REVIEW

The implication of infection with respiratory syncytial virus in pediatric recurrent wheezing and asthma: knowledge expanded post‑COVID‑19 era

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Abstract

Background Respiratory syncytial virus (RSV) infection has been identifed to serve as the primary cause of acute lower respiratory infectious diseases in children under the age of one and a signifcant risk factor for the emergence and development of pediatric recurrent wheezing and asthma, though the exact mechanism is still unknown.

Methods and results In this study, we discuss the key routes that lead to recurrent wheezing and bronchial asthma following RSV infection. It is interesting to note that following the coronavirus disease 2019 (COVID-19) epidemic, the prevalence of RSV changes signifcantly. This presents us with a rare opportunity to better understand the associated mechanism for RSV infection, its efects on the respiratory system, and the immunological response to RSV following the COVID-19 epidemic. To better understand the associated mechanisms in the occurrence and progression of pediatric asthma, we thoroughly described how the RSV infection directly destroys the physical barrier of airway epithelial tissue, promotes infammatory responses, enhances airway hyper-responsiveness, and ultimately causes the airway remodeling. More critically, extensive discussion was also conducted regarding the potential impact of RSV infection on host pulmonary immune response.

Conclusion In conclusion, this study offers a comprehensive perspective to better understand how the RSV infection interacts in the control of the host's pulmonary immune system, causing recurrent wheezing and the development of asthma, and it sheds fresh light on potential avenues for pharmaceutical therapy in the future.

Keywords Respiratory syncytial virus · Recurrent wheezing · Pediatric asthma · Immune escape · COVID-19 pandemic

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Introduction

In young children, respiratory syncytial virus (RSV), which is spread through respiratory droplets and close contact, mostly afects the lower respiratory tract and causes symptoms including fever, coughing, asphyxia, and shortness of breath [[9\]](#page-9-0). One of the most prevalent forms of acute lower respiratory tract infections (LRTI) among children under the age of 1, particularly in infants between the ages of 2 and 6 months, is acute bronchiolitis. RSV re-infection is frequently observed because the immune defense mechanism developed as a result of prior RSV infection does not sustain long. Around 18% of pediatric inpatients under the age of 1 are hospitalized due to acute bronchiolitis triggered by RSV infection [[26\]](#page-10-0). According to a cohort study, between 30 and 40% of infants hospitalized for bronchiolitis, especially the severe variety, may develop recurrent wheezing [[34](#page-10-1)]. RSV is one of the pathogens that frequently give rise to viral illnesses such as severe bronchiolitis [\[59](#page-11-0)]. As a result of the

persistently high incidence rate of RSV bronchiolitis, RSV is a signifcant risk factor for asthma and recurrent wheeze. Hence, a connection between RSV bronchiolitis in infancy and the development of bronchial asthma in children has been proposed [[37](#page-10-2)]. The epidemiological evidence providing the association between viral bronchiolitis in childhood and subsequent recurrent wheeze or asthma was frst brought up in 1959 by Wittig H and Glaser J $[108]$. Supported by some prospective studies, RSV bronchiolitis is currently linked to recurrent wheezing, asthma, and airway remodeling. Sigurs N. et al. [\[88](#page-12-1)] conducted the longest follow-up study to date with a final cut-off age of 18 years old. This compelling evidence showed that severe RSV bronchiolitis in childhood can lead to long-term allergic asthma and airway remodeling. Together, the RSV-LRTI was considered to substantially harm the physical and mental health, especially in early life and was thought to contribute to a predisposition for the development of asthma, and may even bring about the chronic pulmonary disorders in their adult life [[18\]](#page-9-1).

The ongoing COVID-19 pandemic, as a signifcant threat to public health, has caused a surge in patients infected with SARS-CoV-2 and other common respiratory tract viruses that may lead to acute and chronic respiratory illnesses. As a result, the circumstances surrounding the infection of respiratory tract viruses are quite complex, and we need to act promptly to empower ourselves with understanding and insight. Additionally, the development of several groundbreaking public health policies have reshaped our lives and had a big impact on how many other pathogenic viruses, including RSV, were originally transmitted.

Here, this study seeks to update our understanding of the role that RSV infection plays in the occurrence and progression of recurrent wheezing, asthma, and other respiratory disorders in pediatric patients, as well as the mechanism of host-RSV immune response and immune escape, particularly in the COVID-19 pandemic.

The infection with respiratory syncytial virus and its derived pulmonary pathology

Brief introduction to the structure of respiratory syncytial virus

RSV, initially identifed from chimpanzees in 1956, mostly afects newborns and causes catastrophic lower respiratory tract disorders [[9\]](#page-9-0). RSV, as the single-stranded negative RNA envelope virus, belongs to Paramyxoviridae, Pneumoviridae, and Pneumovirus genus. The genome of RSV is composed of two non-structural proteins (NS) named NS1 and NS2, the nucleoprotein (N), phosphoprotein (P), matrix protein (M), small hydrophobic protein (SH), attachment glycoprotein (G), fusion protein (F), M2, and large protein (L). RSV has two subtypes, A and B, and these strains typically co-proliferate even if one of them predominates [[39\]](#page-10-3).

Epidemiological evidence showing lower respiratory tract infection with RSV could cause recurrent wheezing and even asthma in pediatric patients

RSV is a major contributor to pediatric lower respiratory tract infections worldwide, according to epidemiological statistics. RSV can result in 33.1 million acute LRTI cases, 3.2 million hospital admissions, and 118,000 under-5-yearold fatalities annually (mostly in underdeveloped nations). Since it is so widespread, nearly every kid is at risk for LRTI following initial RSV infection in infancy [\[92](#page-12-2)]. The most frequent RSV-related LRTI illness is bronchiolitis, which also accounts for the majority of hospitalizations in children under the age of 2 and baby deaths under the age of 1 [[29\]](#page-10-4). Additionally, RSV-related LRTI is closely linked to the development and occurrence of other chronic respiratory diseases. For instance, in patients with RSV-related LRTI in early childhood (under 3 years old), the incidence of recurrent wheezing ranges from 4 to 47%, while the incidence of asthma ranges from 8 to 76% [\[21\]](#page-9-2). The majority of investigations consistently came to the conclusion that RSV-related LRTI may increase the prevalence of asthma in children [[15\]](#page-9-3). Additionally, RSV-related LRTI in infancy has been linked to lower pulmonary function and increased airway reactivity in kids of school age [\[15](#page-9-3), [21](#page-9-2)].

SARS‑CoV‑2 sharing single‑stranded RNA nature with RSV could cause the resultant respiratory disorders

SARS-CoV-2, which gives rise to COVID-19, is a positivesense, single-stranded RNA virus [[109\]](#page-12-3), as explained in our most recent study [\[43](#page-10-5)], and it has a number of crucial parts, including the membrane (M), nucleocapsid (N), envelope (E), and spike (S) proteins [[51\]](#page-10-6). Since both RSV and SARS-CoV-2 are single-stranded RNA envelope viruses, although RSV is a negative-sense RNA virus, they still share substantial structural similarities. Additionally, SARS-CoV-2 infection may result in serious respiratory conditions. According to reports, SARS-CoV-2 infection can cause acute lung injury (ALI) and acute respiratory distress syndrome (ARDS) by triggering an initial host immune response, intense infammation, and occasionally even a catastrophic cytokine storm [\[101](#page-12-4)].

Importantly, the children population was previously widely recognized with a lower incidence and milder course of COVID-19, possibly due to less infammatory response, while the monitor of the blood cytokine levels in special proinfammatory disorders such as the Kawasaki disease and multisystem infammatory syndrome in children (MIS-C) associated with COVID-19 was recommended (B. M. [\[55](#page-11-1)]. In contrast, the studies in pediatric patient population with underlying primary immunodefciency also demonstrated the favorable outcome among immunosuppressed pediatric patients rather than other comorbidities (B. M. [[54](#page-11-2)], which indicated that a weaker infammatory response might play a protective role in this condition. Hence, in the parts that followed, RSV and SARS-CoV-2 infection were discussed side by side. The major routes to trigger pediatric asthma following RSV infection were discussed, and its efects on immunological response in the respiratory system were especially emphasized.

The major routes of respiratory syncytial virus to trigger pediatric asthma

RSV infection is linked to the majority of acute asthma attacks; it is also thought to be an independent risk factor for recurrent asthma attacks and exacerbations, which can worsen asthma through promoting an infammatory response, airway hyper-responsiveness, and airway remodeling [\[76](#page-11-3), [87](#page-12-5)]. The aforementioned factors are now thought to be the primary means that an RSV infection causes recurrent wheezing and bronchial asthma. The physical barrier of the airway epithelial tissue may be immediately destroyed by RSV invasion, which may also encourage infammatory and hyper-responsive airway responses. As a result, we went into each of these pathogenic episodes individually in this review study.

RSV invasion could directly destruct the physical barrier of airway epithelial tissue

The primary target cells for RSV invasion and the host's initial line of defense against the infection are airway epithelial cells. The airway epithelium is frequently reported to be impaired in various pulmonary illnesses, such as asthma [\[105\]](#page-12-6). RSV infection could cause necrosis and exfoliation of airway epithelial cells, infltration of peripheral lymphocytes, hyperplasia of glands, increased secretion of mucus, or even severe airway obstruction leading to emphysema, atelectasis, etc. The main symptoms of RSV infection are wheezing and dyspnea, which are mostly caused by airway blockage [[25](#page-10-7)].

The surface mucus layer and the apical junction complex between individual airway epithelial cells form a barrier defense against the invasion of harmful bacteria. Tight connections and sticky junctions, which make up the apical junction complex of the epithelium, are crucial for controlling the permeability of the barrier and preserving cell polarization [\[25](#page-10-7)]. RSV infection may cause damage to the airway barrier in asthmatic airway epithelial cells, according to research by Kast J et al. on the infuence of RSV infection on the control of tight junctions [\[44\]](#page-10-8). According to studies conducted by Masaki T. et al. and Tsutsumi H. et al., RSV infection could upregulate the tight junction-related proteins claudin-4 and occluding and ZO-1, raise the cell polarity, thus promote virus budding in human nasal epithelial cells [[63,](#page-11-4) [98\]](#page-12-7).

Mucins, which are signifcant elements of the mucus layer lining the airways, include MUC5AC, MUC1, MUC5B, and others [[8\]](#page-9-4). The innate immune defense is strengthened by the released mucins' interactions with other mucus elements like immunoglobin A (IgA) and lectins [\[6](#page-9-5)]. It was noted that RSV infection resulted in the loss of MUC5AC's negative regulation because its target miRNA, hsa-miR-34b/c, was downregulated [[19\]](#page-9-6). According to other research, RSV infection signifcantly increased the secretion of transforming growth factor (TGF) and extracellular matrix protein, and TGF- triggered epithelial-mesenchymal transition (EMT), which produced extracellular matrix by secreting collagen and fbronectin and increased the expression of MMPs to support airway remodeling [\[114](#page-13-0)]. The homeostasis of epithelial growth and repair is supported by the expression of various mediators, such as matrix metalloproteinase (MMP), cytokines, growth factors, and others by airway epithelial cells themselves $[106]$ $[106]$. These mediators also play a significant role in the secretion of mucin and the control of tight junctions between epithelial cells [[8](#page-9-4), [73,](#page-11-5) [83,](#page-12-9) [95\]](#page-12-10). The repair of the bronchial epithelium was found to be delayed by RSV, similar to many other respiratory viruses. The activity of the matrix degradative enzymes MMP 2 and MMP 9 may rise in response to RSV infection, interfering with the normal repair process and increasing the synthesis of the fbroblast growth factor (FGF) and epidermal growth factor (EGF) [[48](#page-10-9), [106](#page-12-8)].

RSV infection could promote airway infammatory responses

In addition to being the weakened defense after virus infection, airway epithelial cells play a signifcant role in the beginning of infammation. RSV infection primarily afects airway epithelial cells (AECs) at the bronchial, bronchiolar, and alveolar levels [[71](#page-11-6)]. The RSV infection invades the airway epithelial cells, damages the airway mucosal barrier, and causes airway infammation by secreting a number of cytokines, chemokines, and infammatory mediators. It also contributes to the development of bronchial asthma and acute attacks.

Everard M et al. [[20](#page-9-7)] discovered neutrophil and eosinophil infltration in the airway in the nasal secretion of children infected with RSV, while RSV infection could drive the airway epithelial cells to secrete massive cytokines [\[75](#page-11-7)], such as interferon (IFN)-γ, interleukin (IL)-4, IL-5, IL-10, IL-12, and IL-13, chemokines such as CCL2, CCL3, and CCL4, CXC chemokine ligand (CXCL) 8, CXCL10, and CXCL11, and infammatory mediators such as cysteine leukotrienes (CysLTs), which in turn causes multiple infammatory cells to infltrate and generate infammatory response, leading to the condition of local airway infammation, and eventually participating in the occurrence and acute attack of bronchial asthma. RSV infects AECs, recruits and activates neutrophils and NK cells via production of tumor necrosis factor (TNF)- α , CXCL8 (also named as IL-8), and inducible T cell chemokine (ITAC/CXCL11), promotes secretion of chemokines including macrophage infammatory protein (MIP) 1α/CCL3, MIP1β/CCL4, MIP2, interferon-induced protein 10 (IP-10)/CXCL10, and monocyte chemokine (MCP) 1/CCL2, and induce the recruitment of monocytes and memory T cells $(Z, [32, 69]$ $(Z, [32, 69]$ $(Z, [32, 69]$ $(Z, [32, 69]$ $(Z, [32, 69]$. CysLTs, the inflammatory mediators with increased secretion caused by RSV infection, were also implicated in asthma. Hsu C et al. [[40\]](#page-10-11) discovered the signifcantly increased CysLTs in bronchoalveolar lavage fuid (BALF) and lung tissues, together with the RSV-triggered pulmonary infammation in human RSV (A2 strain) infected 6–8 weeks old Balb/c mice. Han J et al. [\[30\]](#page-10-12) observed the effective therapeutic effect of the CysLTs receptor antagonist, Montelukast, on airway infammation triggered by RSV infection. Han Z et al. [\[32\]](#page-10-10) found that concentration of CXCL4 within primary diferentiated airway cells is positively correlated with the clinical severity of RSV infection. Increased CXCL4 can bind to heparan sulfate (HS), the main receptor of RSV, to block RSV attachment to host cells and alleviate pulmonary infammation. Furthermore, Zeng S et al. [[113](#page-13-1)] reported that IL-33 receptor (suppression of tumorigenicity 2, ST2) signaling is crucial in RSV-infected mouse lungs. Th2 and Th17 cytokine levels were also increased, which act on lung stromal cells to recruit more neutrophils to infected local sites [\[46](#page-10-13)].

RSV infection could exacerbate airway hyper‑responsiveness

After inhaling small amounts of irritants or allergens, some individuals may experience abnormal and excessive contractions in their airways, leading to airway stenosis and a signifcant increase in airway resistance. This condition is known as airway hyper-responsiveness (AHR). While the exact mechanism of AHR is not yet fully understood, it is widely accepted that airway infection and infammation play an important role.

It was discovered that neutrophils, which are activated by lipopolysaccharide (LPS) during airway remodeling, play a pivotal role in the augmented response to acetylcholine in the airway, leading to increased airway AHR [[80\]](#page-11-9). Besides, Zhou N et al. [\[116\]](#page-13-2) established a LPS (major component of Gram-negative bacteria, natural ligand of toll-like receptor 4,

TLR4) plus RSV infection animal model in Balb/c mice. The results demonstrated that Gram-negative bacteria–derived LPS could switch the dominant AHR mediator from IFN-γ (the Th1 type cytokine) to extracellular degradative MMP 9 in RSV-infected mice, which exacerbates the pathological AHR and airway remodeling. These pathways present new targets for drug development in the future.

It was discovered that RSV infection could afect AHR through neurological regulation. RSV was observed to induce excess production of neuropeptide substance P by upregulating the mRNA expression of neurokinin-1 (NK-1, the receptor of substance P in the airway), leading to increased AHR phenotypes including bronchoconstriction and excessive secretion of mucus [[93\]](#page-12-11). These clues suggest that balancing neuropeptide production in the airway may help alleviate AHR caused by RSV infection, providing insight into potential clinical therapy.

Various infammatory cells, mediators, and cytokines also contribute to AHR. Skewed T helper (Th) cell profling has long been emphasized in AHR. Compared with its major counterpart Th1 cell type, the Th2 cells mainly secrete interleukins such as IL-4, IL-5, IL-10, and IL-13, along with the IL-9 contributing neutrophils. They contribute to AHR by inducing the production of airway immunoglobulin E (IgE) and eosinophil infltration, increasing infammatory mediators and cells, promoting mucus secretion, afecting ciliated cell diferentiation and cilia movement, triggering smooth muscle cell contraction, and depressing the reactivity of the β adrenergic receptor in airway smooth muscle cells [\[117](#page-13-3)]. Saravia J et al. [[79\]](#page-11-10) discovered increased lung type 2 innate lymphoid cells (ILC2) and IL-33 (another Th2 cytokine) [[33,](#page-10-14) [52](#page-11-11)], along with accumulated eosinophils, resulting in AHR in lung homogenate in acute RSV-infected neonatal mice.

RSV infection could eventually cause airway remodeling

Airway remodeling is a prominent feature of pediatric bronchial asthma, and RSV infection has been found to induce overexpression of vascular endothelial growth factor (VEGF), transforming growth factor β1 (TGF-β1), and insulin-like growth factor 1 (IGF-1) as biomarkers of airway remodeling [[103\]](#page-12-12). The role of VEGF in regulating airway repair and remodeling has been experimentally validated. For example, Lee C et al. found that VEGF induces an asthma-like phenotype characterized by infammation, parenchymal and vascular remodeling, edema, mucus metaplasia, myofbroblast proliferation, and airway hyperresponsiveness in lung-specifc VEGF165 transgenic mice [[49\]](#page-10-15). TGF- β 1, a fibrogenic cytokine, can promote fibroblast transdiferentiation into myofbroblasts, myofbroblast proliferation, and airway smooth muscle cell migration toward the epithelial layer, thereby facilitating the process of tissue remodeling [\[42](#page-10-16)]. IGF-1 can increase the content of collagen and elastin, the major components of pulmonary extracellular matrix, and increase smooth muscle thickness, thus participating in the repair of lung tissue injury and airway remodeling (Y. Y. [\[31](#page-10-17)]. It is therefore understandable that the induced expression of these three remodeling factors following RSV infection in vivo could result in lasting tissue remodeling and physiological consequences.

New situation of RSV infection prevalence and RSV‑related LRTI post‑COVID‑19 era (Fig. [1\)](#page-4-0)

During the past few years of the COVID-19 pandemic, the prevalence of RSV infections has sharply decreased. This can be explained by the fact that RSV and SARS-CoV-2 share the same mode of aerosol transmission, which made it easier to understand how the restrictive measures of public health implemented worldwide had a signifcant impact on the prevalence of RSV [\[17](#page-9-8)]. For example, in Belgium, it was reported that the number of registered RSV cases signifcantly decreased $(>99\%)$ compared to 2019 [\[100](#page-12-13)]. Additionally, RSV, being a seasonal virus, typically peaks between January and February in Poland and other countries in the northern hemisphere [[13\]](#page-9-9). However, during the COVID-19 pandemic, the epidemiological profle of RSV throughout the world underwent a drastic change, and the annual RSV epidemic seemed to be absent in both the winter and spring of 2020–2021 [[14](#page-9-10), [90](#page-12-14), [99\]](#page-12-15). This conclusion was supported by the data from Stamm P et al. [[90\]](#page-12-14), who carried out pointof-care test (POCT)-PCR detecting RSV and SARS-CoV-2 in pediatric emergency rooms during December 1st, 2018, to March 31th, 2021, and compared the RSV incidence before and after COVID-19 epidemic. Moreover, Manti S et al. [\[60\]](#page-11-12) analyzed the epidemiological data of pediatric RSV-LRTI from hospitalized children below 2 years old from October 2018 to December 2021. In summary, it is widely believed that the RSV-LRTI epidemic has been suppressed thanks to COVID-19-related public health measures and changes in population-level social behavior patterns.

To further compare the disease state diferences between hospitalized pediatric patients with COVID-19 and RSV, Fedorczak A et al. [[22\]](#page-9-11) studied children aged 36 months admitted from September 2021 to March 2022 (52 cases with COVID-19 vs. 43 cases with RSV). They compared the clinical manifestations, treatments, and length of hospital stays between the two groups. The RSV group displayed the higher incidence of major clinical manifestation including

Fig. 1 One illustration depicts the current state of the prevalence of RSV infection in the post-COVID-19 period. RSV-LRTI, respiratory syncytial virus–lower respiratory tract infection; COVID-19, corona-

virus disease 2019; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2. Created with BioRender.com

cough, rhinitis, short of breath, moist rales, and wheezing, while the COVID-19 group displayed higher incidence of fever and seizure. Furthermore, the RSV patients required more bronchodilator or oxygen therapy, as well as longer hospital stay. These results suggest that infants infected with RSV may display more severe clinical manifestations than those infected with COVID-19, as the RSV-infected patients required longer hospital stays and more subtle treatment. However, it is important to note that these fndings were unexpected, and it is possible that the lower number of RSV cases during the COVID-19 epidemic could have contributed to a "survivor bias" in the data.

Considering the similar single-stranded RNA nature of RSV and SAR-CoV-2 viruses, it is reasonable to speculate whether the prevalence and severity of respiratory infection caused by RSV might be afected by SARS-CoV-2 infection. Concerns have been raised about the potential coinfection of RSV and SARS-CoV-2 contributing to more severe disease conditions and higher mortality rates. To investigate this possibility, Swets M and colleagues analyzed the clinical outcomes of 6965 hospitalized adults in England between February 6th, 2020, and December 8th, 2021, among whom 220 individuals were co-infected with RSV and SARS-CoV-2. Surprisingly, no signifcant diference was found in disease severity, as indicated by the need for medical support, such as invasive mechanical ventilation, between those with co-infections and those with single SARS-CoV-2 infection [\[91](#page-12-16)]*.*

The mechanism of host‑RSV immune response and immune escape

Innate immune response following RSV infection

During RSV invasion, the innate immune response is quickly activated as the frst line of defense against foreign pathogens, which trigger various types of pattern recognition receptors (PRRs), including TLRs and retinoid inducible gene-1 like receptors (RLRs), to stimulate the production of type I IFNs and recruitment of immune cells for defense action. Type I IFNs play antiviral roles by inhibiting protein synthesis and increasing pro-apoptosis factors, which safeguard the host by preventing the spread of infection by halting the release of newly replicated progeny virus. However, excessive production of type I interferon can also generate pro-infammatory mediators. Therefore, the type I IFN response in respiratory mucosa is linked to recurrent wheezing in the frst 2 years of life in infants with acute RSV infection [\[37\]](#page-10-2).

Among various TLRs, the TLR3 and TLR4 are expected to play a role during RSV infection based on the ligands they sense. During the RSV infection, TLR3 can sense the dsRNA intermediates during RSV replication, while TLR4 recognizes the RSV-F protein. According to Groskreutz D et al. [[27](#page-10-18)], the expression of both TLR3 mRNA and protein, as well as its membrane distribution, increases in A549 cell lines and human bronchial epithelial cells infected with RSV. Both TLR3 and TLR4 could initiate the expression of type I IFN including IFN-α and IFN-β through transcriptional regulation via myeloid diferentiation factor 88 (MyD88)–independent mediation of IFN regulatory factor 3 (IRF3) [\[110](#page-12-17)]. The significant secretion of IFN- α and IFN- β provides local antiviral benefts, efectively restraining virus replication at the infected site [[104\]](#page-12-18). Interferon type I demonstrates antiviral activity by inhibiting protein synthesis and promoting apoptosis factors; however, excessive expression of type I interferon can also generate pro-infammatory mediators [\[97\]](#page-12-19). Compared with rhinovirus infection, pediatric patients with RSV bronchiolitis exhibit greater expression of IFN-I and IFN-III, and increased IFN-I expression in the nasal cavity of RSV-infected individuals correlates with disease severity [[68\]](#page-11-13).

The TLR3/TRIM56/GRP78/TRIF/IFN signaling plays crucial role in the antiviral defense and consequential immunological response potentiated by the sensing of extracellular dsRNA (Y. [[86,](#page-12-20) [107\]](#page-12-21). RSV infection was found to induce the infammatory cytokines and chemokines such as CCL5 production directly through TLR3 signaling pathways, independent on the interferon (IFN) receptor signaling in the human epithelial cells [\[78](#page-11-14)]. Previous studies have also emphasized the critical role of TLR4 in RSV-induced innate immune responses, further validated in TLR4 knockout mice [\[36](#page-10-19)]. TLR4 specifcally recognizes RSV-F protein, activating nuclear factor-kappa B (NF-κB) signaling and the secretion of inflammatory-related molecules [\[47\]](#page-10-20). NF-κB activation via nuclear translocation leads to transcription of downstream pro-infammatory factors and cytokines [\[28](#page-10-21)]. TLR4 senses RSV-F protein to induce human neutrophils to facilitate the formation of the extracellular traps of neutrophils (NETs) to capture RSV, and the NETs will form dense mucus emboli distributed all over the small airways and lungs, leading to airway obstruction and wheezing [\[24](#page-10-22)]. RSV SH protein, which is involved in the pathogenesis of human asthma, can activate TLR4 to promote the release of IL-1β and IL-18, thereby mediating infammatory responses [[62\]](#page-11-15).

RLR, consisting of retinoid inducible gene 1 (RIG1) and MDA5, serves as a critical cytoplasmic receptor for intracellular viral RNA. In children infected with RSV, RLR expressed by airway epithelial cells, dendritic cells (DCs), macrophages, and lymphocytes, facilitates the activation of the transcription factor NF-κB and regulates the expression of infammatory cytokines and type I interferon. RIG1 detects viral RNA products and subsequently interacts with the mitochondrial protein mitochondrial antiviral signaling protein (MAVS). This leads to the phosphorylation of the transcription factor IRF3 by IRF3 kinase and the dimerization and translocation of activated IRF3 to induce the transcription activation of IFN-β [[102](#page-12-22)]. It was reported that the relative gene expression level of melanoma diferentiation–associated protein 5 (MDA5) and RIG1 in nasopharyngeal washes from RSV-infected infants was signifcantly higher than that in RSV free infants. Additionally, RIG1 mRNA expression level is positively correlated with RSV viral load [\[81](#page-11-16)].

Exposure by other respiratory viruses like infuenza (the similar negative-sense RNA virus like RSV) was found to upregulate the expression of various antiviral genes (including RIG1, TLR3, and MDA5, the above-mentioned important players in viral defense) in mouse bone marrow–derived eosinophils [[50\]](#page-10-23). However, other fndings suggest some nonstructural protein components of RSV might play sophisticated "pro-viral" functions by fne-tuning the IFN response following viral infection [[64](#page-11-17)]. Whether RSV infection could boost the innate immune response in its infected and recruited various types of immune cells or not is still called for further investigation.

Furthermore, previous research has revealed that NF-κB is a crucial factor in the activation of NLRP3 infammasomes during RSV infection. These two factors are positively correlated with each other. As we comprehend, during virus infection and lung injury, the excessive activation of infammasomes causes the generation of a large number of infammatory factors in the airway, leading to exacerbation of the existing lung injury (C. [[85](#page-12-23)]. Though the latency of RSV infection is relatively short, it is still recommended to consider both the acute infection and the latent reactivation phase when interpreting RSV infection–induced intersection of innate immune pathways and the interactive immune response [\[16](#page-9-12)].

Adaptive immune response following RSV infection

During an asthmatic attack, the imbalance between Th1/ Th2 [\[84](#page-12-24)] and Th17/Treg [\[74](#page-11-18)] pathways is considered a signifcant mechanism, which also exists in patients infected with RSV. The G protein is crucial to Th2 cytokine bias in RSV-induced pulmonary disorders, promoting the production of IL-4 and IL-5 by Th2 and inhibiting the production of IFN- γ by Th1 [[10\]](#page-9-13). Following RSV infection, the suppressed Th2 population becomes out of control as secreted IFN-γ decreases and highly expressed IL-4 induces recurrent wheezing and even asthma attacks [\[41](#page-10-24)].

The interaction between RSV-G protein and host CX3CR1 is a crucial event during the adaptive immune response of airway epithelial infammation and infection. The G protein can bind with chemokine receptor CX3CR1 to induce a strong cytokine/chemokine response, including the production of IL-6, IL-8, MIP-1 α , and MCP-1, which act on the host-RSV response [[12\]](#page-9-14).

IL-8 is produced by nasal mucosa epithelial cells, HBECs, and recruited neutrophils [[94,](#page-12-25) [112\]](#page-13-4). Besides chemotaxis of neutrophils, IL-8 can induce T lymphocytes to migrate to infammatory sites by itself [[58\]](#page-11-19), and also make the recruited neutrophils release chemotactic substances and mediate the migration of T lymphocytes [[66\]](#page-11-20). Previous studies [\[1](#page-9-15)] have verifed that RSV infection can cause excessive release of IL-8 in airway epithelium, mainly involved in infammatory diseases dominated by neutrophil infltration. This evidence indicates that RSV infection in bronchial epithelial cells induces IL-8 release, which may contribute to susceptibility to RSV infection in asthma.

Interactive immune response following RSV infection

In various TLRs as important player in innate immune system, double-stranded RNA (dsRNA) selectively activates TLR3 and aids in the production of thymic stromal lymphopoietin (TSLP) in epithelial cells [\[45](#page-10-25)]. During the infammatory process, RSV generates numerous dsRNA intermediates during its replication within epithelial cells, stimulating TSLP secretion. Activation of TLR3 by RNA ligands signifcantly increases TSLP mRNA expression (by 66.8 times) [\[65\]](#page-11-21), which plays a crucial role in pulmonary infammation (Y. J. [[57](#page-11-22)] in BALF from both the asthmatic wild-type mice and lung-specific TSLP transgenic mice, causing notable pulmonary pathologies such as airway infammation, goblet cell hyperplasia, and airway hyperresponsiveness. Likewise, TSLP receptor-deficient mice demonstrate a tendency toward Th1 responses, failing to produce the pulmonary infammatory response triggered by allergens [[3,](#page-9-16) [115](#page-13-5)]. TSLP also accumulates in airways of asthmatic patients [\[111](#page-12-26)]. As TSLP is secreted by airway epithelium, it induces the regulation of dendritic cells in differentiating CD4+ Th0 cells into Th2 subtype, consequently leading to the release of IL-4, IL-5, IL-13, and TNF- α , rather than IL-10 and IFN- γ (Y. J. [[56](#page-11-23)]. Besides, the mechanisms such as the killer-cell immunoglobulin-like receptors–HLA interactions indicated that during viral infection, the innate receptors could also modulate adaptive T cell responses and contribute to interactive immune response [[67\]](#page-11-24).

Immune escape of RSV against human host system

The RSV-G protein binds to adhesion molecules on host cells to initiate virus attachment. The protein's variable region leads to diverse antigens, resulting in repeated infections in a single individual. The conserved region contains a CX3C-like cysteine domain, which can compete with CX3C chemokine receptors and dampen immune response mediated by immune cells [[96\]](#page-12-27).

RSV SH protein was discovered to hinder the signaling cascade of TNF-α triggered NF-κB activation $[23]$ $[23]$. The non-structural proteins NS1/NS2 synergistically inhibit the mRNA and protein expression of IFN- α and IFN- β , thus blocking their downstream signaling pathways in macrophages. Spann et al. additionally found that the RSV NS1 and NS2 proteins could obstruct interferon production through the MyD88 independent pathway mediated by TLR3 and TLR4 by interfering with the activation of IRF3, thereby outlasting the potential immune clearance from host cells [[89\]](#page-12-28). NS2 could also bind with RIG1 and inhibit its interaction with MAVS [\[53\]](#page-11-25). Besides, NS1 directly binds with the promoter of IFN- β , and sabotages its binding with calmodulin binding peptide (CBP), eventually afecting the accessibility of IFN-β promoter to transcriptional factor and cofactors such as IRF3 [[77\]](#page-11-26). In vitro experiments also demonstrate that NS1 and NS2 enhance proteasome-mediated degradation by forming a ubiquitin ligase complex, thereby diminishing the level of signal transducer and activator of transcription 2 (STAT2) and its related signaling [\[35](#page-10-27)].

RSV-infected dendritic cells (DCs) exhibit compromised assembly of the immunological synapse, possibly regulated by the N protein of RSV. Expression of N protein in DCs and lung epithelial cells is associated with the decreased antigen determinant of major histocompatibility complex (MHC) on cell surface [\[11](#page-9-17)]. It was found that human leukocyte antigen (HLA, MHC of human beings)–DR expression is suppressed on monocytes, and the reduced HLA-DR expression is correlated with greater disease severity in infected infants [\[2](#page-9-18)].

Potential mechanisms underlying host‑virus immune response and immune escape in both COVID‑19 and RSV‑LRTI (Fig. [2](#page-7-0))

After acute viral infection, pathogen-associated molecular patterns (PAMPs) from invading foreign pathogens and damage-associated molecular patterns (DAMPs) from injured host cells are sensed by PRRs such as TLRs and NLRs. This activates the NF-kB signaling and NLRP3 infammasomes [[72\]](#page-11-27). As we mentioned earlier, TLRs play a major role in RSV defense, and are also found to be activated in COVID-19 [[61\]](#page-11-28). However, TLRs and their downstream signaling can be double-edged swords and dysregulation can lead to excessive production of pro-infammatory mediators, resulting in severe respiratory complications.

Fig. 2 Similar pathways mediated in host-virus immune response and immunological escape following COVID-19 and RSV infection. COVID-19, coronavirus disease 2019; dsRNA, double-stranded

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RNA; HLA, human leukocyte antigen; RSV, respiratory syncytial virus; ssRNA, single-stranded RNA; TLR, toll-like receptor. Created with BioRender.com

The activation of NLRP3 infammasomes plays a signifcant role in the human innate immune response against viral attacks [\[7\]](#page-9-19). It has been observed that such activation of NLRP3 infammasomes is associated with an excessive immune response and a cytokine storm following SARS-CoV-2 infection [\[38\]](#page-10-28). Likewise, this hyperactive immune response can lead to intense inflammation and sometimes even a destructive cytokine storm, resulting in ALI and ARDS in COVID-19 [[70](#page-11-29)]. As previously mentioned, the production of a massive amount of pro-infammatory cytokine resulting from TLRs mediated over activation of NLRP3 infammasome has also been identifed as a major player in pulmonary infammation and tissue injury resulting from RSV infection. These clues indicate that both RSV and COVID-19 can cause severe disease phenotype through the underlying activation of NLRP3 infammasomes.

HLAs as the MHC of human origin play a critical role in the diversity of host immune response and are therefore closely involved in the immune escape process. Recent research has revealed that suppressed HLA-DR expression in monocytes can lead to increased disease severity in infants under 2 years old with RSV and adults aged 20–43 years with COVID-19 [[2,](#page-9-18) [4](#page-9-20)]. Furthermore, the immune-suppressive molecule HLA-G has been found to be induced in lung alveolar epithelial cells in COVID-19 patients aged 36–96 years [[82](#page-12-29)], which is believed to promote immune escape and subsequent disease progression [[5\]](#page-9-21). However, it remains to be investigated whether HLA-G is also implicated in RSV infection. Although the age stratifcation of investigated population is not consistently focused on the infants, these fndings still suggest important clues about the similar HLA interference from RSV and SARS-CoV-2 might help explain their potentially similar pattern of recurrent infection, and more importantly to understand how both viruses might promote aggressive and severe tissue injuries due to compromised immune surveillance.

Conclusion

The evidence presented indicates that RSV infection in infants and young children can lead to RSV bronchitis with recurrent asthmatic symptoms and may be closely related to the later development of bronchial asthma. Current understanding suggests that RSV infection can exacerbate asthma development by damaging airway epithelial tissue, promoting infammation, increasing airway hyper-responsiveness, and ultimately causing airway remodeling.

Given the similar single-stranded RNA nature of SARS-CoV-2 and RSV, we extensively discussed the potential for SARS-CoV-2 to cause similar respiratory disorders. Dysregulation of key TLR signaling, overactive NLRP3 infammasomes, and compromised monocyte HLA-DR were found to be strongly implicated in both RSV and SARS-CoV-2 infections. These similar underlying mechanisms may explain the recurring infections and the viruses' ability to promote aggressive and severe tissue injury due to compromised immune surveillance.

The incidence of RSV was signifcantly reduced and the infection appeared to be competitively suppressed due to the COVID-19 pandemic. This, combined with public health measures and changes in population-level social behavior patterns, may result in a signifcant decrease or even elimination of other respiratory viruses, including RSV. Moreover, this could offer innovative ways to prevent respiratory infections, emphasizing the crucial role of preventive public health interventions. In infants, a single RSV infection was associated with more severe clinical manifestations during a prolonged hospital stay. However, the severity of coinfection between RSV and SARS-CoV-2 did not exhibit a signifcant diference compared to single infections.

Discussion and perspectives

Although previous studies have fed us with lots of information about the strong implication of RSV in pediatric recurrent wheezing and asthma, there are still remaining controversial issues. The systematic in vivo experimental evidence is still in short to conclude whether RSV infection is an independent causative agent of pediatric bronchial asthma, and more detailed molecular mechanism underlying RSV triggered development of asthma. Hopefully, our review could shed some light on how to prevent and cure RSV infection in early life, and more importantly, how to block the triggered recurrent wheezing and asthma after one-time RSV infection.

Despite previous studies providing signifcant data on the correlation between RSV and recurrent wheezing and asthma in children, there are still some controversial issues to be addressed. The current in vivo experimental evidence is insufficient to confirm whether RSV infection is an independent cause of pediatric bronchial asthma, and the molecular mechanism behind RSV-induced asthma development requires further elucidation. This review aims to provide insights into the prevention and treatment of early-life RSV infection and, crucially, how to prevent recurrent wheezing and asthma triggered by one-time RSV infection. In this study, there is an unexpected fnding that infants with single RSV infection experience more severe pulmonary disorders than those with single SARS-CoV-2 infection. However, it is unclear if this conclusion is due to survivor bias and requires further investigation. There is a slight concern that some relevant reports focused on COVID-19 in adults rather than infants, who are the population most susceptible to RSV-LRTI. Hopefully, more studies can be conducted in children to provide us with better insights and more convincing evidence.

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