#### **RESEARCH PAPER**



# **The efects of presence of macrophytes on resource uptake by phytoplankton and zooplankton in a tropical reservoir**

**Anamaria Silva Diniz1 · Watson Arantes Gama1 · Ariadne do Nascimento Moura[1](http://orcid.org/0000-0001-5093-2840)**

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#### **Abstract**

The understanding of factors afecting resource uptake is important to elucidate community dynamics in aquatic ecosystems. Here, we analyzed nutrient–phytoplankton and phytoplankton–zooplankton interactions in open water areas without macrophytes and in areas with foating and submerged macrophytes in the tropical Cajueiro reservoir, semiarid Brazil. Phytoplankton species were classifed into functional groups according to life form and cell size, and zooplankton species were grouped based on functional feeding groups and trophic guilds. Macrophytes favored the efect of nutrients on phytoplankton and the availability of prey for the zooplankton. In open water, unicellular and fagellated phytoplankton were positively infuenced by nitrate and inorganic phosphate, while colonial, flamentous, small, medium and large phytoplankton were positively infuenced by total phosphorus and nitrite. Colonial phytoplankton, mainly flamentous cyanobacteria, was positively associated with zooplankton in areas with macrophytes, while fagellated phytoplankton was negatively related to zooplankton in open water areas. Our results showed that the presence of foating and submerged macrophytes has diferent infuences on the efect of nutrients on phytoplankton and on the efects of phytoplankton on zooplankton in Cajueiro reservoir, and this should be considered when analyzing these communities' dynamics in similar environments, i.e. tropical reservoirs.

**Keywords** Colonial cyanobacteria · Filter-feeding zooplankton · Filter-scraper zooplankton · Flagellate · Nitrogen · Phosphorus

## **Introduction**

Trophic interactions have been studied in aquatic ecology for more than fve decades (Martin [1970;](#page-11-0) McQueen et al. [1986](#page-11-1); Brett and Goldman [1997;](#page-10-0) Sommer [2008](#page-11-2); Diniz et al. [2019](#page-10-1); Severiano et al. [2021](#page-11-3)). The food chain structure can be strongly regulated by bottom-up effects, that is, when the energy fow in the food chain occurs as a function of

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 $\boxtimes$  Ariadne do Nascimento Moura ariadne\_moura@hotmail.com

> Anamaria Silva Diniz anamaria.s.diniz@gmail.com

Watson Arantes Gama watson.arantes@ufrpe.br

<sup>1</sup> Department of Biology, Graduate Program in Biodiversity, Federal Rural University of Pernambuco-UFRPE, Manoel de Medeiros Avenue, Dois Irmãos, Recife, PE CEP 52171-900, Brazil

available resources (McQueen et al. [1989](#page-11-4)). In this sense, nutrients are the main factors controlling phytoplankton growth (Carpenter et al. [1985](#page-10-2)), with nitrogen and phosphorus being the limiting nutrients (Müller and Mitrovic [2015](#page-11-5); Paerl et al. [2018](#page-11-6); Lewis et al. [2020;](#page-11-7) Liu et al. [2021\)](#page-11-8).

The effects of nutrients on phytoplankton have been widely demonstrated in several ecosystems (Smith and Lancelot [2004](#page-11-9); Jeppesen et al. [2012](#page-10-3); Liu et al. [2018](#page-11-10)). More recently, studies have shown that bottom-up efects are more common in phytoplankton communities than top-down regulation by zooplankton (Li et al. [2020](#page-11-11); Frau et al. [2021\)](#page-10-4). For zooplankton, trophic interactions are infuenced by functional feeding groups and tropic guilds, species body size, and selectivity and physiological tolerance to ingested toxins (DeMott [1986;](#page-10-5) Kiørboe et al. [2018;](#page-10-6) Gomes et al. [2019\)](#page-10-7), and by defense characteristics of the prey (Ger et al. [2014](#page-10-8); Lür-ling [2021](#page-11-12)). In addition, the strength of bottom-up effects on zooplankton is dependent on the degree of predation pres-sure by fish as top-down effects (Braun et al. [2021\)](#page-9-0).

Nutrient uptake by phytoplankton is related to the size and structure of organisms, with intermediate-sized phytoplankton being able to exploit available nutrients more efficiently (Marañón [2015;](#page-11-13) Mousing et al. [2018\)](#page-11-14). Along with nutrient availability, water turbulences also infuence phytoplankton metabolism, since they increase nutrient uptake, especially in larger organisms (Naselli-Flores et al. [2021](#page-11-15)).

Throughout evolution, phytoplankton developed diferent and efficient morphological, physiological and behavioral strategies to avoid predation by zooplankton, among which the main morphological strategies are the formation of spines and colonies and the increase in size (Lürling [2021](#page-11-12)). These strategies can be described as functional traits based on their role in regulating predation by zooplankton (Colina et al. [2016\)](#page-10-9). They are informative in explaining the dynamics and structure of the phytoplankton community and its relationship with environmental factors (Chen et al. [2019](#page-10-10); Kruk et al. [2021\)](#page-10-11), such as the climatic, chemical and biological factors of the environment (Reynolds et al. [2002](#page-11-16)).

In freshwater ecosystems, macrophytes have several efects on nutrient cycling, including a reduction in sediment resuspension through water column stabilization (Nurminen and Horppila [2009\)](#page-11-17). Many of the nutrients taken up by macrophytes return to the environment through the decomposition of plant tissues (Banks and Frost [2017;](#page-9-1) Xiao et al. [2017](#page-12-0)), or by the release of nutrients from the reductive sediment through thermal stratifcation of the water column induced by macrophytes (Vilas et al. [2018\)](#page-12-1), or by fuctuation in water level (Keitel et al. [2016\)](#page-10-12). Although little is known about the efects of macrophytes on the availability of resources for aquatic communities in tropical reservoirs, studies have shown that nutrients released by macrophytes favor phytoplankton growth (Vilas et al. [2018](#page-12-1); Wang et al. [2018](#page-12-2)), and also metaphyton, which consists of large sized algae (e.g. *Oedogonium* sp., *Oscillatoria* sp.) that are associated with the presence of macrophytes (Barrow et al. [2019](#page-9-2)). In this sense, macrophytes are also important in the interaction between phytoplankton and zooplankton, as they increase the availability of prey for zooplankton (Fischer and Pusch [2001;](#page-10-13) Kovalenko et al. [2012](#page-10-14)).

The main question of the present study was: do macrophytes enhance resource uptake by phytoplankton and zooplankton communities? To answer this question, we examined the efects of resource availability on phytoplankton and zooplankton in open water areas without macrophytes and in areas with foating and submerged macrophytes in a tropical reservoir, in semiarid Brazil. For this, we analyzed the potential factors controlling phytoplankton (nutrients) and zooplankton (prey availability) in a feld study. We tested the following hypotheses: (i) the interaction between phytoplankton–nutrients and phytoplankton–zooplankton is favored by the presence of macrophytes, by increasing the availability of resources, and (ii) functional groups of phytoplankton and zooplankton respond differently the resource availability in areas with and without the presence of macrophytes.

### **Materials and methods**

#### **Study area and sampling**

The study was carried out in Cajueiro reservoir (8°59′21.3″ S and 36°28′19.9″ W), municipality of Garanhuns, state of Pernambuco, Northeastern Brazil (Fig. [1](#page-2-0)). The regional climate is tropical "As" (Alvares et al. [2013](#page-9-3)), characterized by dry summers and rainy winters. The dry period is concentrated between September and March, and the rainy period between April and August (APAC [2019](#page-9-4)). The reservoir is shallow, with an average depth of  $5.33 \pm 0.88$  m, and has multiple uses, such as water supply for the municipality, irrigation, fshing and recreation for the population. The Cajueiro reservoir is eutrophic, and the banks are colonized by extensive banks of foating macrophytes *Nymphoides indica* (L.) Kuntze, *Eichhornia crassipes* (Mart.) Solms, *Salvinia auriculata* Aubl. and *Salvinia oblongifolia* Martius, and banks of submerged macrophytes *Chara* sp., *Egeria densa* (Planch.) Casp. and *Myriophyllum aquaticum* (Vell.) Verdc.

Samples for analysis of abiotic variables, phytoplankton and zooplankton were taken in open water areas (absence of macrophytes,  $n=3$ ) and in areas with floating macrophyte banks  $(n=3)$  and submerged macrophyte banks  $(n=3)$ , quarterly between November 2018 and August 2019, comprising an annual cycle. The sampling sites were located approximately 400 m equidistant from each other (Fig. [1](#page-2-0)). Water temperature ( $\rm{°C}$ ), dissolved oxygen (mg L<sup>-1</sup>), pH, electrical conductivity ( $\mu$ S cm<sup>-1</sup>) and total dissolved solids  $(mg L^{-1})$  were measured using a HANNA multiparameter probe (HI 9829). The Secchi disk was used to estimate the water transparency, in meters, and the light intensity (µmol photons  $m^{-1} s^{-2}$ ) was measured with a photometer (LI-250A). Reservoir water samples were collected 10 cm below the water surface using graduated buckets with a capacity of 10 L in open water areas and in areas with foating macrophytes, while in areas with submerged macrophytes, water was collected at a depth of  $0.86 \pm 0.13$  m using a van Dorn bottle. The mean depth in the open water sites was 5.33  $\pm$  0.88 m, and 1.06  $\pm$  0.42 m and 0.86  $\pm$  0.12 in the floating and submerged macrophyte banks, respectively.

Water for nutrient analysis was sampled and immediately placed in 300 mL plastic bottles and kept under refrigeration in a Styrofoam box with ice and transported to the laboratory, where samples were frozen until analysis. Samples for phytoplankton counting were taken directly from the water reservoir at the sampling sites and immediately placed in 150 mL amber fasks and fxed with 1% acetic Lugol. For taxonomic analysis of phytoplankton, water was fltered



<span id="page-2-0"></span>**Fig. 1** Location of the Cajueiro reservoir, target of the present study

through a plankton net with 25 µm mesh, placed in 150 mL vials and fxed with 4% formaldehyde. Samples for identifcation and quantifcation of zooplankton were collected by fltering 100 L reservoir water from open water areas and areas with foating and submerged macrophytes, following the procedures for collecting water used for phytoplankton analysis, but using a plankton net with 50 µm mesh. Subsequently, samples were placed in 200 mL plastic bottles and fxed with 4% formaldehyde.

#### **Laboratory analysis**

The concentration of nutrients ( $\mu$ g L<sup>-1</sup>), including nitrite  $(NO_2^-)$ , nitrate  $(NO_3^-)$  and ammoniacal nitrogen  $(NH_4^+)$ was determined according to Golterman et al.  $(1978)$  $(1978)$ , Mackereth et al. ([1978](#page-11-18)) and Koroleff ([1976](#page-10-16)), respectively. Dissolved inorganic nitrogen (DIN) concentrations were obtained by the sum of concentrations of  $NO_2^-$ ,  $NO_3^-$  and  $NH_4^+$ . Orthophosphate  $(PO_4^{3-})$ , inorganic phosphate (Pi) and total phosphorus (TP) were quantifed according to the A.P.H.A. [\(2005\)](#page-9-5).

Phytoplankton species were identifed under an optical microscope using specifc literature (Anagnostidis and Komárek [1988;](#page-9-6) Komárek and Anagnostidis [1999](#page-10-17), [2005](#page-10-18); Komárek and Cronberg [2001](#page-10-19); Prescott et al. [1982;](#page-11-19) Kram-mer and Lange-Bertalot [1991;](#page-10-20) Popovský and Pfiester [1990](#page-11-20); John et al. [2002\)](#page-10-21), and then classifed into morphofunctional groups according to life form (unicellular, colonial, filamentous and flagellar) and cell size (small:  $<$  50  $\mu$ m, medium:  $50-100 \mu m$  and large:  $>100 \mu m$ ). Phytoplankton density (ind mL−1) was estimated according to Utermöhl [\(1958](#page-12-3)) using sedimentation chambers and an inverted microscope (Bioval XDS-1B) at  $400 \times$  magnification. Biovolume  $\text{(mm}^3 L^{-1})$  was estimated based on the volume of species calculated using geometric models by Hillebrand et al. ([1999](#page-10-22)), and multiplied by the population density for each species. Biovolume (mm<sup>3</sup> L<sup>-1</sup>) was converted to biomass  $(\text{mg } L^{-1})$  according to Wetzel and Likens [\(2000](#page-12-4)).

Zooplankton species were identifed using specifc literature (Koste [1978;](#page-10-23) Reid [1985](#page-11-21); Montú and Goeden [1986](#page-11-22); Elmoor-Loureiro [1997;](#page-10-24) Neumann-Leitão et al. [1989\)](#page-11-23). The density of zooplanktonic species (ind  $L^{-1}$ ) was calculated by counting the organisms under an optical microscope (Opton TNB 41B) with samples concentrated to 100 mL and counting three sub-samples of 2 mL in a Sedgewick-Rafter chamber. Species-specifc biomass of zooplankton (μg DW m<sup>-3</sup>, where DW is dry weight) was determined through the density and the average length and body weight of the taxa according to the regression equations of Ruttner-Kolisko [\(1977](#page-11-24)) for rotifers, and of Dumont et al. ([1975\)](#page-10-25) for copepods and cladocerans. Zooplankton species were grouped based on functional feeding groups and trophic guilds: rotifers as microphagous (Obertegger et al. [2011](#page-11-25)), cladocerans as flter-feeders and flter-scrapers (Barnett et al. [2007](#page-9-7)), adult Calanoida copepods as herbivorous (Frau et al. [2019](#page-10-26)) and Copepoda nauplii as microphagous. Only groups that can feed on phytoplankton (herbivorous or omnivorous zooplankton) were considered.

## **Data analysis**

Potential diferences in phytoplankton and zooplankton biomass and nutrient concentrations between sampling sites (open waters, with foating macrophytes and with submerged macrophytes) were tested by an Analysis of Variance (oneway ANOVA) followed by a Tukey's post hoc test. Normality of variance was tested by the Kolmogorov–Smirnov test, and homoscedasticity by the Bartlett test. For heteroscedastic variances, we applied the non-parametric Kruskal-Walllis test, followed by the pairwise Mann–Whitney U-test.

Generalized additive models (GAM) were applied to test the effect of nutrients on phytoplankton and effects of phytoplankton on zooplankton in open waters and in sites with foating and submerged macrophytes. The estimated degree of freedom (e.d.f.) was used to evaluate the smoothing of models, and the ft and signifcance of the model were evaluated using adjusted  $\mathbb{R}^2$  and p values. Dissolved inorganic nitrogen and orthophosphate for phytoplankton, and total phytoplankton biomass for zooplankton, were used as the predictor variable, respectively. Data were log-transformed  $(x+1)$  before statistical analysis.

Redundancy Analysis (RDA) was used to check the infuence of nutrients on phytoplankton groups and the infuence of phytoplankton groups on zooplankton groups. For this, the analysis was applied based on the length of the frst axis of the Detrended Correspondence Analysis (DCA) (Legendre et al. [2011](#page-10-27)). Dependent variables were log transformed and explanatory variables were standardized. Only variables with variance infation factor below 20 and with a significance of  $p < 0.05$  were considered in the final RDA models through the Ordistep function.

Analyses were performed using the *vegan* (Oksanen et al. [2013](#page-11-26)) and *mgcv* (Wood [2004](#page-12-5), [2011\)](#page-12-6) package in R software (R Development Core Team [2019](#page-11-27)) with a signifcance of  $p < 0.05$ .

# **Results**

#### **Nutrients**

The limnological variables found in Cajueiro reservoir in areas without macrophytes and areas with foating and submerged macrophytes throughout the study period are provided in the supplementary material (Table S1). High concentrations of nutrients were found in the foating macrophyte banks; however, no statistical variation was observed between areas without macrophytes and areas with macrophytes for nitrate  $(F=0.03, p=0.962)$ , nitrite  $(F=0.15,$   $p=0.857$ ), ammoniacal nitrogen (F=0.52, p=0.609), dissolved inorganic nitrogen  $(F=0.52, p=0.609)$ , orthophosphate (F=1.38, p=0.298), inorganic phosphate (H=0.47,  $p = 0.788$ ) and total phosphorus (F = 1.92,  $p = 0.201$ ) (Fig. [2\)](#page-4-0).

#### **Biomass of phytoplankton and zooplankton**

Eighty-four phytoplankton taxa were identifed in Cajueiro reservoir, which were grouped into seven functional groups based on cell size and life form (Table S2). The colonial cyanobacteria *Coelomoron tropicale* P.A.C.Senna, A.C.Peres & Komárek and the flamentous cyanobacteria *Raphidiopsis raciborskii* (Wołoszyńska) Aguilera, Berrendero Gómez, Kastovsky, Echenique & Salerno were the dominant species in the phytoplankton community of the reservoir, representing 30.99% and 10.32% of the total biomass of phytoplankton and constituting the colonial and flamentous groups, respectively. In addition to cyanobacteria, the diatom *Cyclotella meneghiniana* Kützing (9.43%) and dinofagellate *Ceratium furcoides* (Levander) Langhans (8.55%), belonging to the unicellular and flagellate groups, respectively, were dominant in Cajueiro reservoir. The group of algae with fagella was mainly represented by dinofagellates and cryptophyceans. Large and medium flamentous species represented 15.20% total biomass of phytoplankton, and were composed of the diazotrophic cyanobacteria *Raphidiopsis* and *Dolichospermum*, and the non-diazotrophic cyanobacteria *Anagnostidinema*, *Oscillatoria*, *Phormidium*, *Planktothrix* and *Pseudanabaena*.

In areas with foating and submerged macrophytes, a signifcant increase was found for the biomass of unicellular  $(H = 9.78, p = 0.007)$ , colonial  $(F = 3.37, p = 0.046)$ , filamentous (F=7.34, p=0.002), small (F=7.63, p=0.001) and large  $(F=3.76, p=0.034)$  functional groups, except for medium-sized  $(H = 4.54, p = 0.103)$  and flagellated phyto-plankton (F=0.73, p=0.489; Fig. [3](#page-5-0)a–g). The total phytoplankton biomass difered signifcantly between the open water areas and sites with foating and submerged macrophytes, with higher biomass found in the submerged macrophyte banks (96.52±65.09 mg L<sup>-1</sup>, H=9.86, p=0.007; Fig. [3h](#page-5-0)).

Fifty zooplankton taxa were identifed, in addition to the class Bdelloidea and order Calanoida and Copepoda nauplii. Zooplankton taxa were grouped into four functional feeding groups and trophic guilds (Table S3). The herbivorous-microphagous group had the highest biomass during the study; however, it did not difer signifcantly between sampling sites  $(H = 5.57, p = 0.061, Fig. 4a)$  $(H = 5.57, p = 0.061, Fig. 4a)$  $(H = 5.57, p = 0.061, Fig. 4a)$ . Higher total zooplankton biomass was observed in foating macrophyte banks  $(183.80 \pm 146.70 \text{ µg DW m}^{-3})$  and in submerged macrophyte banks  $(169.12 \pm 218.12 \,\mu g$  DW m<sup>-3</sup>), significantly differing  $(H=9.46, p=0.008)$  from open water areas <span id="page-4-0"></span>**Fig. 2** Variation in concentrations ( $\mu$ g L<sup>-1</sup>) of nitrate (**a**), nitrite (**b**), ammoniacal nitrogen (**c**), dissolved inorganic nitrogen (DIN, **d**), orthophosphate (**e**), inorganic phosphate (**f**) and total phosphorus (**g**) between open water areas (OW) and areas with foating (FM) and submerged (SM) macrophytes in Cajueiro reservoir, Northeastern Brazil. Lowercase letters represent statistical diferences based on the post-hoc test  $(p < 0.05)$ among sampling sites



 $(37.88 \pm 30.71 \text{ µg DW m}^{-3})$ ; Fig. [4e](#page-6-0)). Floating and submerged macrophytes favored the increase in biomass of the filter-scraper functional group  $(F = 24.18, p < 0.001)$ , and foating macrophytes favored the increase in flter-feeder biomass ( $F = 3.48$ ,  $p = 0.042$ ; Fig. [4c](#page-6-0)–d). Microphagous biomass did not difer signifcantly between sampling sites  $(F=0.58, p=0.565; Fig. 4b).$  $(F=0.58, p=0.565; Fig. 4b).$  $(F=0.58, p=0.565; Fig. 4b).$ 

# **Interaction between nutrients–phytoplankton and phytoplankton–zooplankton**

Generalized additive models (GAM) showed that total phytoplankton biomass had a marginally signifcant positive relationship with dissolved inorganic nitrogen in open water areas ( $p = 0.054$ , Fig. [5a](#page-7-0)) and a significantly

<span id="page-5-0"></span>**Fig. 3** Variation in concentrations (mg  $L^{-1}$ ) of unicellular (**a**), colonial (**b**), flamentous (**c**), fagellated (**d**), small (**e**), medium (**f**) and large (**g**) morphofunctional groups of phytoplankton and total phytoplankton biomass (**h**) between open water areas (OW) and areas with foating (FM) and submerged (SM) macrophytes in Cajueiro reservoir, Northeastern Brazil. Lowercase letters represent statistical diferences based on the post-hoc test  $(p<0.05)$  among sampling sites



negative relationship in sites with foating macrophytes  $(p = 0.0003,$  Fig. [5b](#page-7-0)). No significant relationship was observed between dissolved inorganic nitrogen and total phytoplankton biomass in areas with submerged macrophytes ( $p = 0.654$ , Fig. [5c](#page-7-0)). A positive relationship was found between orthophosphate and total phytoplankton biomass in open water areas ( $p=0.039$ , Fig. [5](#page-7-0)d) and with submerged macrophytes ( $p=0.022$ , Fig. [5](#page-7-0)f), while in sites with foating macrophytes, no signifcant relationship was observed ( $p=0.076$ , Fig. [5](#page-7-0)e). Total zooplankton biomass was not signifcantly related to total phytoplankton biomass (Fig. [5](#page-7-0)g–i).

<span id="page-6-0"></span>**Fig. 4** Variation in concentrations ( $\mu$ g DW m<sup>-3</sup>) of herbivorous (**a**), microphagous (**b**), flter-feeders (**c**) and flter-scrapers (**d**) functional groups of zooplankton and total zooplankton biomass (**e**) between open water areas (OW) and areas with foating (FM) and submerged (SM) macrophytes in Cajueiro reservoir, Northeastern Brazil. Lowercase letters represent statistical diferences based on the post-hoc test  $(p < 0.05)$  among sampling sites



#### **Redundancy analysis**

The RDA model revealed that nutrients explained 59.65% of variation in phytoplankton community compositions  $(F=11.08, p=0.001)$ . Axis 1  $(F=24.64, p=0.001)$  and axis 2 ( $F = 17.62$ ,  $p = 0.001$ ) were significant for the distribution of explanatory variables, with unicellular phytoplankton infuenced by inorganic phosphate (Pi), fagellate phytoplankton by nitrate  $(NO_3^-)$ , and other groups (colonial, filamentous, small, medium and large) infuenced by total phosphorous (TP) and nitrite  $(NO<sub>2</sub>$ . Two groups distinguishing the sites with and without macrophytes were formed on axis 1, with TP and  $NO_2$ <sup> $-$ </sup> related to functional phytoplankton groups in foating and submerged macrophyte banks and Pi and  $NO_3$ <sup> $-$ </sup> in sites without macrophytes (Fig. [6](#page-8-0)a).

For functional feeding groups and trophic guilds of the zooplankton, the RDA indicated that phytoplankton functional groups explained 41.79% of variation in zooplankton community compositions  $(F=11.48, p=0.001)$ , with only axis 1 ( $F = 22.26$ ,  $p = 0.001$ ) showing the signifcant distribution of groups. On the axis 1, there was a separation between open water areas and sites with macrophytes, where colonial phytoplankton biomass, positioned to the right of the axis, positively infuenced herbivores, flter-feeders and flter-scrapers in foating and submerged macrophyte sites, while fagellate phytoplankton were positively related to sites without macrophytes (Fig. [6b](#page-8-0)).

# **Discussion**

In the present study, we showed the nutrient-phytoplankton and phytoplankton–zooplankton interactions in areas with and without macrophytes. The unicellular, colonial,



<span id="page-7-0"></span>**Fig. 5** Generalized additive models (GAM) showing the relationships between nutrients–phytoplankton (**a**– **f**) and phytoplankton– zooplankton (**g**, **h**, **i**) in open water areas (orange line) and areas with foating (blue line) and submerged (green line) macrophytes. The

solid line represents the ftted values of the general model. DIN: dissolved inorganic nitrogen, PO<sub>4</sub>: orthophosphate, TPhytoplankton: total phytoplankton biomass, TZooplankton: total zooplankton biomass

filamentous, small and partly the large functional phytoplankton groups presented higher values of biomass in foating and submerged macrophyte banks. For zooplankton, only cladocerans (flters and scrapers) presented high biomass in foating and submerged macrophyte banks. Phytoplankton was positively correlated with orthophosphate in sites with submerged macrophytes, and negatively correlated with dissolved inorganic nitrogen in sites with foating macrophytes, while no signifcant relationship was found between zooplankton and phytoplankton, partially confrming our frst hypothesis.

Our results showed that foating and submerged macrophytes promoted changes in the phytoplankton community. Macrophytes are responsible for providing heterogeneous ecological niches that support diverse aquatic communities (Barrow et al. [2019;](#page-9-2) Stephan et al. [2019](#page-12-7)). For phytoplankton, macrophytes act as nutrient sources because, through the decomposition of plant tissues, nutrients become bioavailable in water (Wang et al. [2018](#page-12-2)), in addition to serving as substrate for algal colonization due to structural complexity (Nascimento-Filho et al. [2021](#page-11-28)). Algarte et al. [\(2017\)](#page-9-8) showed that both the species richness



<span id="page-8-0"></span>**Fig. 6** Redundancy Analysis (RDA) of (**a**) phytoplankton morphofunctional groups and (**b**) zooplankton functional feeding groups and trophic guilds in relation to variables in open water areas and areas with foating and submerged macrophytes. TP: total phosphorus, Pi: inorganic phosphate,  $NO<sub>2</sub>$ : nitrite,  $NO<sub>3</sub>$ : nitrate

of small-motile algae and the richness of large species were positively related to the presence of macrophytes, so the presence of macrophyte banks may have facilitated the coexistence of diferent species of algae.

For zooplankton, macrophytes serve as a refuge against predators (Figueiredo et al. [2018\)](#page-10-28) and a place for colonization and egg laying (Battauz et al. [2017\)](#page-9-9), and they also increase food availability (Rossa and Bonecker [2003](#page-11-29); Brito et al. [2020](#page-10-29)). In this way, diferent macrophyte species may harbor diferent zooplankton species (Zeng et al. [2017\)](#page-12-8), and this can have different effects on phytoplankton. In the present study, although foating macrophytes harbored higher biomass of herbivorous flter-feeders and scrapers zooplankton (cladocerans), no negative efect was found on phytoplankton biomass (Fig. [5h](#page-7-0)). Differently, the negative efect of zooplankton flter-feeders on phytoplankton was evidenced in other studies (Kozak et al. [2015;](#page-10-30) Gerasimova et al. [2018](#page-10-31)), and this can be explained by their generalist habit, predominant in several species represented by cladocerans.

Our second hypothesis was partially confrmed, as the RDA analysis showed that unicellular and fagellated phytoplankton were favored by nitrate and inorganic phosphate in open water areas, and nitrite and total phosphorus positively infuenced the colonial, flamentous, small, medium and large functional phytoplankton groups in sites with macrophytes. According to Zhang et al. [\(2020](#page-12-9)), some macrophyte species can provide suitable substrates for flamentous algal growth. Similar results were observed by Takamura et al. ([2003\)](#page-12-10), who showed that phytoplankton species responded diferently to the presence of macrophytes, with colonial and flamentous species of medium to large size positively related to lakes with the presence of macrophytes, and fagellated and unicellular species related to both the presence and absence of macrophytes. Our results showed that this relationship can be explained by the efect of nutrients (Fig. [6](#page-8-0)a). Studies performed in semi-arid regions of Brazil showed that submerged macrophytes play an important role in providing nutrients, especially during the dry season and rewetting, common in these regions due to the reduction in water level (Keitel et al. [2016](#page-10-12); Barbosa et al. [2020\)](#page-9-10). Thus, the nutrients available in areas covered by submerged macrophytes have a positive efect on phytoplankton, favoring flamentous, unicellular or colonial species (Monteiro et al. [2021](#page-11-30)).

During the growth period, macrophytes can accumulate nutrients from water or sediment, and when they decompose, the absorbed phosphorus is returned to the aquatic ecosystem (Wang et al. [2018](#page-12-2)). The algae attached to these macrophytes absorb 3.4–8.9% phosphorus released by macrophytes (Carignan and Kalff [1982\)](#page-10-32). High phosphorus concentrations favor the development of phytoplankton, especially cyanobacteria (Simić et al. [2017\)](#page-11-31), and non-diazotrophic cyanobacteria due to their inability to fx atmospheric nitrogen. Reservoirs in Northeastern Brazil are susceptible to cyanobacterial blooms (Moura et al. [2018](#page-11-32)), and eutrophication represents a factor that contributes to the success of these organisms (Amorim and Moura [2021](#page-9-11); Macêdo et al. [2021](#page-11-33)).

In contrast, although no significant difference was observed in the biomass of flagellated species between sampling sites, a positive relationship between fagellated algae and nutrients was observed in the absence of macrophytes (Fig. [6a](#page-8-0)). This suggests that fagellated algae, predominantly represented by *C. furcoides* in the present study, use resources more efficiently in areas without macrophytes. According to Crossetti et al. [\(2019](#page-10-33)), reduction in the relative stability of the water column caused by the absence of macrophytes and the increase in water transparency are decisive for the success of *C. furcoides*. This supports our results, as the fagellated group was related to nitrogen in sites without macrophytes, as shown in the RDA (Fig. [6](#page-8-0)a).

Colonial phytoplankton was positively related to three functional groups of zooplankton in sites with macrophytes. This result accord with Amorim et al. ([2020\)](#page-9-12), which showed a positive correlation between cyanobacteria and zooplankton, such as rotifers and copepod nauplii. This relationship can be explained by ability of zooplankton to coexist with cyanobacteria in eutrophic waters (Amorim et al. [2020\)](#page-9-12), in addition to a poor ability to avoid the ingestion of cyanobacteria (Lürling [2003,](#page-11-34) [2021;](#page-11-12) Colina et al. [2016\)](#page-10-9), leading to the exploitation of alternative food sources, such as other algae (Ger et al. [2016](#page-10-34)). In contrast, a negative relationship was found between fagellated phytoplankton and zooplankton. Herbivorous crustaceans, such as cladocerans and calanoid copepods found in our study, have a feeding preference for medium to large-sized fagellated organisms (Colina et al. [2016](#page-10-9); Titocci et al. [2022](#page-12-11)). Any evidence was found of zooplankton grazing *C. furcoides* (Colina et al. [2016](#page-10-9)).

## **Conclusion**

This study showed that the macrophytes promoted signifcant changes in plankton communities, favoring the growth of small and large-sized algae and unicellular, colonial and flamentous species, in addition to increasing the biomass of flter-feeder and flter-scraper zooplankton. Floating and submerged macrophytes favored nutrient-phytoplankton interactions, while phytoplankton–zooplankton interactions occurred both in the presence of submerged macrophytes and in open water sites, partially confrming our frst hypothesis. Our second hypothesis was partially confrmed too, as colonial, flamentous, small, medium and large phytoplankton were favored by the presence of macrophytes, and fagellated and colonial phytoplankton, represented by cryptophyceans, *C. furcoides* and colonial cyanobacteria, respectively, signifcantly infuenced zooplankton in sites with macrophytes. Our results highlight the importance of evaluating the efects of foating and submerged macrophytes on the relationship between phytoplankton and zooplanktonic communities in tropical reservoirs.

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**Data availability** The datasets generated during and/or analyzed during the current study are available from the corresponding author (ariadne\_moura@hotmail.com) on reasonable request.

### **Declarations**

**Conflicts of interest** The authors declare that they have no confict of interest.

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