REVIEW

Ventilatory and cerebrovascular regulation and integration at high‑altitude

Ryan L. Hoiland1 [·](http://orcid.org/0000-0002-5657-0059) Connor A. Howe1 · Geof B. Coombs1 · Philip N. Ainslie1

Received: 19 October 2017 / Accepted: 9 March 2018 / Published online: 24 March 2018 © Springer-Verlag GmbH Germany, part of Springer Nature 2018

Abstract

Ascent to high-altitude elicits compensatory physiological adaptations in order to improve oxygenation throughout the body. The brain is particularly vulnerable to the hypoxemia of terrestrial altitude exposure. Herein we review the ventilatory and cerebrovascular changes at altitude and how they are both implicated in the maintenance of oxygen delivery to the brain. Further, the interdependence of ventilation and cerebral blood fow at altitude is discussed. Following the acute hypoxic ventilatory response, acclimatization leads to progressive increases in ventilation, and a partial mitigation of hypoxemia. Simultaneously, cerebral blood fow increases during initial exposure to altitude when hypoxemia is the greatest. Following ventilatory acclimatization to altitude, and an increase in hemoglobin concentration—which both underscore improvements in arterial oxygen content over time at altitude—cerebral blood fow progressively decreases back to sea-level values. The complimentary nature of these responses (ventilatory, hematological and cerebral) lead to a tightly maintained cerebral oxygen delivery while at altitude. Despite this general maintenance of global cerebral oxygen delivery, the manner in which this occurs refects integration of these physiological responses. Indeed, ventilation directly infuences cerebral blood fow by determining the prevailing blood gas and acid/base stimuli at altitude, but cerebral blood fow may also infuence ventilation by altering central chemoreceptor stimulation via central $CO₂$ washout. The causes and consequences of the integration of ventilatory and cerebral blood flow regulation at high altitude are outlined.

Keywords Cerebral blood flow · Ventilation · High-altitude · Acclimatization

Introduction

The reduced barometric pressure associated with ascent to high-altitude necessitates compensatory adaptations to mitigate hypoxia-related decrements in physiological functioning. Of the multitude of physiological adaptations that occur, hyperventilation and further ventilatory acclimatization stand alone as the most important $[1, 2]$ $[1, 2]$ $[1, 2]$ $[1, 2]$ (Fig. [1](#page-1-0)). This response reduces the drop in the partial pressure of arterial oxygen $(PaO₂)$ that would occur without any alteration in ventilation (V_E) , while ventilatory acclimatization further and progressively increases $PaO₂$ over time at altitude. Slower hematological changes leading to increased

hemoglobin mass and concentration (Hb) are also important in partially mitigating the drop in arterial oxygen content $(CaO₂)$ associated with altitude. The importance of these two changes is clearly visible in the equation below, whereby PaO₂ primarily influences CaO₂ via its impact on arterial oxyhemoglobin saturation (SaO₂), and (Hb) impacts $CaO₂$ through O_2 binding—both Sa O_2 and (Hb) are directly proportional to $CaO₂$ (Eq. [1\)](#page-0-0):

$$
CaO2(mL · dL-1) = 1.34 \times [Hb](g · dL-1) \times SaO2(%) + 0.003 \times PaO2(mmHg)
$$
 (1)

where 1.34 is the oxygen binding capacity of hemoglobin, and 0.003 is the solubility of $O₂$ in blood. Given the brain's inordinate metabolism for its comparatively small mass and limited oxygen and substrate storage capacity, it is particularly vulnerable in hypoxia and, therefore, at altitude. Testament to this vulnerability is that as little as a 5-s interruption of cerebral blood flow (CBF) can lead to uncon-sciousness [[3](#page-9-2)]. Consequently, the brain is reliant on a consistently adequate cerebral oxygen delivery $(CDO₂)$ to avoid

 \boxtimes Ryan L. Hoiland ryanleohoiland@gmail.com

¹ Centre for Heart, Lung and Vascular Health, School of Health and Exercise Sciences, University of British Columbia-Okanagan Campus, 3333 University Way, Kelowna, BC V1V 1V7, Canada

Fig. 1 Integrative physiological changes during acclimatization to very high altitude (~5000 m). Ascent to altitude stimulates a multitude of physiological adaptations. The immediate response to hypoxia involves a brisk increase in ventilation (V_E) (red line) and cerebral blood flow (CBF) (dark blue line), with a concurrent drop in $PaCO₂$ (light blue line). Hypoxic ventilatory decline ensues leading to a small attenuation of the drop in PaCO₂. Throughout acclimatization, V_E and CBF begin to progressively increase and decrease, respectively. Because of the respiratory alkalosis that follows the hypoxic ventilatory response, arterial and CSF pH (denoted by vertical grey bars) is increased; however, renal bicarbonate $(HCO_3^-$, purple line) excretion increases progressively leading to a stabilization of pH, and potential mitigation of respiratory alkalosis as acclimatization progresses. Finally, hematocrit (HCT) increases following ascent to altitude and throughout acclimatization. This fgure represents general trends based on data from: [\[6](#page-9-5), [51](#page-10-0), [60,](#page-10-1) [65,](#page-10-2) [92](#page-11-0), [93](#page-11-1)]

an energetic crisis and/or cell death. The maintenance of $CDO₂$ is dependent on adequate CBF and $CaO₂$ as demonstrated in Eq. [2:](#page-1-1)

$$
CDO2(mL \cdot min^{-1}) = [CBF (mL \cdot min^{-1}) \times CaO2(mL \cdot dL^{-1})]/1
$$

Thus, cerebral homeostasis at high-altitude hinges upon an increase in CBF, primarily through hypoxic cerebral vasodilation, and a mitigation of hypoxemia, primarily through increased V_E and slower acting hematological changes (Fig. [2](#page-1-2)). However, an additional consequence of elevated V_E is that hypocapnia ensues resulting in cerebral vasoconstriction; therefore, depending on the degree of metabolic compensation for the respiratory alkalosis, this may act to constrain elevations in $CDO₂$. Although acclimatization to high altitude encompasses the integration of multiple physiological responses (Fig. [1](#page-1-0)), these physiological responses may difer slightly from those induced by normobaric hypoxia (even when matched for the same P_1O_2 —reviewed in [[4\]](#page-9-3)). Nevertheless, with particular focus given to high altitude, we provide a brief overview of acid–base balance and then focus our discussion on the causes and consequences of ventilatory and CBF regulation and the potential inter-dependence of these two physiological pathways. The independent and combined influences of these two pathways on $CDO₂$ are further considered.

Fig. 2 Contribution of changes in PaO₂ and $[Hb]$ in the increase in CaO₂ during acclimatization at altitude. Upon exposure to altitude, PaO₂ is reduced, thereby reducing $SaO₂$ and $CaO₂$. Hyperventilation mitigates, but does not alleviate this drop in PaO₂. Ventilatory acclimatization to high altitude contributes to progressive increases in $V_{\rm E}$ and leads to elevations in PaO₂, SaO₂, and CaO₂. Respiratory alkalosis-induced diuresis leads to the early increase in [Hb], with erythropoiesis contributing to later increases in [Hb], both aiding in the increase of $CaO₂$ across acclimatization. Determined using data from [[6\]](#page-9-5)

Acid–base balance at altitude

Following hypoxic stimulation of the carotid bodies and consequent hyperventilation, the partial pressure of arterial carbon dioxide (PaCO₂) is reduced and both arterial and CSF pH are elevated. Laboratory investigations have

 $CDO₂(mL \cdot min^{-1}) = [CBF (mL \cdot min^{-1}) \times CaO₂(mL \cdot dL^{-1})]/100.$ (2) CBF (\times CaO₂ $\frac{1}{100}$.

> demonstrated that within hours of the onset of respiratory alkalosis, excretion of HCO_3^- is upregulated and arterial pH begins to decrease back to normal values [\[5](#page-9-4)]. This excretion of HCO_3^- has been demonstrated to continue for > 2 weeks at a given altitude in concert with the progressive reductions in PaCO₂ that are characteristic of ventilatory acclimatization [[6,](#page-9-5) [7](#page-9-6)]. Although measures of lumbar CSF may not directly refect cisternal and/or extracellular medullary pH [\[8](#page-9-7)], lumbar CSF samples comprise the majority of data assessing central acid–base balance at altitude [[7,](#page-9-6) [9–](#page-9-8)[11](#page-9-9)]. Elimination of $HCO₃⁻$ from CSF follows that of the arterial blood $[9, 11]$ $[9, 11]$ $[9, 11]$ $[9, 11]$ indicating that passive exchange of $CO₂$ across the blood brain barrier and subsequent re-equilibration of the reaction between CO_2 and HCO_3^- leads to changes in CSF $[HCO₃⁻]$ and pH (Eq. [3](#page-1-3)):

$$
CO_2 + H_2O \stackrel{Carbonic Anhydrase}{\rightleftharpoons} H_2CO_3 \rightleftharpoons HCO_3^- + H^+ \tag{3}
$$

How changes in PCO_2 and $[HCO_3^-]$ alter acid-base status of the CSF can be expressed mathematically by the

Henderson-Hasselbalch equation. Using the known pKa (i.e., reaction constant) at 37 $\mathrm{^{\circ}C}$ of 6.1 for the above reac-tion (Eq. [3](#page-1-3)), the relationship between pH, CO_2 and HCO_3^- is as follows $(Eq. 4)$ $(Eq. 4)$ $(Eq. 4)$:

$$
pH = 6.1 + log \frac{[HCO_3^-]}{(0.03 \cdot PCO_2)}
$$
 (4)

Therefore, changes in pH (arterial or CSF) at altitude are contingent on the concurrent changes in PCO_2 and $[HCO_3^-]$. Following renal compensation for the respiratory alkalosis, any given change in $PCO₂$ will result in a larger change in pH (Fig. [3](#page-2-1)). Additional calculations derived from the modifed Stewart Model [\[12\]](#page-9-10) indicate that weakly dissociated protein anions (phosphate and albumin), which increase in concentration at altitude [\[13](#page-9-11), [14](#page-9-12)], are also responsible for some correction of respiratory alkalosis and changes in $[HCO_3^-]$ [[14\]](#page-9-12). The physiological implications of these changes in acid–base balance are discussed in the following sections.

Ventilatory acclimatization

During and following ascent to high altitude, ventilatory acclimatization is evidenced by a progressive increase in $V_{\rm E}$ (and alveolar ventilation) over the first days and/or weeks at altitude. This increase in V_E leads to a progressive reduction in $PaCO₂$ and a partial mitigation of the drop in $PaO₂$ [[2\]](#page-9-1). Thus, acclimatized individuals will have a higher $PaO₂$ and lower PaCO₂ than unacclimatized individuals at any given altitude (Fig. [4](#page-2-2)). The time required to achieve

Fig. 3 Relationship between $PCO₂$ and pH with varying $[HCO₃^-]$. This fgure highlights how, according to the Henderson-Hasselbalch equation, the magnitude of pH changes with varying $PCO₂$ are augmented at altitude following renal compensation for respiratory alkalosis. At sea level, where $[HCO₃⁻]$ approximates 25 meq/L [[43](#page-10-3)], an increase in resting $PaCO₂$ from 40 to 41 mmHg would lead to a reduction in pH of 0.01. Conversely, at altitude where $[HCO₃⁻]$ may approximate 19 meq/L, an increase in resting $PaCO₂$ from 20 to 21 mmHg would lead to a reduction in pH of 0.02

ventilatory acclimatization varies with altitude ranging from approximately 4 days at \sim 3000 m [[2\]](#page-9-1), 8 days at 4000 m [[15](#page-9-13)], \geq 2 weeks at ~5000 m [[6](#page-9-5)], and between 4 and 6 weeks at altitudes above 6000 m $[16]$ $[16]$. It is important to note that, although ventilatory acclimatization might occur, it will never be enough to return $PaO₂$ back to sea-level values. Rather, ventilatory acclimatization refects the near maximal increase in PaO₂ for a given altitude as indicated via the alveolar gas equation—this is limited by P_1O_2 .

Ventilatory acclimatization to altitude occurs in response to combined infuences from central and peripheral chemoreception. Fifty years ago, it was postulated that CSF pH was the primary regulator of ventilatory acclimatization at altitude [\[10](#page-9-15)]; however, it was soon after realised that ventilatory acclimatization occurs irrespective of alkaline CSF and jugular venous blood [\[7](#page-9-6), [9](#page-9-8)]. At present, the role of CSF pH has been more judiciously considered [[18\]](#page-9-16), and data indicate the peripheral chemoreceptors are essential for ventilatory acclimatization $[18–21]$ $[18–21]$. Acid–base changes should, however, not be fully ignored, as reduced HCO_3^- in the blood and CSF mitigates the rise in medullary pH and withdrawal of central chemoreceptor stimulation. Hence, the past 50 years have seen the adoption of carbonic anhydrase inhibitors (e.g., acetazolamide) as efective prophylactic drugs for acute mountain sickness by accelerating renal compensation, thus permitting central chemoreceptor stimulation (via low CSF pH) of ventilation to continue less inhibited [[22](#page-9-18)].

Hypoxic ventilatory response at altitude

The maintenance of $PaO₂$ is paramount for sustaining normal $CDO₂$ at high altitude; therefore, the hypoxic ventilatory

Fig. 4 Effect of altitude acclimatization on alveolar gas composition. Closed (black) symbols are unacclimatized. Open (white) symbols are acclimatized. Redrawn from original data of [[2](#page-9-1), [94](#page-11-2), [95](#page-11-3)] and Wagner et al. [[17](#page-9-19)]. Iso-altitude lines were calculated using the ideal alveolar gas equation and an assumed respiratory exchange ratio of 0.85. With permission from [\[93\]](#page-11-1)

response (HVR) can be considered as one of the most important aspects that mediates ventilatory acclimatization to high altitude (VAH). Under normal conditions, the HVR occurs when PaO₂ falls below \sim 50 mmHg and the resulting hypoxemia is detected by the carotid bodies to stimulate hyperventilation, thereby slightly increasing $PaO₂$. The primary mechanism underlying VAH is a greater sensitivity of the HVR. Well-controlled laboratory data indicate that as little as 8 h (awake) $[23]$ $[23]$ to 24 h (1 day/night) $[24]$ $[24]$ of mild hypoxia (e.g., $P_{I}O_{2} = 127$ mmHg) is adequate to increase isocapnic HVR [[23,](#page-9-20) [24\]](#page-9-21); yet upon arrival to high altitude, HVR appears initially unaltered compared to sea level [\[25](#page-9-22), [26](#page-9-23)]. However, it is well established that the slope of the V_E -SaO₂ response steepens following several days of acclimatization [\[23–](#page-9-20)[28\]](#page-9-24) with this response progressing in a graded manner over time (Fig. [5](#page-3-0)). Potentiation of the HVR can occur with very minor, but sustained, changes in PaO₂ (-10 mmHg $PaO₂$) highlighting that the physiological mechanisms underlying this response are also important at sea level [\[24](#page-9-21)]. The HVR response is apparently similar between men and women [\[27](#page-9-25), [29](#page-9-26)].

Laboratory studies have demonstrated that several hours following an 8 h period of isocapnic or poikilocapnic hypoxia V_E remains elevated during acute hyperoxic breathing, indicating that factors other than acid–base changes (i.e., increased carotid body activity) are responsible for the progressive rise in V_E [\[30](#page-9-27)]. In support of these findings, Barnard et al. observed that $2-3$ h of hypoxia was insufficient to increase the carotid body activity of anaesthetized cats, but that it had increased following chronic exposure [[31\]](#page-9-28). It is also noteworthy that, in line with acute laboratory studies [\[30\]](#page-9-27), a potentiated HVR remains present 5 days after return to sea level from altitude [\[26](#page-9-23)] and that this aspect of acclimatization is partially retained on re-exposure to high altitude [\[32](#page-9-29)]. Hence, it appears that continuous exposure of the carotid bodies to hypoxemia might be required to elicit adjustments of HVR. Interestingly, whereas HVR is acutely

increased in lowlanders after ascending to high altitude, the HVR is approximately halved in long-term (non-native) high altitude residents and is almost completely abolished in lifelong highlanders, suggesting chronic attenuation of the carotid body [[33](#page-9-30)]. The subjects in the aforementioned study lived at high altitude in Colorado, USA (Rocky Mountains); therefore, "native" highlanders may have only populated this region for a couple of generations, indicating that chronic attenuation of HVR can occur independent of genetic adaptation [\[33](#page-9-30)]. The potential infuences of genetic adaptation on ventilatory control at altitude are beyond the scope of this review and outlined elsewhere [\[34](#page-9-31)].

With respect to performance at altitude, a higher HVR measured at sea level has been demonstrated in some successful climbers [\[34–](#page-9-31)[37\]](#page-9-32). However, HVR measurements conducted at sea-level and 5000 m indicate that a lower V_E and HVR—and thus greater ventilatory reserve (i.e., the diference between resting and maximal V_E)—coupled with optimal ventilatory efficiency are predictive of climbing success at extreme altitudes (i.e. summiting Mt. Everest or K2) in those not using supplemental oxygen [[38\]](#page-9-33). These successful climbers are able to ascend to these extreme altitudes without prior maximization of their work of breathing [\[38\]](#page-9-33). These difering fndings are dependent on the altitude at which success is considered and likely also a consequence of the altitude that HVR is measured at, and if hypoxic, the duration of hypoxia prior to measurement. Indeed, as highlighted above, HVR changes with exposure to altitude and thus comparing sea level to high altitude values is difficult, especially given alterations in affer-ent input [\[31](#page-9-28)], central integration [\[39](#page-9-34)] and efferent output [\[40](#page-9-35)] are all likely occurring. The underlying mechanisms infuencing changes in HVR remain difficult to disentangle.

Hypercapnic ventilatory response at altitude

In addition to a direct infuence on ventilation via central chemoreceptor stimulation, there are additional effects

Fig. 5 Changes in HVR over time at high altitude and during laboratory hypoxia. Studies at altitude (flled symbols; solid lines) and in controlled laboratory settings (open symbols; dashed lines) have demonstrated a progressive increase in HVR over time. Further it can be seen the increase in HVR is related to the prevailing level of $P_{ET}O_2$ during hypoxia, as previously noted by Donoghue and colleagues [\[24\]](#page-9-21)

of $PaCO₂$ on the peripheral chemoreceptors, and overall changes in ventilation occur much more readily with alterations in PaCO₂ versus that of PaO₂ only. At high altitude, hyperventilation (i.e., VAH) attenuates the reduction in $PaO₂$ and concomitantly respiratory alkalosis occurs (as described above). Accordingly, another important response implicated in VAH is a leftward shift of the V_E -PaCO₂ relationship to lower PaCO₂ values $[25, 41, 42]$ $[25, 41, 42]$ $[25, 41, 42]$ $[25, 41, 42]$ $[25, 41, 42]$ $[25, 41, 42]$ following a reduction in the ventilatory recruitment threshold at altitude [\[14](#page-9-12)]. This resetting of the hypercapnic ventilatory response (HCVR) occurs due to peripheral chemoreceptor and acid–base adjustments secondary to hypoxia and respiratory alkalosis, respectively [[42\]](#page-9-37). The slope of the V_E -PaCO₂ response also becomes greater at high altitude [[25](#page-9-22), [41](#page-9-36), [43](#page-10-3)], which may facilitate hyperventilation in the face of hypocapnia and suppressed central chemoreceptor stimulation. Importantly, measurement of the efects of central chemoreceptor sensitivity on ventilation at high altitude may be obscured by the logarithmic relationship between PaCO₂ and $[H^+]$ [[41\]](#page-9-36) (see Fig. [3](#page-2-1)). For instance, a 1 mmHg change at a $PaCO₂$ of 20 mmHg following metabolic compensation (i.e., decreased $[HCO_3^-]$) will cause a much greater shift in pH compared to a 1 mmHg change in PaCO₂ at 40 mmHg without any metabolic compensation. Nevertheless, when V_E is indexed against $[H⁺]$ the HCVR is still markedly augmented at altitude [\[43](#page-10-3)]. This latter observation indicates that intrinsic central chemoreceptor sensitivity is increased, and/or a peripheral-central chemoreceptor interaction is augmenting the V_E response to increased $[H^+]$ [\[44](#page-10-4)].

Alterations in peripheral chemoreceptor activity at altitude will also affect the HCVR, which is notionally supported by the observed blunting of the ventilatory response to $CO₂$ in individuals with unilateral and bilateral carotid body resections at sea level [\[45](#page-10-5)]. Furthermore, similar to HVR responses in lowlanders vs. highlanders, HCVR is greater in newcomers to high altitude whereas this response is inhibited in long term residents and high altitude natives [\[15](#page-9-13)]. The HCVR occurs in concert with changes in acid–base balance and cerebrospinal fuid pH and is therefore intimately linked to CBF (see next section "Cerebral blood flow acclimatization"). Collectively, the respiratory responses to reductions in both $PaO₂$ and $PaCO₂$ are extremely important factors underlying VAH. The integration of these responses further facilitates increased respiratory drive, which serves to improve $PaO₂$ values; however, as outlined earlier, full return of $PaO₂$ to sea level values can never occur at high altitude. This fnal point is highlighted by the fact that, despite long-term acclimatization and genetic adaptation, high altitude natives \geq 3000 m above sea-level exhibit severe hypoxemia (PaO₂ < 60 mmHg) with reduced physical and cognitive function compared to sea level populations [[46\]](#page-10-6).

Cerebral blood fow acclimatization

During ascent and initial stay at high altitude, increases in CBF are elicited via reductions in $CaO₂$ and consequent hypoxic cerebral vasodilation (reviewed in [[47\]](#page-10-7)). The magnitude of this increase in CBF is altitude dependent and contingent on the countervailing influences of hypoxia $(\downarrow CaO_2)$ and hypocapnia ($\downarrow PaCO_2$). The resulting cerebral vasodilation occurs throughout the cerebral vasculature from the large extra-cranial cerebral conduit arteries (i.e. ICA and VA) [\[48,](#page-10-8) [49\]](#page-10-9), large intra-cranial arteries (e.g. MCA) [[49–](#page-10-9)[52\]](#page-10-10), through to pial vessels and is adequate to maintain $CDO₂$ (Fig. [6\)](#page-5-0). While hyperventilation mitigates the drop in $CaO₂$ at altitude, it leads to concomitant reductions in PaCO₂ resulting in an increased blood and CSF pH $[53]$ $[53]$ $[53]$ (see "Acid–base balance at altitude"). As the cerebral vasculature is highly sensitive to alterations in pH [\[43](#page-10-3), [54](#page-10-12), [55](#page-10-13)], decreased $PaCO₂$ and increased pH produces a marked vasoconstrictor stimuli [[56\]](#page-10-14) that counteracts the hypoxic vasodilatory stimulus, although the hypoxic stimulus maintains a net vasodilatory outcome $[43]$. Given the interplay of CaO₂ and $PaCO₂$ on CBF regulation at altitude, overall regulation appears dependent on four primary factors: (1) the HVR, (2) the HCVR, (3) hypoxic cerebral vasodilation, and (4) hypocapnic cerebral vasoconstriction. The HVR and HCVR determine the prevailing arterial blood gas stimuli (see "Hypoxic ventilatory response at altitude" and "Hypercapnic ventilatory response at altitude"), while hypoxic cerebral vasodilation and hypocapnic vasoconstriction determine the magnitude by which the cerebral vasculature responds to the arterial blood gas stimuli.

Hypoxic cerebral vasodilation notably occurs via several pathways but appears to be primarily regulated by deoxyhemoglobin-mediated release of ATP and nitric oxide (Fig. [6\)](#page-5-0). These signaling processes help to alleviate cerebral hypoxemia by facilitating increased CBF (reviewed in: [[47](#page-10-7)]). Another important consideration in the regulation of CBF at altitude is sympathetic nervous activity [\[57](#page-10-15)]. However, laboratory study demonstrates no infuence of α_1 -adrenoceptor blockade on CBF during 6 h exposure to hypoxia ($F_1O_2 = 0.11$) [\[58\]](#page-10-16). Conversely, at altitude, combined $α_1$ - and non-selective β-adrenoceptor blockade reduces CBF, yet this is due to a marked drop in mean arterial pressure $\left(\sim 25 \text{ mmHg}\right)$ [\[59](#page-10-17)] and does not appear to have a direct infuence on cerebrovascular tone.

As time at high altitude progresses, CBF stabilizes and starts decreasing toward baseline values within 2–3 days after arrival (Fig. [7\)](#page-6-0) $[60]$. This is a result of both systemic adaptations affecting $CaO₂$ and altered sensitivity of the cerebral vasculature [[6\]](#page-9-5). Comprising the relevant systemic adaptations are: (1) altitude-induced diuresis whereby HCO_3^- is excreted at an elevated rate in an attempt to **Fig. 6** Changes in cerebral blood fow and oxygen delivery following ascent to altitude. Following initial exposure to high altitude, arterial oxygen content $(CaO₂)$ is reduced and cerebral blood fow (CBF) is commensurately increased. Increases and decreases in CaO₂ and CBF, respectively, then mirror each other throughout acclimatization and maintain cerebral oxygen delivery $(CDO₂)$ constant. Alternating vertical bars represent individual days at altitude. Data are labelled based on their corresponding study. 1: [[96](#page-11-4)], 2: [\[97\]](#page-11-5), 3: [\[98\]](#page-11-6), 4: [\[99\]](#page-11-7), 5: [[6](#page-9-5)], 6: [[51](#page-10-0)], 7: [[51](#page-10-0)], 8: [\[48\]](#page-10-8)

compensate for respiratory alkalosis [[61\]](#page-10-18); (2) a substantial loss of plasma volume, decreasing total blood volume, but eliciting a signifcant increase in hematocrit and hemoglobin concentration within days of exposure to altitude [[62\]](#page-10-19). This particular physiological response is a key factor, in addition to ventilatory acclimatization, that drives the early increase in $CaO₂$ from initial hypoxic exposure (Fig. [2\)](#page-1-2) and thus contributes to the progressive decrease in CBF. (3) Following ~ 1 week at altitude, erythropoiesis then increases hemoglobin mass leading to further increases in hematocrit [\[62–](#page-10-19)[65](#page-10-2)]. Another necessary consideration is the infuence of [Hb] on viscosity and the potential implications of viscosity in regulating CBF at altitude. To our knowledge no data have specifcally examined the infuence of viscosity on CBF at altitude; however, the existing data in humans at sea level indicates a likely negligible infuence of viscosity on CBF in hypoxia (reviewed in [[47\]](#page-10-7)). It should be noted, of course, that excessive polycythemia is extremely detrimental to physiological function and health at high altitude in both lowlanders and those with chronic mountain sickness [\[66,](#page-10-20) [67](#page-10-21)].

Alterations in cerebral vascular reactivity to both $O₂$ and $CO₂$ may also be implicated in CBF regulation at high altitude $[6]$ $[6]$. Both increases $[6, 68]$ $[6, 68]$ $[6, 68]$ $[6, 68]$ and no change $[28, 43]$ $[28, 43]$ $[28, 43]$ in hypocapnic cerebral vasoconstriction have been demonstrated upon ascent to and acclimatization at altitude. Therefore, given methodological (technical and logistical) diferences between studies, physiological diferences (e.g., acid–base balance) and the consequent inconsistency of results, it remains relatively unclear how altered hypocapnic vasoconstriction may contribute to the progressively reduced CBF throughout acclimatization. Relative to cerebral sensitivity to O_2 , one study to date has conducted repeated measures indicating that hypoxic cerebral vasodilation is increased at altitude [\[69](#page-10-23)], which is corroborated by more well-controlled laboratory studies [[70\]](#page-10-24). Yet, changes in hypoxic vasodilation with acclimatization have not been examined using volumetric measures of CBF, necessitating judicious interpretation of the currently available data. Nonetheless, the observed increase in hypoxic cerebral vasodilation may counteract the vasoconstrictor stimulus consequent to reduced $PaCO₂$ and underscore the net vasodilatory stimulus and maintained $CDO₂$ observed upon exposure to high altitude. Collectively, global CBF at altitude mirrors changes in $CaO₂$ and although PaCO₂ is a very potent regulator of tone, following initial exposure to altitude, arterial pH is minimally altered (Fig. [1](#page-1-0)). Therefore, it appears $CaO₂$ is the primary factor governing the pattern by which CBF changes following initial exposure to high altitude. Potential alterations in reactivity at altitude may further "fne tune" the observed changes in CBF.

Although global changes in CBF have been shown to maintain $CDO₂$ in hypobaric hypoxia throughout ascent and stay at high altitude, regional diferences in the fow response to altitude have been demonstrated [\[48](#page-10-8), [49](#page-10-9)]. These altitude studies, and those conducted in well-controlled laboratory settings [\[48,](#page-10-8) [49,](#page-10-9) [71,](#page-10-25) [72\]](#page-10-26) display preferential blood fow distribution to the posterior circulation, which perfuses brain regions such as the brainstem, hypothalamus, thalamus and cerebellum [\[71](#page-10-25)]. Despite no relationship between global CBF and AMS $[60]$ $[60]$ and failure of globally maintained CDO₂ to explain cognitive defcits in hypoxia, the aforementioned regionalization of CBF is suggested to be responsible for AMS [[73\]](#page-10-27), as well as cognitive impairment [[72\]](#page-10-26) at altitude. However, given the number of inconsistent studies [\[52,](#page-10-10) [60](#page-10-1)], sufficient data are still lacking with regard to the intricacies

Fig. 7 Putative pathways regulating cerebral blood flow during hypoxia. Increased temperature, erythrocyte deformation, and the conformational change concomitant to transition of oxy- to deoxyhemoglobin all signal erythrocyte mediated release of ATP [[97](#page-11-5)[–103](#page-11-8)]. Released ATP can then bind to the erythrocyte $P2X_7$ receptor in an autocrine fashion to induce erythrocyte mediated EET release [\[104](#page-11-9)], which will increase vascular smooth muscle cell K+ channel con-ductance [[105](#page-11-10)]. The released ATP also binds endothelial P_2Y_2 receptors to initiate a signal cascade involving NO and potentially PGs [[106\]](#page-11-11). Moreover, ATP will breakdown into AMP and subsequently adenosine [\[107\]](#page-11-12) that will also exert a vasodilatory efect on vascular smooth muscle through binding adenosine A_{2A} receptors [\[108](#page-11-13), [109\]](#page-11-14), increasing cAMP levels [\[110](#page-11-15), [111\]](#page-11-16) and also through increasing inward rectify potassium channel conductance [\[108](#page-11-13)]. Prostaglandins, if implicated, bind IP and EP receptors [\[112](#page-11-17)] which increases intra-

of regionalized CBF regulation and its consequent impact(s). Nevertheless, regionally diferential distribution of blood fow likely occurs as a survival mechanism to prioritize delivery to the posterior areas of the brain responsible for controlling functional and homeostatic processes while consequently reducing delivery to areas responsible for higher cognitive function. Although we have intentionally focused on CBF (i.e., infow), it should be noted that a mismatch between cerebral infow and venous outfow is critical in the pathophysiological of AMS [[74\]](#page-10-28) and potentially cerebral

cellular cAMP [[113](#page-11-18)]. NO, derived from the endothelium, through the nitrite reductase activity of erythrocytes [[114\]](#page-11-19), and s-nitrosohemoglobin [\[115,](#page-11-20) [116](#page-12-0)] will lead to increased guanylate cyclase activity and cGMP [[117\]](#page-12-1) as well as directly increase K+ channel conductance [[118\]](#page-12-2). Cyclic nucleotides will upregulate cAMP dependent protein kinase (PKA) and cGMP dependent protein kinase (PKG) activity, which act to inhibit myosin light chain kinase (MLCK; [\[119\]](#page-12-3)), and therefore, reduce smooth muscle tone [\[120](#page-12-4)]. Cyclic nucleotides will also increase potassium channel conductance [\[121\]](#page-12-5), with increased potassium efflux hyperpolarizing cells and reducing activity of voltage gated Ca2+ channels [\[122](#page-12-6)]. Overall, ATP leads to vasodilation that can be conducted through gap junctions [[123](#page-12-7), [124](#page-12-8)]. Green arrows represent activation of a downstream factor, and red arrows represent inhibition of downstream factors. Permission not required. Copyright © 2016, The American Physiological Society

edema [[75](#page-10-29)]. In the latter landmark study, it was reported that $CDO₂$ was maintained via increased arterial inflow (i.e., CBF) and this preceded the development of cerebral edema thus implicating venous outfow restriction as a key mechanism [[75\]](#page-10-29).

Technological advancements in imaging modalities have greatly improved the quantifcation of CBF (e.g., MRI), yet difficulties persist with regards to the portability and feasibility of such equipment during high altitude studies. As such, transcranial Doppler and portable duplex ultrasound devices remain standard tools for assessment of CBF during high altitude expeditions. Although the latter approach can be quite accurate and effective $[51, 76, 77]$ $[51, 76, 77]$ $[51, 76, 77]$ $[51, 76, 77]$ $[51, 76, 77]$ $[51, 76, 77]$, the former fails to provide a complete assessment of CBF [[75,](#page-10-29) [78\]](#page-11-21). Thus, changes in flow as reported through changes in blood velocity using transcranial Doppler ultrasound measurements should be interpreted cautiously in hypoxic environments. This discrepancy between imaging techniques largely underlies the methodological diferences that make comparing high-altitude studies difficult.

Systems integration

Throughout this review we have described the regulation of CBF and V_F at high-altitude in a relatively isolated manner. However, we do not disregard the likely inter-dependence between these two physiological systems at altitude. Although the magnitude of cerebrovascular and ventilatory changes at high altitude may directly impact upon the other it is not as clear which system may lead in a cause/efect relationship, spurring a "chicken or the egg" conundrum. At the simplest level, a brisk HVR will lead to improved $CaO₂$ and theoretically CDO₂; however, this will come at the expense of augmented hypocapnia, which will have a countervailing influence on $CDO₂$ by reducing CBF through hypocapnic vasoconstriction that is dependent on the degree of metabolic acidosis (i.e. $HCO_3^ HCO_3^ HCO_3^-$ excretion; see Fig. 3). Conversely, large increases in CBF will increase $CDO₂$, but may also increase central CO_2/H^+ washout and reduce central chemoreceptor mediated drive to breathe [[79\]](#page-11-22)—a

response that would lead to transient reductions in $CaO₂$ and hence potentially lower $CDO₂$ subsequent to a reduced V_E . Herein, we discuss the animal and human data explicating the interdependence of V_E and CBF, and discuss how this may pertain to regulation at high altitude.

Ultimately, CBF and its reactivity to changes in arterial blood gases and acid/base changes influences V_E by altering the central chemoreceptor stimulation (not sensitivity). This stimulation in the setting of alterations in $PaCO₂$ or $PaO₂$ is contingent upon manipulation of the PaCO₂ to brain tissue $PCO₂$ (PbCO₂) gradient. Irrespective of the type or magnitude of blood gas changes, $PbCO₂$ will be greater than PaCO₂ due to metabolic CO_2 production, resulting in a positive PbCO₂–PaCO₂ gradient (e.g., $[80, 81]$ $[80, 81]$ $[80, 81]$ $[80, 81]$). At a constant PaCO₂, increases in CBF raise central pH by reducing the $PbCO_2$ –PaCO₂ gradient, as greater flow through the arterial system will draw $PbCO₂$ closer to equilibrium with PaCO₂ (Fig. [8](#page-7-0)). While it is the PbCO₂ to PaCO₂ gradient manipulation governing CBF mediated alterations in V_{E} , experimental models have primarily indexed $PbCO₂$ with measures of jugular venous $CO₂$. Previous animal work has demonstrated that increases in CBF [[82\]](#page-11-25), and presumably decreases in CBF, will increase and decrease ventral medullary pH, respectively. The impact of such changes is reflected in the elevated $V_{\rm E}$ sensitivity to hypercapnia and hypoxia during moderate reductions in CBF [[83\]](#page-11-26). However, these increases in V_E sensitivity were dependent upon the indexing of $V_{\rm E}$ against PaCO₂, as $V_{\rm E}$ sensitivity to jugular venous $CO₂$ was unaltered demonstrating the importance of the $PbCO₂-PaCO₂$ gradient in ventilatory control [[84](#page-11-27), [85](#page-11-28)]. Manipulation of cerebrovascular reactivity and

Fig. 8 A theoretical schematic representing the relationship between two fuid compartments separated by a semi permeable membrane. When two fluid compartments separated by a semipermeable membrane are adjacent to one another, a square wave change in the concentration of a substance in one fuid compartment (c1) will be 95% equilibrated with the opposite compartment (c2) over three time constants. A time constant (τ) represents the time required for a volume of fuid equal to that of c1 to fow through c2 (e.g., if c1 is 500 mL, 1.5 L would have to fow through c2 to reach 95% equilibration). This model can be applied to the cerebral vasculature and brain tissue compartment to understand the effect of cerebrovascular reactivity in producing varying stimuli to breathe despite the same value for PaCO₂ (or $P_{ET}CO_2$). The major limitation of this model is that if we use c2 to represent arterial blood vessels proximal to the central chemoreceptors, and c1 to represent the tissue compartment where the central chemoreceptors reside, c1 is in contrast to a steady-state, constantly producing $CO₂$ via metabolism. Therefore, a simple square wave change cannot be assumed between blood and tissue $CO₂$, but for simplicity we can assume that the relationship between arterial and tissue $CO₂$ resides somewhere on the dotted line, based upon the magnitude of flow and metabolic production of $CO₂$ at any given time. Assuming constant $CO₂$ production, higher flow would result in a rightward shift down the line, and an overall reduced blood to tissue gradient. This would then ultimately lead to a reduction in central chemoreceptor stimulation and reduced V_E . Modified with permission from [[90](#page-11-29)]

arterial-jugular venous gradients in humans have corroborated this role of $PbCO_2$ –PaCO₂ gradient changes in ventilatory control [\[85](#page-11-28), [86\]](#page-11-30).

CBF and ventilation in hypoxia

At sea level in the laboratory setting of isocapnic hypoxia, jugular venous $CO₂$ is reduced following hypoxic cerebral vasodilation [[87](#page-11-31), [88\]](#page-11-32) concomitant to increased jugular venous pH [\[87](#page-11-31)]. In our previous study, progressively reduced SaO₂ from 98% at rest to 70% led to an increase in jugular venous pH of 0.02 despite the maintenance of PaCO₂ [\[87](#page-11-31)]. This magnitude of change holds relevance for the control of breathing given that small changes in pH possess a profound infuence on ventilation—a 0.01 change in tissue pH may result in up to a 5 L min⁻¹ change in alveolar ventilation [\[89](#page-11-33)]. Further, our lab has demonstrated that posterior cerebrovascular reactivity to hypoxia is directly correlated to the magnitude of hypoxic ventilatory decline in humans, further supporting the notion that elevated CBF may attenuate $V_{\rm E}$ [[90\]](#page-11-29). While these data are in the setting of isocapnic hypoxia, the greater influence of changes in $PaCO₂$ on pH (Fig. [3](#page-2-1)) [\[43](#page-10-3)] coupled with the markedly increased ventilatory sensitivity to arterial and jugular venous $PCO₂/H⁺$ [\[43](#page-10-3)], should collectively increase the infuence of CBF mediated central CO_2 washout on V_E throughout acclimatization.

Consistent with this notion, it has been suggested that lower posterior CBF reactivity to poikilocapnic hypoxia contributes to the high HVR measured at sea level [[91\]](#page-11-34) that is generally characteristic of successful mountaineers [[35,](#page-9-38) [36](#page-9-39)]. However, as previously noted, a reduced HVR when measured at altitude is related to improved performance at extreme altitude (~ 8000 m) [\[38](#page-9-33)]. At high-altitude, the physiological responses between V_E and CBF occur rapidly with the responses reaching a steady state balance governed by the bidirectional influences of V_E on CBF and vice versa (Fig. [9\)](#page-8-0).

Conclusion

Upon ascent to altitude CBF and V_E increase and are responsible for maintained $CDO₂$. As medullary pH is dependent on arterial pH, medullary metabolism, and medullary blood fow, CBF changes at altitude are poised to directly infuence central chemoreceptor stimulation. On the other hand, the ventilatory response to high-altitude determines the overall arterial blood gas and acid/base stimulus regulating CBF. Thus, these two responses are intimately intertwined in the determination of cerebral homeostasis at altitude.

Fig. 9 Proposed model of the opposing infuences of ventilation and cerebral blood fow at altitude. Red arrows indicate a consequent reduction, while green arrows indicate consequent increases in V_E and/or CBF for the noted physiological changes at altitude. Top box: increased CBF will cause a reduction in $PbCO₂$, due to an increased washout from the brain tissue, and thus reduce central chemoreceptor stimulation. This will lead to a reduction in ventilatory drive. The consequences of reduced V_E are an increased PaCO₂ and reduced $CaO₂$ (relative to normal values at a given altitude), which both act to increase CBF. Bottom box: Increased V_E reduces PaCO₂ and increases $CaO₂$ (relative to normal values at a given altitude), which both lead to reduced CBF. Consequently, central washout for $CO₂$ is reduced and $PbCO₂$ increased, which augments central chemoreceptor stimulation and ventilatory drive. The top box represents a scenario whereby increased CBF and reduced VE occur, while the bottom box portrays the opposite scenario—ultimately these pathways reach a set point, likely governed by ventilatory and cerebrovascular sensitivities

References

- 1. West JB (2006) Human responses to extreme altitudes. Integr Comp Biol 46:25–34.<https://doi.org/10.1093/icb/icj005>
- 2. Rahn H, Otis A (1949) Man's respiratory response during and after acclimatization to high altitude. Am J Physiol 157:445–462
- 3. Rossen R, Kabat H, Anderson JP (1943) Acute arrest of cerebral circulation in man. Arch Neurol Psychiatry 50:510–528
- 4. Conkin J (2016) Equivalent air altitude and the alveolar gas equation. Aerosp Med Hum Perform 87:61–64. [https://doi.](https://doi.org/10.3357/AMHP.4421.2016) [org/10.3357/AMHP.4421.2016](https://doi.org/10.3357/AMHP.4421.2016)
- 5. Gledhill N, Beirne GJ, Dempsey JA (1975) Renal response to short-term hypocapnia in man. Kidney Int 8:376–384. [https://](https://doi.org/10.1023/A:1009017929727) doi.org/10.1023/A:1009017929727
- 6. Lucas SJE, Burgess KR, Thomas KN et al (2011) Alterations in cerebral blood fow and cerebrovascular reactivity during 14 days at 5050 m. J Physiol 589:741–753. [https://doi.org/10.1113/jphys](https://doi.org/10.1113/jphysiol.2010.192534) [iol.2010.192534](https://doi.org/10.1113/jphysiol.2010.192534)
- 7. Dempsey JA, Forster HV, DoPico GA (1974) Ventilatory acclimatization to moderate hypoxemia in man: role of spinal fuid [H+]. J Clin Invest 53:1091–1100
- 8. Blayo MC, Coudert J, Pocidalo JJ (1975) Comparison of cisternal and lumbar cerebrospinal fuid pH in high altitude natives. Pfugers Arch 356:159–167
- 9. Forster HV, Dempsey JA, Dempsey JA (1975) Incomplete during compensation of CSF [H+] in man to high acclimatization. J Appl Physiol 38:1067–1072
- 10. Severinghaus JW, Mitchell RA, Richardson BW, Singer MM (1963) Respiratory suggesting control at high altitude active transport regulation of CSF pH. J Appl Physiol 18:1155–1166
- 11. Weiskopf B, Gabel A, Bent P, Hospital B (1976) Alkaline shift in lumbar and intracranial in man after 5 days at high altitude. J Appl Physiol 41:1–5
- 12. Stewart A (1983) Modern quantitative acid-base chemistry. Can J Physiol Pharmacol 61:1444–1461
- 13. Imoberdorf R, Garlick PJ, McNurlan MA et al (2001) Enhanced synthesis of albumin and fbrinogen at high altitude. J Appl Physiol 90:528–537
- 14. Somogyi RB, Preiss D, Vesely A et al (2005) Changes in respiratory control after 5 days at altitude. Respir Physiol Neurobiol 145:41–52.<https://doi.org/10.1016/j.resp.2004.11.004>
- 15. Chiodi H (1957) Respiratory adaptations to chronic high altitude hypoxia. J Appl Physiol 10:81–87
- 16. Gill MB, Milledge JS, Pugh LGCE, West JB (1962) Alveolar gas composition At 21,000 to 25,700 Ft. (6400–7830 m). J Physiol 163:373–377
- 17. Wagner PD, Sutton JR, Reeves JT, Cymerman A, Groves BM, Malconian MK (1987) Operation Everest II: pulmonary gas exchange during a simulated ascent of Mt. Everest. J Appl Physiol 63(6):2348–2359
- 18. Severinghaus JW (2000) Stumbling over a bias. What happens to spinal fuid pH at high altitude? Am J Respir Crit Care Med 161:3–4
- 19. Bisgard GE, Vogel HK (1971) Hypoventilation and pulmonary hypertension in calves after carotid excision. J Appl Physiol 31:431–437
- 20. Forster HV, Bisgard GE, Klein JP et al (1981) Effect of peripheral chemoreceptor denervation on acclimatization of goats during hypoxia. J Appl Physiol 50:392–398
- 21. Forster HV, Bisgard GE (1976) Ventilatory control in peripheral chemoreceptor denervated ponies during chronic hypoxemia. J Appl Physiol 41:878–885
- 22. Swenson ER (2016) Pharmacology of acute mountain sickness: old drugs and newer thinking. J Appl Physiol 120:204–215. [https](https://doi.org/10.1152/japplphysiol.00443.2015) [://doi.org/10.1152/japplphysiol.00443.2015](https://doi.org/10.1152/japplphysiol.00443.2015)
- 23. Fatemian M, Kim DY, Poulin MJ, Robbins PA (2001) Very mild exposure to hypoxia for 8 h can induce ventilatory acclimatization in humans. Eur J Physiol 441:840–843. [https://doi.](https://doi.org/10.1007/s004240000491) [org/10.1007/s004240000491](https://doi.org/10.1007/s004240000491)
- 24. Donoghue S, Fatemian M, Balanos GM et al (2005) Ventilatory acclimatization in response to very small changes in PO2 in humans. J Appl Physiol 98:1587–1591. [https://doi.org/10.1152/](https://doi.org/10.1152/japplphysiol.01019.2004) [japplphysiol.01019.2004](https://doi.org/10.1152/japplphysiol.01019.2004)
- 25. White P, Gleeson K, Rannels M et al (1987) Altitude acclimatization: infuence on periodic breathing and chemoresponsiveness during sleep. J Appl Physiol 63:401–412
- 26. Sato M, Severinghaus JW, Powell FL et al (1992) Augmented hypoxic ventilatory response in men at altitude. J Appl Physiol 73:101–107
- 27. Bhaumik G, Banerjee PK (2003) Hypoxic ventilatory response changes of men and women 6 to 7 days after climbing from 2100 m to 4350 m altitude and after descent. High Alt Med Biol 4:341–348
- 28. Rupp T, Esteve F, Bouzat P et al (2014) Cerebral hemodynamic and ventilatory responses to hypoxia, hypercapnia, and hypocapnia during 5 days at 4350 m. J Cereb Blood Flow Metab 34:52– 60.<https://doi.org/10.1038/jcbfm.2013.167>
- 29. Muza SR, Rock PB, Fulco CS et al (2001) Women at altitude: ventilatory acclimatization at 4300 m. J Appl Physiol 91:1791–1799
- 30. Tansley JG, Clar C, Pedersen MEF, Robbins PA (1997) Human ventilatory response to acute hyperoxia during and after 8 h of both isocapnic and poikilocapnic hypoxia. J Appl Physiol 82:513–519
- 31. Barnard P, Andronikou S, Pokorski M et al (1987) Time-dependent efect of hypoxia on carotid body chemosensory function. J Appl Physiol 63:685–691
- 32. Subudhi AW, Fan J, Evero O et al (2014) AltitudeOmics: cerebral autoregulation during ascent, acclimatization, and re-exposure to high altitude and its relation with acute mountain sickness. J Appl Physiol 116:724–729. [https://doi.org/10.1152/japplphysi](https://doi.org/10.1152/japplphysiol.00880.2013) [ol.00880.2013](https://doi.org/10.1152/japplphysiol.00880.2013)
- 33. Weil JV, Byrne-Quinn E, Sodal IE et al (1971) Acquired attenuation of chemoreceptor function in chronically hypoxic man at high altitude. J Clin Invest 50:186–195. [https://doi.org/10.1172/](https://doi.org/10.1172/JCI106472) [JCI106472](https://doi.org/10.1172/JCI106472)
- 34. Moore LG (2017) Measuring high-altitude adaptation. J Appl Physiol.<https://doi.org/10.1152/japplphysiol.00321.2017>
- 35. Schoene RB (1982) Control of ventilation to extreme altitude in climbers. J Appl Physiol 53:886–890
- 36. Schoene RB, Lahiri S, Peters RM (1984) Relationship of hypoxic ventilatory response to exercise performance on Mount Everest. J Appl Physiol Respir Env Exerc Physiol 56:1478–1483
- 37. Masuyama S, Kimura H, Sugita T et al (1986) Control of ventilation in extreme-altitude climbers. J Appl Physiol 61:500–506
- 38. Bernardi L, Schneider A, Pomidori L et al (2006) Hypoxic ventilatory response in successful extreme altitude climbers. Eur Respir J 27:165–171. [https://doi.org/10.1183/09031](https://doi.org/10.1183/09031936.06.00015805) [936.06.00015805](https://doi.org/10.1183/09031936.06.00015805)
- 39. Gallman EA, Millhorn DE (1988) Two long-lasting central respiratory responses following acute hypoxia in glomectomized cats. J Physiol 395:333–347
- 40. Dwinell MR, Powell FL (1999) Chronic hypoxia enhances the phrenic nerve response to arterial chemoreceptor stimulation in anesthetized rats. J Appl Physiol 87:817–823
- 41. Tenney SM, Remmers JE, Mithoefer JC (1962) Interaction of CO2 and hypoxic stimuli on ventilation at high altitude. Q J Exp Physiol Cogn Med Sci 48:192–201
- 42. Eger EI, Kellogg RH, Mines AH et al (1968) Influence of CO2 on ventilatory acclimatization to altitude. J Appl Physiol 24:607–615
- 43. Willie CK, MacLeod DB, Smith KJ et al (2015) The contribution of arterial blood gases in cerebral blood fow regulation and fuel utilization in man at high altitude. J Cereb Blood Flow Metab 35:873–881. <https://doi.org/10.1038/jcbfm.2015.4>
- 44. Wilson RJA, Teppema LJ (2016) Integration of central and peripheral respiratory chemorefexes. Compr Physiol 6:1005– 1041. <https://doi.org/10.1002/cphy.c140040>
- 45. Fatemian M, Nieuwenhuijs DJF, Teppema LJ et al (2003) The respiratory response to carbon dioxide in humans with unilateral and bilateral resections of the carotid bodies. J Physiol 549:965– 973. <https://doi.org/10.1113/jphysiol.2003.042259>
- 46. West JB (2017) Are permanent residents of high altitude fully adapted to their hypoxic environment? High Alt Med Biol. [https](https://doi.org/10.1089/ham.2016.0152) [://doi.org/10.1089/ham.2016.0152](https://doi.org/10.1089/ham.2016.0152)
- 47. Hoiland RL, Bain AR, Rieger MG et al (2016) Hypoxemia, oxygen content, and the regulation of cerebral blood fow. Am J Physiol Regul Integr Comp Physiol 310:R398–R413. [https://doi.](https://doi.org/10.1152/ajpregu.00270.2015) [org/10.1152/ajpregu.00270.2015](https://doi.org/10.1152/ajpregu.00270.2015)
- 48. Subudhi AW, Fan J-L, Evero O et al (2014) AltitudeOmics: efect of ascent and acclimatization to 5260 m on regional cerebral oxygen delivery. Exp Physiol 5:772–781. [https://doi.org/10.1113/](https://doi.org/10.1113/expphysiol.2013.075184) [expphysiol.2013.075184](https://doi.org/10.1113/expphysiol.2013.075184)
- 49. Hoiland RL, Bain AR, Tymko MM et al (2017) Adenosine receptor-dependent signaling is not obligatory for normobaric and hypobaric hypoxia-induced cerebral vasodilation in humans. J Appl Physiol 122:795–808. [https://doi.org/10.1152/japplphysi](https://doi.org/10.1152/japplphysiol.00840.2016) [ol.00840.2016](https://doi.org/10.1152/japplphysiol.00840.2016)
- 50. Wilson MH, Edsell MEG, Davagnanam I et al (2011) Cerebral artery dilatation maintains cerebral oxygenation at extreme altitude and in acute hypoxia—an ultrasound and MRI study. J Cereb Blood Flow Metab 31:2019–2029
- 51. Willie CK, Smith KJ, Day TA et al (2014) Regional cerebral blood fow in humans at high altitude: gradual ascent and 2 wk at 5050 m. J Appl Physiol 116:905–910. [https://doi.org/10.1152/](https://doi.org/10.1152/japplphysiol.00594.2013) [japplphysiol.00594.2013](https://doi.org/10.1152/japplphysiol.00594.2013)
- 52. Liu W, Liu J, Lou X et al (2017) A longitudinal study of cerebral blood fow under hypoxia at high altitude using 3D pseudocontinuous arterial spin labeling. Sci Rep 7:43246. [https://doi.](https://doi.org/10.1038/srep43246) [org/10.1038/srep43246](https://doi.org/10.1038/srep43246)
- 53. Lahiri S, Milledge JS (1967) Acid-base in Sherpa altitude residents and lowlanders at 4880 m. Respir Physiol 2:323–334. [https](https://doi.org/10.1016/0034-5687(67)90037-0) [://doi.org/10.1016/0034-5687\(67\)90037-0](https://doi.org/10.1016/0034-5687(67)90037-0)
- 54. Kety SS, Schmidt CF (1948) The effects of altered arterial tensions of carbon dioxide and oxygen on cerebral blood flow and cerebral oxygen consumption of normal young men. J Clin Invest 27:484–492. <https://doi.org/10.1172/JCI101995>
- 55. Willie CK, Macleod DB, Shaw AD et al (2012) Regional brain blood flow in man during acute changes in arterial blood gases. J Physiol 59014:3261–3275. [https://doi.org/10.1113/jphys](https://doi.org/10.1113/jphysiol.2012.228551) [iol.2012.228551](https://doi.org/10.1113/jphysiol.2012.228551)
- 56. Willie CK, Tzeng YC, Fisher JA, Ainslie PN (2014) Integrative regulation of human brain blood fow. J Physiol 592:841–859. <https://doi.org/10.1113/jphysiol.2013.268953>
- 57. Brassard P, Tymko MM, Ainslie PN (2016) Sympathetic control of the brain circulation: appreciating the complexities to better understand the controversy. Auton Neurosci Basic Clin 207:37– 47.<https://doi.org/10.1016/j.autneu.2017.05.003>
- 58. Lewis NCS, Messinger L, Monteleone B, Ainslie PN (2014) Efect of acute hypoxia on regional cerebral blood fow: efect of sympathetic nerve activity. J Appl Physiol 116:1189–1196. <https://doi.org/10.1152/japplphysiol.00114.2014>
- 59. Ainslie PN, Lucas SJE, Fan J-L et al (2012) Infuence of sympathoexcitation at high altitude on cerebrovascular function and ventilatory control in humans. J Appl Physiol 113:1058–1067. <https://doi.org/10.1152/japplphysiol.00463.2012>
- 60. Ainslie PN, Subudhi AW (2014) Cerebral blood fow at high altitude. High Alt Med Biol 15:133–140. [https://doi.org/10.1089/](https://doi.org/10.1089/ham.2013.1138) [ham.2013.1138](https://doi.org/10.1089/ham.2013.1138)
- 61. Ge R-L, Babb TG, Sivieri M et al (2006) Urine acid-base compensation at simulated moderate altitude. High Alt Med Biol 7:64–71.<https://doi.org/10.1089/ham.2006.7.64>
- 62. Pugh LGCE (1964) Blood volume and haemoglobin concentration at altitudes Above 18,000 ft. (5500 m). J Physiol 170:344– 354. <https://doi.org/10.1113/jphysiol.1964.sp007335>
- 63. Siebenmann C, Cathomen A, Hug M et al (2015) Hemoglobin mass and intravascular volume kinetics during and after exposure to 3454 m altitude. J Appl Physiol 119:1194–1201. [https://doi.](https://doi.org/10.1152/japplphysiol.01121.2014) [org/10.1152/japplphysiol.01121.2014](https://doi.org/10.1152/japplphysiol.01121.2014)
- 64. Siebenmann C, Robach P, Lundby C (2017) Regulation of blood volume in lowlanders exposed to high altitude. J Appl Physiol. <https://doi.org/10.1152/japplphysiol.00118.2017>
- 65. Ryan BJ, Wachsmuth NB, Schmidt WF et al (2014) Altitudeomics: rapid hemoglobin mass alterations with early acclimatization to and de-acclimatization from 5260 m in healthy humans. PLoS One 9:e108788.<https://doi.org/10.1371/journal.pone.0108788>
- 66. Penaloza D, Arias-Stella J (2007) The heart and pulmonary circulation at high altitudes: healthy highlanders and chronic mountain sickness. Circulation 115:1132–1146. [https://doi.org/10.1161/](https://doi.org/10.1161/CIRCULATIONAHA.106.624544) [CIRCULATIONAHA.106.624544](https://doi.org/10.1161/CIRCULATIONAHA.106.624544)
- 67. Villafuerte FC, Corante N (2016) Chronic mountain sickness: clinical aspects, etiology, management, and treatment. High Alt Med Biol 17:61–69.<https://doi.org/10.1089/ham.2016.0031>
- 68. Flück D, Siebenmann C, Keiser S et al (2015) Cerebrovascular reactivity is increased with acclimatization to 3454 m altitude. J Cereb Blood Flow Metab 35:1323–1330. [https://doi.org/10.1038/](https://doi.org/10.1038/jcbfm.2015.51) [jcbfm.2015.51](https://doi.org/10.1038/jcbfm.2015.51)
- 69. Jensen JB, Sperling B, Severinghaus JW, Lassen NA (1996) Augmented hypoxic cerebral vasodilation in men during 5 days at 3810 m altitude. J Appl Physiol 80:1214–1218
- 70. Poulin MJ, Fatemian M, Tansley JG et al (2002) Changes in cerebral blood fow during and after 48 h of both isocapnic and poikilocapnic hypoxia in humans. Exp Physiol 87:633–642
- 71. Binks AP, Cunningham VJ, Adams L, Banzett RB (2008) Gray matter blood flow change is unevenly distributed during moderate isocapnic hypoxia in humans. J Appl Physiol 104:212–217. <https://doi.org/10.1152/japplphysiol.00069.2007>
- 72. Lawley JS, Macdonald JH, Oliver SJ, Mullins PG (2017) Unexpected reductions in regional cerebral perfusion during prolonged hypoxia. J Physiol 595:935–947. [https://doi.org/10.1113/](https://doi.org/10.1113/JP272557) JP27255⁷
- 73. Feddersen B, Neupane P, Thanbichler F et al (2015) Regional differences in the cerebral blood fow velocity response to hypobaric hypoxia at high altitudes. J Cereb Blood Flow Metab 35:1–6. <https://doi.org/10.1038/jcbfm.2015.142>
- 74. Wilson MH, Imray CHE (2016) The cerebral venous system and hypoxia. J Appl Physiol 120:244–250. [https://doi.org/10.1152/](https://doi.org/10.1152/japplphysiol.00327.2015) [japplphysiol.00327.2015](https://doi.org/10.1152/japplphysiol.00327.2015)
- 75. Sagoo RS, Hutchinson CE, Wright A et al (2016) Magnetic resonance investigation into the mechanisms involved in the development of high-altitude cerebral edema. J Cereb Blood Flow Metab 37:319–331.<https://doi.org/10.1177/0271678X15625350>
- 76. Hoiland RL, Bain AR, Tymko MM et al (2017) Adenosine receptor dependent signaling is not obligatory for normobaric and hypobaric hypoxia-induced cerebral vasodilation in humans. J Appl Physiol. <https://doi.org/10.1152/japplphysiol.00840.2016>
- 77. Thomas KN, Lewis NCS, Hill BG, Ainslie PN (2015) Technical recommendations for the use of carotid duplex ultrasound for the assessment of extracranial blood fow. Am J Physiol Regul Integr Comp Physiol 309:R707–R720. [https://doi.org/10.1152/](https://doi.org/10.1152/ajpregu.00211.2015) [ajpregu.00211.2015](https://doi.org/10.1152/ajpregu.00211.2015)
- 78. Kellawan MJ, Harrell JW, Roldan-Alzate A et al (2016) Regional hypoxic cerebral vasodilation facilitated by diameter changes primarily in anterior versus posterior circulation. J Cereb Blood Flow Metab 37:2025–2034. [https://doi.org/10.1177/0271678X16](https://doi.org/10.1177/0271678X16659497) [659497](https://doi.org/10.1177/0271678X16659497)
- 79. Ainslie PN, Duffin J (2009) Integration of cerebrovascular CO2 reactivity and chemorefex control of breathing: mechanisms of regulation, measurement, and interpretation. Am J Physiol Regul Integr Comp Physiol 296:R1473–R1495. [https://doi.org/10.1152/](https://doi.org/10.1152/ajpregu.91008.2008) [ajpregu.91008.2008](https://doi.org/10.1152/ajpregu.91008.2008)
- 80. Peebles K, Celi L, McGrattan K et al (2007) Human cerebrovascular and ventilatory CO2 reactivity to end-tidal, arterial and internal jugular vein PCO2. J Physiol 584:347–357. [https://doi.](https://doi.org/10.1113/jphysiol.2007.137075) [org/10.1113/jphysiol.2007.137075](https://doi.org/10.1113/jphysiol.2007.137075)
- 81. Mithoefer JC, Kazemi H (1963) Gas exchange during rebreathing. Ann N Y Acad Sci 109:743–755
- 82. Neubauer JA, Santiago TV, Posner MA, Edelman NH (1985) Ventral medullary pH and ventilatory to hyperperfusion and hypoxia responses. J Appl Physiol 58:1659–1668
- 83. Chapman RV, Santiago TV, Edelman NH (1979) Effects of graded reduction of brain blood fow on chemical control of breathing. J Appl Physiol 47:1289–1294
- 84. Chapman RW, Santiago TV, Edelman NH (1979) Efects of graded reduction of brain blood fow on ventilation in unanesthetized goats. J Appl Physiol Respir Environ Exerc Physiol 47:104–111
- 85. Xie A, Skatrud JB, Morgan B et al (2006) Infuence of cerebrovascular function on the hypercapnic ventilatory response in healthy humans. J Physiol 5771:319–329. [https://doi.](https://doi.org/10.1113/jphysiol.2006.110627) [org/10.1113/jphysiol.2006.110627](https://doi.org/10.1113/jphysiol.2006.110627)
- 86. Fan JL, Burgess KR, Thomas KN et al (2010) Infuence of indomethacin on the ventilatory and cerebrovascular responsiveness to CO2 and breathing stability: the infuence of PCO2 gradients. Am J Physiol Regul Integr Comp Physiol 298:R1648–R1658. <https://doi.org/10.1007/s00421-010-1679-0>
- 87. Ainslie PN, Shaw AD, Smith KJ et al (2014) Stability of cerebral metabolism and substrate availability in humans during hypoxia and hyperoxia. Clin Sci (Lond) 126:661–670. [https://](https://doi.org/10.1042/CS20130343) doi.org/10.1042/CS20130343
- 88. Suzuki A, Nishimura M, Yamamoto H et al (1989) No efect of brain blood fow on ventilatory depression during sustained hypoxia. J Appl Physiol 66:1674–1678
- 89. Pappenheimer JR, Fencl V (1965) Role of cerebral fuids in control of respiration as studied in unanesthetized goats. Am J Physiol 208:436–450
- 90. Hoiland RL, Ainslie PN, Wildfong KW et al (2015) Indomethacin-induced impairment of regional cerebrovascular reactivity: implications for respiratory control. J Physiol 593:1291–1306. <https://doi.org/10.1113/jphysiol.2014.284521>
- 91. Jansen GFA, Kagenaar DA, Basnyat B, Odoom JA (2002) Basilar artery blood fow velocity and the ventilatory response to acute hypoxia in mountaineers. Respir Physiol Neurobiol 133:65–74
- 92. Subudhi AW, Bourdillon N, Bucher J et al (2014) AltitudeOmics: the integrative physiology of human acclimatization to hypobaric hypoxia and its retention upon reascent. PLoS One 9:e92191. <https://doi.org/10.1371/journal.pone.0092191>
- 93. Ainslie PN, Lucas SJE, Burgess KR (2013) Breathing and sleep at high altitude. Respir Physiol Neurobiol 188:233–256
- 94. West JB, Hackett PH, Maret KH, Milledge JS, Peters RM Jr, Pizzo CJ, Winslow RM (1983) Pulmonary gas exchange on the summit of Mount Everest. J Appl Physiol 55:678–687
- 95. Malconian MK, Rock PB, Reeves JT, Cymerman A, Houston CS (1993) Operation Everest II: gas tensions in expired air and arterial blood at extreme altitude. Aviat Space Environ Med 64:37–42
- 96. Severinghaus JW, Chiodi H, Eger EI et al (1966) Cerebral blood fow in man at high altitude. Role of cerebrospinal fuid pH in normalization of fow in chronic hypocapnia. Circ Res 19:274– 282. <https://doi.org/10.1161/01.RES.19.2.274>
- 97. Huang SY, Moore LG, McCullough RE et al (1987) Internal carotid and vertebral arterial fow velocity in men at high altitude. J Appl Physiol 63:395–400
- 98. Jensen JB, Wright AD, Lassen NA et al (1990) Cerebral blood flow in acute mountain sickness. J Appl Physiol 69:430–433
- 99. Baumgartner RW, Bartsch P, Maggiorini M et al (1994) Enhanced cerebral blood fow in acute mountain sickness. Aviat Sp Environ Med 65:726–729
- 100. Kalsi KK, González-Alonso J (2012) Temperature-dependent release of ATP from human erythrocytes: mechasnism for the control of local tissue perfusion. Exp Physiol 97:419–432. [https](https://doi.org/10.1113/expphysiol.2011.064238) [://doi.org/10.1113/expphysiol.2011.064238](https://doi.org/10.1113/expphysiol.2011.064238)
- 101. Bergfeld GR, Forrester T (1992) Release of ATP from human erythrocytes in response to a brief period of hypoxia and hypercapnia. Cardiovasc Res 26:40–47
- 102. Ellsworth ML, Forrester T, Ellis CG, Dietrich HH (1995) The erythrocyte as a regulator of vascular tone. Am J Physiol 269:H2155–H2161
- 103. Sprague RS, Ellsworth ML, Stephenson AH et al (1998) Deformation-induced ATP release from red blood cells requires CFTR activity. Am J Physiol 275:H1726–H1732
- 104. Jiang H, Zhu AG, Mamczur M et al (2007) Stimulation of rat erythrocyte P2X 7 receptor induces the release of epoxyeicosatrienoic acids. Br J Pharmacol 151:1033–1040. [https://doi.](https://doi.org/10.1038/sj.bjp.0707311) [org/10.1038/sj.bjp.0707311](https://doi.org/10.1038/sj.bjp.0707311)
- 105. Gebremedhin D, Ma YH, Falck JR et al (1992) Mechanism of action of cerebral epoxyeicosatrienoic acids on cerebral arterial smooth muscle. Am J Physiol 263:H519–H525
- 106. You J, Johnson TD, Childres WF, Bryan RM (1997) Endothelialmediated dilations of rat middle cerebral arteries by ATP and ADP. Am J Physiol 273:H1472–H1477
- 107. Fuentes E, Palomo I (2015) Extracellular ATP metabolism on vascular endothelial cells: a pathway with pro-thrombotic molecules. Vascul Pharmacol. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.vph.2015.05.002) [vph.2015.05.002](https://doi.org/10.1016/j.vph.2015.05.002)
- 108. Hein TW, Xu W, Ren Y, Kuo L (2013) Cellular signalling pathways mediating dilation of porcine pial arterioles to adenosine A2A receptor activation. Cardiovasc Res 99:156–163. [https://](https://doi.org/10.1093/cvr/cvt072) doi.org/10.1093/cvr/cvt072
- 109. Kalaria RN, Harik SI (1988) Adenosine receptors and the nucleoside transporter in human brain vasculature. J Cereb Blood Flow Metab 8:32–39
- 110. Sattin A, Rall TW (1970) The efect of adenosine and adenine nucleotides on the cyclic adenosine 3′, 5′-phosphate content of guinea pig cerebral cortex slices. Mol Pharmacol 6:13–23
- 111. Nordstrom CH, Rehnrona S, Siesjo BK, Westerberg E (1977) Adenosine in rat cerebral cortex: its determination, normal values, and correlation to AMP and cyclic AMP during shortlasting ischemia. Acta Physiol Scand 101:63–71
- 112. Davis RJ, Murdoch CE, Ali M et al (2004) EP4 prostanoid receptor-mediated vasodilatation of human middle cerebral arteries. Br J Pharmacol 141:580–585. [https://doi.org/10.1038/sj.bjp.07056](https://doi.org/10.1038/sj.bjp.0705645) [45](https://doi.org/10.1038/sj.bjp.0705645)
- 113. Narumiya S, Sugimoto Y, Ushikubi F, Conclusions VI (1999) Prostanoid receptors : structures, properties, and functions. Physiol Rev 79:1193–1226
- 114. Cosby K, Partovi KS, Crawford JH et al (2003) Nitrite reduction to nitric oxide by deoxyhemoglobin vasodilates the human circulation. Nat Med 9:1498–1505. <https://doi.org/10.1038/nm954>
- 115. Stamler JS, Jia L, Eu JP et al (1997) Blood fow regulation by S-nitrosohemoglobin in the physiological oxygen gradient. Science 276:2034–2037
- 116. Lima B, Forrester MT, Hess DT, Stamler JS (2010) S-nitrosylation in cardiovascular signaling. Circ Res 106:633–646. [https://](https://doi.org/10.1161/CIRCRESAHA.109.207381) doi.org/10.1161/CIRCRESAHA.109.207381
- 117. Pearce WJ, Reynier-Rebufel AM, Lee J et al (1990) Efects of methylene blue on hypoxic cerebral vasodilatation in the rabbit. J Pharmacol Exp Ther 254:616–625
- 118. Bolotina VM, Najibi S, Palacino JJ et al (1994) Nitric oxide directly activates calcium-dependent potassium channels in vascular smooth muscle. Nature 368:850–853. [https://doi.](https://doi.org/10.1038/368850a0) [org/10.1038/368850a0](https://doi.org/10.1038/368850a0)
- 119. Adelstein R, Conti M (1978) Phosphorylation of smooth muscle myosin catalytic subunit of adenosine 3′: 5′-monophosphatedependent protein kinase. J Biol Chem 253:8347–8350
- 120. Kerrick WG, Hoar PE (1981) Inhibition of smooth muscle tension by cyclic AMP-dependent protein kinase. Nature 292:253–255
- 121. Song Y, Simard M (1995) Beta-Adrenoceptor stimulation activates large. Conductance Ca2+-activated K+ channels in smooth
- 122. Nelson MT, Patlak JB, Worley JF, Standen NB (1990) Calcium channels, potassium channels, and voltage dependence of arterial smooth muscle tone. Am J Physiol 259:C3–C18
- 123. Kajita Y, Dietrich HH, Dacey RG (1996) Efects of oxyhemoglobin on local and propagated vasodilatory responses induced by adenosine, adenosine diphosphate, and adenosine triphosphate in rat cerebral arterioles. J Neurosurg 85:908–916. [https://doi.](https://doi.org/10.3171/jns.1996.85.5.0908) [org/10.3171/jns.1996.85.5.0908](https://doi.org/10.3171/jns.1996.85.5.0908)
- 124. Dietrich HH, Kajita Y, Dacey RG (1996) Local and conducted vasomotor responses in isolated rat cerebral arterioles. Am J Physiol 271:H1109–H1116