ORIGINAL ARTICLE

Impact of *MIR31HG* **polymorphisms on risk of breast cancer in Chinese women**

Ying Wei1,2 · Xiaolin Wang3 · Zhe Zhang3 · Changtao Zhao2 · Yuwei Chang2 · Zhiqing Bian2 · Xinhan Zhao1

Received: 16 June 2022 / Accepted: 1 March 2023 / Published online: 8 March 2023 © The Author(s) under exclusive licence to Japan Society of Clinical Oncology 2023

Abstract

Background Breast cancer (BC) is one of the leading causes of death worldwide. This study explored the relationship between the *MIR31HG* gene polymorphisms and the risk of BC in Chinese women.

Methods Eight single nucleotide polymorphisms (SNPs) in *MIR31HG* were genotyped among 545 patients with BC and 530 healthy controls using Agena MassARRAY analysis. The PLINK software was used to calculate the odds ratio (OR) and 95% confdence intervals (CIs) via the logistic regression analysis. Multi-factor dimensionality reduction (MDR) analysis was performed to study the impact of SNP-SNP interaction on BC risk.

Results *MIR31HG* rs72703442-AA (OR 0.29, 95% CI 0.10–0.79, *p*=0.026), rs55683539-TT (OR 0.46, 95% CI 0.26–0.80, *p*=0.012) and rs2181559-AA (OR 0.59, 95% CI 0.40–0.89, *p*=0.038) were associated with a reduced risk of BC in Chinese women, as well as stratified results at age ≥52 years. Rs79988146 was correlated with estrogen receptor (ER) and progesterone receptor (PR)in Chinese female BC patients under various genetic models. Age at menarche stratifcation indicated that rs1332184 was associated with increased risk in BC patients, whereas stratifcation by number of births indicated that rs10965064 was associated with reduced risk in BC patients. MDR analysis showed that the best single-locus model for predicting of BC risk are rs55683539, which, rs55683539-CC group was a high risk group and rs55683539-TT group was a low risk group.

Conclusions The results indicated that the *MIR31HG* polymorphisms were associated with a reduced risk of BC in Chinese women.

Keywords *MIR31HG* · Polymorphism · Breast cancer · Risk · Case–control study

Abbreviations

Joint frst authors: Ying Wei and Xiaolin Wang.

 \boxtimes Xinhan Zhao zhaoxinhan@mail.xjtu.edu.cn

- ¹ Department of Internal Medicine Oncology, The First Afliated Hospital of Xi'an Jiaotong University School of Medicine, Xi'an, #227 West Yanta Road, 710061, Shaanxi, China
- ² Department of Internal Medicine Oncology, Yulin No.2 Hospital, Yulin 719000, Shaanxi, China
- Department of General Surgery, Yulin No.2 Hospital, Yulin 719000, Shaanxi, China

Introduction

Breast cancer (BC), which has to be highlighted in China as the same as many other countries, is becoming the heaviest disease burden and the main leading cause of cancer-related death in women [[1–](#page-15-0)[6](#page-15-1)]. In 2015, estimated 272,400 new cases of BC were diagnosed, and estimated 70,700 deaths were expected to occur with incidence increasing year by year in China [\[4](#page-15-2)]. The fact that BC, as a complex disease to tackle, is caused by a combination of various factors such as, age, lifestyle, family history, and serum hormone level [[4,](#page-15-2) [5](#page-15-3), [7](#page-15-4), [8](#page-15-5)]. Among these factors

mentioned above, hereditary factor is considered to be the most important and crucial one, because 5–10% of cases are raised from genetic variation of susceptible genes [[9](#page-15-6)]. Genome-wide association studies (GWAS) have reported signifcant efects of some gene polymorphisms on BC risk [[10](#page-15-7)–[14](#page-15-8)], but many new associations between single nucleotide polymorphisms (SNPs) and BC risk have not been explored.

Long non-coding RNAs (lncRNAs) are non-coding RNAs longer than 200 nucleotides. Recently, several studies have shown a link between genetic variants in lncRNA genes and breast cancer risk. Cui et al. found a SNP 2 kb upstream of H19 transcription start site that was associated with breast cancer risk in estrogen receptor (ER) positive patients in the Chinese population [[15\]](#page-15-9). Wu et al. studied risk associations among 22,977 cases and 105,974 controls of European ancestry and found several novel risk-loci that harbored lncRNA genes [[16](#page-15-10)]. *MIR31HG*, identified as LNCHIFCAR/LOC554202/hsa-lnc-31, is located on chromosome 9 and produces a long non-coding RNA (lncRNA) which acts as a host gene for *MIR-31*. lncRNA with a lack of protein-coding function defned as RNA longer than 200 nucleotides, plays considerable and remarkable role in complex biological activities, including regulating gene expression through chromatin remodeling, controlling gene transcription, participating in post-transcriptional mRNA process, and mediating protein function or localization [\[17\]](#page-15-11). Their dysregulation seems to be contributed to the growth and progression of human tumors [[18](#page-15-12)], and therefore *MIR31HG* is widely reported to be involved in the development of various cancers, such as colorectal cancer [[19](#page-15-13)], bladder cancer [[20\]](#page-15-14), oral cancer [[21](#page-15-15)], lung adenocarcinoma [[17\]](#page-15-11), pancreatic carcinoma [[22](#page-15-16)], esophageal squamous cell carcinoma [[23](#page-15-17)] and BC [\[6,](#page-15-1) [24,](#page-15-18) [25](#page-15-19)]. Studies have shown that as a non-coding oncogene, the down-regulated expression of *MIR31HG* can lead to diminished cell proliferation, migration, invasion and increased apoptosis in BC [[25](#page-15-19)]. Through gene evaluated, a related study also explained that *MIR31HG* was regulated by promoter hypermethylation in triple-negative BC and participated in the regulatory mechanism of BC as an important determination of the invasion metastasis cascade [[24\]](#page-15-18). These studies have shown that *MIR31HG* has an important role in BC, however, there is no report on the relationship between the polymorphisms of *MIR31HG* and the risk of BC.

This study aimed to reveal the impact of *MIR31HG* gene polymorphisms on the risk of BC in Chinese women through a case–control study, and to explore the association of *MIR31HG* polymorphisms with clinical characteristics of BC patients, which may provide a theoretical and experimental basis for further investigating the role of *MIR31HG* on BC carcinogenesis.

Materials and methods

Study population

The study was approved by the ethics committee of Shaanxi Provincial Cancer Hospital, and informed consents were delivered and signed by all participants. A total of 545 patients with BC (mean age 52.00 ± 9.89 years) were recruited and analyzed for the study in 2017 and 2018. The histopathological diagnosis was followed the classifcation of breast tumors by the World Health Organization (WHO5th), and clinical staging was based on American Joint Committeeon Cancer (AJCC) on breast cancer TNM staging system. It is necessary to exclude some patients who had family history of cancer, received radiotherapy, chemotherapy, or other treatments before the period of investigation. BC cases were categorized by estrogen receptor (ER), progesterone receptor (PR), lymph node (LN) metastasis, clinical stage, human epidermal growth factor receptor 2 (HER2), Ki67, tumor size, tumor location and distant metastasis, among which ER and PR test results $>1\%$ were defined as ER-positive and PR-positive, and HER2 3+ and 2+ with fuorescence in situ hybridization (FISH) positive indicated HER2-positive. The Ki67 positive rate 20% was used as the cutoff point to divide patients with BC into low $\left(< 20\% \right)$ and high (>20%) groups. During this period, 530 cancer-free controls (mean age: 51.66 ± 9.67 years) were enrolled from the healthcare of the hospital at the same time. The excluded criteria of controls were as follows: (1) no gynecological neoplasm, (2) no other history of solid cancers, and (3) no immune disorders (Table [1](#page-2-0)). Approximately 3–5 mL of venous blood sample was collected from each participant and then was placed into anti-coagulative tubes stored at − 80 °C until use. Demographic and clinic indicators were recorded by self-administered standardized questionnaires and medical records, respectively.

Extraction of genomic DNA and genotyping

Studies have shown that *MIR31HG* plays an important role in BC, but the correlation between *MIR31HG* polymorphisms and BC risk has not been reported. The selection of the eight candidate SNPs on the *MIR31HG* gene in this study is based on haplotype data or genotype data [[26\]](#page-15-20) and from the 1,000 Genome Projects ([http://www.internatio](http://www.internationalgenome.org/) [nalgenome.org/](http://www.internationalgenome.org/)) to select SNPs with a minor alleles frequency (MAF) greater than 0.05 in the global population.

Following the GoldMag-Mini extraction method (GoldMag Co, Ltd, Xi'an, China) strictly, genomic DNA was extracted from the venous blood. DNA concentration

Table 1 Clinical characteristics in cases and controls

Characteristic	Case $(n = 545)$	Control $(n=530)$	\boldsymbol{p}	
Age mean $\pm SD$	52.00 ± 9.89	51.66 ± 9.67	0.453	
ER(%)				
Positive	371 (68.0%)	-		
Negative	166 (30.5%)			
Missing	8(1.5%)			
$PR(\%)$				
Positive	320 (58.7%)			
Negative	217 (39.8%)			
Missing	$8(1.5\%)$			
LN metastasis				
Yes	267 (49.0%)			
No	278 (51.0%)			
Clinical stage				
I/II	355 (65.1%)			
III/IV	156 (28.6%)			
HER2				
Positive	78 (14.3%)			
Negative	279 (51.2%)			
Missing	188 (34.5%)			
Ki67				
Low $(\leq\!20\%)$	147 (27.0%)			
High $(>20\%)$	363 (66.6%)			
Missing	35 (6.4%)			
Tumor location				
Right	258 (47.3%)			
Left	279 (51.2%)			
between	$8(1.5\%)$			
Tumor size				
\geq 2 cm	305 (56.0%)			
$<$ 2 cm	135 (24.8%)			
Missing	105 (19.3%)			
Number of births				
>1 time	246 (45.1%)			
\leq 1 time	231 (42.4%)			
Missing	68(12.5%)			
Age of menarche				
> 13 years old	332 (60.9%)			
\leq 13 years old	117(21.5%)			
Missing	96 (17.6%)			
Menopausal status				
Post-	280 (51.4%)			
Pre-	164 (30.1%)			
Missing	101 (18.5%)			

P value was calculated by test. *P* < 0.05 indicated a significant difference

SD standard deviation, *ER* estrogen receptor, *PR* progesterone receptor, *LN* lymph node, *HER2*, human epidermal growth factor receptor $\overline{2}$

was measured by spectrometry (DU530 UV/VIS spectrophotometer, Beckman Instruments, Fullerton, CA, USA). The Agena Biosicence Assay Design Suite V2.0 software ([http://agenacx.com/online-tools\)](http://agenacx.com/online-tools) was used to design the extended primer. The MassARRAY Nanodispenser (Agena Bioscience, San Diego, CA, USA) and the MassARRAY iPLEX platform (Agena Bioscience, San Diego, CA, USA) were used to genotype, and then Agena Bioscience TYPER software (version 4.0) was used to analyze the data [\[27\]](#page-15-21).

Statistical analyses

Statistical analysis was set up using Microsoft Excel, SPSS 18.0 statistical package (SPSS, Chicago, IL, USA) and the PLINK 1.07 software. Hardy–Weinberg equilibrium (HWE) for *MIR31HG* genotype distributions of controls were accessed using Fisher's exact test. The demographic and clinical characteristics of study participants were evaluated by chi-squared test to compare the diferences in genotypes and allele frequency distribution between the groups. Welch's T-test was used to compare ages between cases and controls. Logistic regression analysis was used to evaluate the genetic susceptibility of BC under fve genetic models (allele, codominant, recessive, dominant, and additive models). Odds ratios (ORs) and 95% confdence intervals (CIs) from a logistic regression model were performed to analyze the relative risk. All p values were two sided and $p < 0.05$ was considered to be statistically signifcant. Multi-factor dimensionality reduction (MDR) analysis was performed to assess the impact of SNPs interactions on BC risk [[28\]](#page-15-22). We used G*power 3.1.9.2 software to calculate the minimum sample size and actual power values required for this study [[29\]](#page-15-23).

Results

Demographics of study subjects and SNPs information

The minimum sample size of the case population and the control population calculated by G-power software is 107 and 103, respectively, and the actual power value is 0.95. Eight SNPs were genotyped in 545 patients with BC and 530 cancer-free controls. Demographics and clinical characteristics of all study subjects are displayed in Table [1,](#page-2-0) which showed that the sample size of the subject population recruited in this study is completely in line with statistical signifcance. The cases consisted of 371 (68.0%) ER positive tumors, 320 (58.7%) PR positive tumors, 267 (49.0%) LN metastasis positive tumors, 78 (14.3%) HER2 positive tumors, 355 (65.1%) I/II and 156 (28.6%) III/IV clinical stage, 147 (27.0%) low Ki67 status, 258 (47.3%)

right and 279 (51.2%) left in tumor locations. There is no signifcant diference between cases and controls in the distribution of ages $(p=0.453)$. Basic information including position, allele, role, and MAF of all eight *MIR31HG* SNPs between cases and controls is shown in Table [2.](#page-3-0) The genotype distributions of all SNPs were in accordance with HWE $(p > 0.05)$.

Association between MIR131HG polymorphisms and BC risk

In addition, fve multiple genetic models (allele, codominant, dominant, recessive, and additive models) were used to analyze the relationship between the candidate SNPs and the risk of BC in Chinese women. The results showed that under the polygenic model, the three candidate SNPs on the MIR131HG gene were signifcantly associated with BC risk ($p < 0.05$) (see Table [3](#page-4-0)). Specifically, rs72703442 was signifcantly associated with a lower risk of BC under both co-dominant (OR 0.29, 95% CI 0.10–0.79, *p*=0.026) and recessive (OR 0.30, 95% CI 0.11–0.82, *p*=0.011) models. Rs55683539 was signifcantly associated with a lower risk of BC in allelic (OR 0.76, 95% CI 0.62–0.93, *p*=0.007), codominant (OR 0.46, 95% CI 0.26–0.80, *p*=0.012), dominant (OR 0.78, 95% CI 0.61–0.99, *p*=0.040), recessive (OR 0.49, 95% CI 0.29–0.84, *p*=0.008), and additive (OR 0.76, 95% CI 0.62–0.93, *p*=0.007) models. Rs2181559 was signifcantly associated with a lower risk of BC in allelic (OR 0.82, 95% CI 0.69–0.98, *p*=0.028), codominant (OR 0.59, 95% CI 0.40–0.89, *p* =0.038), recessive (OR 0.62, 95% CI 0.43–0.91, $p = 0.014$), and additive (OR 0.82, 95% CI 0.68–0.98, $p = 0.026$) models.

Age‑stratifed analysis

Then, according to the average age of the recruited subjects, we conducted a stratifed analysis with 52 years as the age node to further explore the efect of the *MIR131HG* poly-morphisms on the risk of BC in Chinese women (Table [4](#page-6-0)). In women aged≥52 years, *MIR31HG* rs72703442 was signifcantly associated with reduced risk of BC under both codominant (OR 0.18, 95% CI 0.04–0.81, *p*=0.034) and recessive (OR 0.18, 95% CI 0.04–0.83, *p*=0.010) recessive models. Moreover, we detected rs55683539 in the allele (OR 0.68, 95% CI 0.51–0.90, *p*=0.007) codominant (OR 0.68, 95% CI 0.48–0.98, *p*=0.015), dominant (OR 0.64, 95% CI 0.45–0.91, *p*=0.012), and additive (OR 0.65, 95% CI 0.49–0.88, $p = 0.004$) model and found a significant lower risk result. Meanwhile, women with A allele (compared with those carrying the T allele) and AA genotype (compared with those carrying the TT and TA genotype) for rs2181559 had a reduced risk BC in the allele (OR 0.76, 95% CI 0.56–0.94, *p*=0.014) and co-dominant (OR 0.48, 95% CI 0.27–0.84, *p* =0.032), recessive (OR 0.54, 95% CI 0.32–0.92, $p = 0.021$), and additive (OR 0.72, 95% CI 0.55–0.93, $p = 0.012$) models. However, in women <52 years old, we did not fnd that candidate SNPs have an impact on the risk of BC in Chinese women $(p > 0.05)$.

Stratifed analysis of demographic and clinic indicators in case group

A stratifed analysis of ER, PR, HER2, age at menarche, number of births and menopausal status in the case group was further analyzed. Both ER and PR stratifed analyses indicated that rs79988146 was associated with ER positive and PR positive in BC patients under dominant (ER: OR 2.13, 95% CI 1.04–4.35, *p*=0.028; PR: OR 1.94, 95% CI 1.03–3.62, $p = 0.033$) models (Table [5\)](#page-8-0). Moreover, rs1332184 (allele: OR 0.56, 95% CI 0.35–0.89, *p*=0.014; and additive: OR 0.54, 95% CI 0.33–0.88, *p* = 0.009), rs72703442 (allele: OR 0.39, 95% CI 0.20–0.78, *p*=0.005; codominant: OR 0.29, 95% CI 0.13–0.64, *p*=0.003; dominant OR 0.32, 95% CI 0.15–0.67, *p* = 0.001; and additive: OR 0.37, 95% CI 0.18–0.74, *p*=0.002), rs55683539 (allele: OR 0.59, 95% CI 0.36–0.96, *p*=0.033; dominant OR 0.51, 95% CI 0.29–0.91, *p* = 0.019; and additive: OR 0.60, 95% CI 0.37–0.98, *p* = 0.032) and rs2181559

SNP single nucleotide polymorphism, *HWE* Hardy–Weinberg equilibrium, *A* minor allele, *B* major allele

Table 3 Genotypic model analysis of the relationship between *MIR31HG* SNPs and BC risk

Table 3 (continued)	SNP	Model	Genotype	Case, n $(\%)$	Control, n $(\%)$	OR (95% CI)	\boldsymbol{p}
			A	354 (32.5%)	392 (37.0%)	$0.82(0.69 - 0.98)$	0.028
		Codominant	TT	$212(40.0\%)$	241 (44.2%)	$\mathbf{1}$	0.038
			TA	244 (46.0%)	254 (46.6%)	$0.92(0.71 - 1.18)$	
			AA	74 (14.0%)	50 (9.2%)	$0.59(0.40-0.89)$	
		Dominant	TT	212 (40.0%)	241 (44.2%)	$\mathbf{1}$	0.160
			TA/AA	318 (60.0%)	304 (55.8%)	$0.84(0.66 - 1.07)$	
		Recessive	TT/TA	456 (86.0%)	495 (90.8%)	$\mathbf{1}$	0.014
			AA	74 (14.0%)	50 (9.2%)	$0.62(0.43 - 0.91)$	
		Additive	$\qquad \qquad -$			$0.82(0.68 - 0.98)$	0.026
	rs10965059	Allele	${\bf C}$	983 (90.8%)	903 (91.0%)	1	
			T	99 (9.2%)	89 (9.0%)	$1.02(0.76 - 1.38)$	0.888
		Codominant	CC	410 (82.7%)	450 (83.2%)	1	0.320
			CT	83 (16.7%)	83 (15.3%)	$0.91(0.65 - 1.27)$	
			TT	$3(0.6\%)$	$8(1.5\%)$	$2.43(0.64 - 9.22)$	
		Dominant	CC	410 (82.7%)	450 (83.2%)	1	0.820
			TT/TC	86 (17.3%)	$91(16.8\%)$	$0.96(0.70-1.33)$	
		Recessive	CC/TC	493 (99.4%)	533 (98.5%)	1	0.160
			TT	$3(0.6\%)$	$8(1.5\%)$	$2.47(0.65 - 9.35)$	
		Additive	-			$1.02(0.76 - 1.37)$	0.890
	rs10965064	Allele	$\mathsf C$	684 (62.7%)	660 (62.3%)	$\mathbf{1}$	
			G	406 (37.3%)	400 (37.7%)	$0.98(0.82 - 1.17)$	0.815
		Codominant	CC	206 (38.9%)	218 (40.0%)	1	0.910
			CG	248 (46.8%)	248 (45.5%)	$0.94(0.73 - 1.22)$	
			GG	76 (14.3%)	79 (14.5%)	$0.98(0.68 - 1.42)$	
		Dominant	CC	206 (38.9%)	218 (40.0%)	$\mathbf{1}$	0.700
			CG/GG	324 (61.1%)	327 (60.0%)	$0.95(0.75-1.22)$	
		Recessive	CC/CG	454 (85.7%)	466 (85.5%)	$\mathbf{1}$	0.940
			$\mathbf{G}\mathbf{G}$	76 (14.3%)	79 (14.5%)	$1.01(0.72 - 1.42)$	
		Additive	$\overline{}$	$\overline{}$		$0.98(0.82 - 1.17)$	0.820

P value was calculated by Wald Test adjusted by age

Bold indicated that $p < 0.05$ meant the data was statistically significant

SNP single nucleotide polymorphism, *OR* odds ratio, *95% CI* 95% confdence interval

(allele: OR 0.62, 95% CI 0.41–0.94, *p*=0.025; dominant: OR 0.58, 95% CI 0.35–0.97, *p* = 0.037; and additive: OR 0.64, 95% CI 0.43–0.97, *p* = 0.029) were associated with negative HER2 status. The stratification of patients' menarche age showed that rs1332184 had a signifcant positive correlation with menarche age under allelic (OR 1.50, 95% CI 1.08–2.09, *p* = 0.017), codominant (OR 2.58 95% CI 1.15–5.79, *p* = 0.048), dominant (OR 1.55, 95% CI 1.01–2.37, *p* = 0.044), recessive (OR 2.24, 95% CI 1.03–4.91, $p = 0.049$), and additive (OR 1.52, 95% CI 1.08–2.12, $p = 0.016$) models. In addition, stratification of patients' reproductive times showed that rs55683539 and rs10965064 were signifcantly negatively correlated with patients' age at menarche under both codominant (rs55683539: OR 0.32, 95% CI 0.11–0.93, *p* = 0.031) and recessive (rs55683539: OR 0.29, 95% CI 0.10–0.86, *p*=0.017; and rs10965064: OR 0.52, 95% CI 0.31–0.89,

 $p=0.015$) models (Table [6\)](#page-10-0). We also explore the association of *MIR31HG* SNPs with menopausal status of BC patients. However, no signifcant association was found.

Table [7](#page-12-0) displayed the results of stratifed analysis of LN metastasis, clinical stage, and tumor size in case group. Stratifed analysis of LN metastasis presented a positive relationship between rs79988146 and LN metastasis under allelic (OR 1.79, 95% CI 1.03–3.11, *p*=0.038), dominant (OR 1.88, 95% CI 1.05–3.36, *p* = 0.032), and additive (OR 1.76, 95% CI 1.01–3.08, *p* = 0.042) models. Stratifed analysis of clinical stage displayed that rs1332184 was associated with the higher stage under allelic (OR 1.43, 95% CI 1.06–1.92, *p*=0.020), codominant (OR 2.69, 95% CI 1.27–5.72, *p* = 0.036), recessive (OR 2.50, 95% CI 1.20–5.22, $p = 0.015$), and additive (OR 1.42, 95% CI 1.04–1.92, *p*=0.026) models. Moreover, rs2181559 might **Table 4** Stratifed analysis of the age on association between selected SNPs and BC risk

670 International Journal of Clinical Oncology (2023) 28:664–679

P value was calculated by Wald Test adjusted by age

SNP single nucleotide polymorphism, *OR* odds ratio, *95% CI* 95% confdence interval

**P*<0.05 indicates statistical signifcance

be associated with larger tumor size of BC patients (additive: OR 1.39, 95% CI 1.00–1.94, $p = 0.046$).

MDR analysis for the efect of MIR31HG SNP‑SNP interaction on BC risk

The Dendrogram and the Fruchterman-Reingold describe the interactions between these SNPs (Fig. [1A](#page-14-0), B). Short connections among nodes represent stronger redundant interactions (Fig. [1](#page-14-0)A). A negative value for the two-locus entropy indicates an antagonistic efect, and a positive value indicates a synergistic efect (Fig. [1B](#page-14-0)). MDR analysis showed that candidate SNPs interaction is associated with BC risk (Table [8\)](#page-14-1). The optimal single-locus model for predicting BC risk is rs55683539 [testing accuracy (TA): 0.5038, cross-validation consistency (CVC): 5/10], which, rs55683539-CC group was a high risk group and rs55683539-TT group was a low risk group increase the BC risk. Among the multi-locus models, predicting the best combination of BC risk is through rs79988146, rs1332184, rs72703442, rs2025327, rs55683539, rs2181559, rs10965059, and rs10965064 combination of eight-locus model [TA: 0.5179, CVC: 10/10]. The combination of all high-risk genotypes was associated with an increased risk of BC compared with that of low-risk genotypes.

OR: odds ratio; 95% CI: 95% confdence interval

"/" indicates that the data is not available. *P* was calculated by chi square test

Bold indicated that $p < 0.05$ meant the data was statistically significant

Discussion

In recent years, it has been recognized that most tumor formations are under the combined efects of environmental and genetic factors. According to research, the occurrence of tumors may be the result of the superposition of multiple microscopic susceptibility genes [[30\]](#page-15-24), which may afect the metabolism of carcinogens, repair DNA damages, regulate hormone levels, and protect the immune function. Although numerous studies have been published on genetic associations with BC and, genetic efects of *MIR31HG* on cancer, few studies are concerned with available whether *MIR31HG* could serve as a candidate gene for BC. To the best of our knowledge, this study is the basic and fundamental one to analyze the association between *MIR31HG* gene polymorphisms and BC risk in Chinese women.

Through studies on *MIR31HG* related the functions in a BC mouse model, Augoff et al. confirmed that the changes in the expression level of this gene will facilitate tumor invasion and eventually metastasis [[24\]](#page-15-18). Shi et al. also proved that knocking of *MIR31HG* could inhibit tumor growth in vivo, which showed that the expression level of *MIR31HG* could be referred as a diagnostic and prognostic marker for BC [[25](#page-15-19)]. In this study, we assessed

Table 6 (continued)

SNP	Model	Genotype	Age of menarche		Number of births		Menopausal status	
			OR (95% CI)	\boldsymbol{p}	OR (95% CI)	\boldsymbol{p}	OR (95% CI)	\boldsymbol{p}
		TT	$1.64(0.58 - 4.67)$		$0.29(0.10 - 0.86)$		$2.92(0.71 - 12.00)$	
	Additive		$1.14(0.79-1.65)$	0.490	$0.94(0.68 - 1.30)$	0.690	$1.14(0.71-1.83)$	0.600
rs2181559	Allele	T	$\mathbf{1}$		$\mathbf{1}$		$\mathbf{1}$	
		\mathbf{A}	$1.27(0.93 - 1.74)$	0.136	$1.04(0.79-1.36)$	0.772	$1.13(0.84 - 1.51)$	0.433
	Codominant	TT	$\mathbf{1}$	0.140	$\mathbf{1}$	0.750	$\mathbf{1}$	0.500
		${\rm TA}$	$1.05(0.67-1.64)$		$1.14(0.78-1.69)$		$1.23(0.70-2.17)$	
		${\rm AA}$	$2.05(1.00-4.18)$		$0.96(0.49-1.86)$		$1.79(0.63 - 5.10)$	
	Dominant	TT	$\mathbf{1}$	0.450	$\mathbf{1}$	0.580	$\mathbf{1}$	0.340
		TA/AA	$1.18(0.77-1.81)$		$1.11(0.76 - 1.61)$		$1.30(0.75 - 2.24)$	
	Recessive	$\ensuremath{\mathsf{T}}\ensuremath{\mathsf{T}}\ensuremath{\mathsf{T}}\ensuremath{\mathsf{T}}\ensuremath{\mathsf{A}}$	$\mathbf{1}$	0.051	$\mathbf{1}$	0.730	$\mathbf{1}$	0.360
		${\rm AA}$	$2.00(1.02 - 3.93)$		$0.89(0.47-1.69)$		$1.60(0.58 - 4.36)$	
	Additive	$\overline{}$	$1.29(0.93 - 1.78)$	0.130	$1.04(0.78-1.39)$	0.790	$1.29(0.84 - 1.99)$	0.250
rs10965059	Allele	$\mathbf C$	$\mathbf{1}$		$\mathbf{1}$		$\mathbf{1}$	
		$\mathbf T$	$1.12(0.68 - 1.87)$	0.652	$1.03(0.66 - 1.61)$	0.911	$0.91(0.57-1.47)$	0.707
	Codominant	$\rm CC$	$\mathbf{1}$	0.890	$\mathbf{1}$	0.032	$\mathbf{1}$	0.076
		${\cal C}{\cal T}$	$1.15(0.64 - 2.07)$		$1.43(0.85-2.43)$		$0.68(0.32 - 1.47)$	
		TT	$0.94(0.18 - 4.81)$		$0.12(0.01 - 1.11)$		13.61 (0.69-269.91)	
	Dominant	$\rm CC$	$\mathbf{1}$	0.670	$\mathbf{1}$	0.460	$\mathbf{1}$	0.720
		$\ensuremath{\mathsf{T}\mathsf{T}}\xspace/\!\ensuremath{\mathsf{T}\mathsf{C}}\xspace$	$1.13(0.64 - 1.98)$		$1.21(0.73 - 2.00)$		$0.87(0.42 - 1.81)$	
	Recessive	CC/TC	1	0.920	$\mathbf{1}$	0.024	1	0.040
		$\ensuremath{\mathcal{T}}\ensuremath{\mathcal{T}}$	$0.92(0.18-4.68)$		$0.12(0.01-1.05)$		14.48 (0.73-288.35)	
	Additive	$\overline{}$	$1.09(0.67-1.76)$	0.740	$1.02(0.65 - 1.60)$	0.940	$1.09(0.58 - 2.05)$	0.790
rs10965064	Allele	$\mathsf C$	$\mathbf{1}$		$\mathbf{1}$		$\mathbf{1}$	
		${\bf G}$	$1.21(0.89-1.64)$	0.226	$0.78(0.60 - 1.01)$	0.059	$0.97(0.73 - 1.29)$	0.844
	Codominant	CC	$\mathbf{1}$	0.410	$\mathbf{1}$	0.050	$\mathbf{1}$	0.220
		$\rm CG$	$1.34(0.84 - 2.14)$		$1.04(0.69-1.56)$		$0.77(0.43 - 1.38)$	
		$\rm CC$	$1.38(0.73 - 2.61)$		$0.53(0.30-0.94)$		$1.57(0.68 - 3.59)$	
	Dominant	CC	$\mathbf{1}$	0.180	$\mathbf{1}$	0.510	$\mathbf{1}$	0.760
		CG/GG	$1.35(0.87 - 2.10)$		$0.88(0.60-1.29)$		$0.92(0.53 - 1.59)$	
	Recessive	$\ensuremath{\mathsf{CC}/\mathsf{CG}}$	$\mathbf{1}$	0.600	$\mathbf{1}$	0.015	$\mathbf{1}$	0.130
		$\mathbf{G}\mathbf{G}$	$1.17(0.66 - 2.10)$		$0.52(0.31 - 0.89)$		$1.80(0.83 - 3.89)$	
	Additive	\equiv	$1.20(0.89-1.63)$	0.230	$0.79(0.61 - 1.04)$	0.088	$1.11(0.76-1.64)$	0.580

OR: odds ratio; 95% CI: 95% confdence interval

"/" indicates that the data is not available

P was calculated by chi square test

Bold indicated that $p < 0.05$ meant the data was statistically significant

the relationship between *MIR31HG* gene polymorphisms and the risk of BC, and found some related targets on evaluating BC risk. The results showed that rs72703442, rs55683539 and rs2181559 on *MIR31HG* were signifcantly associated with a reduced risk of BC in Chinese women. And, after age stratifcation, these three SNPs (rs72703442, rs55683539, and rs2181559) on *MIR31HG* were signifcantly associated with a reduced risk of BC in Chinese women ≥ 52 years old. However, no SNPs were found to be associated with BC risk in Chinese women in the $<$ 52-year-old stratification.

By analyzing gene polymorphism, Xia et al. concluded that BC risk was evaluated according to the diferent ER and PR states [\[8](#page-15-5)]. Zhou et al. also believed that the status of ER and PR is still the key to determine the type of BC adjuvant therapy, because estrogen stimulates ER-mediated transcription to increase cell proliferation, thereby increasing the number of DNA replication errors [\[31\]](#page-15-25). Above conclusions proved that clinical indicators especially in the status of ER and PR, have a certain infuence on the development of BC.

ER and PR stratifcation showed that rs79988146 on *MIR31HG* was positively correlated with ER and PR in

Table 7 (continued)

OR: odds ratio; 95% CI: 95% confdence interval

"/" indicates that the data is not available

P was calculated by chi square test

Bold indicated that $p < 0.05$ meant the data was statistically significant

Chinese female BC patients. In addition, stratifcation by age at menarche only found that rs1332184 on *MIR31HG* was associated with age at menarche in BC patients. Number of births stratifcation showed that rs10965064 on *MIR31HG* was negatively correlated with BC patients' Number of births.

BC is a complex disease afected by the interaction of factors such as heredity. Multi-gene or SNP-SNP interaction studies may help to discover the risk factors of BC. Therefore, we performed MDR analysis to determine the potential SNP-SNP interactions among the eight SNPs in the *MIR31HG* gene polymorphisms. SNP-SNP interaction analysis indicated a strong interaction between SNPs on *MIR31HG* for BC sensitivity. In addition, in the multisite model, the combination of rs79988146, rs1332184, rs72703442, rs2025327, rs55683539, rs2181559, rs10965059, and rs10965064 is the best multi-site model for predicting BC sensitivity.

However, there are also some limitations that cannot be neglected in our study. First of all, this is a hospital-based case and control study, which may have some inevitable sample selection bias and the absence of partial sample

Fig. 1 The dendogram (**A**) and fruchterman Rheingold (**B**) of *MIR31HG* SNP-SNP interaction for BC risk. **A** Short connections among nodes represent stronger redundant interactions. **B** Negative percent entropy indicates redundancy

Table 8 SNP–SNP interaction models of the *MIR31HG* gene the predisposition of BC

Model	Testing Bal. Acc	CVC	p
rs55683539	0.5038	5/10	0.020
rs55683539,rs10965059	0.5509	9/10	< 0.001
rs55683539.rs10965059.rs10965064	0.5057	5/10	< 0.001
rs1332184,rs55683539,rs10965059,rs10965064	0.4858	4/10	< 0.001
rs1332184,rs2025327,rs55683539,rs10965059,rs10965064	0.5009	7/10	< 0.001
rs79988146.rs1332184.rs2025327.rs55683539.rs10965059.rs10965064	0.5236	5/10	< 0.001
rs79988146,rs1332184,rs72703442,rs2025327,rs55683539,rs10965059,rs10965064	0.5123	7/10	< 0.001
rs79988146.rs1332184.rs72703442.rs2025327.rs55683539.rs2181559.rs10965059.rs10965064	0.5179	10/10	< 0.001

P values were calculated using χ^2 tests

Bold indicated that $p < 0.05$ meant the data was statistically significant

MDR multifactor dimensionality reduction, *Bal. Acc.* balanced accuracy, *CVC* cross-validation consistency, *OR* odds ratio, *CI* confdence interval

information. Second, our study has a limited generalizability because all participants were Han Chinese. Therefore, further well-designed study with a larger population or other ethnic groups is needed to confrm our fndings. Then, our sample size was too insufficient to support stratified analysis of tumor subtypes. Finally, due to other information was incomplete, we didn't analyze other risk factors for BC, such as lifestyle, family history, and other benign breast lesions. Therefore, population-based studies with a large amount of sample size and more complete information will be needed in the future to improve and enhance the accuracy of assessments and to explore the interaction between genetic variants and these factors.

Conclusion

In conclusion, this study frstly shows that *MIR31HG* gene polymorphisms are associated with a reduced risk of BC in Chinese women, and provides a theoretical basis for future explorations of the relationship between *MIR31HG* gene and BC risk in diferent populations. These fndings can provide new biological insights for understanding the role of *MIR31HG* in the occurrence of BC.

Acknowledgements First of all, we thank all authors for their contributions and supports. Then, we are grateful to all participants for providing blood samples.

Author contributions XZ: conceived and designed the experiments; YW and XW: performed the experiments; CZ and ZZ: analyzed the data; YC: contributed reagents/materials/analysis tools; ZB: prepared the fgures and/or tables; XZ, YW and XW: drafted the work or revised it critically for important content. All authors have read and approved the manuscript.

Funding No.

Availability of data and material All data obtained from the current study are available from the corresponding author upon reasonable request.

Declarations

Conflict of interest The authors have declared that they have no competing interests.

Ethics approval All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Consent for publication Written informed consent was obtained from the patient for publication of this report.

References

- 1. Fan L, Strasser-Weippl K, Li J-J et al (2014) Breast cancer in China. Lancet Oncol 15(7):e279–e289
- 2. Huang BF, Tzeng HE, Chen PC et al (2018) HMGB1 genetic polymorphisms are biomarkers for the development and progression of breast cancer. Int J Med Sci 15(6):580–586
- 3. Kresovich JK, Gann PH, Erdal S et al (2018) Candidate gene DNA methylation associations with breast cancer characteristics and tumor progression. Epigenomics 10(4):367–378
- 4. Wang Y, Zhang H, Lin M et al (2018) Association of FGFR2 and PI3KCA genetic variants with the risk of breast cancer in a Chinese population. Cancer Manag Res 10:1305–1311
- 5. Yan Y, Zhang X (2017) The association between CD28 gene rs3116496 polymorphism and breast cancer risk in Chinese women. Biosci Rep 37(6):BSR20170884
- 6. Zhang M, Jin M, Yu Y et al (2012) Associations of miRNA polymorphisms and female physiological characteristics with breast cancer risk in Chinese population. Eur J Cancer Care (Engl) 21(2):274–280
- 7. Wu L, Shi W, Long J et al (2018) A transcriptome-wide association study of 229,000 women identifes new candidate susceptibility genes for breast cancer. Nat Genet 50(7):968–978
- 8. Xia P, Li B, Geng T et al (2015) FGFR2 gene polymorphisms are associated with breast cancer risk in the Han Chinese population. Am J Cancer Res 5(5):1854–1861
- 9. Tian T, Wang M, Zheng Y et al (2018) Association of two FOXP3 polymorphisms with breast cancer susceptibility in Chinese Han women. Cancer Manag Res 10:867–872
- 10. Zheng W, Zhang B, Cai Q et al (2013) Common genetic determinants of breast-cancer risk in East Asian women: a collaborative study of 23 637 breast cancer cases and 25 579 controls. Hum Mol Genet 22(12):2539–2550
- 11. de Moraes CL, Cruz EMN, Valoyes MAV et al (2021) AGR2 and AGR3 play an important role in the clinical characterization and prognosis of basal like breast cancer. Clin Breast Cancer 22(2):e242–e252
- 12. Zhu Y, Wang T, Tong Y et al (2021) 21-gene recurrence assay associated with favorable metabolic profiles in HR-positive, HER2-negative early-stage breast cancer patients. Front Endocrinol 12:725161
- 13. Invernizzi M, Lopez G, Michelotti A et al (2020) Integrating biological advances into the clinical management of breast cancer related lymphedema. Front Oncol 10:422
- 14. de Sire A, Losco L, Cisari C et al (2020) Axillary web syndrome in women after breast cancer surgery referred to an Oncological Rehabilitation Unit: which are the main risk factors? A retrospective case-control study. Eur Rev Med Pharmacol Sci 24(15):8028–8035
- 15. Suvanto M, Beesley J, Blomqvist C et al (2020) SNPs in lncRNA regions and breast cancer risk. Front Genet 11:550
- 16. Cui P, Zhao Y, Chu X et al (2018) SNP rs2071095 in LincRNA H19 is associated with breast cancer risk. Breast Cancer Res Treat 171(1):161–171
- 17. Qin J, Ning H, Zhou Y et al (2018) LncRNA *MIR31HG* overexpression serves as poor prognostic biomarker and promotes cells proliferation in lung adenocarcinoma. Biomed Pharmacother 99:363–368
- 18. Wang B, Jiang H, Wang L et al (2017) Increased *MIR31HG* lncRNA expression increases geftinib resistance in non-small cell lung cancer cell lines through the EGFR/PI3K/AKT signaling pathway. Oncol Lett 13(5):3494–3500
- 19. Eide PW, Eilertsen IA, Sveen A et al (2019) Long noncoding RNA *MIR31HG* is a bona fde prognostic marker with colorectal cancer cell-intrinsic properties. Int J Cancer 144(11):2843–2853
- 20. He A, Chen Z, Mei H et al (2016) Decreased expression of LncRNA *MIR31HG* in human bladder cancer. Cancer Biomark 17(2):231–236
- 21. Shih J-W, Chiang W-F, Wu ATH et al (2017) Long noncoding RNA LncHIFCAR/*MIR31HG* is a HIF-1α co-activator driving oral cancer progression. Nat Commun 8(1):15874
- 22. Sun K, Zhao X, Wan J et al (2018) The diagnostic value of long non-coding RNA *MIR31HG* and its role in esophageal squamous cell carcinoma. Life Sci 202:124–130
- 23. Ren ZP, Chu XY, Xue ZQ et al (2017) Down-regulation of lncRNA *MIR31HG* correlated with aggressive clinicopathological features and unfavorable prognosis in esophageal squamous cell carcinoma. Eur Rev Med Pharmacol Sci 21(17):3866–3870
- 24. Augof K, McCue B, Plow EF et al (2012) miR-31 and its host gene lncRNA LOC554202 are regulated by promoter hypermethylation in triple-negative breast cancer. Mol Cancer 11:5
- 25. Shi Y, Lu J, Zhou J et al (2014) Long non-coding RNA Loc554202 regulates proliferation and migration in breast cancer cells. Biochem Biophys Res Commun 446(2):448–453
- 26. Patil N, Berno AJ, Hinds DA et al (2001) Blocks of limited haplotype diversity revealed by high-resolution scanning of human chromosome 21. Science (New York, NY) 294(5547):1719–1723
- 27. Ren HT, Li YM, Wang XJ et al (2016) PD-1 rs2227982 polymorphism is associated with the decreased risk of breast cancer in northwest Chinese women: a hospital-based observational study. Medicine 95(21):e3760
- 28. Leem S, Park T (2017) An empirical fuzzy multifactor dimensionality reduction method for detecting gene-gene interactions. BMC Genom 18(Suppl 2):115
- 29. Shahin NN, Abd-Elwahab GT, Tawfq AA et al (2020) Potential role of aryl hydrocarbon receptor signaling in childhood obesity. Biochim Biophys Acta 1865(8):158714
- 30. Curran JE, Weinstein SR, Grifths LR (2000) Polymorphisms of glutathione S-transferase genes (GSTM1, GSTP1 and GSTT1) and breast cancer susceptibility. Cancer Lett 153(1–2):113–120
- 31. Zhou L, He N, Feng T et al (2015). Association of fve single nucleotide polymorphisms at 6q25.1 with breast cancer risk in northwestern China. Am J Cancer Res 5(8):2467–2475

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.