#### **REGULAR PAPER**





# Nicotiana suaveolens accessions with different ploidy levels exhibit different reproductive isolation mechanisms in interspecific crosses with Nicotiana tabacum

Hai He<sup>1</sup>·Takahiro lizuka<sup>1</sup>·Maho Maekawa<sup>2</sup>·Kumi Sadahisa<sup>2</sup>·Toshinobu Morikawa<sup>1</sup>·Masanori Yanase<sup>1,3</sup>·Shuji Yokoi<sup>1,3</sup>·Masayuki Oda<sup>1</sup>·Takahiro Tezuka<sup>1,3</sup>

Received: 22 May 2017 / Accepted: 11 May 2019 / Published online: 21 May 2019 © The Botanical Society of Japan and Springer Japan KK, part of Springer Nature 2019

#### **Abstract**

Reproductive isolation, including prezygotic and postzygotic barriers, is a mechanism that separates species. Many species in the *Nicotiana* section *Suaveolentes* exhibit reproductive isolation in crosses with *Nicotiana tabacum*. In this study, we investigated whether the chromosome numbers and ploidy levels of eight *Nicotiana suaveolens* accessions are related to the reproductive isolation after crosses with *N. tabacum* by flow cytometry and chromosome analyses. Additionally, the internal transcribed spacer (ITS) regions of the eight *N. suaveolens* accessions were sequenced and compared with the previously reported sequences of 22 *Suaveolentes* species to elucidate the phylogenetic relationships in the section *Suaveolentes*. We revealed that four *N. suaveolens* accessions comprised 64 chromosomes, while the other four accessions carried 32 chromosomes. Depending on the ploidy levels of *N. suaveolens*, several types of reproductive isolation were observed after crosses with *N. tabacum*, including decreases in the number of capsules and the germination rates of hybrid seeds, as well as hybrid lethality and abscission of enlarged ovaries at 12–17 days after pollination. A phylogenetic analysis involving ITS sequences divided the eight *N. suaveolens* accessions into three distinct clades. Based on the results, we confirmed that *N. suaveolens* accessions vary regarding ploidy levels and reproductive isolation mechanisms in crosses with *N. tabacum*. These accessions will be very useful for revealing and characterizing the reproductive isolation mechanisms in interspecific crosses and their relationships with ploidy levels.

**Keywords** Internal transcribed spacer region  $\cdot$  Interspecific hybridization  $\cdot$  Phylogenetics  $\cdot$  Polyploidy  $\cdot$  Reproductive isolation  $\cdot$  Tobacco

**Electronic supplementary material** The online version of this article (https://doi.org/10.1007/s10265-019-01114-w) contains supplementary material, which is available to authorized users.

- ☐ Takahiro Tezuka tezuka@plant.osakafu-u.ac.jp
- Graduate School of Life and Environmental Sciences, Osaka Prefecture University, 1-1 Gakuen-cho, Nakaku, Sakai, Osaka 599-8531, Japan
- School of Life and Environmental Sciences, Osaka Prefecture University, Sakai, Osaka 599-8531, Japan
- <sup>3</sup> Education and Research Field, College of Life, Environment, and Advanced Sciences, Osaka Prefecture University, Sakai, Osaka 599-8531, Japan

#### Introduction

Reproductive isolation, including prezygotic and postzygotic barriers, is a mechanism that separates species and plays a crucial role in the evolution of animals and plants (Rieseberg and Blackman 2010; Stebbins 1966). In plants, prezygotic barriers include the inhibition of pollen adhesion to the stigma, pollen germination, pollen tube growth in the style, and pollen tube penetration of the ovule micropyle (Dickinson et al. 2012). Postzygotic barriers, which occur after a successful fertilization, include seed abortion as well as the weakness, inviability, and sterility of the  $F_1$  hybrid plants or their offspring (Bushell et al. 2003; Guo et al. 2016; Ichitani et al. 2007; Kuboyama et al. 2009; Li et al. 1997; Tezuka et al. 2010). These barriers hinder plant breeding programs such as wide hybridizations which incorporates backcrossing of  $F_1$  hybrids between cultivated and related wild species



to the cultivated species. To facilitate wide hybridization breeding, the mechanisms underlying reproductive isolation should be characterized and methods for overcoming the reproductive isolation will need to be developed.

Many species in the *Nicotiana* section *Suaveolentes* have been investigated for reproductive isolation by crosses with the cultivated species N. tabacum (2n = 48), which belongs to the section *Nicotiana* (Tezuka 2012; Tezuka et al. 2010). The section Suaveolentes includes 25 wild species, which are endemic to Australasia except for the African wild species N. africana. All species in section Suaveolentes are allotetraploids (Leitch et al. 2008) and the haploid number of chromosomes within this section can be 16-24, although species carrying 17 pairs of chromosomes have not been identified (Goodspeed 1954; Ladiges et al. 2011). Although N. suaveolens is one of four species with 16 pairs of chromosomes, Wheeler (1935) reported that two of the many N. suaveolens races carry 32 pairs of chromosomes. The section Suaveolentes is a monophyletic group based on internal transcribed spacer (ITS) regions (Chase et al. 2003), plastid genes (Clarkson et al. 2004), and nuclear-encoded chloroplast-expressed glutamine synthetase (Clarkson et al. 2010).

Three types of reproductive isolation have been observed in reciprocal crosses between N. suaveolens JT (this accession name is described in the "Materials and methods") and N. tabacum. Although hybrid seeds that germinate normally can be obtained in the N. suaveolens JT  $(\mathcal{P}) \times N$ . tabacum (d) cross after a conventional hand pollination (Manabe et al. 1989; Yamada et al. 2000), unilateral incongruity (UI) occurs and seeds cannot be obtained in the reciprocal cross after a conventional hand pollination. This incongruity can be overcome with a test-tube pollination and ovule culture to obtain hybrid seedlings (Tezuka and Marubashi 2004). However, hybrid lethality (a phenomenon responsible for the death of hybrid plants) characterized by browning of the hypocotyl and roots (Type II lethality) is observed in reciprocal hybrid seedlings (Marubashi and Onosato 2002; Tezuka and Marubashi 2004, 2006; Yamada et al. 1999). Although the Type II lethality can be overcome or bypassed by several methods, including high temperature cultivation, and tissue culturing, as well as by the lack of the chromosome causing hybrid lethality, the resulting hybrid plants are sterile (Inoue et al. 1994; Manabe et al. 1989; Tezuka and Marubashi 2006). In our previous studies, N. suaveolens JT, which has 16 pairs of chromosomes (Japan Tobacco Inc. 1994), was solely used for crosses with N. tabacum (Tezuka and Marubashi 2004, 2006).

In the present study, we used *N. suaveolens* accessions with an unknown number of chromosomes as well as the JT accession, and investigated types of their reproductive isolation and their phylogenetic relationships. We discuss whether the chromosome numbers and ploidy levels of the

*N. suaveolens* accessions are related to the reproductive isolation, and estimate the evolutionary order and timing of the acquisition of the reproductive isolation in the *N. suaveolens* accessions.

#### **Materials and methods**

#### **Plant materials**

The following eight accessions of wild N. suaveolens were analyzed in this study: PI 230960, PI 555561, PI 555563, PI 555565, PI 555566, PI 555567, PI 555568, and JT. The one accession obtained from the Leaf Tobacco Research Center (Japan Tobacco Inc., Oyama, Japan) was named JT (2n = 4x = 32) to distinguish it from the other accessions because the accession number and line name were not provided. The accessions with Plant Introduction (PI) numbers were obtained from the United States Nicotiana Germplasm Collection (Lewis and Nicholson 2007). Information regarding the seven PI accessions is available in the Genetic Resources Information Network database of the United States National Plant Germplasm System (https://www.ars-grin.gov/npgs/ index.html) (USDA ARS National Genetic Resources Program 2010). The eight accessions were reciprocally crossed with the N. tabacum 'Red Russian' cultivar (2n = 4x = 48). The 'Red Russian' seeds were provided by the Leaf Tobacco Research Center. All plants except plants used for investigation of flower and leaf morphology were cultivated in a greenhouse (natural day length; Osaka Prefecture University, Sakai, Osaka, Japan). Plants used for investigation of flower and leaf morphology were grown under fluorescent lamps (FLR40S·BRN; Toshiba, Tokyo, Japan) in a cultivation room (16-h light/8-h dark; approximately 70  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>; 25 °C).

#### Investigation of flower and leaf morphology

Three plants of each *N. suaveolens* accession were grown in separate plastic pots (13 cm diameter, 12 cm deep; soil volume approximately 1.2 L) filled with a 1:3 mixture of vermiculite (Type GS; Nittai, Osaka, Japan) and culture soil (Sakata Super Mix A; Sakata Seed, Kanagawa, Japan). Flower morphology was analyzed by recording the flower color as well as the corolla-limb diameter and corolla tube length. Ten mature flowers from three plants (3 flowers/2 plants, 4 flowers/1 plant) were examined for all eight *N. suaveolens* accessions. The maximum length and width of cotyledons and 1st to 15th rosette leaves (total two cotyledons and 15 rosette leaves per plant) were measured, with three plants analyzed for each of the eight *N. suaveolens* accessions. The oldest leaf was defined as the 1st rosette leaf. The aspect ratio was calculated by dividing the leaf width by the leaf length.



#### Self-crosses and interspecific crosses

Five plants of each N. suaveolens accession and N. tabacum were used in a cross experiment. The same five N. tabacum plants were used for all reciprocal interspecific crosses and self-crosses, but the number of flowers per plant differed among crosses (Table 2). The plants were pollinated in 2009, 2012, and 2017–2018. In summer, the minimum temperatures in the greenhouse were 20-26 °C and the maximum temperatures were 35-45 °C. In winter, the minimum temperatures were maintained above 15 °C with an air heating system and the maximum temperatures were 30–35 °C. The flowers of plants used as the female parents were emasculated 1 day before anthesis and then pollinated with pollen grains from the male parents. To assess the reproductive isolation after reciprocal crosses between the eight N. suaveolens accessions and N. tabacum, we observed whether pollinated flowers dropped, ovaries enlarged and dropped, capsules were obtained, and seed morphology was normal. Self-crosses of N. suaveolens and N. tabacum were conducted as a control. Seeds obtained from self and interspecific crosses were sterilized with 5% sodium hypochlorite for 15 min. The sterilized seeds were sown in Petri dishes (60 mm diameter, 17 mm deep) containing 8 mL halfstrength Murashige and Skoog medium (Murashige and Skoog 1962) supplemented with 1% sucrose and 0.2% Gelrite (pH 5.8), and then incubated at 28 °C under continuous illumination (approximately 150 µmol m<sup>-2</sup> s<sup>-1</sup>). About 100 seeds per cross were initially sown, but when seeds germinated poorly, additional seeds were sown. We determined whether seeds could germinate, counted the seedlings obtained at 30 days after sowing, and assessed whether the seedlings exhibited lethality at 30 days after germination (Table 2).

#### Statistical analysis

Data were analyzed with the SPSS program (version 22) (IBM Corp., Armonk, NY, USA). Chi square tests were used to compare the number of capsules obtained and seeds that germinated. The tests were performed separately for self-crosses, *N. suaveolens*×*N. tabacum*, and *N. tabacum*×*N. suaveolens*. The corolla-limb diameter, corolla tube length, and maximum length, width and aspect ratio of cotyledons and rosette leaves were compared among the eight *N. suaveolens* accessions using Tukey's multiple comparison test.

#### Flow cytometry

The ploidy levels of the *N. suaveolens* accessions were analyzed by flow cytometry to clarify why diverse cross results were obtained with the various accessions. Young leaves were placed on a Petri dish and then finely chopped with a

sharp razor blade in 400 µL ice-cold Otto I buffer (Doležel and Bartos 2005; Otto 1990). The resulting extract was filtered through a 30-µm nylon mesh, after which 1.6 mL Otto II buffer (Doležel and Bartos 2005; Otto 1990) containing  $2 \mu g \text{ mL}^{-1} 4',6$ -diamino-2-phenylindole was added. For each sample, the DNA content of at least 10,000 nuclei was analyzed with a CyFlow Space flow cytometer (Partec GmbH, Münster, Germany). Previous studies indicated that N. suaveolens JT consists of 32 chromosomes and is an allotetraploid species (Clarkson et al. 2010; Japan Tobacco Inc. 1994). The G1 peak (4C) of N. suaveolens JT was positioned on channel 10<sup>1</sup> on the histogram by adjusting the instrument gain settings in every analysis (Fig. S2h). The same instrument gain settings were used to determine the ploidy levels of the other seven accessions. Two individuals were observed for N. suaveolens JT, but three individuals were observed for the other accessions.

#### **Chromosome analysis**

The *N. suaveolens* root tips were pretreated with distilled water for 24 h at 4 °C and with 2 mM 8-hydroxyquinoline for 4 h at 18 °C, after which they were fixed in ethanol/acetic acid (3:1) overnight to determine chromosome numbers. The root tips were hydrolyzed in 1 N HCl for 8 min at 60 °C, stained with Schiff's reagent, and squashed in 45% acetic acid. For each plant, the number of chromosomes in at least five root tip cells was counted with a BX50 light microscope (Olympus, Tokyo, Japan). Two individuals were observed for *N. suaveolens* JT, but three individuals were observed for the other accessions.

#### ITS amplification and cladistic analysis

The rDNA ITS region was sequenced for all eight N. suaveolens accessions. The DNA was extracted from leaves following a CTAB method (Murray and Thompson 1980). The ITS DNA region was amplified using the primers described by Baldwin (1993) and Sun et al. (1994). The PCR mixtures consisted of 1 x standard buffer (BioAcademia, Osaka, Japan), 0.2 mM each dNTP, 0.2 µM each primer, 20 ng template DNA, and 1.0 U Taq DNA polymerase (BioAcademia) in a total volume of 20 µL. For all reaction mixtures, 3% DMSO was added because GC-rich regions in the ITS sequences cause premature termination of most strands within 100-150 bp of the initiation point (Chase et al. 2003). The PCR program was as follows: 94 °C for 3 min; 35 cycles of 94 °C for 30 s, 50 °C for 1 min, and 72 °C for 1 min; 72 °C for 3 min. The PCR products were separated by 1.5% agarose gel electrophoresis with TBE buffer, and were visualized by staining with ethidium bromide. The target DNA fragments were extracted and purified from the agarose gels with the High Pure PCR Product Purification



Kit (Roche, Mannheim, Germany). The purified DNA fragments were directly sequenced with the BigDye Terminator (version 3.1) Cycle Sequencing kit (Applied Biosystems, USA) and an ABI PRISM 3130xl genetic analyzer (Applied Biosystems). The obtained sequences, which were deposited in the DNA Data Bank of Japan as LC202850–LC202857 (Table 1), were aligned using MUSCLE in the MEGA6 program (Tamura et al. 2013) with other ITS sequences downloaded from GenBank (Table 1, Chase et al. 2003), including sequences from 22 Suaveolentes species, one artificial hybrid N. × didepta (N. debneyi × N. tabacum), N. tabacum, and Petunia axillaris, which served as the ultimate outgroup.

**Table 1** GenBank/DNA Data Bank of Japan accession numbers for rDNA ITS sequences of *Suaveolentes* and related species

Species	Accession no. of ITS <sup>a</sup> AJ492393			
Nicotiana africana Merxm.				
Nicotiana amplexicaulis N. T. Burb.	AJ492394			
Nicotiana benthamiana Domin	AJ492409			
Nicotiana cavicola N. T. Burb.	AJ492395			
Nicotiana debneyi Domin	AJ492439			
Nicotiana eastii Kostoff	AJ492396			
Nicotiana excelsior J. M. Black	AJ492399			
Nicotiana exigua HM. Wheeler	AJ492391			
Nicotiana fragrans Hook.	AJ492397			
Nicotiana goodspeedii HM. Wheeler	AJ492401			
Nicotiana gossei Domin	AJ492390			
Nicotiana hesperis N. T. Burb.	AJ492402			
Nicotiana ingulba J. M. Black	AJ492403			
Nicotiana maritima HM. Wheeler	AJ492404			
Nicotiana megalosiphon Van Huerck and Müll. Arg.	AJ492392			
Nicotiana occidentalis HM. Wheeler	AJ492417			
Nicotiana rosulata (S. Moore) Domin	AJ492405			
Nicotiana rotundifolia Lindl.	AJ492406			
Nicotiana simulans N. T. Burb.	AJ492407			
Nicotiana suaveolens Lehm.	AJ492438			
Nicotiana suaveolens Lehm. PI 230960	LC202850			
Nicotiana suaveolens Lehm. PI 555561	LC202851			
Nicotiana suaveolens Lehm. PI 555563	LC202852			
Nicotiana suaveolens Lehm. PI 555565	LC202853			
Nicotiana suaveolens Lehm. PI 555566	LC202854			
Nicotiana suaveolens Lehm. PI 555567	LC202855			
Nicotiana suaveolens Lehm. PI 555568	LC202856			
Nicotiana suaveolens Lehm. JT	LC202857			
Nicotiana umbratica N. T. Burb.	AJ492400			
Nicotiana velutina HM. Wheeler	AJ492408			
Nicotiana×didepta	AJ492398			
Nicotiana tabacum L.	AJ492447			
Petunia axillaris (Lam.) Britton	AJ492460			

<sup>&</sup>lt;sup>a</sup>AJ accessions are from Chase et al. (2003)

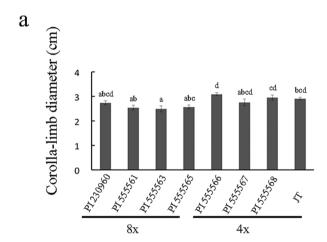


The evolutionary history was inferred using MEGA6 with the maximum-likelihood method based on the General Time Reversible model (Nei and Kumar 2000). Branch support was assessed by the bootstrap resampling with 1,000 replications.

#### **Results**

## Flower and leaf morphology of *N. suaveolens* accessions

The flowers of all accessions were white. The corolla-limb diameter and corolla-tube length was not uniform among the eight *N. suaveolens* accessions (Fig. 1). The corolla-limb diameter ranged from  $2.49 \pm 0.07$  cm (mean and standard error; PI 555563) to  $3.09 \pm 0.06$  cm (PI 555566), and the corolla-tube length ranged from  $3.84 \pm 0.09$  cm (PI 230960) to  $4.5 \pm 0.05$  cm (PI 555566). Accession PI 555566 had the largest corolla-limb diameter and the longest corolla tube.



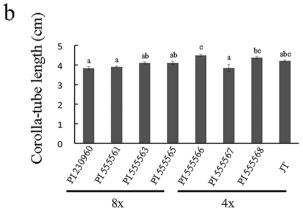
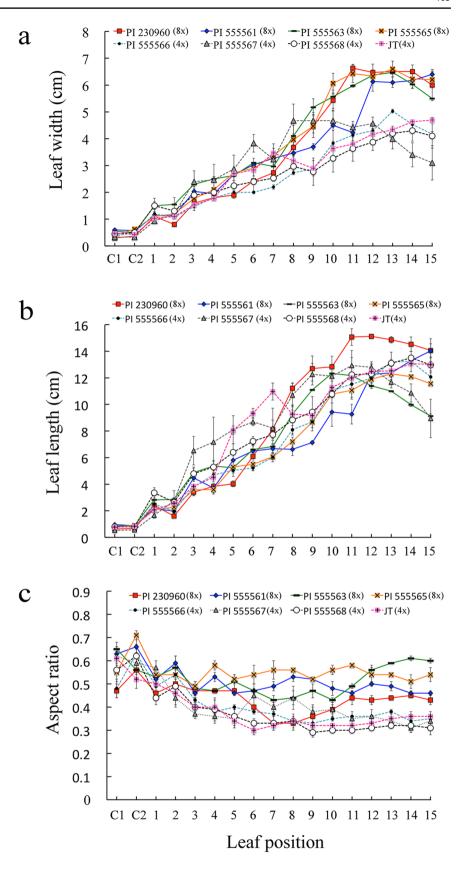


Fig. 1 Flower morphology compared among eight *N. suaveolens* accessions. 4x indicates tetraploid and 8x indicates octoploid. Data are presented as the mean±standard error for ten flowers. Different lower case letters indicate significant differences at the 0.05 level (Tukey's test)

Fig. 2 Sizes of cotyledons and rosette leaves in eight *N. suaveolens* accessions. a Maximum width of cotyledons and rosette leaves. b Maximum length of cotyledons and rosette leaves. c Aspect ratio of the maximum width and the maximum length. C1 and C2 are cotyledons and the leaves at positions 1–15 are 1st to 15th rosette leaves. Error bars indicate the standard error for three individuals





Moreover, the size of the leaves varied among the eight *N. suaveolens* accessions. The maximum leaf widths tended to be greater at the upper leaf positions (10th–15th leaves) in PI 230960, PI 555561, PI 555563, and PI 555565 (Fig. 2a, Table S1). Regarding the maximum leaf lengths, no clear correlation was observed among the eight accessions, although some differences were detected. Accession PI 230960 had the longest leaves (8th–15th leaves) (Fig. 2b, Table S2). The leaf aspect ratios tended to be larger at the upper leaf positions (7th–15th leaves) in PI 230960, PI 555561, PI 555563, and PI 555565 (Fig. 2c, Table S3). Subsequent experiments using flow cytometry and chromosome analyses revealed that the ploidy levels of these four accessions were higher than those of the other four accessions.

## Interspecific crosses between N. suaveolens accessions and N. tabacum

The results of the reciprocal crosses between the eight accessions of N. suaveolens and N. tabacum as well as the selfcrosses of eight N. suaveolens accessions and N. tabacum are presented in Table 2. Selfed seeds were obtained from all accessions, with 91.2-100% of the flowers producing capsules with seeds. Hybrid seeds were obtained from all interspecific crosses between *N. suaveolens* accessions as females and N. tabacum as the male, except for the cross involving PI 555565. Conversely, hybrid seeds were not obtained when N. tabacum was used as the female parent in crosses with eight N. suaveolens accessions, and all flowers dropped at about 7 DAP without the ovaries enlarging, suggesting that fertilization did not occur. Thus, UI was observed in these interspecific crosses. In three interspecific crosses (N. suaveolens PI 230960×N. tabacum, PI 555561×N. tabacum, and PI 555563 × N. tabacum), only 42.1–54.5% of the flowers produced capsules with seeds, which were significantly lower compared with each self-cross. In contrast, in the other four interspecific crosses (N. suaveolens PI  $555566 \times N$ . tabacum, PI555567 × N. tabacum, PI 555568 × N. tabacum, and JT  $\times N$ . tabacum), more than 78.5% of the flowers produced capsules with seeds, which were not significantly different from each self-cross (Fig. 3a). Additionally, in the cross between PI 555565 and N. tabacum, all of the ovaries were enlarged after pollination, suggesting that fertilization had occurred. However, the ovaries dropped at 12-17 days after pollination (DAP) and seeds were never obtained. In all self-crosses and interspecific crosses between N. suaveolens and N. tabacum excluding that involving PI 555565, several flowers did not develop into capsules and dropped at about 7 DAP without the ovaries enlarging, suggesting that fertilization did not occur.

The selfed seeds of eight *N. suaveolens* accessions are presented in Fig. S1, and the germination rates (percentages

of hybrids obtained after sowing) of all accessions were greater than 80% (Fig. 3b). We identified two kinds of hybrid seeds after the interspecific crosses. Most of the hybrid seeds from three crosses (N. suaveolens PI 230960 × N. tabacum, PI 555561  $\times N$ . tabacum, and PI 555563  $\times N$ . tabacum) were fragile and empty, and were obviously different from the seeds resulting from self-crosses (Fig. S1b, d, f). Meanwhile, the hybrid seeds from the other four crosses (N. suaveolens PI 555566 × N. tabacum, PI555567 × N. tabacum, PI 555568 × N. tabacum, and JT × N. tabacum) appeared normal and were similar to the seeds resulting from self-crosses (Fig. S1i, k, m, o). In the crosses between three N. suaveolens accessions (PI 230960, PI 555561 and PI 555563) and N. tabacum, the germination rates of the hybrid seeds were low (0% for PI 230960×N. tabacum, 4.2% for PI 555561 × N. tabacum, and 1.7% for PI 555563 × N. tabacum), and were significantly lower compared with each selfcross. Conversely, the germination rates were high ( $\geq 87.6\%$ ) for the other four interspecific crosses, and were not significantly different from each self-cross (Fig. 3b). Almost all of the hybrid seedlings obtained from all crosses exhibited lethality at 30 days after germination at 28 °C. The characteristic symptom of hybrid lethality in all crosses was the browning of the hypocotyl and roots, which was consistent with Type II hybrid lethality. However, one hybrid seedling from the PI  $555566 \times N$ . tabacum cross and two hybrid seedlings from the JT $\times N$ . tabacum cross exhibited no lethal symptoms after 30 days after germination (Table 2).

## Ploidy levels and chromosome numbers in *N. suaveolens* accessions

The DNA contents of the eight N. suaveolens accessions clearly indicated differences in the ploidy levels (Fig. S2). In N. suaveolens JT, a large G1 (4C) peak and a smaller G2/M (8C) peak were observed (Fig. S2h). Additionally, 4C and 8C peaks were detected in three N. suaveolens accessions (PI 555566, PI 555567 and PI 555568) (Fig. S2e–g). In contrast, 8C (+ 16C) peaks were detected in four N. suaveolens accessions (PI 230960, PI 555561, PI 555563 and PI 555565) (Fig. S2a-d). Thus, four of the N. suaveolens accessions (PI 230960, PI 555561, PI 555563 and PI 555565) were plausibly octoploid, whereas the other three accessions (PI 555566, PI 555567 and PI 555568) were tetraploid, similar to JT. Four N. suaveolens accessions (PI 230960, PI 555561, PI 555563 and PI 555565) comprised 64 chromosomes (Fig. S3a-d), while the other four accessions (PI 555566, PI 555567, PI 555568 and JT) carried 32 chromosomes (Fig. S3e-h). These results were consistent with the flow cytometry analysis.



**Table 2** Interspecific crosses between eight *N. suaveolens* accessions and *N. tabacum* 

$Cross (\stackrel{\bigcirc}{+} \times \stackrel{\frown}{\circlearrowleft})$	Pollination period <sup>a</sup>	No. of flow- ers pollinated	No. of capsules obtained	No. of seeds sown	No. of hybrids obtained <sup>b</sup>	
					Viable	Lethal
PI 230960 (8x) self	4th round	10	10	100	97	0
PI 555561 (8x) self	4th round	14	14	95	83	0
PI 555563 (8x) self	4th round	10	10	111	102	0
PI 555565 (8x) self	4th round	15	14	104	84	0
PI 555566 (4x) self	4th round	12	11	115	101	0
PI 555567 (4x) self	4th round	10	10	110	96	0
PI 555568 (4x) self	4th round	10	10	100	86	0
JT (4x) self	4th round	12	12	100	88	0
N. tabacum self	4th round	11	11	100	98	0
PI 230960 × <i>N. tabacum</i>	1st and 2nd rounds	19	8	214	0	0
PI 555561 × <i>N. tabacum</i>	1st and 2nd rounds	10	5	286	0	12
PI 555563× <i>N. tabacum</i>	1st and 2nd rounds	11	6	525	0	9
PI 555565 × <i>N. tabacum</i>	1st and 2nd rounds	25	$0^{c}$	_	_	_
PI 555566 × <i>N. tabacum</i>	1st and 2nd rounds	14	11	197	1	186
PI 555567 × <i>N. tabacum</i>	1st and 2nd rounds	10	8	185	0	162
PI 555568 × <i>N. tabacum</i>	1st and 2nd rounds	16	16	148	0	143
JT×N. tabacum	3rd round	11	11	164	2	149
N. tabacum×PI 230960	4th round	9	0	_	_	_
N. tabacum×PI 555561	4th round	7	0	_	_	_
N. tabacum×PI 555563	4th round	7	0	_	_	_
N. tabacum×PI 555565	4th round	9	0	_	_	_
N. tabacum×PI 555566	4th round	9	0	_	_	_
N. tabacum×PI 555567	4th round	8	0	_	_	_
N. tabacum×PI 555568	4th round	7	0	_	_	_
N. tabacum×JT	4th round	8	0	_	_	_

<sup>&</sup>lt;sup>a</sup>1st round, Jun.-Sep. 2009, 2nd round, Jun.-Sep. 2012, 3rd round, Jun.-Sep. 2015, 4th round, Oct. 2017–Apr. 2018

## Phylogenetic relationships between *N. suaveolens* accessions and other *Suaveolentes* species inferred from the ITS

The phylogenetic analysis revealed that the eight  $N.\ suaveo-lens$  accessions could be divided into three clades mainly corresponding to their ploidy levels (Fig. 4). The first clade comprising PI 555567,  $N.\ debneyi$ , and  $N.\times didepta$  (bootstrap < 50), was related with other  $N.\ suaveolens$  accessions across two clades. Three tetraploid accessions, PI 555566, PI 555568 and JT, formed a clade that included  $N.\ amplexical$  (bootstrap = 71). This clade was a sister to a clade that included the four octoploid accessions, PI 230960, PI 555561, PI 555563 and PI 555565, as well as the  $N.\ suaveolens$  accession (ITS Accession no. AJ492438) used by Chase et al. (2003) (bootstrap = 70).

#### **Discussion**

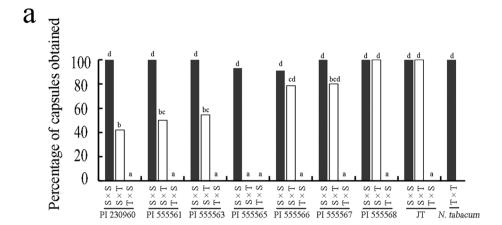
In N. suaveolens, in addition to lines with 16 pairs of chromosomes, lines with 32 pairs of chromosomes were described by Wheeler (1935); however, since then, lines with 32 pairs of chromosomes have not been reported, and it was unclear which lines possess 32 pairs of chromosomes. In the present study, we confirmed that four accessions, PI 230960, PI 555561, PI 555563 and PI 555565, have 32 pairs of chromosomes. Although flower sizes were not correlated with ploidy levels, the maximum widths and the aspect ratios of leaves tended to be larger in these accessions. Our molecular phylogenetic analyses implied that PI 555567 is not grouped with the other three tetraploid N. suaveolens accessions, and instead belongs to the clade with N. debneyi and  $N. \times didepta$ . Although this implies that PI 555567 may not be *N. suaveolens*, the present study is not sufficient to revolve the taxonomy of this accession.

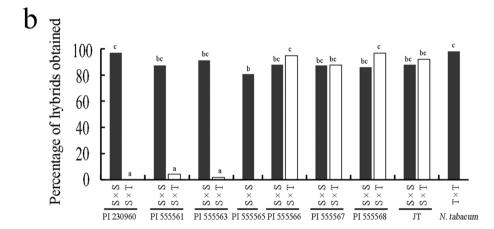


<sup>&</sup>lt;sup>b</sup>Number of hybrids was assessed at 30 days after germination

<sup>&</sup>lt;sup>c</sup>Ovaries enlarged, but dropped at 12-17 days after pollination

Fig. 3 Percentages of capsules obtained after pollination (a) and hybrids obtained after sowing b in self and interspecific crosses. S indicates N. suaveolens and T indicates N. tabacum. Different lower case letters indicate significant differences at the 0.05 level (Chi square test)





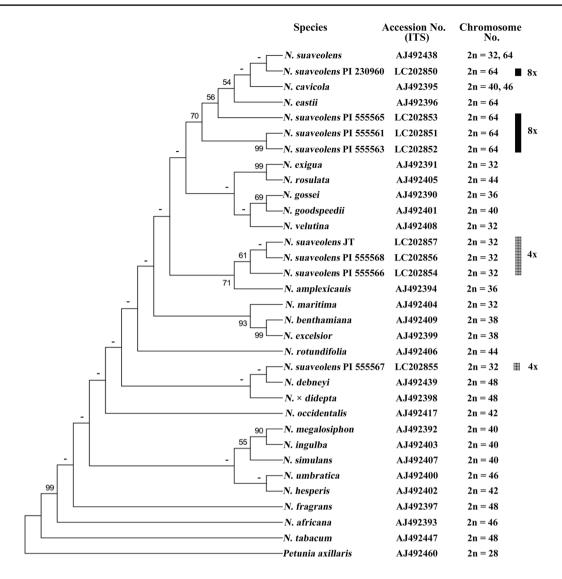
On the basis of our phylogenetic tree constructed from ITS sequences as well as an evaluation of ploidy levels in N. suaveolens accessions, we estimated the evolutionary order and timing of the acquisition of the reproductive isolation mechanisms in the N. suaveolens lineage observed after crosses with N. tabacum. First, because Type II hybrid lethality was observed in all N. suaveolens accessions that produced hybrid seedlings, Type II hybrid lethality was likely acquired in the progenitors of N. suaveolens. Most Suaveolentes species exhibit Type II hybrid lethality in crosses with N. tabacum, and it is thought that the ancestral diploid species of the section Suaveolentes, the allotetraploid ancestor derived from crosses between the ancestral diploid species, or the allotetraploid descendent acquired Type II hybrid lethality during the evolutionary process (Tezuka 2013; Tezuka and Marubashi 2012).

A few hybrid seedlings that did not express lethality were obtained in two crosses in which hybrid seedlings usually express lethality (PI 555566×*N. tabacum* and JT×*N. tabacum*). We previously determined that a few viable hybrids are occasionally obtained in generally lethal crosses because either the whole Q chromosome has been eliminated or the region that includes a hybrid lethality gene or genes on the Q chromosome has been deleted (Tezuka et al. 2010, 2012).

However, other mechanisms are also possible because several viable hybrids appeared to have intact Q chromosomes in the previous study (Tezuka et al. 2010).

Second, UI was observed when N. tabacum was used as the female parent in crosses with eight N. suaveolens accessions. Moreover, UI was also observed when other Suaveolentes species were used for crosses with N. tabacum, although hybrid seeds were obtained relatively easily when N. africana and N. fragrans were used (Tezuka and Marubashi 2012; Tezuka et al. 2010). Therefore, we think that UI may have been acquired by the ancestral species of the section Suaveolentes after the divergence of N. africana and N. fragrans. Additionally, possible prezygotic barriers were observed in crosses between three octoploid N. suaveolens accessions (PI 230960, PI 555561, and PI 555563) and N. tabacum, but not in crosses between tetraploid N. suaveolens accessions and N. tabacum. This implied that the possible prezygotic barriers were acquired during or after the chromosome doubling that occurred in the clade with the octoploid *N. suaveolens* group. However, there is some controversy over the N. suaveolens octoploid accession PI 555565 because prezygotic barriers appeared to be absent in crosses with N. tabacum. Nevertheless, we cannot exclude the possibility that some prezygotic barriers led to





**Fig. 4** Molecular phylogenetic tree based on a maximum-likelihood analysis of rDNA ITS sequences in the section *Suaveolentes. Petunia axillaris* was used as an outgroup. The eight *N. suaveolens* accessions were sequenced. Additionally, the sequences of 22 *Suaveolentes* species, the artificial hybrid *N. didepta*, *N. tabacum*, and *P. axillaris* 

were obtained from the GenBank database. Chromosome numbers were based on published articles by Goodspeed (1954), Purdie et al. (1982), and Japan Tobacco Inc. (1994). Bootstrap = 1,000 replicates (any clade with a hyphen has a BP < 50)

the abscission of the enlarged ovaries in the cross between PI 555565 and *N. tabacum*. Future studies will need to verify or disprove this possibility.

Third, seed abortion was observed after crosses between *N. tabacum* and three octoploid *N. suaveolens* accessions (PI 230960, PI 555561 and PI 555563), but not with the tetraploid *N. suaveolens* accessions. This suggested that seed abortion mechanisms were acquired during or after the chromosome doubling events. In interploidy crosses, endosperm breakdown is widely believed to be responsible for seed failure (Bushell et al. 2003; Kradolfer et al. 2013). Scott et al. (1998) described the seed failure in *Arabidopsis thaliana* interploidy crosses, in which the seeds contained different doses of maternally or paternally expressed imprinted loci

that affected endosperm development. Similar phenomena have also been observed in the seed failure of interspecific crosses between species with the same ploidy level (Ishikawa et al. 2011; Sekine et al. 2013). Additionally, Rebernig et al. (2015) reported that interspecific crosses between *Capsella rubella* and *C. grandiflora*, which have the same ploidy level, resulted in seed abortion due to failed endosperm cellularization, and crosses in the opposite direction resulted in the formation of small seeds (with precociously cellularized endosperm) capable of germinating. Therefore, there are two possible causes of the seed abortion in crosses between octoploid *N. suaveolens* accessions and *N. tabacum*. First, the chromosome doubling event(s) in *N. suaveolens* may be the sole cause of seed abortion. Second, genetic changes



accumulated in the lineage leading to the octoploid lines before or after the chromosome doubling event(s), and the subsequent lines developed the mechanisms responsible for seed abortion.

Fourth, in this study, the abscission of enlarged ovaries (immature fruits) was only observed in crosses between the *N. suaveolens* octoploid accession PI 555565 and *N. tabacum*. Although we did not investigate pollen tube growth, fertilization appeared to have occurred because the ovules in the dropped ovaries were enlarged. Therefore, we speculated that some postzygotic barriers were present during the developmental processes of seeds and/or fruits in this cross. Similar to seed abortion, there are two possible causes for this abscission; i.e., the chromosome doubling event(s) in *N. suaveolens* or the acquisition of genetic factors responsible for the abscission of enlarged ovaries in PI 555565.

In summary, in the present study, we confirmed that *N. suaveolens* accessions vary regarding ploidy levels and reproductive isolation mechanisms in crosses with *N. tabacum*. These accessions will be very useful for revealing and characterizing the reproductive isolation mechanisms in interspecific crosses and their relationships with ploidy levels.

**Acknowledgements** This study was partly supported by JSPS KAK-ENHI Grants (JP20880024 and JP25870627) from the Japan Society for the Promotion of Science.

#### References

- Baldwin BG (1993) Molecular phylogenetics of Calycadenia (Compositae) based on ITS sequences of nuclear ribosomal DNA: chromosomal and morphological evolution reexamined. Am J Bot 80:222–238
- Bushell C, Spielman M, Scott RJ (2003) The basis of natural and artificial postzygotic hybridization barriers in *Arabidopsis* species. Plant Cell 15:1430–1442
- Chase MW, Knapp S, Cox AV et al (2003) Molecular systematics, GISH and the origin of hybrid taxa in *Nicotiana* (Solanaceae). Ann Bot 92:107–127
- Clarkson JJ, Knapp S, Garcia VF, Olmstead RG, Leitch AR, Chase MW (2004) Phylogenetic relationships in *Nicotiana* (Solanaceae) inferred from multiple plastid DNA regions. Mol Phylogenet Evol 33:75–90
- Clarkson JJ, Kelly LJ, Leitch AR, Knapp S, Chase MW (2010) Nuclear glutamine synthetase evolution in *Nicotiana*: phylogenetics and the origins of allotetraploid and homoploid (diploid) hybrids. Mol Phylogenet Evol 55:99–112
- Dickinson GR, Lee DJ, Wallace HM (2012) The influence of pre- and post-zygotic barriers on interspecific *Corymbia* hybridization. Ann Bot 109:1215–1226
- Doležel J, Bartos J (2005) Plant DNA flow cytometry and estimation of nuclear genome size. Ann Bot 95:99–110
- Goodspeed TH (1954) The genus Nicotiana. Chronica Botanica Company, Waltham

- Guo J, Xu X, Li W et al (2016) Overcoming inter-subspecific hybrid sterility in rice by developing indica-compatible *japonica* lines. Sci Rep 6:26878
- Ichitani K, Namigoshi K, Sato M et al (2007) Fine mapping and allelic dosage effect of *Hwc1*, a complementary hybrid weakness gene in rice. Theor Appl Genet 114:1407–1415
- Inoue E, Marubashi W, Niwa M (1994) Simple method for overcoming the lethality observed in the hybrid between *Nicotiana suaveolens* and *N. tabacum*. Breed Sci 44:333–336
- Ishikawa R, Ohnishi T, Kinoshita Y, Eiguchi M, Kurata N, Kinoshita T (2011) Rice interspecies hybrids show precocious or delayed developmental transitions in the endosperm without change to the rate of syncytial nuclear division. Plant J 65:798–806
- Japan Tobacco Inc (1994) The genus *Nicotiana* illustrated. Japan Tobacco Inc, Tokyo
- Kradolfer D, Wolff P, Jiang H, Siretskiy A, Köhler C (2013) An imprinted gene underlies postzygotic reproductive isolation in *Arabidopsis thaliana*. Dev Cell 26:525–535
- Kuboyama T, Saito T, Matsumoto T et al (2009) Fine mapping of *HWC2*, a complementary hybrid weakness gene, and haplotype analysis around the locus in rice. Rice 2:93–103
- Ladiges PY, Marks CE, Nelson G (2011) Biogeography of *Nicotiana* section *Suaveolentes* (Solanaceae) reveals geographical tracks in arid Australia. J Biogeogr 38:2066–2077
- Leitch IJ, Hanson L, Lim KY et al (2008) The ups and downs of genome size evolution in polyploid species of *Nicotiana* (Solanaceae). Ann Bot 101:805–814
- Lewis RS, Nicholson JS (2007) Aspects of the evolution of *Nicotiana tabacum* L. and the status of the United States *Nicotiana* Germplasm Collection. Genet Resour Crop Evol 54:727–740
- Li Z, Pinson SRM, Paterson AH, Park WD, Stansel JW (1997) Genetics of hybrid sterility and hybrid breakdown in an intersubspecific rice (*Oryza sativa L.*) population. Genetics 145:1139–1148
- Manabe T, Marubashi W, Onozawa Y (1989) Temperature-dependent conditional lethality in interspecific hybrids between *Nicotiana suaveolens* Lehm. and *N. tabacum* L. In: Proceedings of the 6th international congress of SABRAO, pp 459–462
- Marubashi W, Onosato K (2002) Q chromosome controls the lethality of interspecific hybrids between *Nicotiana tabacum* and *N. suaveolens*. Breed Sci 52:137–142
- Murashige T, Skoog F (1962) A revised medium for rapid growth and bio assays with tobacco tissue cultures. Physiol Plant 15:473–497
- Murray MG, Thompson WF (1980) Rapid isolation of high molecular weight plant DNA. Nucleic Acids Res 8:4321–4325
- Nei M, Kumar S (2000) Molecular evolution and phylogenetics. Oxford University Press, New York
- Otto F (1990) DAPI staining of fixed cells for high-resolution flow cytometry of nuclear DNA. In: Darzynkiewickz Z, Crissman HA (eds) Methods in cell biology. Academic Press, San Diego, pp 105–110
- Purdie RW, Symon DE, Haegi L (1982) Solanaceae. Flora of Australia 29:1–208
- Rebernig CA, Lafon-Placette C, Hatorangan MR, Slotte T, Köhler C (2015) Non-reciprocal interspecies hybridization barriers in the *Capsella* genus are established in the endosperm. PLoS Genet 11:e1005295
- Rieseberg LH, Blackman BK (2010) Speciation genes in plants. Ann Bot 106:439–455
- Scott RJ, Spielman M, Bailey J, Dickinson HG (1998) Parent-of-origin effects on seed development in *Arabidopsis thaliana*. Development 125:3329–3341
- Sekine D, Ohnishi T, Furuumi H et al (2013) Dissection of two major components of the post-zygotic hybridization barrier in rice endosperm. Plant J 76:792–799



- Stebbins GL (1966) Reproductive isolation and the origin of species. Processes of organic evolution. Prentice-Hall, Upper Saddle River, pp 85–112
- Sun Y, Skinner DZ, Liang GH, Hulbert SH (1994) Phylogenetic analysis of *Sorghum* and related taxa using internal transcribed spacers of nuclear ribosomal DNA. Theor Appl Genet 89:26–32
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. Mol Biol Evol 30:2725–2729
- Tezuka T (2012) Hybrid lethality in the genus *Nicotiana*. In: Mworia JK (ed) Botany. InTech, Rijeka, pp 191–210
- Tezuka T (2013) Hybrid lethality in *Nicotiana*: a review with special attention to interspecific crosses between species in sect. *Suaveolentes* and *N. tabacum*. In: Wallner F (ed) Herbaceous plants: cultivation methods, grazing and environmental impacts. Nova Science Publishers, New York, pp 69–94
- Tezuka T, Marubashi W (2004) Apoptotic cell death observed during the expression of hybrid lethality in interspecific hybrids between *Nicotiana tabacum* and *N. suaveolens*. Breed Sci 54:59–66
- Tezuka T, Marubashi W (2006) Hybrid lethality in interspecific hybrids between *Nicotiana tabacum* and *N. suaveolens*: evidence that the Q chromosome causes hybrid lethality based on Q-chromosome-specific DNA markers. Theor Appl Genet 112:1172–1178
- Tezuka T, Marubashi W (2012) Genes in S and T subgenomes are responsible for hybrid lethality in interspecific hybrids between *Nicotiana tabacum* and *Nicotiana occidentalis*. PLoS One 7:e36204

- Tezuka T, Kuboyama T, Matsuda T, Marubashi W (2010) Seven of eight species in *Nicotiana* section *Suaveolentes* have common factors leading to hybrid lethality in crosses with *Nicotiana tabacum*. Ann Bot 106:267–276
- Tezuka T, Matsuo C, Iizuka T, Oda M, Marubashi W (2012) Identification of *Nicotiana tabacum* linkage group corresponding to the Q chromosome gene(s) involved in hybrid lethality. PLoS One 7:e37822
- USDA ARS National Genetic Resources Program (2010) Germplasm Resources Information Network—(GRIN) [Online Database]. National Germplasm Resources Laboratory, Beltsville, Maryland. https://www.ars-grin.gov/npgs/index.html. Accessed Apr 2015
- Wheeler HM (1935) Studies in *Nicotiana*. II. A taxonomic survey of the Australasian species. Univ Calif Publ Bot 18:45–68
- Yamada T, Marubashi W, Niwa M (1999) Detection of four lethality types in interspecific crosses among *Nicotiana* species through the use of three rescue methods for lethality. Breed Sci 49:203–210
- Yamada T, Marubashi W, Niwa M (2000) Apoptotic cell death induces temperature-sensitive lethality in hybrid seedlings and calli derived from the cross of *Nicotiana suaveolens* × *N. tabacum*. Planta 211:614–622

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

