CORRESPONDENCE

The landscape of genome sequencing and assembling in plants

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Received: 6 October 2022 / Revised: 7 November 2022 / Accepted: 8 November 2022 / Published online: 15 November 2022 © The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2022

Plant genetic information was stored in the DNA sequences, which ultimately control plant traits, including response to diferent environmental stresses genetically or epigenetically (Han et al. [2022](#page-5-0)). Thus, decoding genome sequences will be the frst step and the foundation to understand DNA genetic information. Since the genome sequence of *Arabidopsis thaliana* was fully sequenced and assembled in 2000 (The Arabidopsis Genome Initiative [2000\)](#page-5-1), more and more plant genomes have been sequenced, assembled, and deposited into the public databases. Based on a recent study, a total of 798 plant species have been fully sequenced and assembled (Marks et al. [2021](#page-5-2)). According to this survey, of the currently known 137 land plant orders (Freiberg et al. [2020](#page-5-3)), there are 62 orders that have at least one plant species been sequenced and assembled (Marks et al. [2021](#page-5-2)), whose genomes are currently public available in the databases, such as GenBank, Phytozome, and PlantEnsemble. However, the distribution of these sequenced genomes was unbalanced in diferent plant orders and families, such as 83, 80, and 67 sequenced and assembled genomes were reported in Brassicales, Poales, and Lamiables, respectively. These plant species belong to 146 families, in which 80 species were sequenced and assembled in Brassicaceae, followed by Poaceae and Fabaceae that have 76 and 54 species fully sequenced, respectively. The majority (66) of sequenced families only had one species reported; there are only 16 families in which there are more than 10 species sequenced and assembled (Table [1](#page-1-0)).

The 798 plant species with sequenced and assembled genomes include the following: (1) agriculturally important crops, such as rice (*Oryza sativa*), corn (*Zea mays*), soybeans (*Glycine max*), peanut (*Arachis hypogaea*),

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potato (*Solanum tuberosum*), sesame (*Sesamum indicum*), and cotton (*Gossypium hirsutum*); (2) vegetables, such as tomato (*Solanum lycopersicum*), sweet potato (*Ipomoea batatas*), radish (*Raphanus sativus*), and cabbage (*Brassica oleracea*); (3) fruits, such as apple (*Malus domestica*), pear (*Pyrus communis*), banana (*Musa acuminata*), and strawberry (*Fragaria ananassa*); (4) ornamental flowers and trees, such as *Eschscholzia californica*, *Macleaya cordata*, *Aquilegia coerulea*, *Rosa chinensis*, *Callicarpa americana*, *Rhododendron delavayi*, *Antirrhinum majus*, and *Liriodendron chinense*; (5) architectural and industrial trees, such as *Trema orientale*, *Mesua ferrea*, *Hevea brasiliensis* (rubber tree), *Abies alba*, and *Sequoia sempervirens*; (6) medical plants, such as *Trichopus zeylanicus*, *Glycyrrhiza uralensis*, *Quillaja saponaria*, *Rehmannia glutinosa*, *Salvia miltiorrhiza*, *Andrographis paniculata*, *Coptis chinensis*, *Cannabis sativa*, and *Aquilaria sinensis*, which produce lots of traditional medicines for treating various human diseases; and (7) model plant species, such as *Arabidopsis* and the wild species of plant species mentioned above (Marks et al. [2021](#page-5-2)). The genomes of several special plant species are also sequenced, which include carnivorous plant species, such as *Dionaea muscipula* (Venus fytrap), *Aldrovanda vesiculosa*, *Utricularia reniformis*, and *Utricularia gibba*. From here, we clearly see that the majority of currently sequenced plant genomes are economically important plant species (human related) and their wild species, such as domesticated (135), cultivated (127), and natural commodity (120) plant species (Marks et al. [2021](#page-5-2)). Although there are about half of sequenced species are wild species, many of them are the relatives to agriculturally and economically important plants (Marks et al. [2021](#page-5-2)). To consider the species population of both wild and domesticated species (in which only less than 0.5% of species are domesticated species and more than 99.5% of species are wild species), currently genome sequencing is more preferred to the domesticated plants and their wild relatives.

Due to the cost and technical difficulty, scientists always initially selected species with a small and simple genome size for whole genome sequencing, such as *A. thaliana* was

Family	Order	No. of species	Example species
Brassicaceae	Brassicales	80	Arabidopsis. Rape, mustard, wild cabbage, flax, radish
Poaceae	Poales	76	Corn, sugarcane, foxtail millet, sorghum, bread wheat, durum wheat
Fabaceae	Fabales	54	Peanut, soybean, pigeon pea, chickpea, French bean, pea, cowpea, groundnut
Solanaceae	Solanales	41	Tomato, potato, eggplant, pepper, tobacco
Rosaceae	Rosales	36	Strawberry, apple, apricot, almond, peach, pear, rose
Myrtaceae	Myrtales	34	Brazil cherry, guava, silver mallet wood, smooth-barked coolabah
Malvaceae	Malvales	32	Upland cotton, sea-island cotton, cacao, jute, and tree cotton
Oleaceae	Lamiales	26	Golden bell, fragrant ash, Texas ash
Asteraceae	Asterales	17	Sunflower, lettuce
Lamiaceae	Lamiales	16	Horse mint, American beautyberry, danshen
Salicaceae	Malpighiales	16	Willow tree, white poplar, black cottonwood
Cucurbitaceae	Cucurbitales	15	Bitter melon, winter squash, wild melon, ash gourd
Rutaceae	Sapindales	13	Sweet orange, mandarin orange, clementine
Juglandaceae	Fagales	10	Pecan, walnut
Moraceae	Rosales	10	Breadfruit, breadnut, fig
Pinaceae	Pinales	10	Silver fir, Norway spruce, sugar pine
Vitaceae	Vitales	9	Grape, wine grape
Apiaceae	Apiales	8	Cilantro, fennel
Orchidaceae	Asparagales	8	Dendrobium catenatum, Dendrobium officinale
Amaranthaceae	Caryophyllales	7	Amaranthus hybridus, Amaranthus hypochondriacus
Dioscoreaceae	Dioscoreales	7	Greater yam, bitter yam
Ericaceae	Ericales	7	Strawberry tree, American blueberry
Betulaceae	Fagales	7	European alder, alpine birch
Fagaceae	Fagales	7	Chestnut, beech, oak
Rubiaceae	Gentianales	7	Coffee, David's milkberry
Euphorbiaceae	Malpighiales	7	Rubber tree, cassava
Convolvulaceae	Solanales	7	Sweet potato, glory
Arecaceae	Arecales	6	African oil palm, American oil palm, date palm
Cactineae	Caryophyllales	6	Organ-pipe cactus, saguaro, elephant cactus
Chenopodiaceae	Caryophyllales	6	Quinoa, spinach
Caryophyllaceae	Caryophyllales	5	Starwort, clove pink
Musaceae	Zingiberales	5	Banana
Araceae	Alismatales	4	Elephant ear, duckweed, Wolffia australiana
Droseraceae	Caryophyllales	4	Venus flytrap, waterwheel plant
Polygonaceae	Caryophyllales	4	Buckwheat
Nyssaceae	Cornales	4	Dove tree
Apocynaceae	Gentianales	4	Butterfly flower, Cape periwinkle
Acanthaceae	Lamiales	4	Green chiretta, white mangrove
Orobanchaceae	Lamiales	4	Chinese foxglove
Plantaginaceae	Lamiales	4	Antirrhinum majus
Lauraceae	Laurales	4	Avocado
Ranunculaceae	Ranunculales	4	Colorado blue columbine, Chinese goldthread
Cannabaceae	Rosales	4	Cannabis sativa, charcoal tree
Anacardiaceae	Sapindales	4	Mango, pistachio
Sapindaceae	Sapindales	4	Shantung maple, longan
Crassulaceae	Saxifragales	4	Orpin rose
Anthocerotaceae	Anthocerotales	3	Field hornwort
Marchantiaceae	Marchantiales	3	Common liverwort
Selaginellaceae	Selaginellales	3	Selaginella moellendorffii

Table 1 A total of 798 plant species, belonging to 146 families in 62 orders, with fully genomes sequenced and assembled*

Table 1 (continued)

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* The data was collected based on the recent publication in *Nature Plants* (Marks et al. [2021](#page-5-2))

selected for the frst sequenced plant species (The Arabidopsis Genome Initiative [2000](#page-5-1)). *Gossypium raimondii*, a diploid ancestor of cultivated tetraploid cotton, was frstly sequenced for cotton (Wang et al. [2012;](#page-5-4) Peng et al. [2021](#page-5-5)). Both of them are diploid plant species with small genome size and then move to polyploid species with a more complicated and big genome size (Peng et al. [2021\)](#page-5-5). Currently, all potential genomes can be quickly sequenced and assembled (Marks et al. [2021](#page-5-2)). This is evidenced by the rapid expansion of plant genomes sequenced and assembled in the past one decade. More than 75% of assembled plant genomes were performed in the last 5 years (Marks et al. [2021](#page-5-2)), particularly for the complicated polyploid plant species, such as cotton (*Gossypium hirsutum*) (Li et al. [2015;](#page-5-6) Zhang et al. [2015](#page-5-7)), oilseed rape (*Brassica napus*) (Bancroft et al. [2011\)](#page-5-8), and wheat (*Triticum aestivum*) (Maccaferri et al. [2019;](#page-5-9) Budak et al. [2021](#page-5-10)). The quick development of genome sequencing and assembly is associated with the rapidly developed sequencing technology and associated genome assembling computational tools. In the past two decades, the next-generation deep sequencing (NGS) technologies have been revolutionarily developed (Van Dijk et al. [2018](#page-5-11)), particularly for the long sequence reading NGS, such as PacBio [\(www.pacb.com\)](http://www.pacb.com) and Nanopore [\(www.nanoporetech.com/](http://www.nanoporetech.com/)), which provide more reliable platforms for sequencing more complicated genomes. The mean contig N50 was increased dramatically from 99.5 ± 48.1 kb in 2010 to 3395.2 ± 735.4 kb in 2020 (Marks et al. [2021\)](#page-5-2), majorly contributed by advanced NGS with long read capacity. NGS not only decodes the genomes but also directly detects the diferent types of DNA base modifcation (Van Dijk et al. [2018](#page-5-11)), which provides a powerful tool for studying the genome variance, modifcation, diversity, structure, evolution, and functions at both the genetic and epigenetic levels. Whole genome sequencing also facilitates the study not only on coding sequences but also on the dark matter (Kaur and Zhang [2022\)](#page-5-12), noncoding sequences, such as microRNAs, which has shown signifcant roles in many biological processes in both plants and animals (Li and Zhang [2016;](#page-5-13) Zhang and Unver [2018;](#page-5-14) Gebert and Macrae [2019](#page-5-15)).

Plant genome sequencing was associated with the economic development and the funding investment for scientifc research (Marks et al. [2021](#page-5-2)). Among the 798 sequenced plant species, 77% of sequencing was performed in China (235), the USA (212), and the European nations (168). There are only few species that were sequenced and assembled in other countries. Although there are lots of native species in Africa and South America, including lots of important species, the genome sequencing and assembling of these species were majorly performed by off-continents, mentioned early by China, USA, or the European nations (Marks et al. [2021\)](#page-5-2). As discussed by the authors of *Nature Plant* paper (Marks et al. [2021\)](#page-5-2), more international collaboration should be performed to sequence and assembly more plant genomes for the world's sustainable development.

Although great progress has been made in plant genome sequencing and assembling, plant genome sequencing is just in the start, and it is quickly moving into the next stage for rapid development and application. In the following decade, more and more progresses will be achieved, which include but not limited to the following felds:

The genome sequencing technique and associated computational tools for analyzing genome sequencing data and genome assembly will be further improved. This will include sequencing techniques that can precisely read long sequences, such as current PacBio and Nanopore. It will also signifcantly improve the assembling accuracy if we switch sequencing the entire genome to sequencing an individual chromosome. Chromosome sequencing has many advantages, including avoiding potential repetitive sequence confusions and enhancing the precisely assembling the entire genome sequences. In the next couple of years, chromosome level sequencing technology will enhance the genome sequencing and its application.

Current genome sequencing is majorly focused on agriculturally and economically important plant species and their wild relatives. As further reduced the costs of genome sequencing and assembling, other environmentally important and endangered species will become the new targets for whole genome sequencing. Quick development of the entire world, particularly due to industrialization and its associated global issues, such as global warming and emerging environmental pollutants, is threatening our earth, which further negatively contribute to the biodiversity and sustainable development. How to rescue the endangered species is becoming a big challenge for our entire community. At least, from genome sequencing aspect, we can fully sequence these endangered plant species and store their genetic information that is necessary for the future artifcial intelligence (AI) technology to recover these species.

More research will focus on the temporal and spatial structures and functions of genomes as well as the comparison of multiple genomes in same species and across different species (pan-genome sequencing). These studies will further allow us to understand the fne structures, including 3D structures and the relationship between structures and functions and elucidate how DNA structure changes, even fne change (such as DNA methylation), impact gene function and response to diferent environment changes (ecogenomics and environmental genomics).

Genome sequencing is opening a new window and feld for better usage of plants for our human and environmentally sustainable development. The decoded genetic information provides new targets for improving crop yield, quality, and response to various environmental biotic and abiotic stresses as well as generating new secondary components, such as pharmaceutical drugs. Particularly with the newly developed genome editing tools, such as clustered regularly interspaced short palindromic repeats (CRISPR)/CRISPR-associated protein (Cas) (Jinek et al. [2012](#page-5-16); Cheng et al. [2013](#page-5-17); Cong et al. [2013\)](#page-5-18), it is easier for our human beings to operate an individual genome sequence at a more precise way for various economic and health purposes. Based on the genome sequencing information, by using various versatile CRISPR tools (Jogam et al. [2022;](#page-5-19) Li et al. [2022a](#page-5-20)), scientists already precisely targeted on certain genes for improving crops (Li et al. [2021](#page-5-21), [2022b](#page-5-22); Zhang et al. [2021\)](#page-5-23) and gene therapy (Zhang [2021\)](#page-5-24).

As rapid development of deep sequencing techniques and associated computational programming as well as the decreased cost of these techniques, we believe plant genome sequencing and assembling are stepping into its golden age. Genome sequencing will further facilitate the genome biotechnology and its application in agriculture, environment, and pharmaceuticals as well as industry for the world's food and health safety and security as well as sustainable development.

Author contribution WL and BZ wrote and revised the manuscript.

Data availability Not applicable.

Declarations

Ethical approval Not applicable.

Conflict of interest The authors declare no competing interests.

References

- Bancroft I, Morgan C, Fraser F et al (2011) Dissecting the genome of the polyploid crop oilseed rape by transcriptome sequencing. Nat Biotechnol 29(8):762–766.<https://doi.org/10.1038/nbt.1926>
- Budak H, Appels R, Paux E (2021) Insights on decoding wheat and barley genomes. Funct Integr Genomics 21(2):157–159. [https://](https://doi.org/10.1007/s10142-021-00774-z) doi.org/10.1007/s10142-021-00774-z
- Cheng AW, Wang H, Yang H et al (2013) Multiplexed activation of endogenous genes by crispr-on, an rna-guided transcriptional activator system. Cell Res 23(10):1163–1171. [https://doi.org/10.](https://doi.org/10.1038/cr.2013.122) [1038/cr.2013.122](https://doi.org/10.1038/cr.2013.122)
- Cong L, Ran FA, Cox D et al (2013) Multiplex genome engineering using crispr/cas systems. Science 339(6121):819–823. [https://doi.](https://doi.org/10.1126/science.1231143) [org/10.1126/science.1231143](https://doi.org/10.1126/science.1231143)
- Freiberg M, Winter M, Gentile A et al (2020) Lcvp, the leipzig catalogue of vascular plants, a new taxonomic reference list for all known vascular plants. Sci Data 7(1):416. [https://doi.org/10.1038/](https://doi.org/10.1038/s41597-020-00702-z) [s41597-020-00702-z](https://doi.org/10.1038/s41597-020-00702-z)
- Gebert LFR, Macrae IJ (2019) Regulation of microrna function in animals. Nat Rev Mol Cell Biol 20(1):21–37. [https://doi.org/10.1038/](https://doi.org/10.1038/s41580-018-0045-7) [s41580-018-0045-7](https://doi.org/10.1038/s41580-018-0045-7)
- Han J, Lopez-Arredondo D, Yu G et al (2022) Genome-wide chromatin accessibility analysis unveils open chromatin convergent evolution during polyploidization in cotton. PNAS 119(44):e2209743119. <https://doi.org/10.1073/pnas.220974311>
- Jinek M, Chylinski K, Fonfara I et al (2012) A programmable dualrna-guided DNA endonuclease in adaptive bacterial immunity. Science 337(6096):816–821. [https://doi.org/10.1126/science.](https://doi.org/10.1126/science.1225829) [1225829](https://doi.org/10.1126/science.1225829)
- Jogam P, Sandhya D, Alok A et al (2022) A review on crispr/cas-based epigenetic regulation in plants. Int J Biol Macromol 219:1261– 1271. <https://doi.org/10.1016/j.ijbiomac.2022.08.182>
- Kaur P, Zhang B (2022) New vision on the new era of genome study. Funct Integr Genomics 22(1):1–2. [https://doi.org/10.1007/](https://doi.org/10.1007/s10142-022-00826-y) [s10142-022-00826-y](https://doi.org/10.1007/s10142-022-00826-y)
- Li C, Zhang B (2016) Micrornas in control of plant development. J Cell Physiol 231(2):303–313.<https://doi.org/10.1002/jcp.25125>
- Li F, Fan G, Lu C et al (2015) Genome sequence of cultivated upland cotton (gossypium hirsutum tm-1) provides insights into genome evolution. Nat Biotechnol 33(5):524–530. [https://doi.org/10.1038/](https://doi.org/10.1038/nbt.3208) [nbt.3208](https://doi.org/10.1038/nbt.3208)
- Li C, Brant E, Budak H et al (2021) Crispr/cas: a Nobel Prize awardwinning precise genome editing technology for gene therapy and crop improvement. J Zhejiang Univ-Sci B 22(4):253–284. [https://](https://doi.org/10.1631/jzus.B2100009) doi.org/10.1631/jzus.B2100009
- Li C, Chu W, Gill RA et al (2022) Computational tools and resources for crispr/cas genome editing. Genomics Proteome Bioinforma. <https://doi.org/10.1016/j.gpb.2022.02.006>
- Li S, Lin D, Zhang Y et al (2022) Genome-edited powdery mildew resistance in wheat without growth penalties. Nature. [https://doi.](https://doi.org/10.1038/s41586-022-04395-9) [org/10.1038/s41586-022-04395-9](https://doi.org/10.1038/s41586-022-04395-9)
- Maccaferri M, Harris NS, Twardziok SO et al (2019) Durum wheat genome highlights past domestication signatures and future improvement targets. Nat Genet 51(5):885–895. [https://doi.org/](https://doi.org/10.1038/s41588-019-0381-3) [10.1038/s41588-019-0381-3](https://doi.org/10.1038/s41588-019-0381-3)
- Marks RA, Hotaling S, Frandsen PB et al (2021) Representation and participation across 20 years of plant genome sequencing. Nature Plants 7(12):1571–1578. [https://doi.org/10.1038/](https://doi.org/10.1038/s41477-021-01031-8) [s41477-021-01031-8](https://doi.org/10.1038/s41477-021-01031-8)
- Peng R, Jones DC, Liu F et al (2021) From sequencing to genome editing for cotton improvement. Trends Biotechnol 39(3):221–224. <https://doi.org/10.1016/j.tibtech.2020.09.001>
- The Arabidopsis Genome Initiative A (2000) Analysis of the genome sequence of the fowering plant Arabidopsis thaliana. Nature 408(6814):796–815. <https://doi.org/10.1038/35048692>
- Van Dijk EL, Jaszczyszyn Y, Naquin D et al (2018) The third revolution in sequencing technology. Trends Genet 34(9):666–681. [https://](https://doi.org/10.1016/j.tig.2018.05.008) doi.org/10.1016/j.tig.2018.05.008
- Wang K, Wang Z, Li F et al (2012) The draft genome of a diploid cotton gossypium raimondii. Nat Genet 44(10):1098-1103. [https://](https://doi.org/10.1038/ng.2371) doi.org/10.1038/ng.2371
- Zhang B (2021) Crispr/cas gene therapy. J Cell Physiol 236(4):2459– 2481. <https://doi.org/10.1002/jcp.30064>
- Zhang B, Unver T (2018) A critical and speculative review on microrna technology in crop improvement: current challenges and future directions. Plant Sci 274:193–200. [https://doi.org/10.1016/j.plant](https://doi.org/10.1016/j.plantsci.2018.05.031) [sci.2018.05.031](https://doi.org/10.1016/j.plantsci.2018.05.031)
- Zhang T, Hu Y, Jiang W et al (2015) Sequencing of allotetraploid cotton (Gossypium hirsutum l. Acc. Tm-1) provides a resource for fber improvement. Nat Biotechnol 33(5):531–537. [https://doi.org/](https://doi.org/10.1038/nbt.3207) [10.1038/nbt.3207](https://doi.org/10.1038/nbt.3207)
- Zhang D, Zhang Z, Unver T et al (2021) Crispr/cas: a powerful tool for gene function study and crop improvement. J Adv Res 29:207– 221.<https://doi.org/10.1016/j.jare.2020.10.003>

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