#### **ORIGINAL ARTICLE**



# Variable abnormality of the melanopsin-derived portion of the pupillary light reflex (PLR) in patients with Parkinson's disease (PD) and parkinsonism features

Bruce I. Gaynes <sup>1,2</sup>  $\odot \cdot$  Adnaan Zaffer <sup>1</sup>  $\cdot$  Raman Yousefzai <sup>1</sup>  $\cdot$  Mario Chazaro-Cortes <sup>1</sup>  $\cdot$  Kalea Colletta <sup>1</sup>  $\cdot$  Sandra L. Kletzel <sup>1</sup>  $\cdot$  Mary Beth Jost <sup>1</sup>  $\cdot$  Youngsook Park <sup>1</sup>  $\cdot$  Jasvinder Chawla <sup>1,2</sup>  $\cdot$  Mark V. Albert <sup>3</sup>  $\cdot$  Ting Xiao <sup>4</sup>

Received: 17 December 2020 / Accepted: 10 April 2021 / Published online: 4 May 2021 © This is a U.S. government work and not under copyright protection in the U.S.; foreign copyright protection may apply 2021

#### Abstract

**Objectives** Ascertain and quantify abnormality of the melanopsin-derived portion of the pupillary light reflex (PLR) in patients with Parkinson's disease (PD) and parkinsonism features based on a statistical predictive modeling strategy for PLR classification.

**Methods** Exploratory cohort analysis of pupillary kinetics in non-disease controls, PD subjects, and subjects with parkinsonism features using chromatic pupillometry. Receiver operating characteristic (ROC) curve interpretation of pupillary changes consistent with abnormality of intrinsically photosensitive retinal ganglion cells (ipRGCs) was employed using a thresholding algorithm to discriminate pupillary abnormality between study groups.

**Results** Twenty-eight subjects were enrolled, including 17 PD subjects (age range 64–85, mean 70.65) and nine controls (age range 48–95, mean 63.89). Two subjects were described as demonstrating parkinsonism symptoms due to presumed Lewy body dementia and motor system atrophy (MSA) respectively. On aggregate analysis, PD subjects demonstrated abnormal but variable pupillary dynamics suggestive of ipRGC abnormality. Subjects with parkinsonism features did not demonstrate pupillary changes consistent with ipRGC abnormality. There was no relationship between levodopa equivalent dosage or PD severity and ipRGC abnormality. The pupillary test sensitivity in predicting PD was 0.75 and likelihood ratio was 1.2.

Conclusions ipRGC deficit is demonstrated in PD subjects; however, the degree and constancy of abnormality appear variable.

Keywords Circadian · Retina · Parkinson's disease · Blue light

# Background

Parkinson's disease (PD) is a progressive neurodegenerative disorder characterized by muscle rigidity, tremor, and, in advanced cases, loss of physical movement [1]. Ocular complications are well described in PD and include reduced retinal

Bruce I. Gaynes bgaynes@luc.edu

- <sup>1</sup> Edward Hines Jr. VA Medical Center, Hines, IL, USA
- <sup>2</sup> Loyola University Chicago, Stritch School of Medicine, Maywood, IL, USA
- <sup>3</sup> Biomedical Engineering, Computer Science and Engineering, University of North Texas, Denton, TX, USA
- <sup>4</sup> Computer Science and Engineering, University of North Texas, Denton, TX, USA

dopamine [2], the primary neurotransmitter for retinal phototransduction. [3]. The linkage between retinal dysfunction and PD is an intriguing field of research that has important relevance not only in regard to PD-related co-morbidities but as an etiologic component of the disease process [4, 5]. Mounting evidence supports a link between the retina and nigrostriatal function. For example, bright light therapy for PD has been found to be beneficial in treatment of sleep abnormality through a putative mechanism involving suppression of retinal melatonin [4-7]. Moreover, bright blue treatment demonstrates not only positive effects on sleep and mood but also on PD-related motor function [8]. These findings suggest that functionality of the nigrostriatal system is influenced by a system emanating from the retina to the pineal gland, denoting a key role of circadian entrainment as a component of nigrostriatal operation [9]. The putative link between the retina and nigrostriatal function supports the notion that retinal dysfunction and concomitant circadian

dysregulation are etiologic features in PD that have potential as both biomarkers and treatment targets of PD. The role of circadian dysfunction as an etiologic component of PD is reinforced in studies by Willis et al. that show PD-related symptomatology can be reversed experimentally by direct intravitreal delivery of L-dopa in amounts considered inadequate to result in systemic effects [9, 10]. These studies have implicated the retina not only as an associated neural system affected by PD but also as a component of the disease etiology itself [9, 11]. The importance of appropriate functionality of retinal supported circadian homeostasis is also denoted by exacerbation of PD symptoms in an animal model following enucleation [11].

Light-dark cycles maintained by the retina emanate from a subset of retinal ganglion cells known to be inherently photosensitive and colloquially termed intrinsically photosensitive retinal ganglion cells (ipRGCs) [12]. ipRGCs play a key role in the operation of the suprachiasmatic nucleus in regard to melatonin secretion and circadian homeostasis [13, 14]. The function of ipRGCs is based on the chromophore melanopsin, with peak sensitivity at 470 nm [15]. The response of ipRGC is energy dependent, with higher levels of irradiance leading to greater ipRGC activity [16]. The latter property of ipRGC is intriguing in regard to PD as studies have demonstrated the benefit of bright short-wavelength exposure in ameliorating some of the symptoms of PD-related complications [6]. ipRGCs also mediate other non-image-forming (NIF) visual responses such as the pupillary light reflex (PLR) via corelease of neurotransmitters glutamate and pituitary adenylate cyclase-activating peptide (PACAP) from their synaptic terminals.

Functionality of ipRGC can be reflected in pupillary dynamics [12, 17]. The hallmark finding of pRGC function is persistence of the pupillary response (PPR) denoted by prolonged pupillary constriction following extinguishing a light stimulus of suitable wavelength and intensity to activate the chromophore melanopsin within ipRGCs. A persistent potentiation of the pupil response following cessation of a suitable stimulus is the hallmark of healthy ipRGC function clinically [12]. The circadian dysregulation demonstrated in PD as well as the response to bright blue light in regard to PD therapy suggest a key link between functionality of ipRGC and PD [18, 19]. The purpose of this report is to assess the characteristics of the PPR in PD subjects and those with parkinsonism features versus a control cohort in regard to interpretation of PPR abnormality as a biomarker ipRGC dysfunction indicative of PD pathobiology.

# Methods

This was an exploratory cohort study designed to examine differences in PPR in patients with PD, parkinsonism, and

controls. Patients were enrolled by consecutive sampling. Inclusion criteria include prior diagnosis of PD with treatment of one or more dopaminergic agonists. Patients with parkinsonism features but without definitive PD diagnosis were also enrolled for exploratory analysis. Patients taking drugs known to impact circadian rhythm including lithium, benzodiazepines, steroidal and nonsteroidal anti-inflammatory drugs, and vitamin B12 were excluded from the study. Patients were screened prior to testing for the presence of ocular abnormalities including amblyopia, cataract reducing correctable vision to less than 20/25 in either eye, glaucoma, or any form of retinopathy such as that related to diabetes or age-related macular degeneration. We did not exclude pseudophakic subjects due to the fact retinal irradiance in phakic patients was normalized and corrected for media loss based on the method of van deKraats and van Norren [20] This correction factor obviates confounding of pupillary findings arising from deviation of inter-subject retinal illuminance. Overall, one PD subject and two control subjects were pseudophakic. The degree of PD severity was evaluated by the Hoehn and Yahr grading system [21].

### Pupillometry

The right eye was dilated with 2.5% phenylephrine and 1% tropicamide. Dilation of the right eye was employed in an effort to maintain consistent retinal illumination within and between subjects during stimulation. Following eye dilation, the ipRGC-driven pupil response was measured via the consensual post-illumination pupil response (PIPR) in the left eye. Stimuli presented to the right eye consisted of longwavelength (red) and short-wavelength (blue) narrowband 5s pulses of light. This establishes the adequacy of the irradiance level used in this study to induce ipRGC action. Light stimuli were generated using a custom-built Maxwellian-view optical system consisting of narrow bandpass filters of 25mm-diameter short wavelength; "blue" light,  $\lambda max = 470$ nm; full width half maximum (FWHM) = 10 nm and long wavelength; "red" light,  $\lambda max = 640$  FWHM = 10 nm, imaged to the plane of the pupil in conjunction with appropriate neutral density filters to alter power output to 8 or 30 µW (ILT 9000, International Light Technology). The stimulus pencil was constrained using an aperture of 10 mm in diameter.

The experimental setup included use of an infraredsensitive XIMEA MQ013RG-E2 machine vision camera whose parameters were controlled by the bundled XIMEA CamTool software, which also enables initiation of photo and video capture which was attached to a Haag Streit 900 model slit lamp with a suitable C mount. Patients were placed securely in the biomicroscope to maintain alignment in Maxwellian view while the dilated right eye was stimulated. The stimuli for both long and short wavelengths were based on a spectral irradiance of 8 and 30  $\mu$ W·cm<sup>-2</sup>·nm<sup>-1</sup> resulting in irradiance stimuli of 11.421 and 11.995 log photons  $cm^{-2}$ .  $s^{-1}$  at 640 nm and 14.621 and 15.195 log photons cm<sup>-2</sup> s<sup>-1</sup> at 470 nm determined at the corneal plane respectively. Given the older age of the participants, retinal irradiances were estimated based upon established corrections for age-related changes in the optical density of the media of the eye for stimuli greater than 3° in diameter [20]. The pupillary light reflex was determined by averaging three consensual pupil recordings of 40 s (5-s pre-stimulus, 5-s stimulus, and 30-s post-stimulus). Pupillary dynamics were assessed using the XIMEA CamTool software in conjunction with Adobe Premiere Pro v.2 video capture and Image J imaging software (Rasband, W.S., ImageJ, US National Institutes of Health, Bethesda, MD, USA). Nerve fiber layer thinning has been associated with PD; therefore, assessment of retinal nerve fiber layer thickness and retinal morphology was assessed in the stimulated eye using an automated optical coherence tomography (OCT) instrument (Zeiss Cirrus Model 5000 OCT, Carl Zeiss AG, Oberkochen, Germany) [22].

## **Statistics**

All data were normalized to the average pupillary diameter of the first 5 s of imaging and expressed as percentage baseline units. In order to ascertain the degree of data variability, both control and PD subject pupillary size/time curves were converted to interpretative AUC data for assessment of data scatter and trend. Analysis of the median pupillary area from time zero to time 40 s between high or low energy levels and short vs long-wavelength stimulation was conducted with a Mann-Whitney U test.

In order to illustrate the classification accuracy of the PLRs, we performed receiver operating characteristic (ROC) curve analysis. Given that the ROC-AUC provides information on the discriminability of PPR, a classification can be conducted based on the difference from control subjects. From the overall analysis of ROC-AUC in the control group, a threshold AUC statistic of 0.65 was deemed valid as a cut-off for identifying normal vs abnormal PPR in the established binary model. The relative low AUC findings with shortwavelength high-irradiance stimulation in the control cohort were consistent with normal PPR. Comparative AUC values (and confidence intervals) provided a p value (alpha = 0.05)

that addressed the likelihood of generating an AUC statistic as great or greater than 0.65 assuming no difference in the AUC between control and PD data. To adjust for multiple comparisons, we employed the two-stage linear step-up procedure of Benjamini, Krieger, and Yekutieli using a false discovery rate (Q value) of 6.5% (GraphPad v 9.0). A chi-square analysis was employed to demonstrate sensitivity, specificity, and like-lihood ratio of the PPR in association with PD. Covariance between AUC and levodopa equivalent daily dosage and PD grade severity was determined using a Pearson product-moment correlation coefficient (r). The two subjects with parkinsonian features were assigned to the overall PD subject cohort for the purposes of data analysis as pupillary abnormalities have been previously described in patients with non-PD parkinsonian features [23].

## Results

Overall, 17 PD subjects, 2 parkinsonian subjects, and 9 controls were enrolled. The two parkinsonian patients were described as demonstrating parkinsonism symptoms due to presumed Lewy body dementia and motor system atrophy (MSA) respectively. A description of patient demographics is found in Table 1. Difference in age between PD and control cohorts was not statistically significant (p = 0.0750).

Aggregate AUC analysis of pupillary tracings demonstrated between control and PD cohorts with high-irradiance shortwavelength stimulus demonstrated relative lack of PPR in the PD cohorts (mean AUC of PD cohort = 2684 vs 2436 for controls, p = 0.0144, Mann-Whitney U test, alpha = 0.05, Fig. 1). In contrast to controls, the PD cohort also demonstrated latency in pupillary recovery (Fig. 1). Mean pupillary area vs time data between short (470 nm) and long (640 nm) wavelengths at low irradiance in control and PD cohorts is shown in Fig. 2. Lack of PPR in the control and PD cohorts is consistent with the known high-irradiance photochemical requirement for melanopsin engagement (Fig. 2). Pupillometry data between short and long wavelengths at high irradiance in control and PD cohorts is seen in Fig. 3. Highirradiance short-wavelength stimulus produces a discernable PPR persistent to the 40-s time interval in the control cohort while the long-wavelength high-irradiance stimulus lacks

Table 1	Age and OCT metrics,
PD vs co	ontrol

	Mean (SD) PD $N = 19^*$	Range PD	Mean (SD) control $N = 9$	Range control	p value <sup>+</sup>
Age (years)	70.84 (5.89)	64–85	63.89 (14.81)	48–95	0.1586
OCT (microns)	89.73 (9.93)	78–111	96.80 (12.64)	78–109	0.0750

\* Includes 2 patients with parkinsonism features

<sup>+</sup> Students *t* test, alpha = 0.05

Pupillary area vs time with 470 nm 30 µW stimulus; control aggregate data 140 Percent baseline pupillary area 120 100 80 60 40 20 25 30 35 40 45 0 5 10 15 20 Time (sec.)

Fig. 1 Pupillary tracing between control (left) and PD cohorts. Figures represent the aggregate of pupil area vs time tracings of the overall study cohort. Mean AUC of control and PD data = 2436 and 2684, respectively, p = 0.0144 (Mann-Whitney, alpha = 0.05). The

stimulus on

PPR effect. These observations are consistent with the photochemical action spectrum of melanopsin and ipRGC activity. Lack of PPR in the PD cohort with short-wavelength highirradiance stimuli is consistent with ipRGC abnormality (Fig. 3). A notable aberration in pupil latency at both short- and long-wavelength stimuli is demonstrated in the PD cohort (Figs. 2 and 3). The mean percent baseline pupillary area at the ending 40-s time interval for the high-irradiance shortwavelength stimulus is 66.45% in controls and 80.81% in PD subjects (p = 0.0012, Mann-Whitney, Fig. 3).

Tables 2 outlines the description of the study cohort in terms of levodopa equivalent dosage and PD severity grade. Tables 3, and 4 illustrate the predictive properties of ROC-AUC analysis to discriminate normal and abnormal PPR in PD and control cohorts respectively. Of the 19 subjects in the PD cohort, 8 were found to demonstrate abnormal PPR while one PD subject was deemed as demonstrating "borderline" PPR abnormality (Table 3). Interestingly, in the control cohort, 3 subjects were determined to demonstrate abnormal

Pupillary area vs time with 470 nm 30 µW stimulus; PD aggregate data



overall flat tracings in the control cohort with relative low AUC following extinguishing the stimulus is the hallmark of appropriately functioning ipRGCs. Relative higher AUC data in the PD cohort is indicative of aberrant PPR and ipRGC abnormality

PRR (Table 4). Metrics describing PPR test sensitivity, specificity, and likelihood ratio in identifying PD are shown in Table 5. No relationship was found between levodopa equivalent daily dosage (LEDD) or PD severity grade and pupillary time AUC (r = 0.3551, p = 0.1358; r = 0.2410; p = 0.3184respectively). Although there was no statistical difference in the mean retinal nerve fiber layer thickness between the PD and control cohorts, the PD cohort demonstrated a trend toward decreased retinal ganglion cell layer thickness (Table 1).

# Discussion

Findings of the present study demonstrate variable abnormality in the PLR in a cohort of PD subjects typified by an aberrant PPR following short-wavelength, high-irradiance stimuli. Based on knowledge of the photochemical requirements of melanopsin activation [24], these findings are consistent with ipRGC abnormality as part of the melanopsin-derived



Fig. 2 Mean pupillary area vs time data between short (470 nm) and long (640 nm) wavelengths at low (8 µW) irradiance in control and PD cohorts. Lack of PPR in the control and PD cohorts is consistent with the known high-irradiance photochemical requirement for melanopsin engagement



**Fig. 3** Mean pupillary area vs time data between short (470 nm) and long (640 nm) wavelengths at high (30  $\mu$ W) irradiance in control and PD cohorts. Left: High-irradiance short-wavelength stimuli in the control cohort produces the expected discernable PPR persistent to the 40-s time interval while the long-wavelength high-irradiance stimulus lacks PPR effect. Right: Lack of PPR in the PD cohort with short-wavelength high-

contribution of the PLR. Our findings support that of prior studies which have also disclosed abnormal PPR in PD subjects [25]. In contrast however to prior studies examining ipRGC abnormality in PD patients, the present work included a cohort with a relatively higher Hoehn and Yahr grade indicating an overall higher level of disease severity [26]. Although prior studies have suggested that retinal nerve fiber layer thickness is reduced in relation to disease severity, our findings do not show a similar relationship [25]. Furthermore, our findings did not show a relationship between the PPR and PD severity or levodopa equivalent dose as was noted in prior studies [25]. Additionally, this study supports the notion that PPR abnormality found in PD is not due to morphologic abnormality of the retinal nerve fiber layer as denoted by the similarity in retinal nerve fiber layer thickness between disease and control groups reinforcing findings of prior studies [25]. Retinal nerve fiber compromise has been associated with abnormality in electroretinography findings in PD patients; however, the relationship between electroretinography and pupillary function in PD has not been established [27]. Observations in this study may be explained in several scenarios; disease pathobiological variability; systematic error in the clinical identification of a PD "gold standard"; and methodologic limitations of empirical ROC-AUC analysis for biomarker analysis.

Circadian neurohormonal secretion disrupted by ipRGC abnormality is now regarded as a rational component of neurodegenerative disease pathobiology [28]. Dysfunction of dopaminergic sub-populations in the retina as a result of PD has been shown to correlate with PD-related sleep, locomotor, and visual dysfunction [29]. Curiously, ipRGC are both pre- and postsynaptic to dopaminergic amacrine (DA) cells via reciprocal synapses; inhibitory DA signals are mediated through GABA co-released from DA along with excitatory dopamine



irradiance stimuli is consistent with ipRGC abnormality. Mean percent baseline pupillary area at the ending 40-s time interval for the high-irradiance short-wavelength stimulus = 66.45% in controls vs 80.81% for PD subjects (p = 0.0012, Mann-Whitney). A notable aberration in pupil latency at both short- and long-wavelength stimuli is demonstrated in the PD cohort

signaling [30]. Dopamine has functions in the light-adaptation process by upregulating melanopsin transcription in ipRGCs and thus increasing the photoreceptor's sensitivity [30]. DA releases dopamine extracellularly during daylight, with light

 Table 2
 Description of study cohort identified with PD and/or parkinsonism

Subject no.	LEDD (mg)*	PD grade <sup>+</sup>	
101	750	2	
102	300	3	
103	240	1	
104	600	3	
105	1197	3	
106	450	1	
107	1390	3	
108	580	1	
109	300	1	
110	2760	4	
111	1995	3	
112	1995	2	
113	640	3	
114	600	3	
115-1 <sup>a</sup>	0	1	
116-2 <sup>a</sup>	450	1	
117	214	4	
118	2827	4	
119	300	1	
Mean (SD)	925.7 (864.8)	Median 3	

\* Levodopa equivalent daily dose

+ Hoehn and Yahr grading system at the time of study enrollment

<sup>a</sup> Patients with non-PD parkinsonism features

Table 3 AUC/ROC assessment of PPR abnormality in study cohort identified with PD and/or parkinsonism. Predictive properties of ROC-AUC analysis to discriminate between normal and abnormal PPR. A perfect classification of abnormal PPR = AUC of 1, while a perfect classification of normal PPR = 0.5. In this investigation, we determined a study AUC cut-off of > 0.65 to best reflect abnormal PPR

PD subject	ROC C statistic (AUC)	p value	Confidence interval	PPR assessment
101	0.5244	0.7075	0.3880 to 0.6608	Normal
102	0.7394	0.0002	0.6140 to 0.8648	Abnormal
103^	0.7294	0.0004	0.6071 to 0.8517	Abnormal
104	0.7950	0.0001	0.6829 to 0.9071	Abnormal
105	0.6669	0.0102	0.5432 to 0.7906	Abnormal
106	0.6019	0.1168	0.4728 to 0.7309	Normal
107	0.7725	0.0001	0.6606 to 0.8844	Abnormal
108	0.7494	0.0001	0.6271 to 0.8717	Abnormal
109	0.5794	0.2217	0.4446 to 0.7141	Normal
110	0.5819	0.2075	0.4467 to 0.7170	Normal
111	0.7700	0.0001	0.6591 to 0.8809	Abnormal
112	0.5500	0.4414	0.4127 to 0.6873	Normal
113	0.6275	0.0496	0.4930 to 0.7620	Borderline
114	0.5588	0.3657	0.4274 to 0.6901	Normal
115*	0.5581	0.3708	0.4302 to 0.6861	Normal
116*	0.5844	0.1939	0.4546 to 0.7142	Normal
117	0.6050	0.1060	0.4713 to 0.7387	Normal
118	0.6744	0.0073	0.5460 to 0.8028	Abnormal
119	0.5119	0.8549	0.3817 to 0.6421	Normal

Patients with non-PD parkinsonism features

Pseudophakic

adaptative effects typified by enhancing the activity of cone cells and increasing sensitivity to color and contrast while suppressing rod cell and retinal melatonin activity. Thus, due to diminished retinal dopamine, PD patients are considered to be in a persistent state of dark adaptation [31].

As noted, in this study, there was no correlation between LEDD and PPR abnormality. This finding may be interpreted that either ipRGC dopaminergic dependency acts in a stochastic binary rather than graded fashion or retinal dopamine levels among the PD cohort lacked sufficient variability for statistical significance. Binary expression of ipRGC pupil abnormality in PD may explain the lack of a relationship between LEDD and PD severity seen in this and other studies [25]. It is evident that in aggregate analysis, the identification of PPR abnormality is shown in the PD cohort. However, with the statistical approach in this study, its apparent significant heterogeneity in PPR abnormality exists not only in the PD cohort but also in the control group as well. The presence of abnormal PPR in the control group confounds the sensitivity and specificity of the PPR as a viable biomarker for PD. However, one cannot rule out the possibility that abnormality of PPR in the control cohort is a representative prodromal marker of an evolving neurodegenerative state. Furthermore, the variability in the depth of the PPR abnormality in PD patients is also notable suggesting either a graded degradation in ipRGC function in PD subjects.

Control subject	ROC C statistic (AUC)	Adjusted p value	CI	PPR assessment
201^	0.5238	0.7146	0.3933 to 0.6542	Normal
202	0.5213	0.7435	0.3917 to 0.6508	Normal
203	0.6263	0.0519	0.4954 to 0.7571	Normal
204	0.5756	0.2443	0.4427 to 0.7086	Normal
205^	0.5469	0.4705	0.4172 to 0.6766	Normal
206	0.7631	0.0001	0.6469 to 0.8794	Abnormal
207	0.7000	0.0021	0.5811 to 0.8189	Abnormal
208	0.6719	0.0081	0.5480 to 0.7958	Abnormal
209	0.5900	0.1659	0.4611 to 0.7189	Normal

<sup>^</sup>Pseudophakic

Table 4 AUC/ROC assessment of PRL in the non-PD (control

subjects) cohort

**Table 5** Description of PLR predictive diagnostics. The data indicatemetrics on sensitivity, specificity, and likelihood of PD in presence ofabnormality in the PPR (chi-square with Fisher's exact test, p = 0.6870).Fisher's test defines the probability that random sampling alone results inan association as strong (or stronger) as observed in this study

	Value	Confidence interval
Sensitivity	0.75	0.4677 to 0.9111
Specificity	0.3750	0.1848 to 0.6136
Likelihood ratio	1.20	0.50 to 4.01

The significance of the lack of correlation between PPR and PD disease grade may also lay in the dual nature by which ipRGC and conventional photoreceptors operate in relation to circadian entrainment. Photoreceptors, including ipRGCs as well as rods and cones, contribute to circadian oscillation in two distinct ways, as oscillator components or as mediators of luminous flux that do not involve rhythmicity, a phenomena termed masking [32]. In mammals, light is the most important zeitgeber synchronizing circadian rhythm. Luminous flux can therefore impact human activity in two ways, either by synchronizing an endogenous oscillator, modulated through ipRGCs, which in turn operationalize diurnal activity via a zeitgeber pacemaker, or through masking [33]. Conceptually, masking describes behavioral changes due to exogenous sources of illumination that are non-zeitgeber in origin. For example, exposure to illumination during periods of typical zeitgeber darkness will cause increased activity in a diurnal mammal (positive masking) while decreased illumination during periods of typically zeitgeber daylight results in decreased activity, or negative masking [33]. Positive masking can therefore be considered as a means to enhance activity by light in a diurnal species, while negative masking is used to describe suppression of activity by dark in a diurnal species and by light in a nocturnal species. Therefore, due to masking effects, PD patients with abnormal PPR may indeed display PPR aberration and consequent ipRGC efferent abnormality with resultant sleep and motor abnormality modified by effects of masking in a manner that may not be predictable or expected.

Addressing the lack of PPR abnormality in the two patients with parkinsonian features, previous studies have shown that patients with MSA indeed demonstrate abnormal PLR typified by deficits in constriction and dilation velocities [34]. However, these studies did not examine for PPR abnormality, only dynamics of pupillary constriction and velocity [34]. Although patients with MSA have been shown to have abnormality in measures of dynamic pupillometry, in the present study, neither patient with Lewy body dementia nor MSA demonstrated abnormal PPR typical of ipRGC deficit. Ideally, use of a PLR assessment to distinguish parkinsonian features of MSA or Lewy body dementia from PD would be of significant value. However, based on findings of the present study, it does not appear abnormality of PPR has sufficient sensitivity to be used clinically in this manner

Limitations of this study include a small sample size and the cross-sectional nature of the study design. Assessment of effect size denoted in this study suggests the study is underpowered to determine the true diagnostic accuracy of an abnormal PPR as a rational biomarker for PD. Furthermore, to estimate classification accuracy using standard ROC methods, a gold standard PD PPR status must be ascertained clearly without ambiguity, which is a dilemma as binary PD diagnosis disease classification may be unattainable or inaccurate. Nevertheless, use of ROC-AUC analysis as shown in this study provides a mechanism to both classify and illustrate PPR abnormality in both a qualitative as well as quantitative manner. Future studies that leverage analytical methodology that improve signal strength interpretation in estimating both the presence and extent of PLR abnormality in PD patients are required [34]. In summary, our PD cohort clearly demonstrates aberrant PPR albeit in a variable manner suggesting PD-related disruption of melanopsindriven circadian entrainment with accompanying downstream pathobiological consequences occurs in either a graded fashion, or in a manner which eludes detection by current protocol of pupillary assessment.

**Funding** Partial funding for this project was provided by the Richard Perritt Charitable Trust.

**Data availability** All data associated with this study are freely available online.

Code availability GraphPad Prism version 9.0.0 GraphPad Software, San Diego, CA, USA, www.graphpad.com

#### Declarations

**Ethics approval** This study was reviewed and approved by the Edward Hines Jr. Institutional Review Board and was conducted in accordance with the ethical standards as laid down in the 1964 Declaration of Helsinki and its later amendments.

**Consent to participate and consent for publication** Consent for participating in this study as well as consent for publication was obtained from all study subjects prior to participation in any study-related protocol.

Conflict of interest The authors declare no competing interests.

**Informed consent** Freely provided informed consent documentation acknowledging subject understanding of the purpose, procedures and risks involved in this research study was obtained from all study subjects prior to study participation.

#### References

- Davie CA (2008) A review of Parkinson's disease. Br Med Bull 86: 109–127
- 2. Archibald NK, Clarke MP, Mosimann UP, Burn DJ (2009) The retina in Parkinson's disease. Brain 132:1128–1145
- Harnois C, Di Paolo T (1990) Decreased dopamine in the retinas of patients with Parkinson's disease. Investig Ophthalmol Vis Sci 31: 2473–2475
- Rosenthal NE, Sack DA, Gillin JC, Lewy AJ, Goodwin FK, Davenport Y, Mueller PS, Newsome DA, Wehr TA (1984) Seasonal affective disorder: a description of the syndrome and preliminary findings with light therapy. Arch Gen Psychiatry 41:72– 80. https://doi.org/10.1001/archpsyc.1984.01790120076010
- Terman M, Terman JS (2005) Light therapy for seasonal and nonseasonal depression: efficacy, protocol, safety, and side effects. CNS Spectr 10:647–663
- Willis GL, Turner EJD (2007) Primary and secondary features of Parkinson's disease improve with strategic exposure to bright light: a case series study. Chronobiol Int 24:521–537. https://doi.org/10. 1080/07420520701420717
- Endo T, Matsumura R, Tokuda IT, Yoshikawa T, Shigeyoshi Y, Node K, Sakoda S, Akashi M (2020) Bright light improves sleep in patients with Parkinson's disease: possible role of circadian restoration. Sci Rep 10:7982. https://doi.org/10.1038/s41598-020-64645-6
- Paus S, Schmitz-Hübsch T, Wüllner U, Vogel A, Klockgether T, Abele M (2007) Bright light therapy in Parkinson's disease: a pilot study. Mov Disord 22:1495–1498. https://doi.org/10.1002/mds. 21542
- Willis GL (2008) Intraocular microinjections repair experimental Parkinson's disease. Brain Res 1217:119–131. https://doi.org/10. 1016/j.brainres.2008.03.083
- Willis GL (2008) Parkinson's disease as a neuroendocrine disorder of circadian function: dopamine-melatonin imbalance and the visual system in the genesis and progression of the degenerative process. Rev Neurosci 19:245–316
- Willis GL, Kelly AMA, Kennedy GA (2008) Compromised circadian function in Parkinson's disease: enucleation augments disease severity in the unilateral model. Behav Brain Res 193:37–47. https://doi.org/10.1016/j.bbr.2008.04.017
- Berson DM, Dunn FA, Takao M (2002) Phototransduction by retinal ganglion cells that set the circadian clock. Science 295:1070– 1073. https://doi.org/10.1126/science.1067262
- Skene DJ, Lockley SW, Arendt J (1999) Melatonin in circadian sleep disorders in the blind. NeuroSignals 8:90–95. https://doi. org/10.1159/000014575
- Skene DJ, Arendt J (2007) Circadian rhythm sleep disorders in the blind and their treatment with melatonin. Sleep Med 8:651–655. https://doi.org/10.1016/j.sleep.2006.11.013
- Rodgers J, Hughes S, Pothecary CA, Brown LA, Hickey DG, Peirson SN, Hankins MW (2018) Defining the impact of melanopsin missense polymorphisms using in vivo functional rescue. Hum Mol Genet 27:2589–2603. https://doi.org/10.1093/hmg/ ddy150
- Lucas RJ, Hattar S, Takao M et al (2003) Diminished pupillary light reflex at high irradiances in melanopsin-knockout mice. Science 299:245–247. https://doi.org/10.1126/science.1077293
- Gamlin PDR, McDougal DH, Pokorny J et al (2007) Human and macaque pupil responses driven by melanopsin-containing retinal ganglion cells. Vis Res 47:946–954. https://doi.org/10.1016/j. visres.2006.12.015

- Smilowska K, Van Wamelen DJ, Schoutens AMC et al (2019) Blue light therapy glasses in Parkinson's disease: patients' experience. Parkinson's Dis 2019:1–4. https://doi.org/10.1155/2019/1906271
- Willis GL, Moore C, Armstrong SM (2014) Parkinson's disease, lights and melanocytes: looking beyond the retina. Sci Rep 4. https://doi.org/10.1038/srep03921
- van de Kraats J, van Norren D (2007) Optical density of the aging human ocular media in the visible and the UV. J Opt Soc Am A Opt Image Sci Vis 24:1842. https://doi.org/10.1364/josaa.24.001842
- Hoehn MM, Yahr MD (1967) Parkinsonism: onset, progression, and mortality. Neurology 17:427–441
- Mailankody P, Lenka A, Pal PK (2019) The role of optical coherence tomography in parkinsonism: a critical review. J Neurol Sci 403:67–74. https://doi.org/10.1016/j.jns.2019.06.009
- Park KW, Choi N, Ryu H-S, Kim MS, Lee E-J, Chung SJ (2019) Pupillary dysfunction of multiple system atrophy: Dynamic pupillometric findings and clinical correlations. Parkinsonism Relat Disord 65:234–237
- Adhikari P, Zele AJ, Feigl B (2015) The post-illumination pupil response (PIPR). Investig Ophthalmol Vis Sci 56:3838–3849. https://doi.org/10.1167/iovs.14-16233
- Joyce DS, Feigl B, Kerr G, Roeder L, Zele AJ (2018) Melanopsinmediated pupil function is impaired in Parkinson's disease. Sci Rep 8:1–9. https://doi.org/10.1038/s41598-018-26078-0
- Sengupta P, Dutta K, Ghosh S, Mukherjee A, Pal S, Basu D (2018) Optical coherence tomography findings in patients of Parkinson's disease: an Indian perspective. Ann Indian Acad Neurol 21:150– 155
- Unlu M, Duygu, Sevim G et al (2018) Correlations among multifocal electroretinography and optical coherence tomography findings in patients with Parkinson's disease. Neurol Sci 39:533–541. https://doi.org/10.1007/s10072-018-3244-2
- Lax P, Ortuño-Lizarán I, Maneu V, Vidal-Sanz M, Cuenca N (2019) Photosensitive melanopsin-containing retinal ganglion cells in health and disease: implications for circadian rhythms. Int J Mol Sci 20. https://doi.org/10.3390/ijms20133164
- Korshunov KS, Blakemore LJ, Trombley PQ (2017) Dopamine: a modulator of circadian rhythms in the central nervous system. Front Cell Neurosci 11:1–17. https://doi.org/10.3389/fncel.2017.00091
- Do MTH (2010) Intrinsically photosensitive retinal ganglion cells. Physiol Rev 90:1547–1581. https://doi.org/10.1152/physrev. 00013.2010.Intrinsically
- Klein MO, Battagello DS, Cardoso AR, Hauser DN, Bittencourt JC, Correa RG (2019) Dopamine: functions, signaling, and association with neurological diseases. Cell Mol Neurobiol 39:31–59. https://doi.org/10.1007/s10571-018-0632-3
- Panda S, Sato TK, Castrucci AM et al (2002) Melanopsin (Opn4) requirement for normal light-induced circadian phase shifting. Science 298:2213–2216. https://doi.org/10.1126/science.1076848
- Mrosovsky N (1999) Masking: history, definitions, and measurement. Chronobiol Int 16:415–429
- Park KW, Choi N, Ryu HS, Kim MS, Lee EJ, Chung SJ (2019) Pupillary dysfunction of multiple system atrophy: dynamic pupillometric findings and clinical correlations. Parkinsonism Relat Disord 65:234–237. https://doi.org/10.1016/j.parkreldis. 2019.05.003
- 35. Tabashum T, Zaffer A, Yousefzai R, Colletta K, Jost MB, Park Y, Chawla J, Gaynes B, Albert MV, Xiao T (2021) Detection of Parkinson's Disease Through Automated Pupil Tracking of the Post-illumination Pupillary Response. Front Med 8

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.