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Pyrolysis of agricultural residues for bio-oil production

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Abstract The production of biofuel from biomass waste is of great interest to the scientific community regarding the discovery of solutions to global energy demand and global warming. The pyrolysis of biomass to produce bio-oil is an easy, cheap and promising technology. In the current investigation, the pyrolysis of two different biomasses (cornelian cherry stones and grape seeds) was achieved at temperatures ranging from 300 to 700 °C. The effect of pyrolysis temperatures on the yields of each product was significant. The bio-oil yields were maximized at 500 $^{\circ}$ C for cornelian cherry stones and 700 \degree C for grape seeds. The compositions of bio-oils for both cornelian cherry stones and grape seeds were similar and contained mainly oxygenated hydrocarbons. The compounds observed in this investigation were composed of phenols, alkyl benzenes, alkanes, alkenes, fatty acids, fatty acid esters and a few nitrogen-containing compounds. Bio-char properties were amended in association with both the pyrolysis temperature and biomass type. Bio-chars from cornelian cherry stones

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contained higher carbon and lower oxygen levels than those from grape seeds under identical conditions. Increases in pyrolysis temperatures produced bio-chars containing higher carbon levels and heating values for both carnelian cherry stones and grape seeds.

Keywords Pyrolysis - Cornelian cherry stone - Grape seeds - Bio-oil - Bio-char

Introduction

Biomass is one of the best sources to meet the energy demands of the future as it is not only renewable but also environmentally friendly. The energy obtained from biomass is used mainly in heat production, electricity, and as fuels for vehicles (Houshfar et al. [2014\)](#page-11-0). Various advanced technologies have been developed to obtain biofuels and/or biochemicals from biomass, which include the hydrothermal process (Jin and Enomoto [2011](#page-12-0)), pyrolysis (Torri et al. [2009](#page-12-0)), and combustion (Vassilev et al. [2013\)](#page-12-0).

Pyrolysis of biomass is one of the most efficient technologies used to produce biofuel. The process is carried out at elevated temperatures under an inert atmosphere which is maintained using either argon or nitrogen gases. The process yields bio-oil, solid residue, and gaseous products. Process conditions, as well as the raw materials used, have varying effects on the yields of each product.

Bio-oils and bio-chars are two significant pyrolysis products, and they have their own significant values which can be utilized in a sustainable way. Bio-oils can be used as transportation fuels after they are upgraded using a process such as deoxygenation and dearomatization (Huber et al. [2006](#page-11-0); Amen-Chen et al. [2001](#page-11-0)). Biofuels can be blended with diesel fuels with the assistance of surfactants (Huber et al. [2006](#page-11-0); Bridgwater [2004\)](#page-11-0). They can also be evaluated as a source of value-added chemicals after isolation and purification (Amen-Chen et al. [2001\)](#page-11-0).

Bio-chars from pyrolysis of biomass are also perspective products which can be used to improve soil functions (Marousek [2013a](#page-12-0)). Production and storage of bio-chars in soils have some beneficial aspects such as the mitigation of climate change by sequestering carbon (Marousek et al. [2014a](#page-12-0); Marousek [2013b](#page-12-0)), leading to an increase in the capacity of soil to store water (Marousek [2014](#page-12-0)), an increase in soil microbial biomass (Marousek [2014](#page-12-0)), and an increase in yields of crops (Marousek et al. [2014a\)](#page-12-0).

Fruit wastes and their by-products are important biomass resources for the scientific community to evaluate as potential energy sources. Many studies are available in the literature concerning the pyrolysis of fruit wastes for the production of biofuel via pyrolysis, such as sesame, mustard and neem de-oiled cakes (Volli and Singh [2012](#page-12-0)), tamarind seed (Kader et al. [2013](#page-12-0)), jatropha oil cakes (Raja et al. [2010;](#page-12-0) Jourabchi et al. [2014](#page-12-0)), palm kernel cake (Ngo et al. [2013](#page-12-0)), cottonseed cake (Putun et al. [2006](#page-12-0)), and pomegranate seeds (Ucar and Karagoz [2009](#page-12-0)).

Nayan et al. investigated the production of bio-oil from the pyrolysis of neem seed at the temperature zone from 400 to 500 °C and a heating rate of 20 °C min⁻¹ (Nayan et al. [2013](#page-12-0)). The greatest bio-oil yield (38 wt%) was accomplished at 475 \degree C. Hydrocarbons identified in the bio-oils were numerous and varied. Prominent compounds identified in the bio-oils were octadecanenitrile, oleanitrile, 9-octadecenoic acid methyl ester, and stearic acid methyl ester.

Pyrolysis of karanja seed to produce liquid fuel was carried at temperatures ranging between 500 and 600 °C (Shadangi and Mohanty [2014\)](#page-12-0). The optimal temperature used for the production of the greatest bio-oil yield (55.2 wt\%) was 550 °C. The bio-oil contained a mixture of oxygenated hydrocarbons including phenols, acids, esters, hexane, levoglucosan, amide, nitrile, benzene, furan, and other compounds.

Shadangi and Singh explored the pyrolysis of polanga seed cake at temperatures ranging from 450 to 600 $^{\circ}$ C (Shadangi and Singh [2012](#page-12-0)). The maximal bio-oil yield was approximately 46 %, and it was produced at the temperature of 550 °C. GC–MS analysis showed that the bio-oil contained various compounds. The relative concentrations of the following compounds were high: oleic acid, hexadecanoic acid, octadecanonic acid, octadec-9-enoic acid, hexadecanenitrile, 9-octadecanamide, octadecanamide, phenol, oleanitrile, heptadecane, and pentadecane.

Figueiredo et al. produced bio-oil from the pyrolysis of castor seeds at the temperature of 380 $^{\circ}$ C (Figueiredo and Romeiro [2009](#page-11-0)). The product distribution was as follows: 50 wt% bio-oil, 29 wt% bio-char, and 8 wt% gaseous

a,b By difference

 \degree Higher heating value (HHV) calculated by the Dulong formula $[HHV = 0.338C + 1.428(H - 0/8) + 0.095S]$

products. The fractions of the bio-oils contained long-chain hydrocarbons and polar compounds.

In the current paper, two types of fruit wastes (cornelian cherry stones and grape seeds) were exposed to pyrolysis under identical conditions. Pyrolysis temperatures from 300 to 800 C were studied. Product distributions from the pyrolysis of cornelian cherry stones were compared with the pyrolysis of grape seeds under identical conditions. The bio-oils were analyzed via GC–MS and an elemental analyzer. The bio-chars were studied and classified by means of elemental analysis and scanning electron microscopy techniques.

Materials and methods

Feedstock

Cornelian cherry stones were purchased from a local market in Safranbolu, Turkey. Samples were separated from the skin of the fruits, washed, and air-dried. The airdried samples were grinded in a blender. The samples were milled to a particle size: $\langle 0.5 \text{ mm} \rangle$. Grape seeds were obtained from a local market in Ankara, Turkey. Grape seed samples were used in the same condition of their purchase. Samples were milled to a particle size: $<$ 0.45 mm. The properties of waste biomasses are shown in Table 1.

Pyrolysis procedure

Pyrolysis experiments were completed at different pyrolysis temperatures (from 300 to 800 $^{\circ}$ C) for 1 h. An inert atmosphere was provided by a continuous nitrogen gas

Fig. 1 Pyrolysis setup

1. Nitrogen cyclinder

- 2. Furnace and reactor
- 3. Thermocouple
- 4. Condensors
- 5. Gas outlet

flow of 30 mL min^{-1} . The nitrogen gas flow was introduced before the experiment and continued for 30 min to remove the air inside. A nitrogen purge was injected after the reaction was complete, including the cooling down of the reactor to room temperature. The pyrolysis reactor was made of stainless steel with 6 cm diameter and 21 cm height. The pyrolysis setup is shown in Fig. 1. The pyrolysis reactor was charged with biomass samples of 25 g (dry basis). The system was heated at a rate of $7 \degree C/min$ to the target temperature. The volatile products were swept by nitrogen gas from the reactor to the collection flasks. The first two collections flasks were cooled with a water and ice mixture. The third flask was cooled using water. The pyrolysis experiments were repeated three times with an average standard deviation of 2.5 wt% for bio-oil and biochar.

Analysis of pyrolysis products

The liquid portion from the pyrolysis of biomass was exposed to liquid–liquid extraction using diethyl ether (300 mL) to recover bio-oils for analysis. The bio-oils were analyzed by gas chromatography–mass spectrometry (GC–MS) and elemental analysis. The GC–MS instrument was a 6890 Gas Chromatograph Agilent (30 m \times 0.25 mm i.d. phenyl methyl siloxane capillary column HP-5MS). The injector temperature was held at 250° C. The GC oven temperature was programmed in the following sequence: started at 40 °C; held for 10 min; raised at a rate of 2–170 °C; held for 5 min; raised to 250 °C at a rate of 8 °C; held for 15 min; raised to 300 °C at a rate 15 °C; and held at this final temperature for 10 min.

Fig. 2 TGA curves of biomass samples

The functional groups in the raw materials, liquids (consisting of organic and aqueous phase), and bio-chars were determined with Fourier transform infrared spectrometry for attenuated total reflectance (FTIR-ATR, Perkin Elmer FTIR 100 spectrometer).

The elemental content of the raw samples and bio-char products was established using a LECO CHNS TRuSpec. The surface morphologies of the raw samples and bio-chars were studied with the assistance of a scanning electron microscope (FEI Quanta 450 FEG).

Results and discussion

The TG profiles of the grape seeds and cornelian cherry stones are shown in Fig. 2. Moisture release for both grape

Fig. 3 Product distributions (solid, liquid, and gases) from the pyrolysis of grape seeds and cornelian cherry stones at different temperatures (300, 400, 500, 600, 700, and 800 C)

seeds and cornelian cherry stones took place at approximately 80 °C. Decomposition of the biomasses started just below 200 °C. In the case of grape seeds, the main decomposition occurred at temperatures between 170 and 600 °C and resulted in 67.8 wt% weight losses. The main decomposition of cornelian cherry stones was observed at temperatures between 170 and 600 $^{\circ}$ C which resulted in 71.7 wt% weight losses.

The distributions of products were influenced by both the pyrolysis temperature and biomass type in relation to temperature (as shown in Fig. 3). The trend for solid product yields was similar for both grape seeds and cherry stones. Solid product yields were gradually diminished

with increasing the temperature for both grape seeds and cherry stones. The pyrolysis of cornelian cherry stones yielded less bio-char in comparison with grape seeds under identical conditions. The highest and lowest bio-char yields were 50.30 and 25.72 wt% for cornelian cherry stones and 53.88 and 29.82 wt% for grape seeds, respectively. Bio-oil yields from the pyrolysis of cornelian cherry stones were scaled up when the temperature was increased to 500 °C. After this temperature increase, further boosts in pyrolysis temperatures provided lower bio-oil yields from cornelian cherry stones. In the case of the pyrolysis of grape seeds, bio-oil yields increased as the pyrolysis temperature was raised (up to $700 \degree C$). After further increases in the pyrolysis temperature from 700 to 800 $^{\circ}$ C, the bio-oil yields dropped for both biomass samples. Previous studies showed that bio-oil yields from the pyrolysis of biomass increased incrementally as the temperature was raised to a certain temperature. After this specific temperature that changed depending on the conditions (i.e., type of biomass, type of reactor, and type of pyrolysis), further increases in pyrolysis temperatures generated either lower yields of biooils (Ucar and Karagoz [2009\)](#page-12-0) or the yields remained at almost the same levels (Shadangi and Singh [2012](#page-12-0)). It is believed that each biomass has the potential to provide a maximum bio-oil yield at a certain pyrolysis temperature. After this temperature, further increases in temperature helped to transform the bio-oil to gas products. Accordingly, gas product yields improved for both biomass samples when the pyrolysis temperatures were raised. The greatest gas yield (35 wt% ca.) was obtained at 800 $^{\circ}$ C for both the pyrolysis of grape seeds and cornelian cherry stones.

Figure [4](#page-4-0) presents the FTIR spectra of bio-oil derived from the pyrolysis of grape seeds (Fig. [4](#page-4-0)a) and cornelian cherry stones (Fig. [4b](#page-4-0)) and their corresponding raw materials. Functional groups observed in bio-oils from the grape seeds were similar to those from the cornelian cherry stones. The broad band between $3,200$ and $3,600$ cm⁻¹ represented the OH group. The peak at $2,924$ cm⁻¹ rose due to aliphatic CH₂. The band at 2,855 cm⁻¹ confirmed the stretching of C–H vibrations of the alkyl functional groups (CH₂ and CH₃). The peak at 1,743 cm⁻¹ is assigned to the C=O functional group. The peaks between 1,100 and $1,200 \text{ cm}^{-1}$ are assigned to C–O stretching bonds. The peak at $1,035$ cm⁻¹ represents C-H out-of-plane bending vibrations of the alkene groups. The peaks between 680 and 900 cm^{-1} are assigned to benzene rings (Lievens et al. [2008](#page-12-0)). Previous studies confirmed similar functional groups in bio-oils which were identified by FTIR from the pyrolysis of different biomass resources (Grierson et al. [2011](#page-11-0); Huang et al. [2012;](#page-11-0) Demiral et al. [2012\)](#page-11-0).

Increases in the pyrolysis temperatures did not change the functional groups in the bio-oils. The peak intensities were Fig. 4 FTIR spectrums of raw material and bio-oils from grape seeds (a) and cornelian cherry stones (b) after pyrolysis at 300, 400, 500, 600, 700, and 800 °C

high in bio-oils from cornelian cherry stones in comparison with the raw materials. In the case of bio-oils produced from grape seeds, observed functional groups were similar to those of the raw material. The intensity of the peaks in biooils at between 2,800 and 3,200 cm^{-1} was high in comparison with the raw material. The total ion chromatograms of bio-oils obtained from the pyrolysis of grape seeds and cornelian cherry stones at [5](#page-5-0)00 $^{\circ}$ C are presented in Figs. 5 and [6](#page-6-0), respectively. The main peaks were demonstrated on the chromatograms. Small peaks on the chromatograms or peaks belonging to unidentified compounds are not presented in the figures. The main components in bio-oils for both grape seeds and cornelian cherry stones were oxygenated compounds such as phenol, 2-furanmethonol, 2-methoxyphenol, 4-ethyl-2-methoxyphenol, 2-methoxy-4 propylphenol, 2-ethylphenol, and n-hexadecanoic acid. The formation of phenolic compounds arose predominantly via cleavages of C–O and C–C bond networks in lignophenols (intermediate products of lignin). The following nitrogencontaining compounds were observed in bio-oils from grape

Fig. 5 Total ion chromatograms for the bio-oil obtained from the pyrolysis of grape seeds at 500 °C. I 2-Furanmethanol, 2 p-xylene, 3 1-nonene, 4 2-methyl-2-cyclopenten-1-one, 5 1,2,3-trimethylbenzene, 6 1-ethyl-2-methylbenzene, 7 phenol, 8 decane, 9 butylbenzene, 10 2-methylphenol, 11 3-methylphenol, 12 2-methoxyphenol, 13 undecane, 14 2-ethylphenol, 15 pentylbenzene, 16 naphthalene, 17 2-methoxy-4-methylphenol, 18 dodecane, 19 2,3,5-trimethylphenol, 20 4-ethyl-2-methoxyphenol, 21 tridecane, 22 (E)-2-tetradecene, 23

seeds: (Z)-13-docosenamide and octadecanamide. The compound oleanitrile was observed in bio-oils derived from both grape seeds and cornelian cherry stones. Long-chain hydrocarbons (not including oxygen) such as decane, tridecane, tetradecane, pentadecane, and others were observed in bio-oils produced from grape seeds and cornelian cherry stones. The following compounds were also observed in biooils: o-xylene; p-xylene; butylbenzene; 1,2,3-trimethylbenzene; 1-ethyl; and 2-methylbenzene. Fabbri and co-workers produced bio-char from biomass using pyrolysis gas chromatography (py-GC–MS), and they reported that benzene derivatives mainly originated from the decomposition of char the fraction (Fabbri et al. [2012\)](#page-11-0). The formation of benzene derivates can be explained by the decomposition of bio-chars from the pyrolysis of biomasses. Our results are in agreement with this previous report (Fabbri et al. [2012\)](#page-11-0).

Fatty acids (FA) and fatty acid esters (FAE) were observed in bio-oil from both grape seeds and cornelian cherry stones. The observed FAs from grape seeds were n-hexadecanoic acid, (Z, Z)-9,12-octadecadienoic acid, and oleic acid. FAs from the cornelian cherry stones contained n-hexadecanoic acid and octadecanoic acid. FAEs from the grape seeds were hexadecanoic acid methyl ester and hexadecanoic acid ethyl ester. FAEs from the cornelian cherry stones were 8-octadecanoic acid methyl ester,

Tetradecane, 24 cyclopentadecane, 25 1-pentadecene, 26 1-hexadecene, 27 hexadecane, 28 cyclotetradecane, 29 1-heptadecene, 30 heptadecane, 31 5-octadecyne, 32 octadecane, 33 2-heptadecanone, 34 hexadecanoic acid, methyl ester, 35 n-hexadecanoic acid, 36 hexadecanoic acid, ethyl ester, 37 oleanitrile, 38 2-nonadecanone, 39 (Z,Z)-9,12-octadecadienoic acid, 40 leic acid, 41 (Z)-13-docosenamide, 42 octadecanamide, 43 stigmastan-3,5-diene

octadecanoic acid methyl ester, and hexadecanoic acid bis (2-ethylhexyl) ester. FAs and FAEs in the bio-oils are responsible for the decomposition of extracts.

Elemental contents and heating values of bio-chars obtained from the pyrolysis of cornelian grape seeds and cherry stones are shown in Tables [2](#page-6-0) and [3,](#page-7-0) respectively. Bio-chars included higher carbon contents and lower oxygen contents than those of their corresponding raw materials. Both carbon contents and heating values of biochars from cornelian cherry stones were higher than those from grape seeds under identical conditions. H/C atomic ratios of bio-chars for both of the biomasses were diminished when the pyrolysis temperature increased. The lowest H/C ratio was attained at the highest pyrolysis temperature which indicates that the aromatic contents of bio-chars were the highest. When the aromatic contents of bio-chars were compared, the contents from cornelian cherry stones were greater than those from grape seeds under identical conditions.

The heating value is an important factor which affects the quality of bio-chars. Biomass type (McHenry [2009](#page-12-0)), retention time (Marousek et al. [2014b](#page-12-0)), and heating rates (Angin [2013\)](#page-11-0) are some significant pyrolysis parameters that change the heating values. In addition, new techniques in pyrolysis systems can allow for the production of

Fig. 6 Total ion chromatograms for the bio-oil obtained from the pyrolysis of cornelian cherry stones at 500 $^{\circ}$ C. *I* Toluene, 2 furfural, 3 2-furanmethanol, 4 o-xylene, 5 2-methyl-2-cyclopenten-1-one, 6 5-methylfurfural, 7 phenol, 8 butylbenzene, 9 2-methylphenol, 10 2-methoxyphenol, 11 2-Methoxy-4-methylphenol, 12 1,2-benzenediol, 13 3-methoxy-1,2-benzenediol, 14 4-ethyl-2-methoxyphenol, 15 2-methoxy-4-vinylphenol, 16 2,6-dimethoxyphenol, 17 2-methoxy-4 propylphenol, 18 tetradecane, 19 1-pentadecene, 20 pentadecane, 21

1-hexadecene, 22 8-heptadecene, 23 heptadecane, 24 1-(4-hydroxy-3,5-dimethoxyphenyl)ethanone, 25 pentadecanenitrile, 26 hexadecanoic acid, methyl ester, 27 n-hexadecanoic acid, 28 oleanitrile, 29 8-octadecenoic acid, methyl ester, 30 octadecanoic acid, methyl ester, 31 (Z)-9,17-octadecadienal, 32 octadecanoic acid, 33 hexanedioic acid, bis(2-ethylhexyl) ester, 34 2-(3,4-himethoxyphenyl)-7-hydroxy-4H-1-benzopyran-4-one, 35 5-alpha-ergost-8(14)-ene

Table 2 Elemental composition (wt%) of the bio-chars obtained from grape seeds at different temperatures

Temperature $(^{\circ}C)$		H	N	O ^a	H/C	O/C	N/C	HHV^b (MJ kg ⁻¹)
300	64.81	5.16	2.25	27.79	0.96	0.32	0.030	24.31
400	66.55	5.02	2.26	26.17	0.90	0.29	0.029	24.99
500	74.40	3.57	2.43	19.60	0.58	0.20	0.028	26.75
600	78.39	2.53	2.13	16.95	0.39	0.16	0.023	27.08
700	79.94	1.71	1.87	16.48	0.26	0.15	0.020	26.52
800	82.62	1.37	1.68	14.33	0.20	0.13	0.017	27.32

^a By difference

^b Higher heating value (HHV) calculated by the Dulong formula [HHV = $0.338C + 1.428(H-O/8) + 0.095S$]

profitable bio-chars. In a previous study reported by Marousek and colleagues, it was shown that the production of high-quality bio-chars is possible from anaerobically fermented residues in designated continuous pyrolysis systems (Marousek et al. [2014b](#page-12-0)).

The highest heating values were 32.44 for bio-char from cornelian cherry stones and 27.32 MJ kg⁻¹ for bio-char from grape seeds. The pyrolysis process that was applied to both cornelian cherry stones and grape seeds led to deoxygenation in bio-chars as O/C ratios were lower than their corresponding biomasses.

The functional groups in bio-chars from the pyrolysis of grape seeds and cornelian cherry stones (shown in Fig. [7\)](#page-7-0) revealed that the functional groups in bio-chars were similar and that they started to disappear with the increase in pyrolysis temperatures. Some of the functional groups observed in bio-chars at 300 and 400 $^{\circ}$ C for both grape seeds and cornelian cherry stones are as follows: the peaks at 2,918 and 2,853 cm^{-1} are assigned to C–H vibrations, saturated C–C, and C–H; the peaks at 1,407 and 1,555 cm^{-1} confirmed the aromatic ring C=C stretching vibration. The peak at $1,035$ cm⁻¹ represents C-H out-of-plane bending vibrations

Temperature $(^{\circ}C)$	C	H	N	O ^a	H/C	O/C	N/C	HHV^b (MJ kg ⁻¹)
300	75.15	4.68	0.43	19.74	0.75	0.20	0.005	28.56
400	81.28	3.57	0.38	14.77	0.53	0.14	0.004	29.93
500	90.14	2.62	0.41	6.83	0.35	0.06	0.004	33.00
600	93.05	1.62	0.35	4.98	0.21	0.04	0.003	32.88
700	94.59	0.75	0.12	4.54	0.10	0.04	0.001	32.23
800	96.22	0.36	0.12	3.30	0.04	0.03	0.001	32.44

Table 3 Elemental composition (wt%) of the bio-chars obtained from cornelian cherry stones at different temperatures

 \overline{a} By difference

^b Higher heating value (HHV) calculated by the Dulong formula [HHV = $0.338C + 1.428(H-O/8) + 0.095S$]

Fig. 7 FTIR spectrums of raw material and bio-chars from grape seeds (a) and cornelian cherry stones (b) after pyrolysis at 300, 400, 500, 600, 700, and 800 \degree C

Fig. 8 SEM images of a grape seeds; bio-chars after pyrolysis at b 300 °C, c 400 °C, d 500 °C, e 600 °C f 700 °C, and g 800 °C

of the alkene groups. This peak was also observed at 500 °C. After 500 \degree C, a baseline sloping down to the right was observed and peaks were distorted at the temperatures of 600, 700, and 800 °C. These results demonstrate that the diamond ATR technique is not suitable for the analysis of char samples obtained at high temperatures.

SEM images of bio-chars obtained from grape seeds and cornelian cherry stones are shown in Figs. 8 and [9,](#page-10-0) respectively. The surfaces of bio-chars were altered with the increase in temperatures for both biomass samples. In case of bio-chars from grape seeds, it is clearly seen that the formation of pores started at 300 $^{\circ}$ C which is due to the

Fig. 8 continued

first release of volatiles, mainly cellulose and hemicellulose in the biomass. At 400 $^{\circ}$ C, the pores were distinct. After further increases in temperature to 500 and 600 $^{\circ}$ C, it is believed that the pores were closed by the volatile contents of lignin factions. After $600 \degree C$, the cavities were reformed which can be explained by the complete removal of lignin-derived volatiles in the biomass.

In the case of bio-chars from cornelian cherry stones, crevices and small pores were observed at 300 °C. Likewise, cracks and slits were observed at 400 $^{\circ}$ C. When the temperature rose to 500 °C , pores were observed. A number of pores with several dimensions were observed at 600, 700, and 800 $^{\circ}$ C. It is believed that volatiles were completely removed at temperatures between 600 and 800 °C, and this occurrence caused the formation of cavities.

Conclusion

Pyrolysis of two different biomass samples was carried out at different temperatures from 300 to 800 C. The results show that product distributions under identical conditions were strongly affected by the types of biomass samples. The bio-oil yields reached their peaks at 500 for cornelian cherry stones and 700 $^{\circ}$ C for grape seeds. The highest bioyields were 47.50 wt% for cornelian cherry stones and 41.04 wt% for grape seeds. The contents of bio-oils for both cornelian cherry stones and grape seeds were similar and mainly composed of oxygenated hydrocarbons. Hydrocarbons that did not contain oxygen were also observed. Elemental contents and heating values varied for different raw materials. Both carbon contents and heating values of cornelian cherry stones were higher than those of

Fig. 9 SEM images of a cornelian cherry stones; bio-chars after pyrolysis at b 300 °C, c 400 °C, d 500 °C, e 600 °C, f 700 °C, and g 800 °C

grape seeds at the same pyrolysis temperatures. Bio-chars contained less oxygen than their corresponding raw materials. The surface topography was varied at different pyrolysis temperatures. The formation of many large and small cavities on the surface of the bio-chars from grape seeds was apparent at pyrolysis temperatures of 700 and 800 °C.

In the current paper, it was shown that waste biomasses can be converted into profitable bio-products with high economic value including bio-oils and bio-chars. The

Fig. 9 continued

production of valuable bio-products from waste biomasses is important in terms of environmental concerns and economical factors.

References

- Amen-Chen C, Pakdel H, Roy C (2001) Production of monomeric phenols by thermochemical conversion of biomass: a review. Bioresour Technol 79:277–299. doi[:10.1016/S0960-8524\(00\)00180-2](http://dx.doi.org/10.1016/S0960-8524(00)00180-2)
- Angin D (2013) Effect of pyrolysis temperature and heating rate on biochar obtained from pyrolysis of safflower seed press cake. Bioresour Technol 128:593–597. doi:[10.1016/j.biortech.2012.10.150](http://dx.doi.org/10.1016/j.biortech.2012.10.150)
- Bridgwater AV (2004) Biomass fast pyrolysis. Therm. Sci. 8(2):21–49. doi:[10.2298/TSCI0402021B](http://dx.doi.org/10.2298/TSCI0402021B)
- Demiral I, Eryazıcı A, Sensoz S (2012) Bio-oil production from pyrolysis of corncob (Zea mays L.). Biomass Bioenerg 36:43–49. doi[:10.1016/j.biombioe.2011.10.045](http://dx.doi.org/10.1016/j.biombioe.2011.10.045)
- Fabbri D, Torri C, Spokas KA (2012) Analytical pyrolysis of synthetic chars derived from biomass with potential agronomic application (biochar). Relationships with impacts on microbial carbon dioxide production. J Anal Appl Pyrol 93:77–84. doi:[10.](http://dx.doi.org/10.1016/j.jaap.2011.09.012) [1016/j.jaap.2011.09.012](http://dx.doi.org/10.1016/j.jaap.2011.09.012)
- Figueiredo MK-K, Romeiro GA (2009) Low temperature conversion (LTC) of castor seeds—a study of the oil fraction (pyrolysis oil). J Anal Appl Pyrol 86:53–57. doi:[10.1016/j.jaap.2009.04.006](http://dx.doi.org/10.1016/j.jaap.2009.04.006)
- Grierson S, Strezov V, Shah P (2011) Properties of oil and char derived from slow pyrolysis of Tetraselmis chui. Bioresour Technol 102:8232–8240. doi[:10.1016/j.biortech.2011.06.010](http://dx.doi.org/10.1016/j.biortech.2011.06.010)
- Houshfar E, Wang L, Vaha- Savo N, Brink A, Lovas T (2014) Characterisation of CO/NO/SO2 emission and ash-forming elements from the combustion and pyrolysis process. Clean Technol Environ Policy. doi:[10.1007/s10098-014-0762-3](http://dx.doi.org/10.1007/s10098-014-0762-3)
- Huang Y, Wei Z, Qiu Z, Yin X, Wu C (2012) Study on structure and pyrolysis behavior of lignin derived from corncob acid hydrolysis residue. J Anal Appl Pyrol 93:153–159. doi:[10.1016/j.jaap.](http://dx.doi.org/10.1016/j.jaap.2011.10.011) [2011.10.011](http://dx.doi.org/10.1016/j.jaap.2011.10.011)
- Huber GW, Iborra S, Corma A (2006) Synthesis of transportation fuels from biomass: chemistry, catalysts, and engineering. Chem Rev 106:4044–4098. doi[:10.1021/cr068360d](http://dx.doi.org/10.1021/cr068360d)
- Jin F, Enomoto H (2011) Rapid and highly selective conversion of biomass into value-added products in hydrothermal conditions: chemistry of acid/basecatalysed and oxidation reaction. Energy Environ Sci 4:382–397. doi[:10.1039/C004268D](http://dx.doi.org/10.1039/C004268D)
- Jourabchi SA, Gan S, Ng HK (2014) Pyrolysis of Jatropha curcas pressed cake for bio-oil production in a fixed-bed system. Energy Convers Manage 78:518–526. doi[:10.1016/j.enconman.2013.11.](http://dx.doi.org/10.1016/j.enconman.2013.11.005) [005](http://dx.doi.org/10.1016/j.enconman.2013.11.005)
- Kader MA, Islam MR, Parveen M, Haniu H, Takai K (2013) Pyrolysis decomposition of tamarind seed for alternative fuel. Bioresour Technol 149:1–7. doi:[10.1016/j.biortech.2013.09.032](http://dx.doi.org/10.1016/j.biortech.2013.09.032)
- Lievens C, Yperman J, Cornelissen T, Carleer R (2008) Study of the potential valorisation of heavy metal contaminated biomass via phytoremediation by fast pyrolysis: part II: characterisation of the liquid and gaseous fraction as a function of the temperature. Fuel 87:1906–1916. doi[:10.1016/j.fuel.2007.10.023](http://dx.doi.org/10.1016/j.fuel.2007.10.023)
- Marousek J (2013a) Two-fraction anaerobic fermentation of grass waste. J Sci Food Agric 93:2410–2414. doi:[10.1002/jsfa.6046](http://dx.doi.org/10.1002/jsfa.6046)
- Marousek J (2013b) Removal of hardly fermentable ballast from the maize silage to accelerate biogas production. Ind Crop Prod 44:253–257. doi:[10.1016/j.indcrop.2012.11.022](http://dx.doi.org/10.1016/j.indcrop.2012.11.022)
- Marousek J (2014) Significant breakthrough in biochar cost reduction. Clean Technol Environ Policy. doi[:10.1007/s10098-014-0730-y](http://dx.doi.org/10.1007/s10098-014-0730-y)
- Marousek J, Haskova S, Zeman R, Vanickova R (2014a) Managerial preferences in relation to financial indicators regarding the mitigation of global change. Sci Eng Ethics. doi:[10.1007/s11948-014-9531-2](http://dx.doi.org/10.1007/s11948-014-9531-2)
- Marousek J, Zeman R, Vanickova R, Haskova S (2014b) New concept of urban green management. Clean Technol Environ Policy. doi:[10.1007/s10098-014-0736-5](http://dx.doi.org/10.1007/s10098-014-0736-5)
- McHenry MP (2009) Agricultural bio-char production, renewable energy generation and farm carbon sequestration in Western Australia: certainty, uncertainty and risk. Agric Ecosyst Environ 129:1–7. doi:[10.1016/j.agee.2008.08.006](http://dx.doi.org/10.1016/j.agee.2008.08.006)
- Nayan NK, Kumar S, Singh RK (2013) Production of the liquid fuel by thermal pyrolysis of neem seed. Fuel 103:437–443. doi:[10.](http://dx.doi.org/10.1016/j.fuel.2012.08.058) [1016/j.fuel.2012.08.058](http://dx.doi.org/10.1016/j.fuel.2012.08.058)
- Ngo T-A, Kim J, Kim S–S (2013) Fast pyrolysis of palm kernel cake using a fluidized bed reactor: design of experiment and characteristics of bio-oil. J Ind Eng Chem 19:137–143. doi:[10.](http://dx.doi.org/10.1016/j.jiec.2012.07.015) [1016/j.jiec.2012.07.015](http://dx.doi.org/10.1016/j.jiec.2012.07.015)
- Putun E, Uzun BB, Putun AE (2006) Fixed-bed catalytic pyrolysis of cotton-seed cake: effects of pyrolysis temperature, natural zeolite content and sweeping gas flow rate. Bioresour Technol 97:701–710. doi:[10.1016/j.biortech.2005.04.005](http://dx.doi.org/10.1016/j.biortech.2005.04.005)
- Raja SA, Kennedy ZR, Pillai BC, Lee C (2010) Flash pyrolysis of jatropha oil cake in electrically heated fluidized bed reactor. Energy 35:2819–2823. doi:[10.1016/j.energy.2010.03.011](http://dx.doi.org/10.1016/j.energy.2010.03.011)
- Shadangi KP, Mohanty K (2014) Thermal and catalytic pyrolysis of Karanja seed to produce liquid fuel. Fuel 115:434–442. doi:[10.](http://dx.doi.org/10.1016/j.fuel.2013.07.053) [1016/j.fuel.2013.07.053](http://dx.doi.org/10.1016/j.fuel.2013.07.053)
- Shadangi KP, Singh RK (2012) Thermolysis of polanga seed cake to bio-oil using semi batch reactor. Fuel 97:450–456. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.fuel.2012.02.058) [fuel.2012.02.058](http://dx.doi.org/10.1016/j.fuel.2012.02.058)
- Torri C, Lesci IG, Fabbri D (2009) Analytical study on the production of a hydroxylactone from catalytic pyrolysis of carbohydrates with nanopowder aluminium titanate. J Anal Appl Pyrolysis 84:25–30. doi[:10.1016/j.jaap.2008.10.002](http://dx.doi.org/10.1016/j.jaap.2008.10.002)
- Ucar S, Karagoz S (2009) The slow pyrolysis of pomegranate seeds: the effect of temperature on the product yields and bio-oil properties. J Anal Appl Pyrol 84:151–156. doi:[10.1016/j.jaap.](http://dx.doi.org/10.1016/j.jaap.2009.01.005) [2009.01.005](http://dx.doi.org/10.1016/j.jaap.2009.01.005)
- Vassilev SV, Baxter D, Vassileva CG (2013) An overview of the behaviour of biomass during combustion: part I. Phase-mineral transformations of organic and inorganic matter. Fuel 112: 391–449. doi:[10.1016/j.fuel.2013.05.043](http://dx.doi.org/10.1016/j.fuel.2013.05.043)
- Volli V, Singh RK (2012) Production of bio-oil from de-oiled cakes by thermal pyrolysis. Fuel 96:579–585. doi[:10.1016/j.fuel.2012.](http://dx.doi.org/10.1016/j.fuel.2012.01.016) [01.016](http://dx.doi.org/10.1016/j.fuel.2012.01.016)