



Singular cases of Alzheimer's disease disclose new and old genetic “acquaintances”

Cinzia Coppola^{1,2} · Dario Saracino¹ · Mariano Oliva¹ · Lorenzo Cipriano¹ · Gianfranco Puoti¹ · Sabina Pappatà^{3,4} · Giuseppe Di Fede⁵ · Marcella Catania⁵ · Martina Ricci⁵ · Sara Cimini⁵ · Giorgio Giaccone⁵ · Simona Bonavita¹ · Giacomina Rossi⁵

Received: 18 June 2020 / Accepted: 25 September 2020 / Published online: 2 October 2020

© The Author(s) 2020

Abstract

Background Alzheimer's disease (AD) is the most common age-related dementia. Besides its typical presentation with amnesic syndrome at onset, atypical AD cases are being increasingly recognized, often in presenile age.

Objectives To provide an extensive clinical and genetic characterization of six AD patients carrying one or more singular features, including age of onset, atypical phenotype and disease progression rate. By reviewing the pertinent literature and accessing publicly available databases, we aimed to assess the frequency and the significance of the identified genetic variants.

Methods Biomarkers of amyloid- β deposition and neurodegeneration were used to establish the in vivo diagnosis of probable AD, in addition to neurological and neuropsychological evaluation, extensive laboratory assays and neuroradiological data. Considering the presenile onset of the majority of the cases, we hypothesized genetically determined AD and performed extensive genetic analyses by both Sanger sequencing and next generation sequencing (NGS).

Results We disclosed two known missense variants, one in *PSEN1* and the other in *PSEN2*, and a novel silent variant in *PSEN2*. Most notably, we identified several additional variants in other dementia-related genes by NGS. Some of them have never been reported in any control or disease databases, representing variants unique to our cases.

Conclusions This work underlines the difficulties in reaching a confident in vivo diagnosis in cases of atypical dementia. Moreover, a wider genetic analysis by NGS approach may prove to be useful in specific cases, especially when the study of the so-far known AD causative genes produces negative or conflicting results.

Keywords Alzheimer's disease · Mutation · Dementia · Amyloid · Genetics · Biomarkers

Introduction

Alzheimer's disease (AD) is the most common age-related degenerative dementia. From a clinical perspective, AD typically displays an amnesic syndrome of the hippocampal type that can be associated with various cognitive or behavioural deficits during disease evolution [1]. Atypical forms of AD present with relative preservation of memory at onset and generally occur at an earlier age. They include posterior, logopenic and frontal variant of AD [1]. According to the revised international criteria, at least one biomarker of in vivo Alzheimer's pathology must be positive: a cerebrospinal fluid (CSF) profile consisting of decreased amyloid- β 1–42 ($A\beta_{42}$) together with increased total tau (T-tau) or 181-phosphorylated tau (P-tau) concentrations, or an increased retention on amyloid tracer PET (AMY-PET) [1]. In addition to the use of single CSF markers, the combination of multiple

✉ Cinzia Coppola
cinzia.coppola@unicampania.it

¹ Department of Advanced Medical and Surgical Sciences, University of Campania “L. Vanvitelli”, Naples, Italy

² Second Division of Neurology, University of Campania “Luigi Vanvitelli”, Isola 8 – Edificio 10 Policlinico “Federico II” via Pansini 5, 80131 Naples, Italy

³ Institute of Biostructure and Bioimaging, National Council of Research, Naples, Italy

⁴ Department of Advanced Biomedical Sciences, Federico II University, Naples, Italy

⁵ Division of Neurology V – Neuropathology, Fondazione IRCCS Istituto Neurologico Carlo Besta, Milan, Italy

CSF markers in the form of ratios further increases the diagnostic accuracy [2]. AD is usually sporadic, with age of onset most often being > 65 years, thus qualifying for late onset AD (LOAD). In no more than 5% of all patients, a positive familial history for dementia or a clear-cut autosomal dominant pattern of inheritance can be found. These familial AD cases (FAD) arise before age 65 more frequently than sporadic cases, hence the definition of early onset AD (EOAD) [3]. Approximately 50% of FAD patients carry a mutation in presenilin 1 (*PSEN1*), presenilin 2 (*PSEN2*) or amyloid- β protein precursor (*APP*) genes, with more than 350 variants collectively identified so far [4, 5]. However, some of them are not pathogenic or their significance remains uncertain, as they may qualify as genetic risk factors or disease-modifying alterations.

Here, we describe a case series of cognitive disorders with in vivo biomarker positivity for A β deposition, showing various clinical atypical aspects together with peculiar genetic features. We disclosed two known missense variants, one in *PSEN1* and the other in *PSEN2*, and a novel silent variant in *PSEN2*. Moreover, additional variants in dementia-related genes have been identified by next generation sequencing (NGS). Our results are intriguing as they raise the question of the role of genetic risk burden in AD.

Patients and methods

Subjects

We describe 6 unrelated cases affected by cognitive disorders who underwent a complete diagnostic protocol including neurological and neuropsychological evaluation, extensive laboratory assays, EEG, structural (CT or MR) and functional (^{18}F FDG-PET) neuroimaging. The research for AD pathophysiological biomarkers, either CSF A β_{42} , T-tau and P-tau assay or amyloid tracer PET, was performed in all patients.

Case 1

This patient insidiously presented at age 55 with short-term memory impairment and apathy. Familial history and neurological examination were negative, except for Epstein sign; MMSE was 22/30. An extensive neuropsychological evaluation showed deficits of long- and short-term memory, language, abstract reasoning, executive functions and a marked anosognosia. Brain MRI disclosed diffuse cortical atrophy, while ^{18}F FDG-PET (Fig. 1) revealed bilateral hypometabolism in the frontal dorso-lateral, superior parietal, temporo-parietal cortices, with prevalent involvement of the left hemisphere, and posterior cingulate cortex (PCC). AMY-PET showed increased uptake mainly in the frontal and lateral temporal regions.

Case 2

In this patient, the onset of cognitive impairment was approximately at 75 years and characterized by short-term memory deficits, anomia, subtle behavioural changes (mild disinhibition) and in the following months psychomotor slowing. Familial history was negative. Neurological examination disclosed an asymmetric parkinsonian syndrome (R > L). MMSE was 22/30. Neuropsychological testing revealed deficits in verbal memory, attention, abstract reasoning and semantics. Brain MRI showed moderate atrophy in frontal, lateral temporal and temporo-mesial cortex prevailing on the left side. AMY-PET evidenced a massive and diffuse burden of amyloid- β . His extrapyramidal syndrome showed satisfying response to L-Dopa administration.

Case 3

This woman presented with apathy and short-term memory deficit at the age of about 62. Family history evidenced memory disturbances in her mother and grandmother. MMSE was 27/30. Neuropsychological assessment detected long-term verbal memory and attentional deficits. Brain CT revealed diffuse supratentorial white matter hypodensity, while ^{18}F FDG-PET (Fig. 1) showed mild hypometabolism mainly affecting the left hemisphere and involving the mesial and lateral temporal cortex, the dorsolateral/medial frontal cortex and to a lesser extent the PCC. AMY-PET evidenced a diffuse amyloid deposition.

Case 4

This patient, without family history of dementia, around the age of 59 developed apathy with a language disorder characterized by word-finding problems and slow, hesitating speech, followed by psychomotor agitation, delusional ideation, clumsiness of his upper left limb and generalized motor slowness. His language got significantly worse, with agrammatism and telegraphic sentences, but with only mild impairment in comprehension. Neurological examination at age 61 disclosed mixed pyramidal and extrapyramidal syndrome, prevailing on the left side, left cortical sensory loss and frontal release signs. His MMSE score was 7/30, being non-fluent aphasia with features of apraxia of speech and dressing apraxia among the most significant cognitive deficits. Brain MRI revealed discrete atrophy mainly in temporo-insular cortices bilaterally, whereas ^{18}F FDG-PET (Fig. 1) disclosed severe and diffuse cortical hypometabolism more marked in temporo-parietal cortices, precuneus and PCC bilaterally, with prevalent involvement of the right side and slight striatal metabolic asymmetry (R < L). AMY-PET detected diffuse burden of amyloid- β .

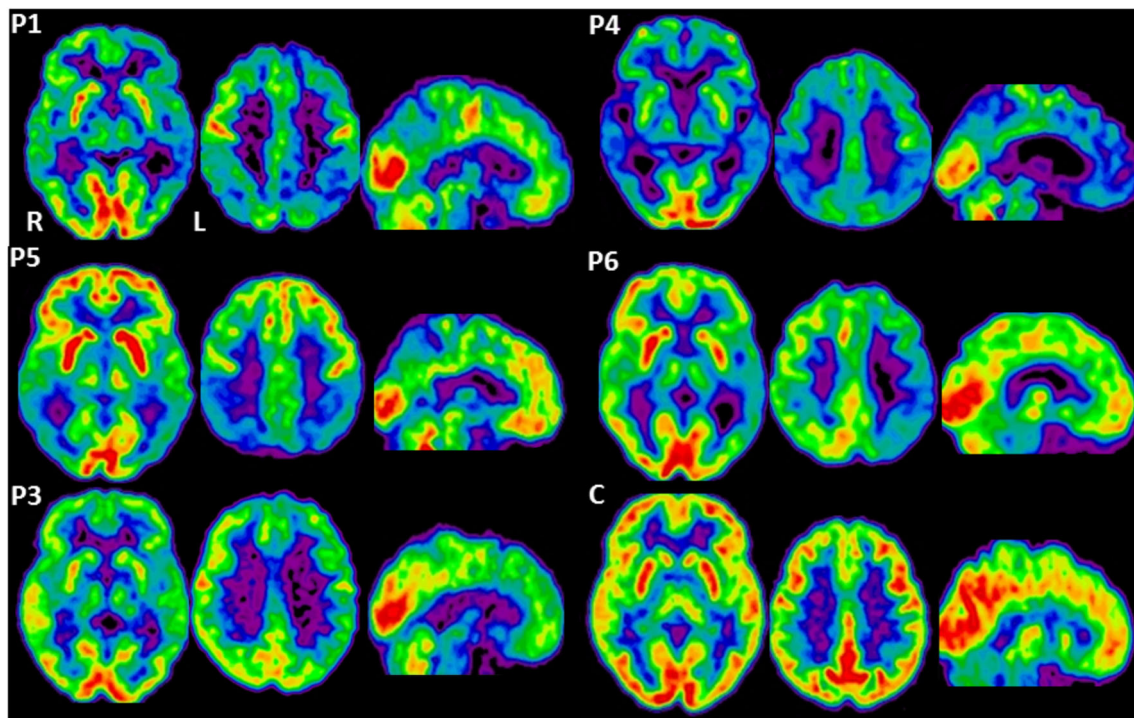


Fig. 1 PET transaxial images at two axial and one sagittal section show ^{18}F FDG uptake in the patients (P1, P4, P5, P6, P3) and in a control subject (C). Compared with control, P1, P4, and P5 showed a relative hypometabolism involving mainly the posterior cortical regions

including the PCC with some spread to the frontal cortex in P1, P4 and to the striatum (R < L) in P4. P6 and P3 show less marked asymmetric hypometabolism (L < R) involving also the striatum (L < R)

Case 5

This subject developed at age 45 a complex behavioural syndrome characterized by apathy, social withdrawal, and eating and sleep disorders. Familial history was negative. Over the next 5 years, there was a clinical worsening with word-finding difficulties, dyscalculia, memory deficits and motor clumsiness in both hands. At age 50, his MMSE score was 16/30 and he presented ideo-motor apraxia, anomia, verbal memory deficits and dysexecutive syndrome. EEG showed diffuse slowing of cerebral electric activity, while brain MRI detected atrophy in parietal regions with slight right prevalence. Subsequently, he also manifested limb myoclonus and psychomotor agitation with complex visual hallucinations. Seven years after the symptom onset, the patient came to our observation and underwent a more extensive diagnostic protocol. Neurological examination showed left pyramidal and bilateral asymmetric (L > R) extrapyramidal syndrome, action-induced limb myoclonus and Epstein sign. MMSE score was 8/30. Brain MRI evidenced marked and diffuse atrophy, with posterior predominance. ^{18}F FDG-PET (Fig. 1) demonstrated bilateral hypometabolism in the parietal, occipital and temporal lobes and in the PCC with relative sparing of frontal lobes and subcortical structures. The occipital hypometabolism mostly involved the associative visual regions with relative sparing of the primary visual cortex. CSF biomarkers assay revealed reduced $\text{A}\beta_{42}$ (456 pg/mL; normal values – n.v. – >

500 pg/mL) [6] and a massive increase of both T-tau (3435 pg/mL; n.v. < 300 pg/mL) [6] and P-tau (470 pg/mL, n.v. < 61 pg/mL) [7]. T-tau/ $\text{A}\beta_{42}$ ratio was 7.533 (n.v. ≤ 0.52), whereas P-tau/ $\text{A}\beta_{42}$ was 1.031 (n.v. ≤ 0.08) [2].

Case 6

This case, without family history for cognitive disorders, insidiously presented at age 52 with a language disorder characterized by anomia and apraxia of speech which progressively worsened until mutism. Neurological examination showed a “worried” facial expression, asymmetric (R > L) mixed pyramidal and extrapyramidal syndrome, focal and segmental myoclonus, exaggerated startle reaction and frontal release signs. A neuropsychological examination showed severe non-fluent aphasia with almost complete mutism and slightly impaired comprehension, severe bucco-lingual and ideomotor apraxia. EEG showed marked slowing in cerebral electric activity. Brain MRI revealed asymmetrical cortico-subcortical atrophy prevailing in left fronto-temporal areas. ^{18}F FDG-PET (Fig. 1) showed asymmetric cortical hypometabolism characterized by a prevalent involvement of the left temporo-parietal cortex and, to a lesser extent, of the left premotor-motor and sensorimotor regions. In addition, there was also a mild left striatal and thalamic hypometabolism. CSF $\text{A}\beta_{42}$ was decreased (235 pg/mL; n.v. > 500 ng/mL), whereas T-tau and P-tau were normal; T-tau/

$A\beta_{42}$ and P-tau/ $A\beta_{42}$ ratios were 1.247 and 0.145 respectively. The research of 14.3.3 protein in CSF was negative.

Patient consents

Written informed consent was acquired from all patients for genetic analysis, processing data and permission to publish data in respect of privacy.

Biochemical analysis

CSF levels of T-tau, P-tau and $A\beta_{42}$ were determined with human specific ELISA kits (Innogenetics). Plasma level of progranulin was measured using an ELISA kit (Human Progranulin ELISA kit, Adipogen Inc., Seoul, Korea).

Genetic analysis

Sanger Sequencing of *APP*, *PSEN1* and *PSEN2* genes [8, 9] and *APOE* genotyping [10] was performed in all cases. Additionally, a gene panel of 48 dementia-related genes was analysed by NGS techniques. Nextera Rapid Capture system for enrichment (Illumina) coupled with gene-specific probes (Integrated DNA Technologies) was used to sequence the following genes: *APP*, *PSEN1*, *PSEN2*, *PRNP*, *GRN*, *MAPT*, *CHMP2B*, *FUS*, *TARDBP*, *VCP*, *TREM2*, *ABCA7*, *APOE*, *BINI*, *CALH1*, *CCL2*, *CCNF*, *CD33*, *CHCHD10*, *CLU*, *CSF1R*, *CST3*, *CTSF*, *DCTN1*, *FLNC*, *hnRNPA1*, *hnRNPA2B1*, *ITM2B*, *LRRK2*, *NCSTN*, *NOS3*, *NOTCH3*, *OPTN*, *PFN1*, *PLD3*, *PRKAR1B*, *SERPINI1*, *SIGMAR1*, *SNCA*, *SNCB*, *SORL1*, *SQSTM1*, *STH*, *TBK1*, *TMEM106B*, *TUBA4A*, *TYROBP*, *UBQLN2*. Sequencing was performed on the Illumina MiSeq instrument using 2X150 bp paired-end read cycles. MiSeq Reporter software (Illumina) was used for alignment (reference human genome UCSC hg19) and variant calling. Variants were annotated using Variant Studio software (Illumina). Low-quality variants were filtered out using the Illumina Qscore threshold of 30; in addition, variants with a minor allele frequency higher than 2% in GnomAD (Genome Aggregation Database, <http://gnomad.broadinstitute.org/>) were filtered out. Variants of interest were confirmed using standard Sanger sequencing.

Sorting Intolerant From Tolerant (SIFT) and Polymorphism Phenotyping (PolyPhen) softwares were used to predict pathogenicity of missense mutations. Combined annotation-dependent depletion (CADD) score (<https://cadd.gs.washington.edu/>) was used to predict the pathogenicity of a truncating variant (*SORL1* Ser10STOP). NetGene2 (<http://www.cbs.dtu.dk/services/NetGene2/>) and BDGP (http://www.fruitfly.org/seq_tools/splice.html) splice site prediction tools were used to predict the effect on the splice site of the *DCTN1* c.3529 + 5G > A variant.

Results

Clinical, instrumental and CSF findings

Our series consists of six cases whose clinical features are summarized in Table 1. Disease onset was in the presenile period in all patients (mean age of onset: 54.6 ± 6.6), except for case 2. Only case 3 showed family history for dementia. The onset was typical in two patients (cases 1 and 3) and atypical in the others. Moreover, an extrapyramidal syndrome complicated all these atypical cases. The clinical diagnosis was AD in cases 1, 2, 3 and 4. In case 5, there was an important discrepancy between clinical findings, suggestive of behavioural variant of frontotemporal dementia (bvFTD) with parkinsonism, and MRI and ^{18}F FDG-PET data, expression of atypical AD. In case 6, the clinical diagnosis was corticobasal syndrome (CBS). Given the peculiarity of disease onset, plasma progranulin dosage was performed in cases 4, 5 and 6, with normal values. In addition, all patients underwent an *APOE* genotyping, which only in cases 2 and 3 showed a $\epsilon 3/\epsilon 4$ heterozygosity. The diagnosis of probable AD was supported by at least one positive pathophysiological biomarker in all cases: AMY-PET in cases 1, 2, 3 and 4 and CSF biomarkers in cases 5 and 6. In case 6, although T-tau and P-tau values were not increased, T-tau/ $A\beta_{42}$ and P-tau/ $A\beta_{42}$ ratios both resulted well above the standardized cut-offs [2], thus strongly suggesting an underlying AD pathology.

Genetic findings

Diagnostic genes (*APP*, *PSEN1*, *PSEN2*, *PRNP*, *GRN*, *MAPT*, *CHMP2B*, *FUS*, *TARDBP*, *VCP*, *TREM2*) were sequenced at 100% by NGS (read depth $\geq 20\text{X}$) or, in some cases with incomplete coverage, by standard Sanger technique. Genetic results are described in Table 2 and Table 3. Population frequency, in silico pathogenicity prediction (SIFT and Polyphen) and classification in Human Gene Mutation Database (HGMD) are presented. Briefly, concerning AD-causative genes (Table 2), we identified two known missense variants, Glu318Gly in *PSEN1* (patients 1 and 2) and Arg71Trp in *PSEN2* (patient 3), and a novel silent variant, Ser236Ser in *PSEN2* (patient 4). Moreover, thanks to NGS approach, we disclosed other variants in dementia-related genes, in particular *FUS*, *ABCA7*, *CSF1R*, *DCTN1*, *SERPINI1* and *SORL1* (Table 3).

Some variants have never been reported in any control (GnomAD) or disease (HGMD) databases, representing variants unique to our cases. CADD analysis of the *SORL1* Ser10STOP variant predicted pathogenicity, as well as NetGene2 and BDGP predictions of the *DCTN1* c.3529 + 5G > A splice variant.

Table 1 Clinical, instrumental and laboratory data of patients

Case	Age at onset (y)	Cognitive symptoms at onset	Motor syndrome	MRI/CT	¹⁸ F-DG-PET	Amyloid tracer PET	CSF biomarkers				Plasma progranulin	APOE	Diagnosis	
							Aβ ₄₂ tau	T-tau	P-tau	P-tau/Aβ ₄₂				
1	55	Memory and attention disorders	Absent	Diffuse cortical atrophy	Bilateral temporo-parietal, precuneus, PCC and frontal dorso-lateral cortical hypometabolism (L < R) n.a.	Increased cortical uptake in the frontal and lateral temporal regions Marked and diffuse uptake	n.a.	n.a.	n.a.	n.a.	n.a.	3/3	EOAD	
2	75	Memory deficit and behavioural syndrome	Asymmetric parkinsonism (R > L) L-Dopa responsive	Asymmetric fronto-temporal atrophy with left prevalence	Asymmetric temporo-parietal, precuneus and PCC hypometabolism (R < L); slight striatal hypometabolism (R < L)	Increased cortical uptake in the frontal, lateral temporal and parietal regions	n.a.	n.a.	n.a.	n.a.	n.a.	3/4	Atypical LOAD	
3	62	Apathy and memory deficit	Absent	Diffuse supratentorial white matter hypodensity	Mild left mesial and lateral temporal and frontal cortices hypometabolism; very mild left PCC and putamen hypometabolism	Increased cortical uptake in the frontal, lateral temporal and parietal regions	n.a.	n.a.	n.a.	n.a.	n.a.	3/4	Familial EOAD	
4	59	Apathy, apraxia, non-fluent aphasia	Mixed pyramidal and extrapyramidal syndrome (L > R)	Temporo-insular atrophy	Marked diffuse bilateral cortical hypometabolism more evident in the temporo-parietal cortex, precuneus and PCC (R < L); slight striatal hypometabolism (R < L)	Diffuse uptake	n.a.	n.a.	n.a.	n.a.	148.9	3/3	Atypical EOAD	
5	45	Behavioural syndrome, sleep disorder	Left pyramidal and asymmetric extrapyramidal syndrome (L > R); myoclonus	Bilateral posterior (mainly parietal) cortical atrophy	Marked bilateral temporal, parietal, associative occipital cortex and PCC hypometabolism.	n.a.	456	3435	470	7.533	1.031	135.6	3/3	Atypical EOAD
6	52	Speech disorders	Mixed pyramidal and extrapyramidal syndrome (R > L); myoclonus	Asymmetrical cortico-subcortical atrophy with left fronto-temporal prevalence	Cortical temporal, parietal and frontal premotor and sensorimotor hypometabolism (L < R); mild left striatal, thalamic and PCC hypometabolism	n.a.	235	293	34	1.247	0.145	100.3	3/3	Atypical EOAD presenting as CBS

EOAD, early-onset Alzheimer’s disease; LOAD, late-onset Alzheimer’s disease; n.a., not available; PCC, posterior cingulate cortex; y, years; CBS, cortico-basal syndrome

Table 2 DNA variants found in genes causative for AD

Case	Gene	Coordinates	Transcript	DNA variant	Amino acid variant	Sift	PolyPhen	GnomAD Freq %	HGMD classification
1	<i>PSEN1</i>	73,673,178	NM_000021.3	gAa/gGa	Glu318Gly	tol	ben	1.485	Disease-associated polymorphism with supporting functional evidence
2	<i>PSEN1</i>	73,673,178	NM_000021.3	gAa/gGa	Glu318Gly	tol	ben	1.485	Disease-associated polymorphism with supporting functional evidence
3	<i>PSEN2</i>	227,071,475	NM_000447.2	Cgg/Tgg	Arg71Trp	del	ben	0.3836	Disease causing mutation?
4	<i>PSEN2</i>	227,076,671	NM_000447.2	agT/agC	Ser236Ser	-	-	1.331	No

Sift, Sorting Intolerant From Tolerant software; *PolyPhen*, Polymorphism Phenotyping software; *GnomAD*, Genome Aggregation Database; *HGMD*, Human Gene Mutation Database; *ben*, benign; *del*, deleterious; *tol*, tolerated

Discussion

Alzheimer's disease is mainly distinguished in a typical presentation with hippocampal amnesic syndrome and atypical forms with different cognitive or behavioural deficits.

In this paper, we describe a series of 6 unrelated patients affected by dementing syndromes characterized by one or more “atypical” features including age at onset, clinical presentation and disease progression rate. Case 5 presented a complex syndrome indicative of bvFTD with parkinsonism and additional atypical features. The severity of clinical picture and the high levels of CSF tau might suggest the possibility of a prion disease. However, the long course, the MRI features, the neuroimaging findings (parieto-temporal atrophy and hypometabolism) and CSF Aβ₄₂ reduction made presenile AD the most likely diagnosis. Case 6 was classified as possible CBS, a clinical syndrome

with different underlying pathological substrates [11, 12]. In vivo AD pathophysiological biomarkers and ¹⁸F-DG-PET hypometabolic pattern suggested an underlying AD pathology (CBS-AD), in agreement with the results of a recent combined ¹⁸F-DG-PET/neuropathological study [13]. Notably, in all patients, the in vivo AD pathophysiological biomarkers supported the diagnosis of probable AD. Indeed, these biomarkers should always be looked for, together with the downstream degenerative topographical biomarkers (¹⁸F-DG-PET, MRI), in atypical dementia cases.

The results of genetic analyses were, in our opinion, very interesting. The variant found in cases 1 and 2, *PSEN1* Glu318Gly, was first identified in patients with EOAD [14]. Studies performed to define its effects on amyloid-β metabolism gave conflicting results [15, 16], and association studies were inconclusive [16, 17]. The variant disclosed in case 3,

Table 3 DNA variants found in other dementia-related genes

Case	Gene	Coordinates	Transcript	DNA variant	Amino acid variant	Sift	PolyPhen	GnomAD Freq %	HGMD classification
1	<i>ABCA7</i>	1047345	NM_019112.3	Gac/Tac	Asp679Tyr	del	prob dam	0	No
	<i>SORL1</i>	121425954	NM_003105.5	aCa/aTa	Thr833Ile	del	poss dam	0	No
2	<i>FUS</i>	31196452	NM_004960.3	tAt/tGt	Tyr239Cys	tol	prob dam	0.001698	No
5	<i>DCTN1</i>	74598723	NM_004082.4	Atc/Gtc	Ile196Val	tol	ben	0.4519	Functional polymorphism
	<i>SORL1</i>	121323069	NM_003105.5	tCg/tAg	Ser10STOP*	-	-	0	No
6	<i>CSF1R</i>	149456911	NM_005211.3	Gcc/Acc	Ala273Thr	tol	prob dam	0	No
	<i>DCTN1</i>	74590116	NM_004082.4	c.3529 + 5G > A [†]		-	-	0.59160	No
	<i>SERPINI1</i>	167512569	NM_001122752.1	Gca/Aca	Ala280Thr	tol	ben	1.125	No

Sift, Sorting Intolerant From Tolerant software; *PolyPhen*, Polymorphism Phenotyping software; *GnomAD*, Genome Aggregation Database; *HGMD*, Human Gene Mutation Database; *ben*, benign; *del*, deleterious; *dam*, damaging; *poss*, possibly; *prob.*, probably; *tol*, tolerated

*Combined annotation dependent depletion (CADD) score (<https://cadd.gs.washington.edu/>) was > 35

[†]NetGene2 (<http://www.cbs.dtu.dk/services/NetGene2/>) and BDGP (http://www.fruitfly.org/seq_tools/splice.html) splice site prediction tools predicted loss of splice site

PSEN2 Arg71Trp, probably involved in protein stability and signalling pathways [18], has been found in patients with EOAD or LOAD, as well as in healthy subjects and Parkinson's disease dementia [19, 20], and only in one large AD family it seemed to clearly segregate with the disease [21, 22]. It is possible that, by interacting with other factors, *PSEN1* Glu318Gly and *PSEN2* Arg71Trp increase disease risk and modulate clinical phenotype. *PSEN2* Ser236Ser, present in case 4, is a silent variant whose pathogenicity is not predictable.

Among the relevant findings of NGS analysis, *ABCA7* and *SORL1* are well-known AD risk genes [23, 24]. The *ABCA7* transporter is involved in A β clearance and its mutations accelerate amyloidosis in a mouse model of AD [25]. A strong association was demonstrated between *ABCA7* variations and amyloidosis in AD patients [26]. A reduced expression of *SORL1*, promoter of the APP non-amyloidogenic pathway [27], has been demonstrated in human AD brains, and its genetic variants increase risk of both LOAD and EOAD [28]. In patient 1, we identified the *ABCA7* Asp679Tyr and the *SORL1* Thr833Ile variants. They had never been reported before but are predicted to be deleterious by in silico analyses, therefore possibly exerting a synergistic effect with the *PSEN1* Glu318Gly variant in amyloidogenic process.

Patient 2, affected by LOAD with parkinsonism, harboured the Tyr239Cys variant in *FUS*, a gene implicated in ALS and FTD cases [29]. This variant is present in GnomAD with a very low frequency and is predicted to be deleterious by some in silico analyses.

In patient 5, we found the Iso196Val variant in *DCTN1* gene. Several *DCTN1* mutations have been described in association with ALS, degenerative parkinsonisms and Perry syndrome [30, 31]. Interestingly, our patient displayed some features of Perry syndrome at disease onset, such as personality change, and eating and sleep disturbances, while parkinsonism occurred thereafter. However, in vivo biomarkers more likely predicted amyloid- β rather than TDP-43 pathology, which is Perry syndrome's substrate. Despite some evidence of pathogenicity from in vitro studies [32], *DCTN1* Iso196Val variant has been reported both in patients and in several healthy controls, making it a possible risk factor rather than a causative mutation. This patient also presented the Ser10STOP variant in *SORL1*, which is a truncating variant absent in ExAc (Exome Aggregation Consortium, <http://exac.broadinstitute.org/>) and GnomAD databases, with a CADD score of 35: these types of variant are considered as definitely pathogenic and associated with a significant 12-fold increased AD risk, which is comparable with the *APOE*- ϵ 4 homozygosity effect [33]. Rare pathogenic *SORL1* mutations segregate with disease in LOAD families, and their pathological mechanism is likely to be haploinsufficiency [34].

In case 6, we found variants in other dementia-related genes. The novel Ala273Thr variant, predicted as damaging by in silico analysis, was identified in *CSF1R*. *CSF1R*

mutations are causative of adult-onset leukoencephalopathy with axonal spheroids and pigmented glia [35], and have recently been reported in pathologically confirmed AD subjects [36]. Noteworthy, one of these cases exhibited a clinical picture very similar to that of our case. We can therefore hypothesize that rare variants of *CSF1R* may influence the susceptibility to AD, as already shown for other adult-onset leukodystrophy causative genes, such as *TREM2* and *NOTCH3* [37, 38]. Mutations in *SERPINI1* are responsible for familial encephalopathy with neuroserpin inclusion bodies [39]. Though rapidly progressive dementia and myoclonus belong to the clinical spectrum of *SERPINI1* mutations [40], the Ala280Thr variant found in patient 6 is predicted as tolerated by in silico analyses. Finally, the splicing mutation c.3529 + 5G > A identified in *DCTN1* gene is predicted as potentially capable of altering the splicing site by in silico analyses; therefore, a possible pathogenic effect cannot be excluded.

In conclusion, two relevant aspects emerge from the observations made on this case series. First, some of the patients here presented are paradigmatic of the difficulties in reaching a confident in vivo diagnosis due to the "atypical" clinical aspects, despite the application of very extensive diagnostic protocols. Therefore, post-mortem neuropathological examination remains the gold standard to definitely elucidate the nature of the neurodegenerative process in the single patient with atypical dementia.

Second, in this series of cases, it is also possible to highlight the very interesting aspects emerging from a wider than standard genetic analysis. We found the coexistence of more than one rare non-causative genetic variant in 4 out of 6 patients, suggesting an additive contribution of them to develop dementia, whereas each single variant may not be sufficient. This raises a crucial question: what is the role of these non-causative mutations that are increasingly found in different neurological disorders, particularly in dementias? One hypothesis is that they could act as risk or modifier factors to the disease. Further studies adding evidence from NGS data to the current knowledge will be necessary to support this hypothesis and to define the individual risk associated to each variant.

Acknowledgements Open access funding provided by Università degli Studi della Campania Luigi Vanvitelli within the CRUI-CARE Agreement.

Author contribution All authors have reviewed the contents of the manuscript being submitted, approved its contents and validated the accuracy of the data.

Data availability There are no figures, videos or other data which could allow the identification of the subjects.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflicts of interest.

Ethical approval No experimental procedure was performed. All the investigations carried out were part of the diagnostic protocol. Therefore, we had no need to submit this study to the approval of the Ethics Committee. Instead, written informed consent was acquired for the diagnostic procedures to reach the diagnosis and for the use of data for research purposes in respect of privacy.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Dubois B, Feldman HH, Jacova C, Hampel H, Molinuevo JL, Blennow K, DeKosky ST, Gauthier S, Selkoe D, Bateman R, Cappa S, Crutch S, Engelborghs S, Frisoni GB, Fox NC, Galasko D, Habert M-O, Jicha GA, Nordberg A, Pasquier F, Rabinovici G, Robert P, Rowe C, Salloway S, Sarazin M, Epelbaum S, de Souza LC, Vellas B, Visser PJ, Schneider L, Stern Y, Scheltens P, Cummings JL (2014) Advancing research diagnostic criteria for Alzheimer's disease: the IWG-2 criteria. *Lancet Neurol* 13:614–629
- Duits FH, Teunissen CE, Bouwman FH, Visser P-J, Mattsson N, Zetterberg H, Blennow K, Hansson O, Minthon L, Andreasen N, Marcusson J, Wallin A, Rikkert MO, Tsolaki M, Parnetti L, Herukka S-K, Hampel H, De Leon MJ, Schröder J, Aarsland D, Blankenstein MA, Scheltens P, van der Flier WM (2014) The cerebrospinal fluid "Alzheimer profile": easily said, but what does it mean? *Alzheimers Dement* 10:713–723.e2
- Wu L, Rosa-Neto P, Hsiung G-YR, Sadovnick AD, Masellis M, Black SE, Jia J, Gauthier S (2012) Early-onset familial Alzheimer's disease (EOFAD). *Can J Neurol Sci* 39:436–445
- Bertram L, McQueen MB, Mullin K, Blacker D, Tanzi RE (2007) Systematic meta-analyses of Alzheimer disease genetic association studies: the AlzGene database. *Nat Genet* 39:17–23
- Stenson PD, Mort M, Ball EV, Evans K, Hayden M, Heywood S, Hussain M, Phillips AD, Cooper DN (2017) The Human Gene Mutation Database: towards a comprehensive repository of inherited mutation data for medical research, genetic diagnosis and next-generation sequencing studies. *Hum Genet* 136:665–677
- Sjögren M, Vanderstichele H, Agren H, Zachrisson O, Edsbacke M, Wikkelso C, Skoog I, Wallin A, Wahlund LO, Marcusson J, Nägga K, Andreasen N, Davidsson P, Vanmechelen E, Blennow K (2001) Tau and Abeta42 in cerebrospinal fluid from healthy adults 21–93 years of age: establishment of reference values. *Clin Chem* 47:1776–1781
- Vanderstichele H, De Vreese K, Blennow K, Andreasen N, Sindic C, Ivanou A, Hampel H, Bürger K, Parnetti L, Lanari A, Padovani A, DiLuca M, Bläser M, Olsson AO, Pottel H, Hulstaert F, Vanmechelen E (2006) Analytical performance and clinical utility of the INNOTEST PHOSPHO-TAU(181P) assay for discrimination between Alzheimer's disease and dementia with Lewy bodies. *Clin Chem Lab Med* 44:1472–1480
- Mullan M, Crawford F, Axelman K, Houlden H, Lilius L, Winblad B, Lannfelt L (1992) A pathogenic mutation for probable Alzheimer's disease in the APP gene at the N-terminus of beta-amyloid. *Nat Genet* 1:345–347
- Cruts M, van Duijn CM, Backhovens H, Van den Broeck M, Wehnert A, Semeels S, Sherrington R, Hutton M, Hardy J, St George-Hyslop PH, Hofman A, Van Broeckhoven C (1998) Estimation of the genetic contribution of presenilin-1 and -2 mutations in a population-based study of presenile Alzheimer disease. *Hum Mol Genet* 7:43–51
- Wenham PR, Price WH, Blandell G (1991) Apolipoprotein E genotyping by one-stage PCR. *Lancet* 337:1158–1159
- Josephs KA, Hodges JR, Snowden JS, Mackenzie IR, Neumann M, Mann DM, Dickson DW (2011) Neuropathological background of phenotypic variability in frontotemporal dementia. *Acta Neuropathol* 122:137–153
- Armstrong MJ, Litvan I, Lang AE, Bak TH, Bhatia KP, Borroni B, Boxer AL, Dickson DW, Grossman M, Hallett M, Josephs KA, Kertesz A, Lee SE, Miller BL, Reich SG, Riley DE, Tolosa E, Troster AI, Vidailhet M, Weiner WJ (2013) Criteria for the diagnosis of corticobasal degeneration. *Neurology* 80:496–503
- Pardini M, Huey ED, Spina S, Kreis WC, Morbelli S, Wassermann EM, Nobili F, Ghetti B, Grafman J (2019) FDG-PET patterns associated with underlying pathology in corticobasal syndrome. *Neurology* 92:e1121–e1135
- Sandbrink R, Zhang D, Beyreuther K, Schaeffer S, Bauer J, Masters CL, Förstl H (1996) Missense mutations of the PS-1/S182 gene in German early-onset Alzheimer's disease patients. *Ann Neurol* 40:265–266
- Dermaut B, Cruts M, Slooter AJC, Van Gestel S, De Jonghe C, Vanderstichele H, Vanmechelen E, Breteler MM, Hofman A, van Duijn CM, Van Broeckhoven C (1999) The Glu318Gly substitution in presenilin 1 is not causally related to Alzheimer disease. *Am J Hum Genet* 64:290–292
- Albani D, Roiter I, Artuso V, Batelli S, Prato F, Pesaresi M, Galimberti D, Scarpini E, Bruni A, Franceschi M, Piras MR, Confaloni A, Forloni G (2007) Presenilin-1 mutation E318G and familial Alzheimer's disease in the Italian population. *Neurobiol Aging* 28:1682–1688
- Jin S, Pastor P, Cooper B, Cervantes S, Benitez BA, Razquin C, Goate A, Ibero-American Alzheimer Disease Genetics Group Researchers, Cruchaga C (2012) Pooled-DNA sequencing identifies novel causative variants in PSEN1, GRN and MAPT in a clinical early-onset and familial Alzheimer's disease Ibero-American cohort. *Alzheimers Res Ther* 4:34
- To MD, Gokgoz N, Doyle TG, Donovan DB, Knight JA, Hyslop PS, Bernstein A, Andrulis IL (2006) Functional characterization of novel presenilin-2 variants identified in human breast cancers. *Oncogene* 25:3557–3564
- Nicolas G, Wallon D, Charbonnier C, Quenez O, Rousseau S, Richard A-C, Rovelet-Lecrux A, Coutant S, Le Guennec K, Bacq D, Garnier J-G, Olaso R, Boland A, Meyer V, Deleuze J-F, Munter HM, Bourque G, Auld D, Montpetit A, Lathrop M, Guyant-Maréchal L, Martinaud O, Pariente J, Rollin-Sillaire A, Pasquier F, Le Ber I, Sarazin M, Croisile B, Boutoleau-Bretonnière C, Thomas-Antérion C, Paquet C, Sauvée M, Moreaud O, Gabelle A, Sellal F, Ceccaldi M, Chamard L, Blanc F, Frebourg T, Campion D, Hannequin D (2016) Screening of dementia genes by whole-exome sequencing in early-onset Alzheimer disease: input and lessons. *Eur J Hum Genet* 24:710–716
- Schulte EC, Fukumori A, Mollenhauer B, Hor H, Arzberger T, Perneczky R, Kurz A, Diehl-Schmid J, Hüll M, Lichtner P, Eckstein G, Zimprich A, Haubenberger D, Pirker W, Brücke T, Bereznai B, Molnar MJ, Lorenzo-Betancor O, Pastor P, Peters A, Gieger C, Estivill X, Meitinger T, Kretzschmar HA, Trenkwalder C, Haass C, Winkelmann J (2015) Rare variants in β -amyloid precursor protein (APP) and Parkinson's disease. *Eur J Hum Genet* 23:1328–1333

21. Wallon D, Rousseau S, Rovelet-Lecrux A, Quillard-Muraine M, Guyant-Maréchal L, Martinaud O, Pariente J, Puel M, Rollin-Sillaire A, Pasquier F, Le Ber I, Sarazin M, Croisile B, Boutoleau-Bretonnière C, Thomas-Antérion C, Paquet C, Moreaud O, Gabelle A, Sella F, Sauvée M, Laquerrière A, Duyckaerts C, Delisle M-B, Streichenberger N, Lannes B, Frebourg T, Hannequin D, Campion D (2012) The French series of autosomal dominant early onset Alzheimer's disease cases: mutation Spectrum and cerebrospinal fluid biomarkers. *J Alzheimers Dis* 30:847–856
22. Cruchaga C, Chakraverty S, Mayo K, FLM V, Mitra RD, Faber K, Williamson J, Bird T, Diaz-Arrastia R, Foroud TM, Boeve BF, Graff-Radford NR, St. Jean P, Lawson M, Ehm MG, Mayeux R, Goate AM, for the NIA-LOAD/NCRAD Family Study Consortium (2012) Rare variants in APP, PSEN1 and PSEN2 increase risk for AD in late-onset Alzheimer's disease families. *PLoS One* 7:e31039
23. Selkoe DJ, Hardy J (2016) The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol Med* 8:595–608
24. Bellenguez C, Charbonnier C, Grenier-Boley B, Quenez O, Le Guennec K, Nicolas G, Chauhan G, Wallon D, Rousseau S, Richard AC, Boland A, Bourque G, Munter HM, Olaso R, Meyer V, Rollin-Sillaire A, Pasquier F, Letenneur L, Redon R, Dartigues J-F, Tzourio C, Frebourg T, Lathrop M, Deleuze J-F, Hannequin D, Genin E, Amouyel P, Debette S, Lambert J-C, Campion D, Hannequin D, Campion D, Wallon D, Martinaud O, Zarea A, Nicolas G, Rollin-Sillaire A, Bombois S, Mackowiak M-A, Deramecourt V, Pasquier F, Michon A, Le Ber I, Dubois B, Godefroy O, Etchary-Bouyx F, Chauviré V, Chamard L, Berger E, Magnin E, Dartigues J-F, Auriacombe S, Tison F, de la Sayette V, Castan D, Dionet E, Sella F, Rouaud O, Thauvin C, Moreaud O, Sauvée M, Formaglio M, Mollion H, Roullet-Solignac I, Vighetto A, Croisile B, Didic M, Félician O, Koric L, Ceccaldi M, Gabelle A, Marelli C, Labauge P, Jonveaux T, Vercelletto M, Boutoleau-Bretonnière C, Castelnuovo G, Paquet C, Dumurgier J, Hugon J, De Boissgueheneuc F, Belliard S, Bakchine S, Sarazin M, Barrellon M-O, Laurent B, Blanc F, Pariente J, Jurici S (2017) Contribution to Alzheimer's disease risk of rare variants in TREM2, SORL1, and ABCA7 in 1779 cases and 1273 controls. *Neurobiol Aging* 59:220.e1–220.e9
25. Kim WS, Li H, Ruberu K, Chan S, Elliott DA, Low JK, Cheng D, Karl T, Garner B (2013) Deletion of Abca7 increases cerebral amyloid- β accumulation in the J20 mouse model of Alzheimer's disease. *J Neurosci* 33:4387–4394
26. Apostolova LG, Risacher SL, Duran T, Stage EC, Goukasian N, West JD, Do TM, Grotts J, Wilhalme H, Nho K, Phillips M, Elshoff D, Saykin AJ (2018) Associations of the top 20 Alzheimer disease risk variants with brain amyloidosis. *JAMA Neurol* 75:328–341
27. Willnow TE, Andersen OM (2013) Sorting receptor SORLA - a trafficking path to avoid Alzheimer disease. *J Cell Sci* 126:2751–2760
28. Andersen OM, Rudolph I-M, Willnow TE (2016) Risk factor SORL1: from genetic association to functional validation in Alzheimer's disease. *Acta Neuropathol* 132:653–665
29. Svetoni F, Frisone P, Paronetto MP (2016) Role of FET proteins in neurodegenerative disorders. *RNA Biol* 13:1089–1102
30. Münch C, Rosenbohm A, Sperfeld A-D, Uttner I, Reske S, Krause BJ, Sedlmeier R, Meyer T, Hanemann CO, Stumm G, Ludolph AC (2005) Heterozygous R1101K mutation of the DCTN1 gene in a family with ALS and FTD. *Ann Neurol* 58:777–780
31. Wider C, Dachselt JC, Farrer MJ, Dickson DW, Tsuboi Y, Wszolek ZK (2010) Elucidating the genetics and pathology of Perry syndrome. *J Neurol Sci* 289:149–154
32. Stockmann M, Meyer-Ohendorf M, Achberger K, Putz S, Demestre M, Yin H, Hendrich C, Linta L, Heinrich J, Brunner C, Proepper C, Kuh GF, Baumann B, Langer T, Schwalenstöcker B, Braunstein KE, von Arnim C, Schneuwly S, Meyer T, Wong PC, Boeckers TM, Ludolph AC, Liebau S (2013) The dynactin p150 subunit: cell biology studies of sequence changes found in ALS/MND and Parkinsonian syndromes. *J Neural Transm* 120:785–798
33. Holstege H, van der Lee SJ, Hulsman M, Wong TH, van Rooij JG, Weiss M, Louwersheimer E, Wolters FJ, Amin N, Uitterlinden AG, Hofman A, Ikram MA, van Swieten JC, Meijers-Heijboer H, van der Flier WM, Reinders MJ, van Duijn CM, Scheltens P (2017) Characterization of pathogenic SORL1 genetic variants for association with Alzheimer's disease: a clinical interpretation strategy. *Eur J Hum Genet* 25:973–981
34. Verheijen J, Van den Bossche T, van der Zee J, Engelborghs S, Sanchez-Valle R, Lladó A, Graff C, Thonberg H, Pastor P, Ortega-Cubero S, Pastor MA, Benussi L, Ghidoni R, Binetti G, Clarimon J, Lleó A, Fortea J, de Mendonça A, Martins M, Grau-Rivera O, Gelpi E, Bettens K, Mateiu L, Dillen L, Cras P, De Deyn PP, Van Broeckhoven C, Sleegers K (2016) A comprehensive study of the genetic impact of rare variants in SORL1 in European early-onset Alzheimer's disease. *Acta Neuropathol* 132:213–224
35. Rademakers R, Baker M, Nicholson AM, Rutherford NJ, Finch NC, Soto-Ortolaza A, Lash J, Wider C, Wojtas A, DeJesus-Hernandez M, Adamson J, Kouri N, Sundal C, Shuster EA, Aasly J, MacKenzie J, Roeber S, Kretzschmar HA, Boeve BF, Knopman DS, Petersen RC, Cairns NJ, Ghetti B, Spina S, Garbern J, Tselis AC, Uitti R, Das P, Gerpen V, Jan A, Meschia JF, Levy S, Broderick DF, Graff-Radford N, Ross OA, Miller BB, Swerdlow RH, Dickson DW, Wszolek ZK (2012) Mutations in the colony stimulating factor 1 receptor (CSF1R) gene cause hereditary diffuse leukoencephalopathy with spheroids. *Nat Genet* 44:8
36. Sassi C, Nalls MA, Ridge PG, Gibbs JR, Lupton MK, Troakes C, Lunnon K, Al-Sarraj S, Brown KS, Medway C, Lord J, Turton J, Bras J, Blumenau S, Thielke M, Josties C, Freyer D, Dietrich A, Hammer M, Baier M, Dirnagl U, Morgan K, Powell JF, Kauwe JS, Cruchaga C, Goate AM, Singleton AB, Guerreiro R, Hodges A, Hardy J, Passmore P, Craig D, Johnston J, McGuinness B, Todd S, Heun R, Kölsch H, Kehoe PG, Vardy ERLC, Hooper NM, Mann DM, Pickering-Brown S, Brown K, Lowe J, Morgan K, Smith AD, Wilcock G, Warden D, Holmes C (2018) Mendelian adult-onset leukodystrophy genes in Alzheimer's disease: critical influence of CSF1R and NOTCH3. *Neurobiol Aging* 66:179.e17–179.e29
37. Guerreiro RJ, Lohmann E, Kinsella E, Brás JM, Luu N, Gurunlian N, Dursun B, Bilgic B, Santana I, Hanagasi H, Gurvit H, Gibbs JR, Oliveira C, Emre M, Singleton A (2012) Exome sequencing reveals an unexpected genetic cause of disease: NOTCH3 mutation in a Turkish family with Alzheimer's disease. *Neurobiol Aging* 33:1008.e17–1008.e23
38. Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogava E, Majounie E, Cruchaga C, Sassi C, Kauwe JSK, Younkin S, Hazrati L, Collinge J, Pocock J, Lashley T, Williams J, Lambert J-C, Amouyel P, Goate A, Rademakers R, Morgan K, Powell J, St. George-Hyslop P, Singleton A, Hardy J (2013) TREM2 variants in Alzheimer's disease. *N Engl J Med* 368:117–127
39. Belorgey D, Crowther DC, Mahadeva R, Lomas DA (2002) Mutant Neuroserpin (S49P) that causes familial encephalopathy with Neuroserpin inclusion bodies is a poor proteinase inhibitor and readily forms polymers in vitro. *J Biol Chem* 277:17367–17373
40. Roussel BD, Lomas DA, Crowther DC (2016) Progressive myoclonus epilepsy associated with neuroserpin inclusion bodies (neuroserpinosis). *Epileptic Disord* 18:103–110

We assure that the data contained in the manuscript being submitted have not been previously published, have not been submitted elsewhere and will not be submitted elsewhere while under consideration at Neurological Sciences.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.