hindi M, Malaeb L, Hamdan HZ (2020) Nanotechnology to remove polychlorinated biphenyls and polycyclic aromatic hydrocarbons from water: a review. Environ Chem Lett 18:729–746. <https://doi.org/10.1007/s10311-020-00979-x>
- <span id="page-6-0"></span>Campana AL, Florez SL, Nogeura MJ, Fuentes OP, Puentes PR, Cruz JC, Osma JF (2019) Enzyme based electrochemical biosensors for microfuidic platforms to detect pharmaceutical residues in wastewater. Biosensors 9:41. [https://doi.org/10.3390/bios901004](https://doi.org/10.3390/bios9010041) [1](https://doi.org/10.3390/bios9010041)
- <span id="page-6-11"></span>Centi S, Rozum B, Laschi S, Palchetti I, Mascini M (2006) Disposable electrochemical magnetic beads-based immunosensors. Chem Anal 51:963–975
- <span id="page-6-20"></span>Centi S, Silva E, Laschi S, Palchetti I, Mascini M (2007) Polychlorinated biphenyls (PCBs) detection in milk samples by an electrochemical magneto-immunosensor (EMI) coupled to solid-phase extraction (SPE) and disposable low-density arrays. Anal Chim Acta 594:9–16. <https://doi.org/10.1016/j.aca.2007.04.064>
- <span id="page-6-21"></span>Chamundeeswari M, Jeslin J, Verma ML (2019) Nanocarriers for drug delivery applications. Environ Chem Lett 17:849–865. [https://](https://doi.org/10.1007/s10311-018-00841-1) [doi.org/10.1007/s10311-018-00841-1](https://doi.org/10.1007/s10311-018-00841-1)
- <span id="page-6-9"></span>Chee GJ, Nomura Y, Karube I (1999) Biosensor for the estimation of low biochemical oxygen demand. Anal Chim Acta 379:185–191. [https://doi.org/10.1016/S0003-2670\(98\)00680-1](https://doi.org/10.1016/S0003-2670(98)00680-1)
- <span id="page-6-6"></span>Chee GJ, Nomura Y, Ikebukuro K, Karube I (2000) Optical fber biosensor for the determination of low biochemical oxygen demand. Biosens Bioelectron 15:371–376
- <span id="page-6-10"></span>Chen H, Mousty C, Cosnier S, Silveira C, Moura JJG, Almeida MG (2007) Highly sensitive nitrite biosensor based on the electrical wiring of nitrite reductase by [ZnCr-AQS] LDH. Electrochem Commun 9:2240–2245. [https://doi.org/10.1016/j.eleco](https://doi.org/10.1016/j.elecom.2007.05.030) [m.2007.05.030](https://doi.org/10.1016/j.elecom.2007.05.030)
- <span id="page-6-15"></span>Cock LS, Arenas AMZ, Aponte AA (2009) Use of enzymatic biosensors as quality indices: a synopsis of present and future trends in the food industry. Chilean J Agric Res 69:270–280
- <span id="page-6-18"></span>Compagnone D, Ricci A, Del Carlo M, Chiarini M, Pepe A, Sterzo CL (2010) New poly(aryleneethynylene)s as optical active platforms in biosensing. Selective fuorescent detection of Hg(II) obtained by the use of amino acidic groups anchored on conjugated backbones. Microchim Acta 170:313–319. [https://doi.org/10.1007/](https://doi.org/10.1007/s00604-010-0322-4) [s00604-010-0322-4](https://doi.org/10.1007/s00604-010-0322-4)
- <span id="page-6-5"></span>De Benedetto GE, Di Masi S, Penetta A, Malitesta C (2019) Response surface methodology for the optimisation of electrochemical biosensors for heavy metals detection. Biosensors 9:26. [https://doi.](https://doi.org/10.3390/bios9010026) [org/10.3390/bios9010026](https://doi.org/10.3390/bios9010026)
- <span id="page-6-16"></span>DeFrank JJ, Beaudry WT, Cheng TC, Harvey SP, Stroup AN, Szafraniec LL (1993) Screening of halophilic bacteria and Alteromonas species for organophosphorus hydrolyzing enzyme activity. Chem Biol Interact 87:141
- <span id="page-6-17"></span>Dhull V, Gahlaut A, Dilbaghi N, Hooda V (2013) Acetylcholinesterase biosensors for electrochemical detection of

 $\circled{2}$  Springer

- <span id="page-7-7"></span>Dubey S, Shri M, Gupta A, Rani V, Chakrabarty D (2018) Toxicity and detoxifcation of heavy metals during plant growth and metabolism. Environ Chem Lett 16:1169–1192. [https://doi.](https://doi.org/10.1007/s10311-018-0741-8) [org/10.1007/s10311-018-0741-8](https://doi.org/10.1007/s10311-018-0741-8)
- <span id="page-7-10"></span>Durrieu C, Tran-Minh C (2002) Optical algal biosensor using alkaline phosphatase for determination of heavy metals. Environ Res Sect B 51:206–209. [https://doi.org/10.1006/](https://doi.org/10.1006/eesa.2001.2140) [eesa.2001.2140](https://doi.org/10.1006/eesa.2001.2140)
- <span id="page-7-13"></span>Farre M, Pasini O, Carmen Alonso M, Castillo M, Barcelo D (2001) Toxicity assessment of organic pollution in wastewaters using a bacterial biosensor. Anal Chim Acta 426:155–165. [https://doi.](https://doi.org/10.1016/S0003-2670(00)00826-6) [org/10.1016/S0003-2670\(00\)00826-6](https://doi.org/10.1016/S0003-2670(00)00826-6)
- <span id="page-7-31"></span>Fernández H, Arévalo FJ, Granero AM, Robledo SN, Nieto CHD, Riberi WI, Zon MA (2017) Electrochemical biosensors for the determination of toxic substances related to food safety developed in South America: mycotoxins and herbicides. Chemosensors 5:23.<https://doi.org/10.3390/chemosensors5030023>
- <span id="page-7-21"></span>Gavlasova P, Kuncova G, Kochankova L, Mackova M (2008) Whole cell biosensor for polychlorinated biphenyl analysis based on optical detection. Int Biodeterior Biodegrad 62:304–312. [https](https://doi.org/10.1016/j.ibiod.2008.01.015) [://doi.org/10.1016/j.ibiod.2008.01.015](https://doi.org/10.1016/j.ibiod.2008.01.015)
- <span id="page-7-24"></span>Granek V, Rishpon J (2002) Detecting endocrine-disrupting compounds by fast impedance measurements. Environ Sci Technol 36:1574–1578.<https://doi.org/10.1021/es015589w>
- <span id="page-7-5"></span>Hart JP, Abass AK, Cowell D (2002) Development of disposable amperometric sulfur dioxide biosensors based on screen printed electrodes. Biosens Bioelectron 17:389
- <span id="page-7-6"></span>Herschkovitz Y, Eshkenazi I, Campbell CE, Rishpon J (2000) An electrochemical biosensor for formaldehyde. J Electroanal Chem 491:182. [https://doi.org/10.1016/S0022-0728\(00\)00170-4](https://doi.org/10.1016/S0022-0728(00)00170-4)
- <span id="page-7-28"></span>Hondred JA, Breger JC, Alves NJ, Trammell SA, Walper SA, Medintz IL, Clausen JC (2018) Printed graphene electrochemical biosensors fabricated by Inkjet Maskless Lithography for rapid and sensitive detection of organophosphates. ACS Appl Mater Interfaces 10:11125–11134. <https://doi.org/10.1021/acsami.7b19763>
- <span id="page-7-34"></span>Hwang E, Song J, Zhang J (2019) Integration of nanomaterials and bioluminescence resonance energy transfer techniques for sensing biomolecules. Biosensors 9:42. [https://doi.org/10.3390/bios9](https://doi.org/10.3390/bios9010042) [010042](https://doi.org/10.3390/bios9010042)
- <span id="page-7-9"></span>Ion AC, Ion I, Culetu A (2010) Carbon-based nanomaterials: environmental applications. Univ Politehn Bucharest 38:129–132
- <span id="page-7-2"></span>Jain Y, Goel A, Rana C, Sharma N, Verma ML, Jana AK (2010) Biosensors, types and applications. International conference on biomedical engineering and assistive technologies at National Institute of Technology, Jalandhar, India, December 17–19, 2010. [https://www.researchgate.net/publication/281204612\\_Biosensors](https://www.researchgate.net/publication/281204612_Biosensors_types_and_applications) [\\_types\\_and\\_applications](https://www.researchgate.net/publication/281204612_Biosensors_types_and_applications)
- <span id="page-7-14"></span>Jouanneau S, Recoules L, Durand MJ, Boukabache A, Picot V, Primault Y, Lakel A, Sengelin M, Barillon B, Thouand G (2014) Methods for assessing biochemical oxygen demand (BOD): a review. Water Res 49:62–82. [https://doi.org/10.1016/j.watre](https://doi.org/10.1016/j.watres.2013.10.066) [s.2013.10.066](https://doi.org/10.1016/j.watres.2013.10.066)
- <span id="page-7-33"></span>Jyoti A, Tomar RS (2017) Detection of pathogenic bacteria using nanobiosensors. Environ Chem Lett 15:1–6. [https://doi.org/10.1007/](https://doi.org/10.1007/s10311-016-0594-y) [s10311-016-0594-y](https://doi.org/10.1007/s10311-016-0594-y)
- <span id="page-7-0"></span>Kalyani N, Goel S, Jaiswal S (2020) On-site sensing of pesticides using point-of-care biosensors: a review. Environ Chem Lett. [https://](https://doi.org/10.1007/s10311-020-01070-1) [doi.org/10.1007/s10311-020-01070-1](https://doi.org/10.1007/s10311-020-01070-1)
- <span id="page-7-15"></span>Kara S, Keskinler B, Erhan E (2008) A novel microbial BOD biosensor developed by the immobilization of *P. Syringae* in micro-cellular polymers. J Chem Technol Amp Biotechnol 84:511–518. [https://](https://doi.org/10.1002/jctb.2071) [doi.org/10.1002/jctb.2071](https://doi.org/10.1002/jctb.2071)
- <span id="page-7-20"></span>Khadro B, Namour P, Bessueille F, Leonard D, Jafrezic-Renault N (2008) Enzymatic conductometric biosensor based on PVC

membrane containing methyl viologen/nafon®/nitrate reductase for determination of nitrate in natural water samples. Sens Mater 20:267–279

- <span id="page-7-30"></span>Kim HS, Devarenne TP, Han A (2018) Microfudic systems for microalgal biotechnology: a review. Algal Res 30:149–161. [https://doi.](https://doi.org/10.1016/j.algal.2017.11.020) [org/10.1016/j.algal.2017.11.020](https://doi.org/10.1016/j.algal.2017.11.020)
- <span id="page-7-32"></span>Kurosawa S, Aizawa H, Park JW (2005) Quartz crystal microbalance immunosensor for highly sensitive 2,3,7,8-tetrachlorodibenzo-pdioxin detection in fy ash from municipal solid waste incinerators. Analyst 130:1495–1501.<https://doi.org/10.1039/b506151b>
- <span id="page-7-3"></span>Kuswandi B (2018) Nanobiosensors for detection of micropollutants. Environ Nanotechnol. [https://doi.org/10.1007/978-3-319-76090](https://doi.org/10.1007/978-3-319-76090-2_4)  $-2^4$
- <span id="page-7-17"></span>Kuswandi B (2019) Nanobiosensor approaches for pollutant monitoring. Environ Chem Lett 17:975–990. [https://doi.org/10.1007/](https://doi.org/10.1007/s10311-018-00853-x) [s10311-018-00853-x](https://doi.org/10.1007/s10311-018-00853-x)
- <span id="page-7-18"></span>Kwok NY, Dongb S, Loa W (2005) An optical biosensor for multi-sample determination of biochemical oxygen demand (*BOD)*. Sensors Actuators B Chem 110:289–298. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.snb.2005.02.007) [snb.2005.02.007](https://doi.org/10.1016/j.snb.2005.02.007)
- <span id="page-7-23"></span>Lin SH, Juang RS (2009) Adsorption of phenol and its derivatives from water using synthetic resins and low-cost natural adsorbents: a review. J Environ Manag 90:1336–1349. [https://doi.](https://doi.org/10.1016/j.jenvman.2008.09.003) [org/10.1016/j.jenvman.2008.09.003](https://doi.org/10.1016/j.jenvman.2008.09.003)
- <span id="page-7-22"></span>Luong JHT, Male KB, Glennon JD (2008) Biosensor technology: technology push versus market pull. Biotechnol Adv 26:492–500. <https://doi.org/10.1016/j.biotechadv.2008.05.007>
- <span id="page-7-4"></span>Malhotra S, Vaerma A, Tyagi N, Kumar V (2017) Biosensors: principle, types and applications. IJARIIE 3:3639–3644
- <span id="page-7-8"></span>Malik LA, Bashir A, Qureashi A, Pandith AH (2019) Detection and removal of heavy metal ions: a review. Environ Chem Lett 17:1495–1521.<https://doi.org/10.1007/s10311-019-00891-z>
- <span id="page-7-11"></span>Mallat E, Barzen C, Abuknesha R, Gauglitz G, Barcelo D (2001) Fast determination of paraquat residues in water by an optical immunosensor and validation using capillary electrophoresisultraviolet detection. Anal Chim Acta 427:165–171. [https://doi.](https://doi.org/10.1016/S0003-2670(00)01016-3) [org/10.1016/S0003-2670\(00\)01016-3](https://doi.org/10.1016/S0003-2670(00)01016-3)
- <span id="page-7-16"></span>Marrazza G, Chianella I, Mascini M (1999) Disposable DNA electrochemical biosensors for environmental monitoring. Anal Chim Acta 387:297. [https://doi.org/10.1016/S0003-2670\(99\)00051-3](https://doi.org/10.1016/S0003-2670(99)00051-3)
- <span id="page-7-29"></span>Matejovsky L, Pitschmann V (2018) New carrier made from glass nanofbers for the colorimetric biosensor of cholinesterase inhibitors. Biosensors 8:1–10. <https://doi.org/10.3390/bios8020051>
- <span id="page-7-12"></span>Mazhari BBZ, Agsar D, Prasad MVNA (2017) Development of paper biosensor for the detection of phenol from industrial effluents using bioconjugate of Tyr-AuNPs mediated by novel isolate *Streptomyces tuirus*DBZ39. J Nanomater 2017:1–8. [https://](https://doi.org/10.1155/2017/1352134) [doi.org/10.1155/2017/1352134](https://doi.org/10.1155/2017/1352134)
- <span id="page-7-1"></span>Mishra GK, Sharma V, Mishra RK (2018) Elecrochemical aptasensors for food and environmental safeguarding: a review. Biosensors 8:28.<https://doi.org/10.3390/bios8020028>
- <span id="page-7-19"></span>Moorcroft MJ, Davis J, Compton RG (2001) Detection and determination of nitrate and nitrite: a review. Talanta 54:785–803. [https://](https://doi.org/10.1016/S0039-9140(01)00323-X) [doi.org/10.1016/S0039-9140\(01\)00323-X](https://doi.org/10.1016/S0039-9140(01)00323-X)
- <span id="page-7-25"></span>Moraes NV, Grando MD, Valério DAR, Oliveira DP (2008) Exposiçãoambiental a desreguladoresendócrinos: alteraçõesnahomeostase dos hormôniosesteroidais e tireoideanos. Braz J Toxicol 21:1–8
- <span id="page-7-27"></span>Mulchandani A, Chen W, Mulchandani P, Wang J, Rogers KR (2001) Biosensors for direct determination of organophosphate pesticides. Biosens Bioelectron 16:225–230. [https://doi.org/10.1016/](https://doi.org/10.1016/S0956-5663(01)00126-9) [S0956-5663\(01\)00126-9](https://doi.org/10.1016/S0956-5663(01)00126-9)
- <span id="page-7-26"></span>Muller M, Rabenoellina F, Balaguer P, Patureau D, Lemenach K, Budzinski K (2008) Chemical and biological analysis of endocrinedisruptors hormones and estrogenic activity in an advanced

sewage treatment plant. Environ Toxicol Chem 27:1649–1658. <https://doi.org/10.1897/07-519>

- <span id="page-8-19"></span>Nakamura H, Karube I (2003) Current research activity in biosensors. Anal Bioanal Chem 377:446–468. [https://doi.org/10.1007/s0021](https://doi.org/10.1007/s00216-003-1947-5) [6-003-1947-5](https://doi.org/10.1007/s00216-003-1947-5)
- <span id="page-8-32"></span>Nasiri N, Clarke C (2019) Nanostructured gas sensors for medical and health applications: low to high dimensional materials. Biosensors 9:43.<https://doi.org/10.3390/bios9010043>
- <span id="page-8-11"></span>Nistor C, Rose A, Farré M, Stoica L, Ruzgas T, Wollenberger U, Pfeifer D, Barceló D, Gorton L, Emnéus J (2002) In-feld monitoring of cleaning efficiency in wastewater treatment plants using two phenol-sensitive biosensors. Anal Chim Acta 456:3–17. [https://doi.org/10.1016/S0003-2670\(01\)01015-7](https://doi.org/10.1016/S0003-2670(01)01015-7)
- <span id="page-8-13"></span>Nomura Y, Ikebukuro K, Yokoyama K, Takeuchi T, Arikawa Y, Ohno S, Karube I (1998) Application of a linear alkylbenzene sulfonate biosensor to river water monitoring. Biosens Bioelectron 13:1047. [https://doi.org/10.1016/S0956-5663\(97\)00077-8](https://doi.org/10.1016/S0956-5663(97)00077-8)
- <span id="page-8-12"></span>Philp JC, Balmand S, Hajto E, Bailey MJ, Wiles S, Whiteley AS, Lilley AK, Hajto J, Dunbar SA (2003) Whole cell immobilised biosensors for toxicity assessment of a wastewater treatment plant treating phenolics containing waste. Anal Chim Acta 487:61–74. [https://doi.org/10.1016/S0003-2670\(03\)00358-1](https://doi.org/10.1016/S0003-2670(03)00358-1)
- <span id="page-8-14"></span>Ponomareva ON, Arlyapov VA, Alferov VA, Reshetilov AN (2011) Microbial biosensors for detection of biological oxygen demand: a review. Appl Biochem Microbiol 47:1–11. [https://](https://doi.org/10.1134/S0003683811010108) [doi.org/10.1134/S0003683811010108](https://doi.org/10.1134/S0003683811010108)
- <span id="page-8-22"></span>Power B, Liu X, Germaine KJ, Ryan D, Brazil D, Dowling DN (2011) Alginate beads as a storage, delivery and containment system for genetically modified PCB degrader and PCB biosensor derivatives of *Pseudomonas forescence* F113. J Appl Microbiol 110:1351–1358. [https://doi.org/10.1111/j.1365-2672.2011.04993](https://doi.org/10.1111/j.1365-2672.2011.04993.x) [.x](https://doi.org/10.1111/j.1365-2672.2011.04993.x)
- <span id="page-8-30"></span>Prasad R, Kumar V, Prasad KS (2014) Nanotechnology in sustainable agriculture: present concerns and future aspects. Afr J Biotechnol 13:705–713. <https://doi.org/10.3389/fmicb.2017.01014>
- <span id="page-8-31"></span>Prasad R, Pandey R, Barman I (2016) Engineering tailored nanoparticles with microbes: quo vadis. WIREs Nanomed Nanobiotechnol 8:316–330.<https://doi.org/10.1002/wnan.1363>
- <span id="page-8-0"></span>Prasad R, Bhattacharya A, Nguyan QD (2017) Nanotechnology in sustainable agriculture: recent developments, challenges and perspectives. Front Microbiol 8:1014. [https://doi.org/10.3389/](https://doi.org/10.3389/fmicb.2017.01014) [fmicb.2017.01014](https://doi.org/10.3389/fmicb.2017.01014)
- <span id="page-8-21"></span>Pribyl J, Hepel M, Skládal P (2006) Piezoelectric immunosensors for polychlorinated biphenyls operating in aqueous and organic phases. Sensors Actuators B Chem 113:900–910. [https://doi.](https://doi.org/10.1016/j.snb.2005.03.077) [org/10.1016/j.snb.2005.03.077](https://doi.org/10.1016/j.snb.2005.03.077)
- <span id="page-8-3"></span>Rani V, Verma ML (2020) Biosensor applications in the detection of heavy metals, polychlorinated biphenyls, biological oxygen demand, endocrine disruptors, hormones, dioxin, and phenolic and organophosphorus compounds. In: Tuteja SK, Arora D, Dilbaghi N, Lichtfouse E (eds) Nanosensors for environmental applications: Environmental Chemistry for a Sustainable World, vol 43. Springer, Cham, pp 1–28. [https://doi.org/10.1007/978-3-](https://doi.org/10.1007/978-3-030-38101-1_1) [030-38101-1\\_1](https://doi.org/10.1007/978-3-030-38101-1_1)
- <span id="page-8-10"></span>Rathnayake IVN, Megharaj M, Bolan N, Naidu R (2009) Tolerance of heavy metals by gram positive soil bacteria. World Acad Sci Eng Technol 53:1185–1189
- <span id="page-8-2"></span>Rodriguez-Mozaz S, Marco MP, Alda MJL, Barceló D (2004) Biosensors for environmental applications: future development trends. Pure Appl Chem 76:723–752. [https://doi.org/10.1351/pac20](https://doi.org/10.1351/pac200476040723) [0476040723](https://doi.org/10.1351/pac200476040723)
- <span id="page-8-18"></span>Rodriguez-Mozaz S, Marco MP, Alda MJL, Barceló D (2005) A global perspective: biosensors for environmental monitoring. Talanta 65:291–297. <https://doi.org/10.1016/j.talanta.2004.07.006>
- <span id="page-8-23"></span>Rodriguez-Mozaz S, Alda MJL, Barceló D (2006) Biosensors as useful tools for environmental analysis and monitoring. Anal

Bioanal Chem 386:1025–1041. [https://doi.org/10.1007/s0021](https://doi.org/10.1007/s00216-006-0574-3) [6-006-0574-3](https://doi.org/10.1007/s00216-006-0574-3)

- <span id="page-8-4"></span>Rogers KR (2006) Recent advances in biosensor techniques for environmental monitoring. Anal Chim Acta 568:222–231. [https://](https://doi.org/10.1016/j.aca.2005.12.067) [doi.org/10.1016/j.aca.2005.12.067](https://doi.org/10.1016/j.aca.2005.12.067)
- <span id="page-8-6"></span>Rogers KR, Gerlach CL (1996) Environmental biosensors: a status report. Environ Sci Technol 30:486–491. [https://doi.org/10.1021/](https://doi.org/10.1021/es962481l) [es962481l](https://doi.org/10.1021/es962481l)
- <span id="page-8-15"></span>Salehi ASM, Ookyang S, Earl CC, Tang MJS, Hunt P, Smith MT, Wood W, Bundy BC (2018) Biosensing estrogenic endocrine disruptors in human blood and urine: a RAPID cell-free protein synthesis approach. Toxicol Appl Pharmacol 345:19–25. [https](https://doi.org/10.1016/j.taap.2018.02.016) [://doi.org/10.1016/j.taap.2018.02.016](https://doi.org/10.1016/j.taap.2018.02.016)
- <span id="page-8-25"></span>Samsonova JV, Uskova NA, Andresyuk AN, Franek M, Elliott CT (2004) Biacorebiosensor immunoassay for 4-nonylphenols: assay optimization and applicability for shellfsh analysis. Chemosphere 57:975–985. [https://doi.org/10.1016/j.chemospher](https://doi.org/10.1016/j.chemosphere.2004.07.028) [e.2004.07.028](https://doi.org/10.1016/j.chemosphere.2004.07.028)
- <span id="page-8-1"></span>Saravanan A, Kumar PS, Hemavathy RV, Jeevantham S, Kamalesh R, Sneha S, Yaashikaa PR (2020) Methods of detection of foodborne pathogens: a review. Environ Chem Lett 1:1. [https://doi.](https://doi.org/10.1007/s10311-020-01072-z) [org/10.1007/s10311-020-01072-z](https://doi.org/10.1007/s10311-020-01072-z)
- <span id="page-8-8"></span>Sayago I, Aleixandre M, Santos JP (2019) Development of tin oxidebased nanosensors for electronic nose environmental applications. Biosensors 9:21. <https://doi.org/10.3390/bios9010021>
- <span id="page-8-24"></span>Scognamiglio V, Pezzotti I, Pezzotti G, Cano J, Manfredonia I, Buonasera K (2012) Towards an integrated biosensor array for simultaneous and rapid multi-analysis of endocrine disrupting chemicals. Anal Chim Acta 751:161–170. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.aca.2012.09.010) [aca.2012.09.010](https://doi.org/10.1016/j.aca.2012.09.010)
- <span id="page-8-16"></span>Seifert M, Haindl S, Hock B (1999) Development of an enzyme linked receptor assay (ELRA) for estrogens and xenoestrogens. Anal Chim Acta 386(3):191–199. [https://doi.org/10.1016/S0003](https://doi.org/10.1016/S0003-2670(99)00044-6) [-2670\(99\)00044-6](https://doi.org/10.1016/S0003-2670(99)00044-6)
- <span id="page-8-5"></span>Sharpe M (2003) It's a bug's life: biosensors for environmental monitoring. J Environ Monit 5:109–113
- <span id="page-8-20"></span>Shpigun LK, Andryukhina EY (2019) A new electrochemical sensor for direct detection of purine antimetabolites and DNA degradation. J Anal Methods Chem 2019:1–8. [https://doi.](https://doi.org/10.1155/2019/1572526) [org/10.1155/2019/1572526](https://doi.org/10.1155/2019/1572526)
- <span id="page-8-26"></span>Simonian AL, Flounders AW, Wild JR (2004) FET-based biosensors for the direct detection of organophosphate neurotoxins. Electroanalysis 16:1896–1906.<https://doi.org/10.1002/elan.200403078>
- <span id="page-8-27"></span>Singh S, Kumar V, Chauhan A, Datta S, Wani AB, Singh N, Singh J (2018) Toxicity, degradation and analysis of the herbicide atrazine. Environ Chem Lett 16:211–237. [https://doi.org/10.1007/](https://doi.org/10.1007/s10311-017-0665-8) [s10311-017-0665-8](https://doi.org/10.1007/s10311-017-0665-8)
- <span id="page-8-9"></span>Srivastava NK, Majumder CB (2008) Novel biofltration methods for the treatment of heavy metals from industrial wastewater. J Hazard Mater 151:1–8. [https://doi.org/10.1016/j.jhazm](https://doi.org/10.1016/j.jhazmat.2007.09.101) [at.2007.09.101](https://doi.org/10.1016/j.jhazmat.2007.09.101)
- <span id="page-8-29"></span>Srivastava AK, Dev A, Karmakar S (2018) Nanosensors and nanobiosensors in food and agriculture. Environ Chem Lett 16:161–182. <https://doi.org/10.1007/s10311-017-0674-7>
- <span id="page-8-28"></span>Starodub NF, Dzantiev BB, Starodub VM, Zherdev AV (2000) Immunosensor for the determination of herbicide simazine based on an ion selective feld efect transistor. Anal Chem Acta 424:37–43. [https://doi.org/10.1016/S0003-2670\(00\)01143-0](https://doi.org/10.1016/S0003-2670(00)01143-0)
- <span id="page-8-17"></span>Sticher P, Jaspers MC, Stemmler K, Harms H, Zehnder AJ, van der Meer JR (1997) Development and characterization of a wholecell bioluminescent sensor for bioavailable middle-chain alkanes in contaminated groundwater samples. Appl Environ Microbiol 63:4053–4060
- <span id="page-8-7"></span>Tayanc M (2000) An assessment of spatial and temporal variation of sulfur dioxide levels over Istanbul, Turkey. Environ Pollut 107:61–69. [https://doi.org/10.1016/S0269-7491\(99\)00131-1](https://doi.org/10.1016/S0269-7491(99)00131-1)
- <span id="page-9-3"></span>Tijani JO, Fatoba OO, Babajide OO, Petrik LF (2016) Pharmaceuticals, endocrine disruptors, personal care products, nanomaterials and perfuorinated pollutants: a review. Environ Chem Lett 14:27–49. <https://doi.org/10.1007/s10311-015-0537-z>
- <span id="page-9-7"></span>Tunesi MM, Kalwer N, Abbas MW, Karakus S, Soomro RA, Kilislioglu A, Abro MI, Hallam AR (2018) Functionalised CuO nanostructures for the detection of organophosphorus pesticides: a non-enzymatic inhibition approach coupled with nano-scale electrode engineering to improve electrode sensitivity. Sensors Actuators B Chem 260:480–489. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.snb.2018.01.084) [snb.2018.01.084](https://doi.org/10.1016/j.snb.2018.01.084)
- <span id="page-9-9"></span>Velasco-García MN, Mottram T (2003) Biosensor technology addressing agricultural problems. Biosyst Eng 84:1–12. [https://doi.](https://doi.org/10.1016/S1537-5110(02)00236-2) [org/10.1016/S1537-5110\(02\)00236-2](https://doi.org/10.1016/S1537-5110(02)00236-2)
- <span id="page-9-0"></span>Verma ML (2017a) Nanobiotechnology advances in enzymatic biosensors for the agri-food industry. Environ Chem Lett 15:555–560. <https://doi.org/10.1007/s10311-017-0640-4>
- <span id="page-9-2"></span>Verma ML (2017b) Enzymatic nanobiosensors in the agricultural and food industry. In: Ranjan S, Dasgupta N, Lichfouse E (eds) Nanoscience in food and agriculture 4, Sustainable agriculture reviews, vol 24. Springer, Cham, pp 229–245. ISBN 978-3-319-53111-3
- <span id="page-9-4"></span>Verma N, Singh M (2005) Biosensors for heavy metals. Biometals 18:121–129. <https://doi.org/10.1007/s10534-004-5787-3>
- <span id="page-9-1"></span>Verma ML, Kanwar SS, Jana AK (2010) Bacterial biosensors for measuring availability of environmental pollutants. In: BEATS 2010 proceedings of 2010 international conference on biomedical engineering and assistive Technol Jalandhar India, 2010, pp 1–7. [http://www.bmeindia.org/paper/BEATs2010\\_149](http://www.bmeindia.org/paper/BEATs2010_149)
- <span id="page-9-13"></span>Verma ML, Kumar S, Das A, Randhawa JS, Chamundeeswari M (2019) Chitin and chitosan-based support materials for enzyme immobilization and biotechnological applications. Environ Chem Lett 18:315–323. <https://doi.org/10.1007/s10311-019-00942-5>
- <span id="page-9-12"></span>Verma ML, Dhanya BS, Sukriti Thakur M, Jeslin J, Kushwaha R (2020) Carbohydrate and protein based biopolymeric nanoparticles: current status and biotechnological applications. Int J Biol Macromol 154:390–412. [https://doi.org/10.1016/j.ijbio](https://doi.org/10.1016/j.ijbiomac.2020.03.105) [mac.2020.03.105](https://doi.org/10.1016/j.ijbiomac.2020.03.105)
- <span id="page-9-10"></span>Vismara C, Garavaglia A (1997) 4-chloro-2methylphenoxyaceticacid containing compounds. Genotoxicity evaluation by Mutatox assay and comparison with acute (Microtox) and embryo (FETAX) toxicities. Bull Environ Contam Toxicol 58:582–588
- <span id="page-9-5"></span>Waring RH, Harris RM (2005) Endocrine disrupters: a human risk? Mol Cell Endocrinol 244:2–9. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.mce.2005.02.007) [mce.2005.02.007](https://doi.org/10.1016/j.mce.2005.02.007)
- <span id="page-9-6"></span>Xu YF, Velasco-Garcia M, Mottram TT (2005) Quantitative analysis of the response of an electrochemical biosensor for progesterone in milk. Biosens Bioelectron 20:2061–2070. [https://doi.](https://doi.org/10.1016/j.bios.2004.09.009) [org/10.1016/j.bios.2004.09.009](https://doi.org/10.1016/j.bios.2004.09.009)
- <span id="page-9-11"></span>Yulaev MF, Sitdikov RA, Dmitrieva NM, Yazynina EV, Zherdev AV, Dzantiev BB (2001) Development of a potentiometric immunosensor for herbicide simazine and its application for food testing. Sensors Actuators 75:129–135. [https://doi.org/10.1016/](https://doi.org/10.1016/S0925-4005(01)00551-2) [S0925-4005\(01\)00551-2](https://doi.org/10.1016/S0925-4005(01)00551-2)
- <span id="page-9-8"></span>Zhang Y, Arugula MA, Wales M, Wild J, Simonian AL (2015) A novel layer-by-layer assembled multi-enzyme/CNT biosensor for discriminative detection between organophosphorus and non-organophosphorus pesticides. Biosens Bioelectron 67:287–295. [https](https://doi.org/10.1016/j.bios.2014.08.036) [://doi.org/10.1016/j.bios.2014.08.036](https://doi.org/10.1016/j.bios.2014.08.036)

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