### **REVIEW ARTICLE**

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# Mechanisms of immune-deposit formation and the mediation of immune renal injury

Received: January 31, 2005 / Accepted: March 17, 2005

Abstract The passive trapping of preformed immune complexes is responsible for some forms of glomerulonephritis that are associated with mesangial or subendothelial deposits. The biochemical characteristics of circulating antigens play important roles in determining the biologic activity of immune complexes in these cases. Examples of circulating immune complex diseases include the classic acute and chronic serum sickness models in rabbits, and human lupus nephritis. Immune deposits also form "in situ". In situ immune deposit formation may occur at subepithelial, subendothelial, and mesangial sites. In situ immune-complex formation has been most frequently studied in the Heymann nephritis models of membranous nephropathy subepithelial with immune deposits. While the autoantigenic target in Heymann nephritis has been identified as megalin, the pathogenic antigenic target in human membranous nephropathy had been unknown until the recent identification of neutral endopeptidase as one target. It is likely that there is no universal antigen in human membranous nephropathy. Immune complexes can damage glomerular structures by attracting circulating inflammatory cells or activating resident glomerular cells to release vasoactive substances, cytokines, and activators of coagulation. However, the principal mediator of immune complexmediated glomerular injury is the complement system, especially C5b-9 membrane attack complex formation. C5b-9 inserts in sublytic quantities into the membranes of glomerular cells, where it produces cell activation, converting normal cells into resident inflammatory effector cells that cause injury. Excessive activation of the complement system is normally prevented by a series of circulating and cellcomplement regulatory proteins. bound Genetic deficiencies or mutations of these proteins can lead to the

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W.G. Couser Division of Nephrology, University of Washington, Seattle, WA, USA spontaneous development of glomerular disease. The identification of specific antigens in human disease may lead to the development of fundamental therapies. Particularly promising future therapeutic approaches include selective immunosuppression and interference in complement activation and C5b-9-mediated cell injury.

**Key words** Complement · Glomerulonephritis · Membranous nephropathy · C5b-9 · Complement inhibitor · Hemolytic uremic syndrome

### Introduction

In most forms of glomerulonephritis, injury occurs secondary to the formation of immune deposits, containing immunoglobulins, complement, and other proteins, at various intraglomerular locations. The type of injury that results is dependent on four factors: the mechanism of the deposit formation, the site of deposit formation, the composition of the deposits and the amount of the deposits. It must be understood that an immune complex reflects a dynamic interaction between antigen and antibody, rather than a permanent physical structure or unit. The composition of the immune complex can change, either in the circulation or within tissue. The purpose of this review is to summarize mechanisms of immune-deposit formation and their consequences in immune renal injury.

#### **Circulating immune-complex trapping**

The passive trapping of preformed immune complexes was thought to be responsible for many forms of human glomerulonephritis, because the kidney is a particularly susceptible target for immune deposit formation. Immune complexes in the circulation are delivered at a high rate because the kidney receives 25% of the cardiac output. Intraglomerular pressure is higher than in other capillary

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**Fig. 1.** Circulating immune formation. Circulating immune complexes deposit primarily in the mesangium or in the subendothelial space. The clinical and morphologic consequences of immune-complex deposition depend on the location and biochemical characteristics of the deposits. *GEn*, glomerular endothelial cell; *GEpi*, glomerular epithelial cell; *MC*, mesangial cell; *Ag*, antigen; *Ab*, antibody

beds, and more protein than usual may be forced across the glomerular capillary wall. The glomerular capillaries provide a large and highly permeable surface through which circulating immune complexes circulate. The capillary walls are also highly negatively charged structures that facilitate the localization of positively charged macromolecules. All of these properties facilitate the nonspecific trapping of circulating immune complexes in glomeruli.

Circulating-complex trapping occurs primarily in the mesangium and/or on the subendothelial, surface of the capillary wall (Fig. 1). Circulating preformed complexes are too large to cross the capillary wall intact to localize in the subepithelial space, although, in some circumstances, subendothelial immune complexes may dissociate to cross the glomerular basement membrane (GBM) and reform in situ in a subepithelial distribution.

The biochemical characteristics of the circulating antigen play important roles in determining the biologic activity of immune complexes. The size and blood clearance kinetics influence the properties of the macromolecular immune complexes.<sup>1</sup> When an antigen bears few epitopes (oligovalent) or when there is excess antigen, immune complexes tend to be small and soluble. Small immune complexes are inefficient at complement activation and attachment to Fc receptors; therefore, they tend to be relatively inert. As the antigen-to-antibody ratio nears equivalence, cross-linking is maximized and large insoluble immune complexes are formed. Large complexes have a high affinity for binding to Fc receptors and activate complement, which facilitates rapid removal from the circulation by phagocytosis. Medium-sized complexes that are less well-cleared but can activate complement and bind to Fc receptors tend to deposit in tissues and initiate glomerular injury.

The net charge and ability to bind to glomerular structures are important as well. Immune complexes containing cationic proteins tend to form glomerular deposits.<sup>2,3</sup> This is explained by the charge interactions between such soluble molecules and anionic sites in the basement membrane.<sup>4,5</sup> Furthermore, immune complexes containing cationic antigens exhibit a higher potential to activate complement compared to those of native or anionic antigens.<sup>6,7</sup> This property of cationic immune complexes may enhance their capacity to induce inflammatory reactions.

The best example of circulating immune complex disease is the classic acute serum sickness model in rabbits, induced by a single, large bolus injection of the foreign serum protein, bovine serum albumin (BSA). Binding of antibody to BSA results in circulating immune complexes, which accumulate in the glomerulus and other vessels and initiate acute glomerulonephritis and vasculitis. It is unclear whether there is an equivalent of acute serum sickness involving the kidney in humans, although post-infectious nephritis is felt by some to involve an analogous mechanism. Systemic lupus erythematosus (SLE) is an excellent example of an immune-complex disease in which the antigen is endogenous.<sup>8,9</sup> Circulating immune complexes are readily detected in patients with SLE, and contain a variety of specificities, including antibodies to double-stranded (ds) DNA, ribonucleoproteins, and others. Levels of circulating immune complexes as well as those of anti-ds DNA antibodies correlate with disease activity,<sup>10</sup> and anti-DNA and DNA immune complexes were found to be enriched in kidney eluates from lupus patients.<sup>11</sup> Thus, it has been speculated that immune deposits in the glomeruli in SLE reflect preformed immune-complex trapping. However, experimental evidence suggests the involvement of other pathogenic mechanisms in lupus nephritis as well. Immune complexes may also be formed in situ via charge interactions between anti-DNA/histone antibodies and DNA/ histones already deposited in the glomerulus,<sup>12-15</sup> and some lupus autoantibodies also bind directly to intrinsic glomerular antigens, leading to in situ immune-complex formation, as described below.

Despite the extensive literature on immune-complex disease, written over many decades, there is little direct evidence that the passive trapping of preformed immune complexes formed in the circulation leads to significant tissue injury at sites where the complexes become localized, including the glomerulus.<sup>16</sup>

#### In situ immune-complex formation

Immune deposits may form "in situ" when an antibody binds specifically either to antigens that are constituents of normal glomerular structures or to soluble antigens that become localized independently in the glomerulus, on the basis of nonspecific uptake by the mesangium or charge interactions with anionic sites in the glomerular capillary wall.

In the early 1980s, there was considerable debate regarding the relative importance of circulating immune complexes versus the in situ formation of immune deposits. The Heymann nephritis model of membranous nephropathy, which is described in more detail below, served an important role in clarifying this question. Initially it had been believed that immune-complex deposition in glomeruli was due to the glomerular trapping of circulating immune complexes formed by circulating tubular antigens and the corresponding antibodies. However, between 1978 and 1983, groups led by Hoedemaeker and Couser,17-21 almost simultaneously reported studies that showed subepithelial immune complex formation following the direct perfusion of bloodless kidneys with pathogenic IgG antibody, thus establishing that subepithelial immune deposits in this classic model of membranous nephropathy developed through a novel mechanism, referred to as in situ immune-complex formation.<sup>16</sup>

The location of immune deposits that form in situ is determined by the site of expression of the relevant intrinsic glomerular antigen or the site where a nonimmunologic biochemical interaction leads to the trapping, or "planting" of an antigen. As a consequence, in situ immune-deposit formation may occur at subepithelial, subendothelial, or mesangial sites (Fig. 2). The clinical and morphologic consequences of this process depend on where the deposits form and which glomerular cells are targeted. However, in situ immune-complex formation has been most extensively studied in the Heymann models of membranous nephropathy involving subepithelial immune deposits, as discussed in the next section.

#### Lessons about in situ immune-complex formation learned from membranous nephropathy

Subepithelial deposits are the characteristic features of membranous nephropathy, which is the most common cause of nephrotic syndrome in adults. Central to the pathogenesis of membranous nephropathy is the formation of immune complex deposits in the subepithelial space and the lamina rara externa of the GBM that cause a membranelike thickening of the capillary wall.

The antibody constituent of immune complexes in membranous nephropathy consists of IgG (often IgG4) and so far largely unidentified antigens. Elution of pathogenic antibody has been largely unsuccessful in human membranous nephropathy, due to many problems, including the limited



Fig. 2. In situ immune complex formation. In situ immune-deposit formation may occur at subepithelial, subendothelial, and mesangial sites. Immune complexes in mesangial or subendothelial sites lead to the recruitment of inflammatory cells, with subsequent inflammatory changes, i.e., nephritis. In contrast, circulating cells do not have access to subepithelial deposits, leading to resident glomerular cell injury mediated by noninflammatory mechanisms, i.e., nephropathy

availability of suitable tissue and the need to employ relatively harsh elution procedures. Hepatitis B, hepatitis C, and *Helicobacter pylori* antigens; tumor antigens; and thyroglobulin have all been detected in the subepithelial deposits,<sup>22-24</sup> but there is no proof that these antigens are pathogenic, and autoimmune mechanisms may also be involved.

Our understanding of the molecular and kinetic concepts of subepithelial immune complex formation derives mainly from studies of Heymann nephritis in the rat,<sup>25</sup> which faithfully reproduces human membranous nephropathy. Following the discovery that immune deposits in Heymann nephritis form by antibody IgG binding to a native glomerular antigen, the autoantigenic target was identified as the podocyte-membrane protein, now called megalin.<sup>26</sup> Megalin, which was previously called glycoprotein (gp) 330, is a member of the low-density lipoprotein (LDL)-receptor family and is expressed both on the proximal tubular brush border and the soles of podocyte foot processes where immune complexes are initially formed.<sup>19</sup>

In rats, megalin seems to be expressed exclusively in glomeruli and the tubular border; it is not expressed on human podocytes or detected in subepithelial immune deposits in patients with membranous nephropathy.<sup>27</sup> Therefore, studies of this model should be interpreted, as a more general principle, to suggest that podocyte-membrane proteins are likely to serve as targets for immune-complex formation in situ and this results formation in pathogenic cascades downstream essentially similar to those defined experimentally.

Although the quest to identify the pathogenic antigens in human membranous nephropathy has been notoriously difficult and unproductive, despite many attempts, Ronco's group<sup>28</sup> recently provided the first breakthrough by identifying the neutral endopeptidase (NEP, or metallomembrane endopeptidase; MME) as a pathogenic antigenic target in one unusual form of human membranous nephropathy. Determination of the pathogenic mechanisms of immunedeposit formation was based on the finding that a genetically NEP-deficient mother developed antibodies to this enzyme during a first pregnancy; these antibodies were then transplacentally transferred into the fetal circulation of a second infant, who was not deficient in this enzyme. These antibodies localized in the glomerulus by binding to the normally expressed NEP antigen on the podocyte surface, causing the in situ formation of subepithelial immune deposits and neonatal membranous nephropathy.<sup>28</sup> Recent studies, by the same group, of three families, each with a newborn with neonatal membranous nephropathy, identified truncating mutations of the MME gene (coding for NEP) as the cause of alloimmunization during pregnancy.<sup>29</sup> However, NEP apparently serves as a target only in a very small number of patients; namely, the offspring of those rare mothers who lack a functional gene for this enzyme. It is likely that there is no universal antigen for membranous nephropathy in the human disease, but that other podocyte antigens are probably involved.

## Mechanisms of immune-complex-mediated tissue injury

When immune complexes form *in situ* or passively deposit in glomeruli from the circulation, they can attract and activate infiltrating leukocytes or intrinsic glomerular cells to release many local mediators of inflammation, including complement, growth factors, vasoactive substances, cytokines, and activators of coagulation.

Receptors for the Fc region of immunoglobulins are expressed on many cells of the immune system and kidney. Fc receptors initiate a number of responses, including Fc receptor-mediated phagocytosis and antibody-dependent cell-mediated cytotoxicity. However, antibodies specific for a given antigen make up only a very small proportion of the total amount of immunoglobulin in plasma. Therefore, the Fc receptors distinguish antibody molecules bound to something from the vast majority of free antibody molecules. This condition is met by two changes that occur when antibodies bind to antigen: the aggregation of antibodies

and conformational changes in the Fc portion of the molecule. The cross-linking of Fc receptors by immunoglobulins in immune complexes initiates inflammation, which is amplified by cytokines and activated complement components.

The Fcy receptor, which binds the Fc part of the constant region of IgG, is one of the most important classes of immune complex-binding receptors. Two general types of Fcy receptor are recognized - the activator receptors, characterized by the presence of a cytoplasmic immunotyrosine activator motif responsible for cell signaling, and the inhibitor receptors, characterized by the presence of a cytoplasmic immunotyrosine inhibitory motif. In NZB/NZW F1 mice, a model of lupus nephritis, the animals lacking activator Fcy receptor on circulating leukocytes were protected from immune complex-induced glomerular injury.<sup>30</sup> Protection of the kidney by activator Fcy receptor deficiency was also observed in a model of anti-GBM disease.<sup>31,32</sup> Furthermore, mice lacking inhibitor Fcy receptor on a C57BL/6 background developed a spontaneous lupus-like disease with glomerular sclerosis.<sup>33</sup> Recent studies by Alpers' group (Muhlfeld et al.<sup>34</sup>) showed exacerbation of cryoglobulinassociated membranoproliferative glomerulonephritis in a mouse model without inhibitor Fcy receptor. Despite the presence of Fcy receptors on intrinsic renal cells, experimental studies show that glomerular damage is mediated entirely through Fcy receptors on circulating bone marrowderived cells.<sup>35,36</sup> All of these studies emphasize a crucial role for Fcy receptors in immune-complex diseases and potentially demonstrate that immunoglobulin and complement deposition could be uncoupled from inflammatory consequences in some models.

However, another group has reported the successful induction of crescentic nephrotoxic nephritis in antibodydeficient,  $\mu$ -chain-deficient mice.<sup>37</sup> It is, therefore, likely that both Fcy receptor-dependent and Fcy receptor-independent mechanisms involving T cells may be operative in different phases of various diseases.

#### Complement activation associated with immunecomplex formation

The principal mediator of immunoglobulin-induced glomerular injury is the complement system. When immune deposits form, local complement activation occurs, leading to subsequent tissue injury.

Subendothelial deposits are readily accessible to circulating inflammatory cells, such as neutrophils and platelets, and cause severe damage, with endothelial cell injury, the activation of pro-coagulant cascades, fibrin deposition, and exudative lesions, as seen in type I mesangioproliferative glomerulonephritis and post-infectious glomerulonephritis.

Mesangial deposits injure and activate resident mesangial cells, which then de-differentiate to express  $\alpha$ -smooth muscle actin; the cells then proliferate and overproduce growth factors, cytokines, and extracellular matrix, as seen in IgA nephropathy and other diseases.

Subepithelial deposits, as observed in membranous nephropathy, are characterized by complement-mediated podocyte injury and the lack of any apparent inflammatory reaction. This is because subepithelial immune complexes are separated from the circulation by the GBM, and the mediators released by activated podocytes are carried by filtration forces into the urine, rather than toward the circulation.

The complement system, which is an important mediator of inflammation and tissue injury, is a family of more than 20 serum and cell-surface proteins that function as a cascade. Immune complexes formed by IgG and nephritogenic antigens bind to complement factor C1q and activate the C1 complex, leading to the formation of C3 convertase and the enzymatic cleavage of the central complement component C3. Activation of C3, the most abundant of the serum complement proteins, results in the release of the chemotactic factor C3a and covalent attachment of the C3b fragment to host cells, which is an important step for amplification through the alternative pathway and for continued activation of the terminal membrane attack complex, C5b-9.

Local complement activation by the classical, lectin, or alternative pathways also generates chemotactic factors such as C3a and C5a. C3a and C5a function in a synergistic manner with Fc-receptor crosslinking to activate mast cells and stimulate inflammatory cells, thereby leading to tissue injury.

However, the most nephritogenic effect of complement activation appears to be the generation of the membrane attack complex, or C5b-9. C5b-9 inserts in sublytic quantities into the membranes of glomerular cells, where it produces cell activation, converting normal cells into resident effector cells to cause injury. In 1980, Salant and colleagues<sup>38</sup> were the first to show, using generalized complement depletion with cobra venom factor, that podocyte injury in the Heymann nephritis model was complement-dependent. Later studies showed that the same effect could be achieved with C6 depletion alone, thus implicating the C5b-9 membrane attack complex in this process.<sup>39-42</sup>

C5b-9 formation damages podocytes via various mechanisms. Sublethal C5b-9 attack in glomerular epithelial cells triggers metabolic changes and the induction of the de novo synthesis of prostaglandins and proteases.<sup>43,44</sup> Complement activation induces a massive increase in the biosynthesis of nicotinamide adenine dinucleotide phosphate, reduced (NADPH)-oxido-reductase in podocytes, which, in turn, causes local production of oxygen radicals.<sup>45,46</sup> These products facilitate the degradation of the GBM.

C5b-9 activates phospholipase A2 and induces phospholipid hydrolysis in podocytes, resulting in the impairment of endoplasmic reticulum (ER) membrane integrity and subsequent ER stress.<sup>47</sup> Sublytic C5b-9 attack induces DNA damage in podocytes both in vitro and in vivo.<sup>48</sup> Furthermore, complement activation leads to the reversible disruption of actin microfilaments in cultured podocytes, and the dissociation of nephrin from the actin cytoskeleton in experimental membranous nephropathy.<sup>49,50</sup> Thus, proteinuria in membranous nephropathy is a consequence of GBM damage caused by oxidants and proteases derived from podocytes, as well as podocyte cytoskeletal changes induced by C5b-9. For more details of the podocyte response to complement attack which leads to proteinuria, readers are referred to our recent review of membranous nephropathy.<sup>51</sup>

C5b-9 formation is crucial in the injury of other glomerular cell types as well. Complement attack activates mesangial cells and induces the production of various disease mediators.<sup>52,53</sup> Furthermore, complement attack directly stimulates mesangial cell proliferation.<sup>54</sup> Cell proliferation and mesangial expansion in experimental glomerulonephritis models resembling IgA nephropathy also reflect C5b-9-induced cell activation, with the overproduction of platelet-derived growth factor (PDGF) and transforming growth factor (TGF)- $\beta$ , which are representative mediators of mesangial injury.<sup>55,56</sup> In a mesangioproliferative glomerulonephritis model of anti-Thy1 nephritis in rats, C6 deficiency resulted in lack of initial mesangial damage and subsequent renal-disease phenotypes.<sup>57</sup>

The relevance of complement activation in human mesangioproliferative glomerulonephritis was demonstrated in IgA nephropathy. Zwirner and colleagues<sup>58</sup> showed that the presence of complement activation was associated with more severe renal disease and suggested that activated C3 could serve as a clinical predictor of the disease course. In IgA nephropathy, it is speculated that IgA activates the complement cascade via the lectin pathway.<sup>59-63</sup>

An essential role of C5b-9 formation in glomerular endothelial injury has also been demonstrated in a variety of animal models, including a thrombotic microangiopathy model induced by immune-complex formation on glomerular endothelial cells.<sup>64,65</sup> Endothelial injury in lupus occurs when C5b-9 causes the upregulation of leukocyte adhesion molecules, as well as cell lysis, detachment, and apoptosis.<sup>66</sup>

It should be noted that the balance between the harmful effects of complement activation and the beneficial effects of immune-complex processing and apoptotic debris disposal is important. While the complement system is considered to be a major mediator of tissue injury in SLE, a genetic deficiency of the early complement components is also associated with SLE. This is called the "lupus paradox",<sup>67</sup> because, on the one hand, complement is pathogenic but, on the other, it is protective. This is a good example to illustrate the necessity for a fine balance of the complement system in vivo. In the MRL/lpr mouse model of lupus nephritis, complement inhibition by soluble Crry,<sup>68,69</sup> or deficiency of factor B,<sup>70</sup> protected the animals from renal disease. In contrast, C3 -/- MRL/lpr mice were not protected from renal disease, and this lack of protection was associated with evidence of elevated immune-complex deposition in the glomerulus.<sup>71</sup>

# Complement regulatory proteins as a protective mechanism

Complement activation is normally closely regulated by a series of circulating and cell-bound complement regulatory proteins that protect cells from injury.<sup>72–74</sup>

Our group<sup>75,76</sup> demonstrated that the overexpression of complement regulatory proteins in cultured glomerular cells served to protect against complement attack. Furthermore, the blockade of complement regulatory proteins by neutralizing antibodies resulted in aggravation of various renal disease models.<sup>77–79</sup> Quigg and colleagues<sup>80,81</sup> showed that the neutralization of a complement regulatory protein by pathogenic antibodies, which bound to podocyte surfaces and formed immune deposits, was essential to induce proteinuria in a Heymann nephritis model in rats. A protective role for complement regulatory proteins in renal disease has also been demonstrated, utilizing a gene-targeting approach, by our group and others.<sup>82–86</sup>

One may also ask whether a lack of complement regulatory proteins might result in renal disease. The answer is yes. Recent molecular biological analysis has demonstrated that genetic abnormalities of factor H, a soluble complement regulatory protein at the hit point of C3b, are associated with atypical hemolytic uremic syndrome (HUS).<sup>87-89</sup> It is estimated that 15% to 30% of atypical HUS can be ascribed to factor H mutations.<sup>90,91</sup> Recent studies employing surface plasmon resonance analysis have revealed a reduction of the binding affinity of mutated factor H to C3, demonstrating the functional importance of the mutations.<sup>92</sup> Furthermore, another membrane-bound complement regulatory protein, which works at the same step as factor H, has also been associated with atypical HUS.93,94 All these findings suggest that inability to control the intravascular activation of the complement cascade can lead to endothelial injury, the formation of thrombus, and eventual HUS, emphasizing the substantial biological importance of complement regulatory proteins, especially in renal disease.

One question remains to be answered. While patients with heterozygous and some homozygous factor-H deficiencies develop atypical HUS, other homozygous factor H-deficient subjects suffer from membranoproliferative glomerulonephritis.<sup>95</sup> Both factor H-knockout mice and pigs with congenital deficiency of factor H develop membranoproliferative glomerulonephritis but not HUS.<sup>96,97</sup> This finding may be partly explained by the relative amounts of factor H, but it is still unknown why complete factor H deficiency in humans sometimes leads to atypical HUS, while, at other times, it results in membranoproliferative glomerulonephritis.

### **Future therapeutic approaches**

The identification of specific antigens in human disease can lead to the development of new and specific therapies. In the field of neurology, Campylobacter has recently been identified to be responsible for the development of Guillain Barre syndrome.<sup>98,99</sup> Identification of NEP as a target in membranous nephropathy is a break-through in nephrology. Recent studies by Yoshizawa and colleagues<sup>100,101</sup> have shown that nephritis-associated plasmin receptor is the likely pathogenic antigen in acute post-streptococcal glomerulonephritis. This is a rapidly advancing area and one of the most promising frontiers in immune-complex disease research, because it offers the possibility of selectively targeting disease-specific elements of the immune response, or restoring tolerance to self antigens, without inducing generalized immunosuppression.

Other essential approaches to be attempted include the design of ways to interfere with complement activation and C5b-9-mediated cell injury. The utilization of complement regulatory proteins in various ways has considerable potential as a therapeutic tool in both glomerular and interstitial disease.

Acknowledgments We thank Dr. Reiko Inagi (Tokai University School of Medicine, Kanagawa, Japan) for her helpful advice and support. Portions of the work reviewed here were supported by research grants from the Kato Memorial Bioscience Foundation (to Dr. Nangaku) and grants from the United States National Institutes of Health (DK 34198 and DK 74067; to Dr. Couser).

#### References

- Doekes G, van Es LA, Daha MR. Binding and activation of the first complement component by soluble immune complexes: effect of complex size and composition. Scand J Immunol 1984; 19:99–110.
- Gallo GR, Caulin-Glaser T, Lamm ME. Charge of circulating immune complexes as a factor in glomerular basement membrane localization in mice. J Clin Invest 1981;67:1305–13.
- Gauthier VJ, Mannik M, Striker GE. Effect of cationized antibodies in preformed immune complexes on deposition and persistence in renal glomeruli. J Exp Med 1982;156:766–77.
- Kanwar YS, Farquhar MG. Anionic sites in the glomerular basement membrane. In vivo and in vitro localization to the laminae rarae by cationic probes. J Cell Biol 1979;81:137–53.
- Rennke HG, Cotran RS, Venkatachalam MA. Role of molecular charge in glomerular permeability. Tracer studies with cationized ferritins. J Cell Biol 1975;67:638–46.
- Coimbra TM, Gouveia MA, Ebisui L, Barbosa JE, Lachat JJ, de Carvalho IF. Influence of antigen charge in the pathogenicity of immune complexes in rats. Br J Exp Pathol 1985;66:595–603.
- Michelin MA, Crott LS, Assis-Pandochi AI, Coimbra TM, Teixeira JE, Barbosa JE. Influence of the electric charge of the antigen and the immune complex (IC) lattice on the IC activation of human complement. Int J Exp Pathol 2002;83:105–10.
- Ponticelli C, Moroni G. Lupus nephritis. J Nephrol 2000;13:385– 99.
- Shlomchik MJ, Madaio MP. The role of antibodies and B cells in the pathogenesis of lupus nephritis. Springer Semin Immunopathol 2003;24:363–75.
- Izui S, McConahey PJ, Theofilopoulos AN, Dixon FJ. Association of circulating retroviral gp70-anti-gp70 immune complexes with murine systemic lupus erythematosus. J Exp Med 1979;149:1099– 116.
- Koffler D, Shur PG, Kunkel HG. Immunological studies concerning nephritis of systemic lupus erythematosus. J Exp Med 1967;126:607–24.
- Schmiedeke TM, Stockl FW, Weber R, Sugisaki Y, Batsford SR, Vogt A. Histones have high affinity for the glomerular basement membrane. Relevance for immune complex formation in lupus nephritis. J Exp Med 1989;169:1879–94.

- Morioka T, Woitas R, Fujigaki Y, Batsford SR, Vogt A. Histone mediates glomerular deposition of small size DNA anti-DNA complex. Kidney Int 1994;45:991–7.
- 14. Kramers C, Hylkema MN, van Bruggen MC, van de Lagemaat R, Dijkman HB, Assmann KJ, et al. Anti-nucleosome antibodies complexed to nucleosomal antigens show anti-DNA reactivity and bind to rat glomerular basement membrane in vivo. J Clin Invest 1994;94:568–77.
- van Bruggen MC, Kramers C, Walgreen B, Elema JD, Kallenberg CG, van den Born J, et al. Nucleosomes and histones are present in glomerular deposits in human lupus nephritis. Nephrol Dial Transplant 1997;12:57–66.
- Couser WG, Salant DJ. In situ immune complex formation and glomerular injury. Kidney Int 1980;17:1–13.
- Couser WG, Steinmuller DR, Stilmant NM, Salant DJ, Lowenstein LM. Experimental glomerulonephritis in the isolated perfused rat kidney. J Clin Invest 1978;62:1275–87.
- van Damme BJC, Fleuren GJ, Bakker WW, Vernier RL, Hoedemaeker Ph J. Experimental glomerulonephritis in the rat induced by antibodies directed against tubular antigens. V. Fixed glomerular antigens in the pathogenesis of heterologous immune complex glomerulonephritis. Lab invest 1978;38:502–10.
- Kerjaschki D, Farquhar MG. Immunochemical localization of the Heymann nephritis antigen (gp330) in glomerular epithelial cells of normal Lewis rats. J Exp Med 1983;157:667–86.
- Madaio MP, Salant DJ, Cohen AJ, Adler S, Couser WG. Comparative study of in situ immune deposit formation in active and passive Heymann nephritis. Kidney Int 1983;23:498–505.
- Neale TJ, Wilson CB. Glomerular antigens in Heymann's nephritis: reactivity of eluted and circulating antibody. J Immunol 1982; 128:323–30.
- Horl WH, Kerjaschki D. Membranous glomerulonephritis (MGN). J Nephrol 2000;113:291–316.
- Ronco PM. Paraneoplastic glomerulopathies: new insights into an old entity. Kidney Int 1999;56:355–77.
- 24. Debiec H, Guigonis V, Mougenot B, Haymann J-P, Bensman A, Deschenes G, et al. Antenatal membranous glomerulonephritis with vascular injury induced by anti-neutral endopeptidase antibodies: toward new concepts in the pathogenesis of glomerular diseases. J Am Soc Nephrol 2003;14:S27–32.
- 25. Heymann W, Lund HZ, Hackel DB. The nephrotic syndrome in the rats; with special reference to the progression of the glomerular lesion and to the use of nephrotoxic sera obtained from ducks. J Lab Clin Med 1952;39:218–24.
- Kerjaschki D, Farquhar MG. The pathogenic antigen of Heymann nephritis is a membrane glycoprotein of the renal proximal tubule brush border. Proc Natl Acad Sci USA 1982;79: 5557–61.
- Farquhar M, Saito A, Kerjaschki D, Orlando RA. The Heymann nephritis antigenic complex: megalin (gp330) and RAP. J Am Soc Nephrol 1995;6:35–47.
- Debiec H, Guigonis V, Mougenot B, Decobert F, Haymann JP, Bensman A, et al. Antenatal membranous glomerulonephritis due to anti-neutral endopeptidase antibodies. N Engl J Med 2002; 346:2053–60.
- Debiec H, Nauta J, Coulet F, van der Burg M, Guigonis V, Schurmans T, et al. Role of truncating mutations in *MME* gene in fetomaternal alloimmunisation and antenatal glomerulonephritis. Lancet 2004;364:1252–9.
- Clynes R, Dumitru C, Ravetch JV. Uncoupling of immune complex formation and kidney damage in autoimmune glomerulonephritis. Science 1998;279:1052–4.
- Park SY, Ueda S, Ohno H, Hamano Y, Tanaka M, Shiratori T, et al. Resistance of Fc receptor-deficient mice to fatal glomerulonephritis. J Clin Invest 1998;102:1229–38.
- Suzuki Y, Shirato I, Okumura K, Ravetch JV, Takai T, Tomino Y, et al. Distinct contribution of Fc receptors and angiotensin IIdependent pathways in anti-GBM glomerulonephritis. Kidney Int 1998;54:1166–74.
- Bolland S, Ravetch JV. Spontaneous autoimmune disease in Fc(gamma)RIIB-deficient mice results from strain-specific epistasis. Immunity 2000;13:277–85.
- 34. Muhlfeld AS, Segerer S, Hudkins K, Carling MD, Wen M, Farr AG, et al. Deletion of the Fc · receptor IIb in thymic stromal lymphopoietin transgenic mice aggravates membrano-

proliferative glomerulonephritis. Am J Pathol 2003;163:1127-36.

- Tarzi RM, Davies KA, Robson MG, Fossati-Jimack L, Saito T, Walport MJ, et al. Nephrotoxic nephritis is mediated by Fc receptors on circulating leukocytes and not intrinsic renal cells. Kidney Int 2002;62:2087–96.
- 36. Suzuki Y, Gomez-Guerrero J, Shirato I, Lopez-Franco O, Gallego-Delgado J, Sanjuan G, et al. Pre-existing glomerular immune complexes induce polymorphonuclear cell recruitment through an Fc receptor-dependent respiratory burst: potential role in the perpetuation of immune nephritis. J Immunol 2003; 170:3243–53.
- Li S, Holdsworth SR, Tipping PG. Antibody independent crescentic glomerulonephritis in mu chain deficient mice. Kidney Int 1997;51:672–8.
- Salant DJ, Belok S, Madaio MP, Couser WG. A new role for complement in experimental membranous nephropathy in rats. J Clin Invest 1980;66:1339–50.
- Perkinson DT, Baker PJ, Couser WG, Johnson RJ, Adler S. Membrane attack complex deposition in experimental glomerular injury. Am J Pathol 1985;120:121–8.
- Cybulsky AV, Quigg RJ, Salant DJ. The membrane attack complex in complement-mediated glomerular epithelial cell injury: formation and stability of C5b-9 and C5b-7 in rat membranous nephropathy. J Immunol 1986;137:1511–6.
- Baker PJ, Ochi RF, Schulze M, Johnson RJ, Campbell C, Couser WG. Depletion of C6 prevents development of proteinuria in experimental membranous nephropathy in rats. Am J Pathol 1989;135:185–94.
- Savin VJ, Johnson RJ, Couser WG. C5b-9 increases albumin permeability of isolated glomeruli in vitro. Kidney Int 1994; 46:382–7.
- Quigg RJ, Cybulsky AV, Jacobs JB, Salant DJ. Anti-Fx1A produces complement-dependent cytotoxicity of glomerular epithelial cells. Kidney Int 1988;34:43–52.
- 44. McMillan JI, Riordan JW, Couser WG, Pollock AS, Lovett DH. Characterization of a glomerular epithelial cell metalloproteinase as matrix metalloproteinase-9 with enhanced expression in a model of membranous nephropathy. J Clin Invest 1996;97:1094– 101.
- 45. Neale TJ, Ullrich R, Ojha P, Poczewski H, Verhoeven AJ, Kerjaschki D. Reactive oxygen species and neutrophil respiratory burst cytochrome b558 are produced by kidney glomerular cells in passive Heymann nephritis. Proc Natl Acad Sci USA 1993;90: 3645–9.
- 46. Shah SV. Evidence suggesting a role for hydroxyl radical in passive Heymann nephritis in rats. Am J Physiol Renal Fluid Electrolyte Physiol 1988;254:F337–44.
- 47. Cybulsky AV, Takano T, Papillon J, Khadir A, Liu J, Peng H. Complement C5b-9 membrane attack complex increases expression of endoplasmic reticulum stress proteins in glomerular epithelial cells. J Biol Chem 2002;277:41342–51.
- Pippin JW, Durvasula R, Petermann A, Hiromura K, Couser WG, Shankland SJ. DNA damage is a novel response to sublytic complement C5b-9-induced injury in podocytes. J Clin Invest 2003;111:877–85.
- Topham PS, Haydar SA, Kuphal R, Lightfoot JD, Salant DJ. Complement-mediated injury reversibly disrupts glomerular epithelial cell actin microfilaments and focal adhesions. Kidney Int 1999:55:1763–75.
- Yuan H, Takeuchi E, Taylor GA, McLaughlin M, Brown D, Salant DJ. Nephrin dissociates from actin, and its expression is reduced in early experimental membranous nephropathy. J Am Soc Nephrol 2002;13:946–56.
- 51. Nangaku M, Shankland SJ, Couser WG. Cellular response to injury in membranous nephropathy. J Am Soc Nephrol.
- Lovett DH, Haensch GM, Goppelt M, Resch K, Gemsa D. Activation of glomerular mesangial cells by the terminal membrane attack complex of complement. J Immunol 1987;138:2473– 80.
- 53. Schonermark M, Deppisch R, Riedasch G, Rother K, Haensch GM. Induction of mediator release from human glomerular mesangial cells by the terminal complement components C5b-9. Int Arch Allergy Appl Immunol 1991;96:331– 7.

- Couser WG, Pippin J, Shankland SJ. Complement (C5b-9) induces DNA synthesis in rat mesangial cells in vitro. Kidney Int 2001;59:905–12.
- 55. Iida H, Seifert R, Alpers CE, Gronwald RG, Phillips PE, Pritzl P, et al. Platelet-derived growth factor (PDGF) and PDGF receptor are induced in mesangial proliferative nephritis in the rat. Proc Natl Acad Sci USA 1991;88:6560–4.
- 56. Isaka Y, Fujiwara Y, Ueda N, Kaneda Y, Kamada T, Imai E. Glomerulosclerosis induced by in vivo transfection of transforming growth factor-beta or platelet-derived growth factor gene into the rat kidney. J Clin Invest 1993;92:2597–601.
- 57. Brandt J, Pippin J, Schulze M, Hansch GM, Alpers CE, Johnson RJ, et al. Role of the complement membrane attack complex (C5b-9) in mediating experimental mesangioproliferative glomerulonephritis. Kidney Int 1996;49:335–43.
- Zwirner J, Burg M, Schulze M, Brunkhorst R, Gotze O, Koch KM, et al. Activated complement C3: a potentially novel predictor of progressive IgA nephropathy. Kidney Int 1997;51:1257–64.
- Roos A, Bouwman LH, van Gijlswijk-Janssen DJ, Faber-Krol MC, Stahl GL, Daha MR. Human IgA activates the complement system via the mannan-binding lectin pathway. J Immunol 2001;167:2861–8.
- Hisano S, Matsushita M, Fujita T, Endo Y, Takebayashi S. Mesangial IgA2 deposits and lectin pathway-mediated complement activation in IgA glomerulonephritis. Am J Kidney Dis 2001;38:1082–8.
- Endo M, Ohi H, Satomura A, Hidaka M, Ohsawa I, Fujita T, et al. Regulation of in situ complement activation via the lectin pathway in patients with IgA nephropathy. Clin Nephrol 2001;55:185– 91.
- 62. Matsuda M, Shikata K, Wada J, Sugimoto H, Shikata Y, Kawasaki T, et al. Deposition of mannan binding protein and mannan binding protein-mediated complement activation in the glomeruli of patients with IgA nephropathy. Nephron 1998;80: 408–13.
- 63. Endo M, Ohi H, Ohsawa I, Fujita T, Matsushita M, Fujita T. Glomerular deposition of mannose-binding lectin (MBL) indicates a novel mechanism of complement activation in IgA nephropathy. Nephrol Dial Transplant 1998;13:1984–90.
- Nangaku M, Alpers CE, Pippin J, Shankland SJ, Adler S, Kurokawa K, et al. Renal microvascular injury induced by antibody to glomerular endothelial cells is mediated by C5b-9. Kidney Int 1997;52:1570–8.
- Hughes J, Nangaku M, Alpers CE, Shankland SJ, Couser WG, Johnson RJ. The C5b-9 membrane attack complex mediates endothelial cell apoptosis in experimental glomerulonephritis. Am J Physiol 2000;278:F747–57.
- 66. Belmont HM, Buyon J, Giorno R, Abramson S. Up-regulation of endothelial cell adhesion molecules characterizes disease activity in systemic lupus erythematosus. The Shwartzman phenomenon revisited. Arthritis Rheum 1994;37:376–83.
- Carroll MC. A protective role for innate immunity in systemic lupus erythematosus. Nat Rev Immunol 2004;4:825–31.
- Bao L, Haas. M, Boackle SA, Kraus DM, Cunningham PN, Alexander JJ, Anderson RK, et al. Transgenic expression of a soluble complement inhibitor protects against renal disease and promotes survival in MRL/lpr mice. J Immunol 2002;168:3601– 7.
- 69. Bao L, Haas M, Kraus DM, Hack BK, Rakstang JK, Holers VM, et al. Administration of a soluble recombinant complement C3 inhibitor protects against renal disease in MRL/lpr mice. J Am Soc Nephrol 2003;14:670–9.
- Watanabe H, Garnier G, Circolo A, Wetsel RA, Ruiz P, Holers VM, et al. Modulation of renal disease in MRL/lpr mice genetically deficient in the alternative complement pathway factor B. J Immunol 2000;164:786–94.
- Sekine H, Reilly CM, Molano ID, Garnier G, Circolo A, Ruiz P, et al. Complement component C3 is not required for full expression of immune complex glomerulonephritis in MRL/lpr mice. J Immunol 2001;166:6444–51.
- Nangaku M. Complement regulatory proteins in glomerulonephritis. Kidney Int 1998;54:1419–28.
- Nangaku M, Johnson RJ, Couser WG. Complement regulatory proteins and glomerulonephritis. Exp Nephrol 1997;5: 345–54.

- Nangaku M. Complement regulatory proteins: are they important in disease? J Am Soc Nephrol 2003;14:2411–3.
- Nangaku M, Meel RL, Pippin J, Gordon KL, Morgan BP, Johnson RJ, et al. Transfected CD59 protects mesangial cells from injury induced by antibody and complement. Kidney Int 1996;50:257–66.
- Nangaku M, Quigg RJ, Shankland SJ, Okada N, Johnson RJ, Couser WG. Overexpression of Crry protects mesangial cells from complement mediated injury. J Am Soc Nephrol 1997;8:223– 33.
- Nangaku M, Alpers CE, Pippin J, Shankland SJ, Kurokawa K, Adler S, et al. CD59 protects glomerular endothelial cells from immune-mediated thrombotic microangiopathy in rats. J Am Soc Nephrol 1998;9:590–7.
- Nishikage H, Baranyi L, Okada H, Okada N, Isobe K, Nomura A, et al. The role of a complement regulatory protein in rat mesangial glomerulonephritis. J Am Soc Nephrol 1995;6:234–41.
- Matsuo S, Nishikage H, Yoshida F, Nomura A, Piddlesden SJ, Morgan BP. Role of CD59 in experimental glomerulonephritis in rats. Kidney Int 1994;46:191–200.
- Schiller B, He C, Salant DJ, Lim A, Alexander JJ, Quigg RJ. Inhibition of complement regulation is key to the pathogenesis of active Heymann nephritis. J Exp Med 1998;188:1353–8.
- Cunningham PN, Hack BK, Ren G, Minto AW, Morgan BP, Quigg RJ. Glomerular complement regulation is overwhelmed in passive Heymann nephritis. Kidney Int 2001;60:900–9.
- Yamada K, Miwa T, Liu J, Nangaku M, Song WC. Critical protection from renal ischemia reperfusion injury by CD55 and CD59. J Immunol 2004;172:3869–75.
- Sogabe H, Nangaku M, Ishibashi Y, Wada T, Fujita T, Sun X, et al. Increased susceptibility of decay-accelerating factor (DAF) deficient mice to anti-GBM glomerulonephritis. J Immunol 2001;167:2791–7.
- Hanafusa N, Sogabe H, Yamada K, Wada T, Fujita T, Nangaku M. Contribution of genetically engineered animals to the analyses of complement in the pathogenesis of nephritis. Nephrol Dial Transplant 2002;17(Suppl 9):34–6.
- Lin F, Salant DJ, Meyerson H, Emancipator S, Morgan BP, Medof ME. Respective roles of decay-accelerating factor and CD59 in circumventing glomerular injury in acute nephrotoxic serum nephritis. J Immunol 2004;172:2636–42.
- Lin F, Emancipator SN, Salant DJ, Medof ME. Decayaccelerating factor confers protection against complementmediated podocyte injury in acute nephrotoxic nephritis. Lab Invest 2002;82:563–9.
- Warwicker P, Goodship TH, Donne RL, Pirson Y, Nicholls A, Ward RM, et al. Genetic studies into inherited and sporadic hemolytic uremic syndrome. Kidney Int 1998;53:836–44.
- Rougier N, Kazatchkine MD, Rougier JP, Fremeaux-Bacchi V, Blouin J, Deschenes G, et al. Human complement factor H deficiency associated with hemolytic uremic syndrome. J Am Soc Nephrol 1998;9:2318–26.
- 89. Noris M, Ruggenenti P, Perna A, Orisio S, Caprioli J, Skerka C, et al. Hypocomplementemia discloses genetic predisposition to hemolytic uremic syndrome and thrombotic thrombocytopenic purpura: role of factor H abnormalities. Italian Registry of Familial and Recurrent Hemolytic Uremic Syndrome/Thrombotic Thrombocytopenic Purpura. J Am Soc Nephrol 1999;10:281–93.
- 90. Caprioli J, Castelletti F, Bucchioni S, Bettinaglio P, Bresin E, Pianetti G, et al. International Registry of Recurrent and Familial HUS/TTP. Complement factor H mutations and gene polymorphisms in haemolytic uraemic syndrome: the C-257T, the A2089G and the G2881T polymorphisms are strongly associated with the disease. Hum Mol Genet 2003;12:3385–95.
- Richards A, Buddles MR, Donne RL, Kaplan BS, Kirk E, Venning MC, et al. Factor H mutations in hemolytic uremic syndrome cluster in exons 18–20, a domain important for host cell recognition. Am J Hum Genet 2001;68:485–90.
- 92. Manuelian T, Hellwage J, Meri S, Caprioli J, Noris M, Heinen S, et al. Mutations in factor H reduce binding affinity to C3b and heparin and surface attachment to endothelial cells in hemolytic uremic syndrome. J Clin Invest 2003;111:1181–90.
- Richards A, Kemp EJ, Liszewski MK, Goodship JA, Lampe AK, Decorte R, et al. Mutations in human complement regulator, membrane cofactor protein (CD46), predispose to development

of familial hemolytic uremic syndrome. Proc Natl Acad Sci USA 2003;100:12966–71.

- 94. Noris M, Brioschi S, Caprioli J, Todeschini M, Bresin E, Porrati F, et al. International Registry of Recurrent and Familial HUS/TTP. Familial haemolytic uraemic syndrome and an MCP mutation. Lancet 2003;362:1542–7.
- 95. Dragon-Durey MA, Fremeaux-Bacchi V, Loirat C, Blouin J, Niaudet P, Deschenes G, et al. Heterozygous and homozygous factor H deficiencies associated with hemolytic uremic syndrome or membranoproliferative glomerulonephritis: report and genetic analysis of 16 cases. J Am Soc Nephrol 2004;15:787–95.
- Pickering MC, Cook HT, Warren J, Bygrave AE, Moss J, Walport MJ, et al. Uncontrolled C3 activation causes membranoproliferative glomerulonephritis in mice deficient in complement factor H. Nat Genet 2002;31:424–8.
- Hogasen K, Jansen JH, Mollnes TE, Hovdenes J, Harboe M. Hereditary porcine membranoproliferative glomerulonephritis type II is caused by factor H deficiency. J Clin Invest 1995;95: 1054–61.

- 98. Yuki N, Suzuki K, Koga M, Nishimoto Y, Odaka M, Hirata K, et al. Carbohydrate mimicry between human ganglioside GM1 and *Campylobacter jejuni* lipooligosaccharide causes Guillain-Barre syndrome. Proc Natl Acad Sci USA 2004;101:11404–9.
- 99. Godschalk PC, Heikema AP, Gilbert M, Komagamine T, Ang CW, Glerum J, et al. The crucial role of *Campylobacter jejuni* genes in anti-ganglioside antibody induction in Guillain-Barre syndrome. J Clin Invest 2004;114:1659–65.
- 100. Yoshizawa N, Yamakami K, Fujino M, Oda T, Tamura K, Matsumoto K, et al. Nephritis-associated plasmin receptor and acute poststreptococcal glomerulonephritis: characterization of the antigen and associated immune response. J Am Soc Nephrol 2004;15:1785–93.
- Oda T, Yamakami K, Omasu F, Suzuki S, Miura S, Sugisaki T, et al. Glomerular plasmin-like activity in relation to nephritisassociated plasmin receptor in acute poststreptococcal glomerulonephritis. J Am Soc Nephrol 2005;16:247–54.