

## Promoter methylation of *DAPK1*, *FHIT*, *MGMT*, and *CDKN2A* genes in cervical carcinoma

Chiaki Banzai · Koji Nishino · Jinhua Quan · Kosuke Yoshihara ·  
Masayuki Sekine · Tetsuro Yahata · Kenichi Tanaka ·  
Gynecological Cancer Registry of Niigata

Received: 31 August 2012 / Accepted: 26 January 2013 / Published online: 14 March 2013  
© Japan Society of Clinical Oncology 2013

### Abstract

**Background and objectives** Aberrant DNA methylation contributes to the malignant phenotype in virtually all types of human cancer. This study explored the relationship between promoter methylation and inactivation of the *DAPK1*, *FHIT*, *MGMT*, and *CDKN2A* genes in cervical cancer.

**Methods** The promoter methylation of *DAPK1*, *FHIT*, *MGMT*, and *CDKN2A* was investigated by using a methylation-specific polymerase chain reaction in 53 specimens of cervical cancer (42 squamous cell carcinoma, 11 adenocarcinoma), 22 specimens of intraepithelial neoplasia tissues, and 24 control normal cervical tissue specimens. The correlation of promoter methylation with the clinicopathological features of cervical cancer was analyzed. The expressions of *DAPK1*, *FHIT*, *MGMT*, and *CDKN2A* were detected by measuring relative mRNA levels.

**Results** The promoter methylation of *DAPK1*, *FHIT*, *MGMT*, and *CDKN2A* in cervical cancer vs. intraepithelial neoplasia vs. normal cervical tissue was 75.5 vs. 31.8 vs. 4.2 % ( $p < 0.0001$ ), 66.0 vs. 59.1 vs. 25.0 % ( $p = 0.0033$ ), 34.0 vs. 27.3 vs. 20.8 % ( $p = 0.76$ ), and 17.0 vs. 31.8 vs. 8.3 % ( $p = 0.11$ ), respectively. The methylation of the promoter region significantly decreased the expression of only *DAPK1* ( $p = 0.03$ ). The methylation rate of the *DAPK1* gene promoter was significantly higher in cervical cancer tissues than in cervical intraepithelial neoplasia and normal cervical tissues.

**Conclusion** Promoter methylation may therefore lead to the inactivation of the *DAPK1* gene, and may be related to the progression of cervical oncogenesis.

**Keywords** Cervical cancer · Methylation · *DAPK1*

### Introduction

Cervical cancer is the fourth most common cancer in Japanese females, with an estimated 24,240 cases in 2003, including 8,674 cases of invasive cervical cancer and 6,955 cases of carcinoma in situ [1]. Multiple epidemiology and molecular biological studies indicate that human papillomavirus (HPV) is the major factor associated with the development of cervical cancer [2–5]. It is clear that other factors are involved in cervical carcinogenesis because the majority of patients with HPV-associated lesions, such as cervical intraepithelial neoplasia, do not progress to invasive cancer but remain stable or spontaneously regress over time [6]. Therefore, it is likely that host genetic and epigenetic events play an important role in cervical carcinogenesis. The CpG islands of a large number of genes that are unmethylated in normal tissues are methylated to various degrees in several types of human cancer and particularly in gynecological cancer [7–11]. The aberrant methylation of CpG islands within the promoter regions of several tumor suppressor genes has been reported in cervical cancer [11–23]. The extent of aberrant promoter hypermethylation and its association with loss of gene function in cancer suggests that CpG island methylation is an important mechanism in inactivating tumor suppressor genes. Many studies have previously reported the promoter hypermethylation of *DAPK1* (45 %), *FHIT* (11–50 %), *MGMT* (5–60 %), *CDKN2A* (7–57 %), and other genes during the progression

C. Banzai (✉) · K. Nishino · J. Quan · K. Yoshihara ·  
M. Sekine · T. Yahata · K. Tanaka  
Division of Obstetrics and Gynecology, Graduate School  
of Medical and Dental Sciences, Niigata University,  
1-757 Asahimachi-dori, Chuo Ward, Niigata 951-8510, Japan  
e-mail: chiaki-b@med.niigata-u.ac.jp

of cervical cancer [12, 13, 17, 18, 25, 30]. The purpose of the present study was to examine whether or not similar tendencies exist in Japanese patients. This study analyzed the relationship between the clinicopathological parameters and the methylation of the *DAPK1*, *FHIT*, *MGMT*, and *CDKN2A* genes in normal, premalignant, and malignant cervical samples to determine whether it is a useful molecular marker for monitoring cervical cancer.

## Materials and methods

### Clinical samples and DNA preparation

Samples were obtained from 53 primary cervical carcinoma patients, 22 cervical intraepithelial neoplasia and carcinomas in situ (CIN3-CIS), and 24 normal cases that were treated at Niigata University Medical and Dental Hospital, Ryukyu University Hospital, Niigata Cancer Center Hospital, and Nagaoka Red Cross Hospital, between 2004 and 2010. We extracted DNA from the tumor cells to assess the presence or absence of methylation, and performed a cytological examination of the lesion before or after surgery to confirm the presence of carcinoma in situ of the cervix in relation to cases of cervical dysplasia. All diagnoses were confirmed by pathological examination.

The clinicopathological characteristics of the samples are given in Tables 1 and 2. Thirty-seven of the tumors were stage 1, and 16 were stage 2–4. The stages of each cancer were established according to the 1995 staging system of the International Federation of Gynecology and Obstetrics (FIGO) criteria. 81 % (17/21) of CIN3 and 72 % (26/36) of cervical carcinoma patients were infected with HPV (Table 2).

Forty-two of 53 tumors were squamous cell carcinomas and 11 were adenocarcinomas. None of the patients had received radiotherapy or chemotherapy before the samples were obtained. The specimens were immediately processed in the laboratory for DNA extraction, HPV typing, and quantitative RT-PCR analysis.

### Bisulfite modification and methylation-specific PCR

The bisulfite conversion of genomic DNA was performed using a CpGenome DNA modification kit (Chemicon International, Temecula, CA, USA). The promoter methylation of *DAPK1*, *FHIT*, *MGMT*, and *CDKN2A* was investigated by using a methylation-specific polymerase chain reaction (PCR).

### Quantitative real-time PCR analysis

Total RNA was extracted from tissue samples using TRIzol reagent (Invitrogen). Total RNA (1 µg) from cervical

**Table 1** Clinical characteristics in normal, CIN3, and cancer samples

	Normal	CIN3	Cancer	<i>p</i> value*
No.	24	22	53	
Age (mean ± SD)	48.1 ± 10.5	38.9 ± 13.5	49.3 ± 13.3	0.66
Smoking				
Yes	12	13	21	
No	12	9	32	0.27

CIN3 cervical intraepithelial neoplasia, grade 3

\* Chi-squared test

**Table 2** Clinical characteristics of CIN3 and cancer samples

	CIN3	Cancer
No.	22	53
HPV high-risk	17	26
Others	2	4
Negative	2	6
Unknown	1	17

HPV high-risk types (16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 68)

**Table 3** Frequency of promoter methylation in normal, CIN3, and cancer samples

Gene	Normal	CIN3	Cancer	<i>p</i> value*
<i>CDKN2A</i>	2/24 (8.3 %)	7/22 (31.8 %)	9/53 (17.0 %)	0.11
<i>DAPK1</i>	1/24 (4.2 %)	7/22 (31.8 %)	40/53 (75.5 %)	<0.0001
<i>FHIT</i>	6/24 (25.0 %)	13/22 (59.1 %)	35/53 (66.0 %)	0.0033
<i>MGMT</i>	5/24 (20.8 %)	6/22 (27.3 %)	18/53 (34.0 %)	0.76

Values are number of samples positive for hypermethylation out of the total number of samples tested that produced a valid result (percent)

\* Chi-squared test

**Table 4** Frequency of promoter methylation in SCC and AD samples

Gene	SCC	AD	<i>p</i> value*
<i>CDKN2A</i>	7/42 (2.4 %)	2/11 (18.2 %)	1
<i>DAPK1</i>	33/42 (78.6 %)	7/11 (63.4 %)	0.31
<i>FHIT</i>	28/42 (66.7 %)	7/11 (63.4 %)	1
<i>MGMT</i>	15/42 (35.7 %)	3/11 (27.3 %)	0.73

Values are number of samples positive for hypermethylation out of the total number of samples

tested that produced a valid result (percent)

SCC squamous cell carcinoma, AD adenocarcinoma

\* Chi-squared test

cancer was used as a template in first-strand cDNA synthesis with the SuperScript III First-Strand Synthesis System (Invitrogen).

The cDNA was diluted 1:10 for subsequent real-time PCR, which was carried out using TaqMan Gene Expression

Assays (Applied Biosystems) with the TaqMan Universal PCR Master Mix II (Applied Biosystems) on a 7900HT Sequence Detection System (Applied Biosystems) according to the manufacturer’s instructions.

The relative quantification method was used to measure the amounts of the respective genes in cervical cancer samples, which were normalized to *GAPDH*.

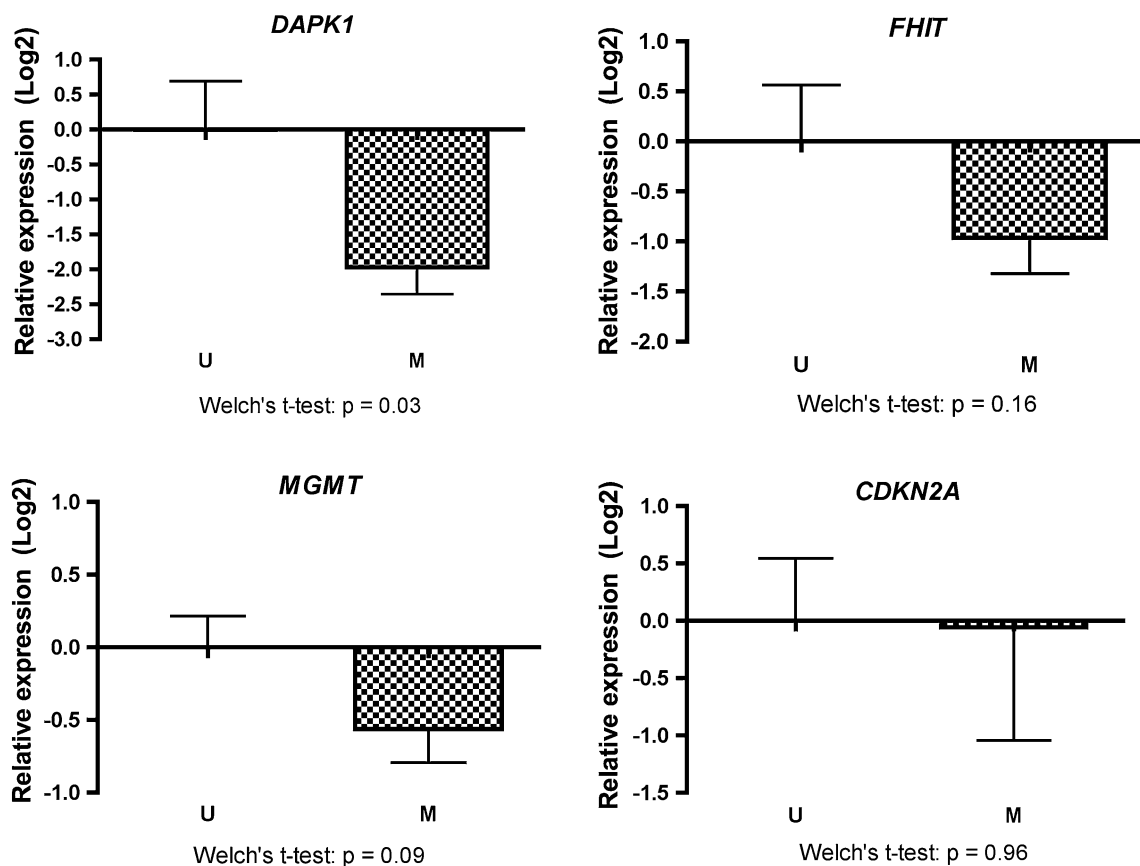
**Table 5** The association between *DAPK1* methylation status and staging in squamous cell carcinoma

	Methylation (-)	Methylation (+)	Frequency of methylation (%)	<i>p</i> value*
Stage				
CIS ( <i>n</i> = 14)	10	4	28.6	
I ( <i>n</i> = 29)	7	22	75.9	
II, III, IV ( <i>n</i> = 13)	2	11	84.6	0.0024

\* Chi-squared test

**Results**

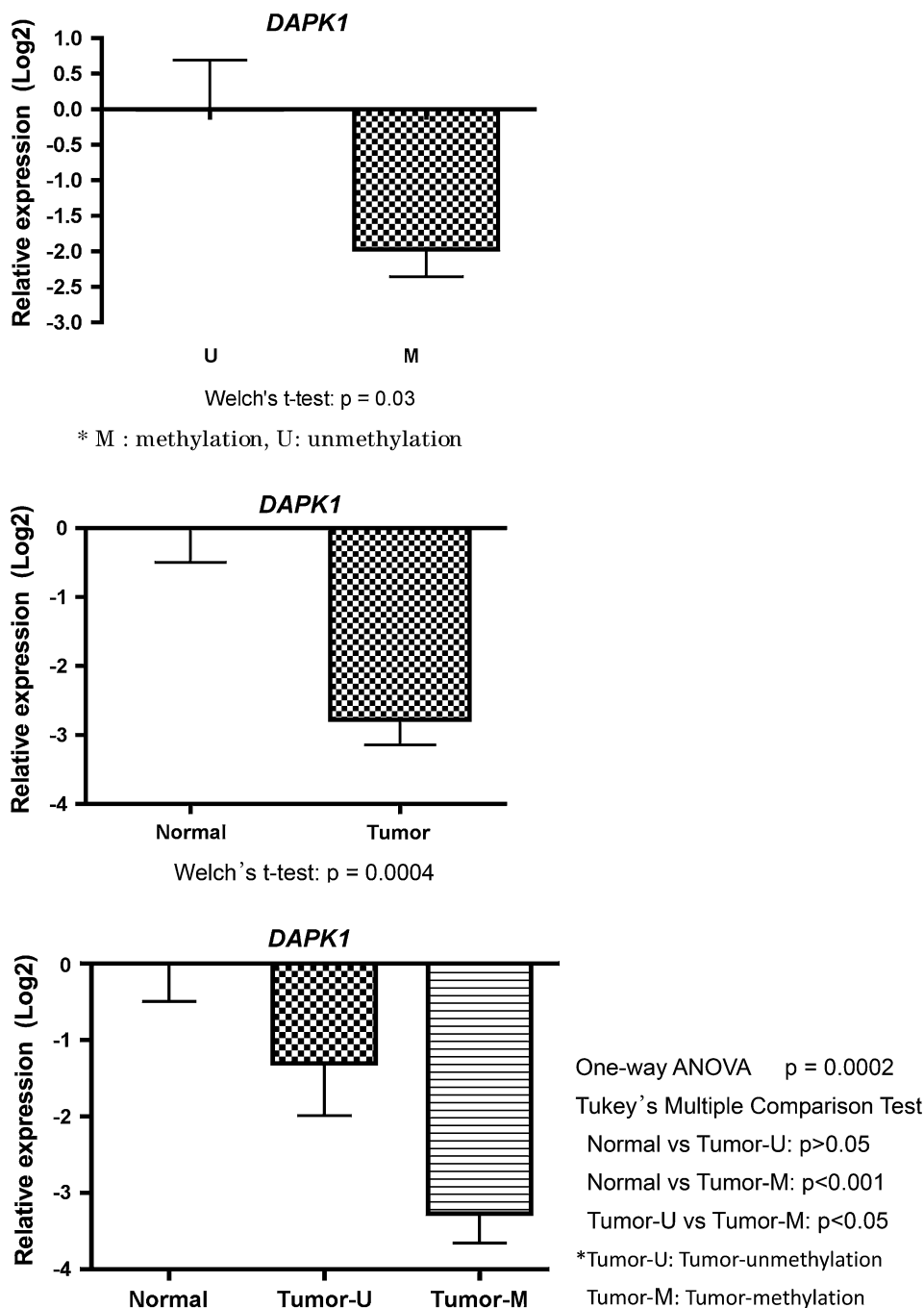
The promoter methylation of *DAPK1*, *FHIT*, *MGMT* and *CDKN2A* in cervical cancer vs. intraepithelial neoplasia vs. normal cervical tissue was 75.5 vs. 31.8 vs. 4.2 % (*p* < 0.0001), 66.0 vs. 59.1 vs. 25.0 % (*p* = 0.0033), 34.0 vs. 27.3 vs. 20.8 % (*p* = 0.76), and 17.0 vs. 31.8 vs. 8.3 % (*p* = 0.11), respectively (Table 3). There were no significant differences in the methylation rate of *DAPK1*, *FHIT*, *MGMT*, and *CDKN2A* between squamous cell carcinoma and adenocarcinoma (Table 4). There was a trend toward DAPK promoter hypermethylation among normal cervical tissues and cervical cancer (squamous cell carcinoma and adenocarcinoma). There was an increasing trend toward DAPK promoter methylation among carcinoma in situ, cervical cancer stage 1, and cervical carcinoma stage 2–4, with methylation rates of 28.6 % (4/14), 75.9 % (22/29), and 84.6 % (11/13), respectively (*p* = 0.0024; Table 5). The association between promoter methylation status and relative mRNA expression of *DAPK1*, *FHIT*, *MGMT*, and *CDKN2A* genes in cervical cancer is shown in Fig. 1. The comparison of the methylation of a promoter region and the



\* M: methylation, U: unmethylation

**Fig. 1** The association between promoter methylation status and relative mRNA expression of 4 genes in cancer tissues

**Fig. 2** The association between promoter methylation status and relative mRNA expression of DAPK in cancer tissues



relative level of mRNA between normal cervical tissue and cervical cancer revealed that only *DAPK1* showed a significant difference ( $p = 0.03$ ). The association between promoter methylation status and relative mRNA expression of DAPK in cancer tissues and normal tissues showed decreased mRNA expression when *DAPK1* was methylated (Fig. 2).

**Discussion**

High-risk HPV infection is associated with cervical carcinoma [2–5]. In Japanese females, HPV infection was

examined using restriction fragment length polymorphism (RFLP), and the rate of high-risk HPV infection was 94.8 % in CIN2/CIN3 cases and 93.4 % in patients with invasive cancer. In addition, in patients with cervical cancer, the rate of high-risk HPV infection was 90 % for patients in their twenties, 75.9 % for those in their 30s, 65.9 % for those in their 40s and 64.0 % for those in their 50s, indicating a decreasing trend in the rate of infection with age [29]. This study found that 81 % of CIN3 and 72 % of cervical carcinoma was associated with high-risk HPV infection. It was found that the average age of

patients with cervical cancer was 49 years, and this relatively older age may have been responsible for the low rate of HPV infection in our study. Other factors are also involved in cervical carcinoma. The inactivation of tumor suppressor genes has been investigated. Methylation plays an important role in tumorigenesis. The aberrant methylation of the normally unmethylated CpG islands of many tumor suppressor genes is associated with transcriptional inactivation and loss of expression [17, 26, 27]. Many studies have investigated the promoter hypermethylation of *DAPK1*, *FHIT*, *MGMT*, *CDKN2A*, and other genes during the progression of cervical oncogenesis [12, 13, 17, 18, 25, 30]. One study showed a correlation between tobacco use and methylation of *CDKN2A* [28].

The current study investigated the methylation profiles in the promoter region of *DAPK1*, *FHIT*, *MGMT*, and *CDKN2A* genes in normal, CIN3, and cervical carcinoma tissues.

No relationship was found between the methylation of *MGMT* and *CDKN2A* and cervical carcinoma in the current study.

*FHIT* has been identified as a candidate tumor suppressor gene, and re-expression of *FHIT* in a variety of human cell lines results in growth inhibition and induction of apoptosis. *FHIT* is hypermethylated in 40–50 % of cervical carcinomas [25]. The current study found that there is increased methylation of *FHIT* in cervical carcinoma tissue, but this was not associated with a significant decrease in the mRNA expression of *FHIT*.

*DAPK1* is a pro-apoptotic serine/threonine protein kinase that is dysregulated in a wide variety of cancers. In addition, *DAPK1* is involved in the control of autophagy [24].

There was a significant decrease in the expression of mRNA associated with the methylation of the promoter region of *DAPK1*, thus suggesting that *DAPK1* was involved in the occurrence of cervical carcinoma. Although *DAPK1* is reported to show a higher level of methylation in squamous cell carcinoma than in adenocarcinoma [13], there was no significant difference in the current study. The increased methylation of *DAPK1* suggested the possibility that *DAPK1* was associated with the progression of the disease.

**Conflict of interest** The authors do not have any conflict of interests.

## References

- Center for Cancer Control and Information Services, National Cancer Center, Japan. <http://ganjoho.jp/professional/statistics/statistics.html>
- Walboomers JM, Jacobs MV, Manos MM et al (1999) Human papillomavirus is a necessary cause of invasive cervical cancer worldwide. *J Pathol* 189:12–19
- Schiffman MH, Castle P (2003) Epidemiologic studies of a necessary causal risk factor: human papillomavirus infection and cervical neoplasia. *J Natl Cancer Inst* 95:E2
- Bosch FX, Manos MM, Munoz N et al (1995) Prevalence of human papillomavirus in cervical cancer: a worldwide perspective. International biological study on cervical cancer (IBSCC) study group. *J Natl Cancer Inst* 87:796–802
- Waggoner SE (2003) Cervical cancer. *Lancet* 361:2217–2225
- Holowaty P, Miller AB, Rohan T et al (1999) Natural history of dysplasia of the uterine cervix. *J Natl Cancer Inst* 91:252–258
- Jones PA, Baylin SB (2002) The fundamental role of epigenetic events in cancer. *Nat Rev Genet* 3:415–428
- Baldwin RL, Nemet E, Tran H et al (2004) BRCA1 promoter region hypermethylation in ovarian carcinoma: a population-based study. *Cancer Res* 60:5329–5333
- Salvesen HB, Macdonald N, Ryan A et al (2001) PTEN methylation is associated with advanced stage and microsatellite instability in endometrial carcinoma. *Int J Cancer* 91:22–26
- Zyaman M, Saka A, Millar A et al (2002) Methylation of adenomatous polyposis coli in endometrial cancer occurs more frequently in tumors with microsatellite instability phenotype. *Cancer Res* 62:3663–3666
- Yang HJ, Lin VWS, Wang Y et al (2004) Detection of hypermethylated genes in tumor and plasma of cervical cancer patients. *Gynecol Oncol* 93:435–440
- Dimitrios I, Pangona O, Ioannis M et al (2009) Correlation of promoter hypermethylation in hTERT, DAPK and MGMT genes with cervical oncogenesis progression. *Oncol Rep* 22:199–204
- Zhao XL, Meng ZY, Qiao YH et al (2008) Promoter methylation of DAPK gene in cervical carcinoma. *Chin J Cancer* 27(9): 212–215
- Dong SM, Kim HS, Rha SH et al (2001) Promoter methylation of multiple genes in carcinoma of the uterine cervix. *Clin Cancer Res* 7:1982–1986
- Virmani AK, Muller C, Rathi A et al (2001) Aberrant methylation during cervical carcinogenesis. *Clin Cancer Res* 7:584–589
- Narayan G, Arias-Pulido H, Koul S et al (2003) Frequent promoter methylation of CDH1, DAPK, RARB, and HIC1 genes in carcinoma of cervix uteri: its relationship to clinical outcome. *Mol Cancer* 2:24
- Jeong DH, Youm MY, Kin YN et al (2006) Promoter methylation of p16, DAPK, CDH1, and TIMP-3 genes in cervical cancer: correlation with clinicopathologic characteristics. *Int J Gynecol Cancer* 16:1234–1240
- Widschwendter A, Gatringer C, Ivarsson L et al (2004) Analysis of aberrant DNA methylation and human papillomavirus DNA in cervicovaginal specimens to detect invasive cervical cancer and its precursors. *Clin Cancer Res* 10:3396–3400
- Lea JS, Coleman R, Kurien A et al (2004) Aberrant p16 methylation is a biomarker for tobacco exposure in cervical squamous cell carcinogenesis. *Am J Obstet Gynecol* 190:674–679
- Wong YF, Chung TK, Cheung TH et al (1999) Methylation of p16INK4A in primary gynecologic malignancy. *Cancer Lett* 136:231–235
- Widschwendter A, Muller HM, Fiegl H et al (2004) DNA methylation in serum and tumors of cervical cancer patients. *Clin Cancer Res* 10:565–571
- Ivanova T, Vinokurova S, Petrenko A et al (2004) Frequent methylation of 5' flanking region of TIMP-2 gene in cervical cancer. *Int J Cancer* 108:882–886
- Widschwendter A, Ivarsson L, Blassnig A et al (2004) CDH1 and CDH13 methylation in serum is an independent prognostic marker in cervical cancer patients. *Int J Cancer* 109:163–166
- Michie AM, McCaig AM, Nakagawa R et al (2010) Death-associated protein kinase (DAPK) and signal transduction: regulation in cancer. *FEBS J* 277:74–80

25. Ki KD, Lee SK, Tong SY et al (2008) Role of 5'-CpG island hypermethylation of the FHIT gene in cervical carcinoma. *J Gynecol Oncol* 19(2):117–122
26. Baylin SB, Herman JG, Graff JR et al (1998) Alterations in DNA methylation: a fundamental aspect of neoplasia. *Adv Cancer Res* 72:141–196
27. Jones PA, Laird PW (1999) Cancer epigenetic comes of age. *Nat Genet* 21:163–167
28. Lea JS, Coleman R, Kurien A et al (2004) Aberrant p16 methylation is a biomarker for tobacco exposure in cervical squamous cell carcinogenesis. *Am J Obstet Gynecol* 190:674–679
29. Onuki M et al (2009) Human papillomavirus infections among Japanese women: age-related prevalence and type-specific risk for cervical cancer. *Cancer Sci* 100(7):1312–1316
30. Lin Z, Gao M, Zhang X et al (2005) The hypermethylation and protein expression of p16 INK4A and DNA repair gene O6-methylguanine-DNA methyltransferase in various uterine cervical lesions. *J Cancer Res Clin Oncol* 131(6):364–370