ORIGINAL ARTICLE



Molecular seasonal, age and gender distributions of *Cryptosporidium* in diarrhoeic Egyptians: distinct endemicity

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Received: 31 August 2015 / Accepted: 28 September 2015 / Published online: 6 October 2015 © Springer-Verlag Berlin Heidelberg 2015

Abstract Cryptosporidiosis is a worldwide gastrointestinal disease caused by the protozoan Cryptosporidium parasite. It has a broad range of seasonal and age-related prevalence. We aimed to study the molecular prevalence and seasonality of Cryptosporidium over a period of 1 year in a cohort of Egyptian diarrhoeic patients. Stool samples were collected from 865 diarrhoeic patients attending outpatient clinics of Cairo University hospitals, from all age groups over a 12-month period, examined microscopically for faecal Cryptosporidium oocysts by the acid-fast staining method and for copro-DNA detection using nested polymerase chain reaction (nPCR) assays. PCR-positive samples were characterised molecularly by nPCR-restriction fragment length polymorphism (RFLP) to determine Cryptosporidium genotypes. Cryptosporidium copro-DNA was detected in 19.5 % of the collected samples throughout the year, with a major peak in summer (August) and a small rise in spring (April). Infection was mainly C. hominis (95.8 %) followed by C. parvum (3.0 %), affecting all age groups, with predominance in the pre-school age group, and decrease with age. There were statistically significant associations between the detection of Cryptosporidium and season, diarrhoea, patient age and drinking water, while gender, contact with animals and presence of mucus in stool showed no association. Cryptosporidium in diarrhoeic Egyptians was of distinct endemicity, with the bi-model mostly influenced by population dynamics, with a clear high

A. A. El-Badry aelbadry@kasralainy.edu.eg prevalence in pre-school children and predominating anthroponotic (*C. hominis*) transmission throughout the year. The obtained results highlight *Cryptosporidium* as a water contaminant and an important cause of health problems in Egypt, necessitating further studies of the risk factors.

Introduction

Cryptosporidium is a worldwide enteric zoonotic protozoan parasite infecting a wide range of hosts, including mammals, birds, reptiles and fish [1]. Cryptosporidiosis was identified as a worldwide health problem and included in the World Health Organization (WHO) Neglected Diseases Initiative in 2004 [2]. It is a chief cause of diarrhoeal diseases in both developing and developed countries [3]. In Egypt, it is reported as a virulent agent of diarrhoea, especially in childhood, with varied prevalence [4]. Routine diagnosis is by coproscopy and coproimmunoassay is limited, as some Cryptosporidium infections escaped detection and species identification. Polymerase chain reaction (PCR)-based methods, beside having high diagnostic performance, have been used for the identification of species and genotypes. PCR followed by restriction fragment length polymorphism (RFLP) analysis or sequencing were required to understand the epidemiology and study outbreaks [5, 6]. Several Cryptosporidium species cause human infection, with zoonotic C. parvum and anthroponotic C. hominis being the main species, contributing to the complexity of cryptosporidiosis epidemiology [7].

Seasonality is a character of many infectious enteric diseases, including cryptosporidiosis. Environmental influences and population socio-demographic and behavioural characteristics are the main drivers of enteric disease seasonality. They affect parasite transmission and spread [8]. Some researchers have reported seasonal variation of cryptosporidiosis in their

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studies in developing countries, including Egypt; however, they didn't include all months of the year or they didn't study the molecular identification of *Cryptosporidium* species or studies were done on small sample sizes, which are considered limitations to health studies and statistical power.

This study aimed to determine the molecular prevalence of *Cryptosporidium* in diarrhoeic Egyptians over a 12-month period to assess its true seasonal pattern. Also, patients' age and gender distributions were determined.

Materials and methods

Study population and ethical considerations

This was a cross-sectional study. Stool samples submitted for parasite examination from 862 diarrhoeic patients attending outpatient clinics of Cairo University hospitals, from all age groups over a 12-month period from March 2013 to March 2014 were collected. Their related data were recorded.

The study was ethically approved by the ethical committee of Faculty of Medicine, Cairo University and informed consent was obtained from patients or their relatives and parents of young children, and they responded to questionnaires.

Sample collection and processing

A single faecal sample was obtained from each case. Collected stool samples were examined microscopically for faecal Cryptosporidium oocyst by the acid-fast staining method prior to and after concentration. The remaining part of the specimen was stored at -20 °C for molecular studies. Genomic DNA was extracted from the remaining part of fresh frozen faecal samples using the FavorPrep Stool DNA Isolation Mini Kit (Favorgen Biotech Corporation, Ping-Tung 908, Taiwan), according to the manufacturer's instructions after thermal shock of samples (five cycles of deep freezing and boiling in a water bath, each for 5 min), with prolongation of incubation for 1 h at 95 °C after 56 °C at 10 min. Extracted copro-DNA was amplified by nested PCR (nPCR) targeting the COWP gene, using two sets of primers: external primers, BCOWPF (5'-ACCGCTTCTCAACAACCATCTTGTCCTC-3') and BCOWPR (5'-CGCACCTGTTCCCACTC AATGTAAACCC-3'), which amplify a 796-bp fragment [9], and nested primers, cry-15 (5'-GTAGATAATGGAAGAGAGATTGTG-3') and cry-9 (5'-GGACTGAAATACAGGCATTATCTTG-3'), which amplify a 553-bp fragment [10]. The reaction mixture and conditions were done in a total volume of 25 µL, according to Spano et al. [10]. The amplified products were visualised with 1.5 % agarose gel electrophoresis after ethidium bromide staining. PCR products were digested by RsaI (Fermentas UAB, V.Graiciuno 8, LT-02241 Vilnius, Lithuania). Digestion of **Table 1**Diagnostic yield of the used nested polymerase chain reaction(nPCR) assay for the detection and genotyping of *Cryptosporidium*within the study group

nPCR-RFLP Negative		Frequency 694	% 80.5 %	
	C. parvum	5	0.6 % (3.0 % within positive group)	
	Non-typed	2	0.2 % (1.2 % within positive group)	
	Total	168	19.5 %	
Total		862	100 %	

Data are presented as frequency and %

N-COWP fragments was resolved by electrophoresis in 3.2 % typing-grade agarose gels containing ethidium bromide. The fragments were visualised by UV light to determine the *Cryptosporidium* genotype.

Coproscopy was carried out in the Diagnostic & Research Unit of Parasitic Diseases (DRUP) and the copro-nPCR assay was held in the Lab of Molecular Medical Parasitology (LMMP), Department of Medical Parasitology, Faculty of Medicine, Cairo University, Egypt.

Statistical analysis

Data were tabulated and processed by the Statistical Package for the Social Sciences (SPSS) version 17 (Chicago, IL, USA) for statistical analysis. Positive rates were expressed as percentages. Differences in prevalence rates among groups of the



Fig. 1 Agarose gel electrophoresis showing: L: 100-bp DNA molecular weight marker; lanes 1–3: restriction fragment length polymorphism (RFLP) products after digestion with *Rsa*I endonuclease with *Cryptosporidium parvum* genotype 2 digestion products at 34, 106 and 410 bp (the 34 band is very small, faint and difficult to see); lanes 4 and 5: RFLP products after digestion with *Rsa*I endonuclease with *C. hominis* digestion products at 34, 106 and 285 bp (the 34 band is very small, faint and difficult to see); lanes 6–7: products of the nested polymerase chain reaction (nPCR) targeting the COWP gene of *Cryptosporidium* at 553 bp

(95.8 %) samples, followed by C. parvum in 5 (3.0 %) sam-

ples; the remaining two cases were non-typed. The diagnostic yield of the used nPCR assay for the detection and genotyping of *Cryptosporidium* within the study group is represented in

Table 1. Cryptosporidium oocysts were detected in 64 (7.4 %)

ples throughout the year, with a large increase in June to Sep-

Cryptosporidium was detected in the collected study sam-

samples using MZN-stained stool smears.

studied variables were compared by the Chi-square test. Data were considered significant for a p-value<0.05.

Results

Out of 862 examined stool samples with nPCR, copro-DNA was detected in 168 (19.5 %) stool samples of diarrhoeic patients. Among them, infection was mainly *C. hominis* in 161

Fig. 2 Seasonal distribution of cases of diarrhoea (**a**), *Cryptosporidium* (**b**) and its species (**c**) among diarrhoeic patients positive by nPCR (*p*-values for the seasonal distribution of diarrhoea [**a**] was 0.0001 and for *Cryptosporidium*positive cases [**b**] was 0.003)



Fig. 3 Age distribution of *Cryptosporidium* among diarrhoeic patients positive for *Cryptosporidium* by nPCR (*p*-value for the age distribution was 0.0001)



to April, peaking in April. It affects both sex and all age groups, with predominance in the pre-school age group (Figs. 1, 2, 3 and 4).

Among the studied variables, there were statistically significant associations between season (month), diarrhoea seasonality, patient age and drinking water and detection of *Cryptosporidium*, while gender, contact with animals and presence of mucus in stool showed no association (Figs. 2, 3 and 4 and Table 2).

Discussion

In our study, *Cryptosporidium* was a prevailing protozoan with distinct endemicity among diarrhoeic Egyptians. *Cryptosporidium* transmission occurred throughout the year and was due to sporadic rather than outbreak-associated infections. The bi-model seasonal pattern was identified with a major seasonal summer peak, preceded by a small spring peak and both peaks showed *Cryptosporidium* human strain (*C. hominis*) predominance. There was a close association between cryptosporidial infection and the occurrence of diarrhoea. The period with an increase in *Cryptosporidium*

prevalence was associated with higher prevalence in diarrhoea during the same period.

A wide range of *Cryptosporidium* molecular prevalences (4.6-25 %) was reported in Egypt, and most of the studies reported a high prevalence [6, 11–13]. We reported a prevalence of 7.4 % using MZN-stained stool smear. Coproscopy had specificity; all of them were positive by PCR. However, it was of limited sensitivity, with many cases that escaped diagnosis.

The UK, oceanic countries (Australia and New Zealand), Northern Americas (USA and Canada) and European countries had clear bi-modal peaks, with one major peak in spring (UK, oceanic countries) or late summer and early autumn (Northern Americas) attributed to the human strain and an additional smaller second peak related to the bovine strain with an increase in animal contact; the lowest cases were in winter [14]. In Brazil [15], Ethiopia [16], the Philippines [17] and many tropical countries, transmission was associated with rainy seasons [18, 19].

In Egypt, there is a little winter rainfall; however, cryptosporidiosis was more prevalent in summer, as also reported in some tropical countries with little rainfall, such as Peru, which also shows the prevalence of cryptosporidiosis in warm



Fig. 4 Pattern of seasonal age group distribution of *Cryptosporidium* species among diarrhoeic patients positive by nPCR (*p*-value for the age group distribution was 0.0001)

 Table 2 nPCR-positive cases of cryptosporidiosis in association with the different studied variables other than season and age group

		nPCR				
		Negative	Positive	Total	p-Value*	
Child/adult	Child Adult	401 (46.5 %) 293 (34.0 %)	125 (14.5 %) 43 (5.0 %)	526 (61.0 %) 336 (39.0 %)	0.0001	
Gender	Male Female	368 (42.7 %) 326 (37.8 %)	87 (10.1 %) 81 (9.4 %)	455 (52.8 %) 407 (47.2 %)	0.773	
Type of water	Tape Filter	674 (78.2 %) 12 (1.4 %)	150 (17.4 %) 12 (1.4 %)	824 (95.6 %) 24 (2.8 %)	0.0001	
	Mineral	8 (0.9 %)	6 (0.7 %)	14 (1.6 %)		
Animal contact	Yes No	222 (25.8 %) 472 (54.8 %)	54 (6.3 %) 114 (13.2 %)	276 (32.0 %) 586 (68.0 %)	0.969	
Mucus	Yes No	604 (70.1 %) 90 (10.4 %)	152 (17.6 %) 16 (1.9 %)	756 (87.7 %) 106 (12.3 %)	0.223	
Total		694 (80.5 %)	168 (19.5 %)	862 (100.0 %)		

Data are presented as frequency and %

**p*-Value<0.05 is significant

seasons [20], while in Kuwait, it occurred during the cool season [3].

The difference in *Cryptosporidium* genotypes distribution was attributed to differences in the influences of infection sources. The bovine strain *Cryptosporidium* peak was attributed to the land use pattern with contamination of the water supply from young livestock (main reservoir) by hydrological phenomena in areas with rainfall/flood events and agricultural practices related to calving, while human strain predominance was related to water contamination from human activities with person-to-person transmission [8, 21].

This global *Cryptosporidium* species shift seems not to be applied in our study in Cairo, Egypt, with both peaks being attributed to the predominating human strain, with distinct endemicity

Similar to our results, Abd El Kader et al. [11] showed an outcome of 80 % for *C. hominis* and 40 % for *C. parvum*, with *C. parvum* predominating throughout the year and *C. hominis* mainly in August. Helmy et al. [22] found that *C. hominis* was 1.6 times more prevalent than *C. parvum* in children in Ismailia, Egypt. Contradictory to our results, Eida et al. [23] reported *C. parvum* predominance in Egypt. El-Shazly et al. [24] and Abd El Kader et al. [11] reported that the highest prevalence was in summer but the latter recorded, besides the major summer peak, another smaller one in spring in Cairo and attributed it to bovine strains.

C. hominis predominance in this report is similar to studies conducted in Uganda [19], Malawi, Kenya [25], South Africa [26], Australia, Canada, Japan, USA and developing countries [27].

Contradictory to our results, *C. parvum* was more often detected in humans than *C. hominis* in studies in Middle Eastern countries [3, 4, 28–31] and European countries, such as

Portugal [32], UK [33] and Ireland [34]. Spain and Wales showed a relatively balanced relation of *C. hominis/C. parvum* [35].

Having anthroponotic Cryptosporidium strain predominance with no association between Cryptosporidium prevalence and animal contact in our study population reveals that population dynamics influence the transmission pattern of *Cryptosporidium* in Cairo. Water contamination may be the key determinant of distinct seasonality in Egypt and may be coupled with population dynamics that increase person-toperson transmission in hot months and spring due to outdoor activities, including recreational water use [36]. In addition, attendees of Cairo university hospitals come from urban and peri-urban areas of higher population densities and compromised infrastructures that favour person-to-person transmission and had low socio-economic class populations, depending mainly on chlorinated water, in which Cryptosporidium can survive. All cases were endemic, as none of the study population had history of travel. The lack of proper sanitation and infrastructure may be the origin of water contamination by faecal materials or they may become contaminated by storage in dirty containers [37].

Socio-demographic and behavioural differences between our study populations and those in other studies may explain the differences in the prevalence of the most frequent *Cryptosporidium* species. Also, these studies were products of reduced sample sizes and sample collection was not included in all months of the year.

Our study deduced statistically significant differences for *Cryptosporidium* between children and adults. Cryptosporidiosis affects all age groups, with the highest prevalence level among pre-school children aged 2– 6 years (32.5 %), followed by older children aged 6– 12 years (20.8 %), and it decreases with age. Similarly, Abdel-Messih et al. [38], in Egypt, confirmed that 61.9 % of infected cases were related to this age group. The high prevalence of cryptosporidiosis in children has been reported in many countries, including Canada, USA, New Zealand, Ireland, England and France [14, 20, 34, 39, 40]. This high incidence of the disease in children may be related to the lack of pre-existing immunity, as older people may get exposed to *Cryptosporidium* infection in their lifetime. Moreover, children were more exposed to water during playing, increasing the chance of getting infected, and there was a more frequent attendance by physicians of diarrhoeic children than adults [11, 40].

In this study, *Cryptosporidium* was detected in males (n=455) more than females (n=407); however, the difference in sex distribution was statistically insignificant. This is in accordance with the results of many studies [11, 34, 39, 40], suggesting significant male infection predominance. In contrast, Yoder et al. [14] found that most of the reported cases in 2005 occurred among females. However, they stated that their data on race and ethnicity were incomplete.

Conclusion

There was a distinct endemicity of *Cryptosporidium* seasonality in diarrhoeic Egyptians, with a clear high true prevalence and predominance of anthroponotic (*C. hominis*) transmission throughout the year, and cases were due to sporadic rather than outbreak-associated infections. Differences in sex distribution for *Cryptosporidium* was not significant but was significant for age distribution, and pre-school children showed the highest level. Molecular tools are a must in *Cryptosporidium* prevalence studies to identify species and sub-genotypes.

The obtained results are important for the development of public health strategies, and improvement of disease prediction, prevention and control in the absence of treatment or reliable vaccine. The results also highlight *Cryptosporidium* as a water contaminant and an important cause of health problems in Egypt necessitating further studies of the risk factors.

Contribution of each author All manuscript authors contributed to every activity of it; idea of paper, study design, collection of materials, methodology, writing the paper and revising it.

Conflict of interest The authors declare that they have no competing interests

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References

- Smith HV, Cacciò SM, Cook N, Nichols RA, Tait A (2007) *Cryptosporidium* and *Giardia* as foodborne zoonoses. Vet Parasitol 149:29–40
- 2. Neglected Diseases Initiative of the World Health Organization. Home page at: http://www.who.int/neglected_diseases/en/
- Sulaiman IM, Hira PR, Zhou L, Al-Ali FM, Al-Shelahi FA, Shweiki HM, Iqbal J, Khalid N, Xiao L (2005) Unique endemicity of cryptosporidiosis in children in Kuwait. J Clin Microbiol 43(6): 2805–2809
- Youssef FG, Adib I, Riddle MS, Schlett CD (2008) A review of cryptosporidiosis in Egypt. J Egypt Soc Parasitol 38:9–28
- Xiao L (2010) Molecular epidemiology of cryptosporidiosis: an update. Exp Parasitol 124(1):80–89
- Ghallab MMI, Abdel-Aziz IZ, Shoeib EY, El-Badry AA (2014) Laboratory utility of coproscopy, copro immunoassays and copro nPCR assay targeting Hsp90 gene for detection of *Cryptosporidium* in children, Cairo, Egypt. J Parasit Dis 8(2):1–5
- Peng MM, Xiao L, Freeman AR, Arrowood MJ, Escalante AA, Weltman AC, Ong CS, Mac Kenzie WR, Lal AA, Beard CB (1997) Genetic polymorphism among *Cryptosporidium parvum* isolates: evidence of two distinct human transmission cycles. Emerg Infect Dis 3:567–573
- Lal A, Hales S, French N, Baker MG (2012) Seasonality in human zoonotic enteric diseases: a systematic review. PLoS One 7(4): 31883
- 9. Pedraza-Díaz S, Amar C, Nichols GL, McLauchlin J (2001) Nested polymerase chain reaction for amplification of the cryptosporidium oocyst wall protein gene. Emerg Infect Dis 7(1):49–56
- Spano F, Putignani L, McLauchlin J, Casemore DP, Crisanti A (1997) PCR-RFLP analysis of the *Cryptosporidium* oocyst wall protein (COWP) gene discriminates between *C. wrairi* and *C. parvum*, and between *C. parvum* isolates of human and animal origin. FEMS Microbiol Lett 150:209–217
- Abd El Kader NM, Blanco MA, Ali-Tammam M, Abd El Ghaffar Ael R, Osman A, El Sheikh N, Rubio JM, de Fuentes I (2011) Detection of *Cryptosporidium parvum* and *Cryptosporidium hominis* in human patients in Cairo, Egypt. Parasitol Res 110(1): 161–166
- El-Settawy MA, Fathy GM (2012) Evaluation and comparison of PCR, coproantigen ELISA and microscopy for diagnosis of *Cryptosporidium* in human diarrheic specimens. J Am Sci 8(12): 1378–1385
- Fathy MM, Abdelrazek NM, Hassan FA, El-Badry AA (2014) Molecular copro-prevalence of *Cryptosporidium* in Egyptian children and evaluation of three diagnostic methods. Indian Pediatr 51(9):727–729
- Yoder JS, Beach MJ; Centers for Disease Control and Prevention (CDC) (2007) Giardiasis surveillance—United States, 2003–2005. MMWR Surveill Summ 56(7):11–18
- Newman RD, Sears CL, Moore SR, Nataro JP, Wuhib T, Agnew DA, Guerrant RL, Lima AAM (1999) Longitudinal study of *Cryptosporidium* infection in children in Northeastern Brazil. J Infect Dis 180:167–175
- Adamu H, Petros B, Hailu A, Petry F (2010) Molecular characterization of *Cryptosporidium* isolates from humans in Ethiopia. Acta Trop 115(1–2):77–83
- Natividad FF, Buerano CC, Lago CB, Mapua CA, De Guzman BB, Seraspe EB, Samentar LP, Endo T (2008) Prevalence rates of *Giardia* and *Cryptosporidium* among diarrheic patients in the Philippines. Southeast Asian J Trop Med Public Health 39(6): 991–999
- Peng MM, Meshnick SR, Cunliffe NA, Thindwa BD, Hart CA, Broadhead RL, Xiao L (2003) Molecular epidemiology of

cryptosporidiosis in children in Malawi. J Eukaryot Microbiol 50: 557–559

- Tumwine JK, Kekitiinwa A, Nabukeera N, Akiyoshi DE, Rich SM, Widmer G, Feng X, Tzipori S (2003) *Cryptosporidium parvum* in children with diarrhea in Mulago Hospital, Kampala, Uganda. Am J Trop Med Hyg 68:710–715
- Bern C, Ortega Y, Checkley W, Roberts JM, Lescano AG, Cabrera L, Verastegui M, Black RE, Sterling C, Gilman RH (2002) Epidemiologic differences between cyclosporiasis and cryptosporidiosis in Peruvian children. Emerg Infect Dis 8:581–585
- Learmonth JJ, Ionas G, Ebbett KA, Kwan ES (2004) Genetic characterization and transmission cycles of *Cryptosporidium* species isolated from humans in New Zealand. Appl Environ Microbiol 70:3973–3978
- Helmy YA, Krücken J, Nöckler K, von Samson-Himmelstjerna G, Zessin KH (2013) Molecular epidemiology of *Cryptosporidium* in livestock animals and humans in the Ismailia province of Egypt. Vet Parasitol 193:15–24
- Eida AM, Eida MM, El-Desoky A (2009) Pathological studies of different genotypes of human *Cryptosporidium* Egyptian isolates in experimentally mice. J Egypt Soc Parasitol 39(3):975–990
- El-Shazly AM, El-sheikha HM, Soltan DM, Mohammad KA, Morsy TA (2007) Protozoal pollution of surface water sources in Dakahlia Governorate, Egypt. J Egypt Soc Parasitol 37(1):51–64
- 25. Gatei W, Greensill J, Ashford RW, Cuevas LE, Parry CM, Cunliffe NA, Beeching NJ, Hart CA (2003) Molecular analysis of the 18S rRNA gene of *Cryptosporidium* parasites from patients with or without human immunodeficiency virus infections living in Kenya, Malawi, Brazil, the United Kingdom, and Vietnam. J Clin Microbiol 41:1458–1462
- Leav BA, Mackay MR, Anyanwu A, O'Connor RM, Cevallos AM, Kindra G, Rollins NC, Bennish ML, Nelson RG, Ward HD (2002) Analysis of sequence diversity at the highly polymorphic Cpgp40/ 15 locus among *Cryptosporidium* isolates from human immunodeficiency virus-infected children in South Africa. Infect Immun 70: 3881–3890
- Xiao L, Fayer R (2008) Molecular characterisation of species and genotypes of *Cryptosporidium* and *Giardia* and assessment of zoonotic transmission. Int J Parasitol 38(11):1239–1255
- Al-Brikan FA, Salem HS, Beeching N, Hilal N (2008) Multilocus genetic analysis of *Cryptosporidium* isolates from Saudi Arabia. J Egypt Soc Parasitol 38:645–658

- Hijjawi N, Ng J, Yang R, Atoum MF, Ryan U (2010) Identification of rare and novel *Cryptosporidium* GP60 subtypes in human isolates from Jordan. Exp Parasitol 125:161–164
- Iqbal J, Khalid N, Hira PR (2011) Cryptosporidiosis in Kuwaiti children: association of clinical characteristics with *Cryptosporidium* species and subtypes. J Med Microbiol 60(Pt 5): 647–652
- Alyousefi NA, Mahdy MA, Lim YA, Xiao L, Mahmud R (2013) First molecular characterization of *Cryptosporidium* in Yemen. Parasitology 140(6):729–734
- Alves M, Xiao L, Sulaiman I, Lal AA, Matos O, Antunes F (2003) Subgenotype analysis of *Cryptosporidium* isolates from humans, cattle, and zoo ruminants in Portugal. J Clin Microbiol 41:2744– 2747
- 33. McLauchlin J, Amar C, Pedraza-Díaz S, Nichols GL (2000) Molecular epidemiological analysis of *Cryptosporidium* spp. in the United Kingdom: results of genotyping *Cryptosporidium* spp. in 1,705 fecal samples from humans and 105 fecal samples from livestock animals. J Clin Microbiol 38:3984–3990
- Garvey P, McKeown P (2009) Epidemiology of human cryptosporidiosis in Ireland, 2004–2006: analysis of national notification data. Euro Surveill 14(8). pii: 19128
- Llorente MT, Clavel A, Goñi MP, Varea M, Seral C, Becerril R, Suarez L, Gómez-Lus R (2007) Genetic characterization of *Cryptosporidium* species from humans in Spain. Parasitol Int 56(3):201–205
- Cama VA, Bern C, Roberts J, Cabrera L, Sterling CR, Ortega Y, Gilman RH, Xiao L (2008) *Cryptosporidium* species and subtypes and clinical manifestations in children, Peru. Emerg Infect Dis 14: 1567–1574
- El-Sherbini GT, Abosdera MM (2013) Risk factors associated with intestinal parasitic infections among children. J Egypt Soc Parasitol 43(1):287–294
- Abdel-Messih IA, Wierzba TF, Abu-Elyazeed R, Ibrahim AF, Ahmed SF, Kamal K, Sanders J, Frenck R (2005) Diarrhea associated with *Cryptosporidium parvum* among young children of the Nile River Delta in Egypt. J Trop Pediatr 51:154–159
- Hlavsa MC, Watson JC, Beach MJ (2004) Cryptosporidiosis surveillance—United States, 1999–2002. In: Surveillance Summaries. MMWR 54(1):1–8
- ANOFEL Cryptosporidium National Network (2010) Laboratorybased surveillance for *Cryptosporidium* in France, 2006–2009. Euro Surveill 15(33):19642