# Resurrecting the Lost Flames of American Chestnut

## Jeffrey M. Kane, $^{1\star}$  J. Morgan Varner, $^{2}$  and Michael R. Saunders $^{3}$

<sup>1</sup>Department of Forestry and Wildland Resources, Wildland Fire Laboratory, Humboldt State University, 1 Harpst Street, Arcata, California 95521, USA; <sup>2</sup>Pacific Wildland Fire Sciences Laboratory, United States Department of Agriculture, Forest Service, 400 N 34th Street, Suite 201, Seattle, Washington 98103, USA; <sup>3</sup>Department of Forestry and Natural Resources, Purdue University, 715 W State Street, West Lafayette, Indiana 47907, USA

### ABSTRACT

Loss of native foundation tree species to introduced pests profoundly alters the structure and function of many forest ecosystems. Recent advances to resurrect or prevent the loss of species by developing resistant hybrids hold promise, but uncertainty remains about the potential impacts of introducing a novel genotype on ecological processes, such as fire. A classic example of a nonnative, pathogen-caused loss of a foundation species is American chestnut (Castanea dentata), a species now functionally extinct from the eastern US but undergoing experimental trials to resurrect the species with a putatively disease-resistant genotype. We compared the litter flammability among American chestnut, Chinese chestnut (C. mollisima), and a population of first intercross, third-generation backcrossed hybrid  $(BC_3F_2)$  using laboratory burning experiments. Litter flammability of American chestnut was consistently greater than Chinese chestnut, whereas the hybrid had an intermediate flammability or more closely resembled Chinese chestnut flammability by some mea-

sures. Greater flammability in American chestnut was associated with a longer leaf structure with greater curling when dry. American chestnut had flammability properties consistent with other pyrophytic species present in contemporary fireprone ecosystems. The loss of American chestnut may have altered litter flammability of some eastern US forest ecosystems, a result more commonly associated with compositional changes in conjunction with fire exclusion and other disturbances. Resurrecting American chestnut with a hybrid genotype may mitigate this change in areas where less flammable species have replaced American chestnut. Resurrection of lost foundation species through introduction of resistant genotypes may represent a resounding ecological success story, but unanticipated changes to ecological processes, such as fire, should be considered.

CrossMark

Key words: fire ecology; disease resistance; global change; litter flammability; mesophication; restoration; resurrection ecology.

#### Received 24 February 2018; accepted 14 October 2018; published online 18 December 2018

Data will be hosted at the Humboldt State University data sharing repository.

\*Corresponding author; e-mail: jkane@humboldt.edu

#### **HIGHLIGHTS**

- American chestnut litter is remarkably flammable based on laboratory experiments.
- The backcross hybrid (BC<sub>3</sub>F<sub>2</sub>) generally had similar or slightly lower litter flammability than American chestnut.
- Loss of American chestnut may have contributed to reduced fire frequency.

Author Contributions JMK, JMV, and MRS all contributed to the study concept and design and writing of the manuscript, JMK and MRS performed the research, and JMK analyzed the data.

#### **INTRODUCTION**

Numerous forest ecosystems have experienced substantial reduction or loss related to non-native insect and pathogen outbreaks (Ellison and others [2005;](#page-10-0) Aukema and others [2010](#page-9-0)). Many of these events have long been a strong driver of change in forests locally and globally (Elton [1958](#page-10-0)), dramatically altering forest structure and composition that can directly and indirectly disrupt key ecosystem functions, such as nutrient cycling, primary productivity, and natural disturbance processes (Liebhold and others [1995;](#page-10-0) Lovett and others [2006](#page-10-0); Metz and others [2013\)](#page-10-0). These impacts are especially concerning when forest loss impacts foundation species whose traits define and structure ecosystem dynamics and promote locally stable conditions (Dayton [1972;](#page-9-0) Ellison and others [2005\)](#page-10-0). Given the rapid increases in global trade and travel (Levine and D'Antonio [2003](#page-10-0); Aukema and others [2010\)](#page-9-0) and increasing global temperatures related to climate change (Sturrock and others [2011;](#page-11-0) Weed and others [2013\)](#page-11-0), forest loss from nonnative, invasive pests and pathogens are projected to continue over the coming decades and may result in the extirpation or extinction of some tree species.

These events have prompted forest scientists and mangers to seek creative solutions to recover lost or declining species through promoting resistance. Recent successes in developing resistant genotypes, through phenotypic selection, hybridization with related species, or genetic engineering, hold promise to resurrect functionally extirpated or extinct species to their native ecosystems (Sniezko [2006](#page-11-0)). Introducing resistant genotypes for a species that has long been absent can serve as a successful example of resurrection ecology (also referred to ''de-extinction'') and provide potential insight into other species that are or may go extinct in the near future. Yet, successful resurrection of a species is contingent on the replacement genotype serving as an adequate ecological proxy of the lost species (Shapiro [2017](#page-11-0)), requiring examination of the broader ecological impacts of reintroduction (Sherkow and Greely [2013](#page-11-0)).

Resurrecting species through the introduction of novel genotypes may alter ecological processes in unintended or undesired ways (Whitham and others [2003](#page-11-0), [2006](#page-11-0)). Numerous studies on terrestrial ecosystems have found that intraspecific differences in plant genotypes can have cascading impacts on higher-order ecological processes, such as community assembly, nutrient dynamics, and disturbance regimes (Bailey and others [2004;](#page-9-0) Schweitzer and others [2004\)](#page-11-0). Given the preponderance of fire-prone forests globally, there is a need to consider the influence of novel genotypes on fire behavior and effects in these ecosystems.

Feedbacks between vegetation traits and fire regimes regulate ecological processes in many fireprone ecosystems (Beckage and others [2009](#page-9-0); Veldman and others [2013](#page-11-0)). In most forested ecosystems, the predominant driver of surface fire behavior is senesced leaf litter and fine woody fuels (Rothermel [1983\)](#page-11-0). Many studies have determined that tree litter from frequent-fire forest types often has greater flammability (Fonda [2001;](#page-10-0) Kane and others [2008](#page-10-0); Engber and Varner [2012\)](#page-10-0), which has been closely linked to individual leaf traits and litter fuelbed properties. Typically, tree species with larger or longer leaves (among other traits) will have fuelbed densities conducive to greater oxygen permeation that promote greater flame lengths, faster flaming times, and greater fuel consumption (Scarff and Westoby [2006;](#page-11-0) Engber and Varner [2012;](#page-10-0) Kreye and others [2013;](#page-10-0) Varner and others [2015b](#page-11-0)). Additionally, leaf traits and associated litter fuelbed traits can influence the moisture content and drying rates that can affect litter flammability (Kreye and others [2013,](#page-10-0) [2018](#page-10-0)). Changes in the leaf traits of litter fuelbeds may have implications on the flammability of fire-prone ecosystems (Mola and others [2014](#page-10-0); Varner and others [2017](#page-11-0); Kreye and others [2018\)](#page-10-0), and shifts in composition have been linked with altered fire regimes (Brooks and others [2004\)](#page-9-0).

One of the earliest examples of extensive tree mortality in the USA was chestnut blight, a disease caused by the fungus Cryphonectria parasitica. Since the pathogen was first detected in the early twentieth century (Murrill [1906](#page-10-0)), American chestnut (Castanea dentata) has been eliminated from the overstory throughout its historical range and almost exclusively persists as small understory resprouts (Anagnostakis [1987](#page-9-0)), rendering the species as functionally extinct (Saterberg and others [2013](#page-11-0)). Prior to its loss, American chestnut comprised as much as 25% of the trees and 36% of the basal area across large areas of the eastern hardwood forest (Foster and others [2002](#page-10-0); Elliott and Swank [2008](#page-9-0); Wang and others [2013](#page-11-0)). This species also regulated numerous ecological processes (for example, productivity, decomposition, and nutrient cycling; Paillet [2002\)](#page-11-0) and provided important ecosystem services (for example, lumber, fence post, and food resources; Youngs [2000](#page-11-0)), contributing to its characterization as a foundation species (Ellison and others [2005;](#page-10-0) Jacobs and others [2013](#page-10-0)).

Forest managers are considering large-scale reforestation and restoration plantings of putatively disease-resistant American chestnut seedlings produced through backcross hybridization with blightresistant Chinese chestnut (C. mollissima) (Steiner and Carlson [2006;](#page-11-0) Jacobs [2007\)](#page-10-0). Although ongoing work holds much promise, the effectiveness and feasibility of a large-scale effort still presents challenges (Clark and others [2014](#page-9-0), [2016;](#page-9-0) Steiner and others [2017\)](#page-11-0). Recent research has advanced our understanding of the performance of disease-resistant chestnut genotypes (Clark and others [2012](#page-9-0), [2016;](#page-9-0) Brown and others [2014](#page-9-0); Knapp and others [2014;](#page-10-0) Pinchot and others [2015](#page-11-0)), although the potential impacts of these novel genotypes on ecological processes and other factors have been largely unexamined (Jacobs [2007](#page-10-0); Jacobs and others [2013\)](#page-10-0).

Although the fire ecology of American chestnut is not well resolved (Foster and others [2002](#page-10-0)), multiple lines of evidence suggest that many eastern hardwood forests that contained American chestnut had historically experienced litter-driven surface fires, with relatively low incidences of stand-replacing crown fires. Pre-blight fire return intervals conservatively ranged from 3 to 13 years (Guyette and others [2012](#page-10-0); Brose and others [2014](#page-9-0); Stambaugh and others [2015](#page-11-0); Varner and others [2016;](#page-11-0) Lafon and others [2017\)](#page-10-0). As a result, managers continue to promote the use of prescribed fire in many of the forests where American chestnut could be reintroduced (Brose and others [2014](#page-9-0)). However, any strong differences in litter flammability between American chestnut hybrids and the functionally extinct species could promote undesired conditions that alter the flammability of the ecosystem with potential negative ecological or economic impacts.

The aim of this study was to examine differences in chestnut litter flammability of a backcross hybrid genotype  $(BC_3F_2)$  in comparison with a pure American and a pure Chinese chestnut through laboratory drying and burning trials. The specific objectives were: (1) to determine whether leaf and flammability characteristics differed among the three chestnut types; (2) to determine the leaf traits most associated with litter flammability characteristics among the three chestnut types; and (3) to examine the litter flammability of American chestnut in context with other species from fireprone forests. The results of this study will provide critical information into the past and future fire ecology of American chestnut and eastern North American hardwood forests, and the potential ecological consequences of resurrecting a functionally extinct species that holds insights for other resurrected species in fire-prone ecosystems.

#### **METHODS**

We collected litter samples from eight replicate trees per chestnut type: American chestnut (C. dentata); Chinese chestnut (C. mollissima); and third-generation backcrossed hybrid  $(BC_3F_2)$ . Trees were part of the Purdue University Hardwood Tree Improvement and Regeneration Center chestnut breeding program in partnership with the American Chestnut Foundation. The  $BC_3F_2$  trees originated from open-pollinated  $BC_3F_1$  trees from multiple families within a single plantation. Litter was collected in September 2015 from several planting sites within a 10 km radius of West Lafayette, Indiana, USA. Plantations were composed of either widely spaced (3 m) open-canopy trees or more narrowly spaced (1 m), closed-canopy trees. All sampled trees were 9 years old at the time of litter collection. The backcross hybrid and most pure American chestnut collections were from small, healthy trees ranging from 10 to 15 cm in diameter at breast height (1.3 m; dbh), whereas one American chestnut and all Chinese chestnut were 15–35 cm in dbh.

All approximately  $-100 \text{ g}$  samples were collected from recently fallen or loosely attached, sun and shade leaves on each individual tree and placed in paper bags. Samples were oven-dried at  $40^{\circ}$ C for 24 h. Leaf trait measurements were taken from 15 leaves randomly selected from each replicate. Leaf edge thickness was measured using electronic calipers, and curl depth was measured as the height of a dry leaf above a flat surface to the nearest millimeter (Engber and Varner [2012](#page-10-0)). These same sample leaves were then wetted and flattened between two heavy flat surfaces and allowed to dry. Afterward, leaf length, maximum width, area, and perimeter were measured using WinFOLIA image processing software (Regent Instruments, Quebec, Canada). Leaf volume was calculated by multiplying leaf thickness by the surface area of one side. Surface area-to-volume ratio was calculated by dividing leaf thickness by two (Mauseth [2000](#page-10-0)), and specific leaf area was calculated by dividing leaf area by dry mass.

Litter burning trials were conducted in the Humboldt State University Wildland Fire Laboratory using standard methods (Fonda [2001\)](#page-10-0). Relative humidity ranged from 58.0% to 70.3% and air temperatures ranged from  $20^{\circ}$ C to  $26^{\circ}$ C at the time of the burning. Burning trials consisted of approximately 15 g of litter from each replicate per chestnut type (24 total trials). Each litter sample was evenly distributed as a fuelbed over a 35 cm  $\times$  35 cm lattice of xylene-soaked cotton strings on a stainless-steel platform. A  $2.75 \text{ m} \times 2.75 \text{ m}$  fume hood above the platform generated a constant draw of  $15-20$  cm s<sup>-1</sup>, though we detected no airflow at the fuelbed. Prior to ignition, fuelbed depth was measured to the nearest mm from four locations (7 cm from each corner of the fuelbed). Strings were ignited from all sides and a timer was started at litter ignition. Maximum flame height was visually estimated to the nearest cm using a vertical ruler placed just behind the burning platform. Flaming time was measured as the duration from litter ignition to flaming extinction, and smoldering time was measured as the duration from flaming extinction to smoldering extinction. Both flaming time and smoldering time were recorded to the nearest second. Percent litter consumption was calculated as the proportion of the initial litter mass consumed  $(Eq. 1).$ 

$$
Consumption(\%) = \frac{(litter mass - residual mass)}{litter mass}
$$
  
 
$$
\times 100
$$
 (1)

Fuelbed drying was measured over a 72-h period under laboratory conditions. After drying at  $40^{\circ}$ C for 24 h, 15 g from five of the replicates per chestnut type was used to create litter beds for the desorption trials. Litter beds were soaked in a water bath for 24 h, then removed, and consistently agitated to remove excess surface water on the leaves before being placed in  $24 \times 24 \times 4$  cm preweighed aluminum pans with 36 holes (5 mm diameter) spaced 25 mm apart. Each pan was elevated on wooden slats to allow drainage of any residual water. At this time, we recorded the wet weight of the litter to calculate the initial moisture content (Kreye and others [2013](#page-10-0)). Litter beds were allowed to air-dry under laboratory conditions that were controlled by a dehumidifier (Frigidaire FFAD7033R1, Charlotte, North Carolina, USA). Relative humidity and air temperature throughout the drying experiments were somewhat constant, ranging from 43% to 50% and 23 $^{\circ}$ C to 24 $^{\circ}$ C, respectively. Litter beds were weighed at half hour intervals for the first 4 h, every hour for the next 20 h, and intermittently until the moisture content stabilized over multiple intervals. Percent moisture content was calculated for each time step (Eq. 2):

$$
m_t = \frac{\text{(litter mass}_t - \text{litter mass}_{od})}{\text{litter mass}_{od}} \times 100, \qquad (2)
$$

where  $m_t$  is moisture content at time t, litter mass<sub>t</sub> is fuel mass at time  $t$ , and litter mass<sub>od</sub> is oven-dried fuel mass. Based on percent moisture content, we calculated the relative moisture content of a litter bed for each time step (Eq. 3; Fosberg and others [1970\)](#page-10-0):

$$
E = \frac{(m_t - m_f)}{(m_i - m_f)},
$$
\n(3)

where E is relative moisture content,  $m_i$  is initial moisture content, and  $m_f$  is final moisture content.

Response time  $(\tau)$  of each litter bed was determined using piecewise regression with the seg-mented package (Muggeo [2008\)](#page-10-0). Response time represents the time a fuel particle or fuelbed takes to reach 63% of the equilibrium moisture content. Shorter response times indicate fuels that dry and gain moisture faster than fuels with longer response times. The natural log of relative moisture content  $(E)$  was separated into two linear sections: initial and final time lag (Nelson and Hiers [2008](#page-10-0)) based on the calculated break point determined by the piecewise regression. Response time for each litter bed was calculated algebraically using the regression equation of the relationship between time and  $log E$  for the initial time lag, where  $E = 0.368$  (Eq. 4).

$$
\tau = \frac{\log(E)}{\text{slope}} - \text{intercept} \tag{4}
$$

#### Data Analysis

Leaf traits, drying characteristics (initial moisture content, response time), and flammability measurements were compared among the three chestnut types using a one-way analysis of variance (ANOVA) followed by Bonferroni (all pair-wise) multiple comparison tests when significant differences were detected. Chestnut types were compared using the nonparametric Kruskal–Wallis one-way ANOVA on ranks followed by the Dunn test to determine differences among chestnut types because data did not meet parametric test assumptions. Univariate correlations among all leaf traits and flammability measures were completed using simple linear regression and assessed for violations of test assumptions. We also conducted a multivariate analysis of all flammability measurements among chestnut types using principal com<span id="page-4-0"></span>ponents analysis with the prcomp function in the vegan package (Oksanen and others [2017](#page-11-0)). A permutation multivariate analysis of variance test was conducted to detect a chestnut type effect followed by a Tukey HSD test with a Bonferroni correction factor to detect pair-wise differences among types. Although flammability measures have been correlated in other studies (Engber and Varner [2012](#page-10-0)), only flaming time and consumption were significantly correlated in our dataset  $(r^2 = 0.59)$ ,  $p < 0.0001$ ). Subsequently, we compared the relationship between leaf traits and flammability scores using the *envfit* function in the *vegan* package (Oksanen and others [2017\)](#page-11-0) and report all significant relationships ( $\alpha = 0.1$ ) based on 999 permutations. Analyses were all completed using R statistical software (R Development Core Team [2017\)](#page-11-0).

#### **RESULTS**

Although the backcross hybrid is expected to be 94% genetically similar to American chestnut, there were significant differences in litter flammability during the laboratory experiment (Table 1). American chestnut had 18% taller flames  $(p = 0.005;$  Figure 1) and 32% shorter flaming times ( $p = 0.006$ ) than Chinese chestnut. The hybrid litter flammability was either similar to Chinese chestnut (for example, maximum flame height) or had intermediate values between American and Chinese chestnut (for example, flaming time). Chinese chestnut and the backcross hybrid had approximately 10% lower initial moisture content than American chestnut but did not differ significantly ( $p = 0.204$ ). Response time also did not differ significantly among chestnut types



Figure 1. Laboratory burn demonstrating the flame heights of American chestnut (Castanea dentata). The average maximum flame height for all observations was 98.9 cm (range =  $91-106$  cm).

 $(p = 0.206)$ , but Chinese chestnut had more than a 50% lower value than either American chestnut or the hybrid.

Differences in leaf traits among chestnut types were also observed (Table [2](#page-5-0)), some of which related to variation in flammability. Chinese chestnut





Comparisons are based on a Kruskal–Wallis nonparametric one-way ANOVA (df = 2, 21), using the Dunn test to determine differences among chestnut types. Significant differences in characteristics are depicted in bold, with differences among types denoted with superscripted letters.

Characteristics	American chestnut (C. dentata)	<b>Backcross</b> hybrid	Chinese chestnut (C. mollissima)	$\boldsymbol{F}$	p
Dry mass $(g)$	0.52(0.03)	0.48(0.04)	0.52(0.04)	0.97	0.617
Max leaf length (cm)	18.8 $(0.6)^a$	17.2 $(0.4)^{ab}$	16.6 $(0.6)^b$	4.71	0.020
Max leaf width (cm)	6.3 $(0.3)^a$	5.5 $(0.2)^{b}$	6.7 $(0.2)^a$	7.62	0.003
Leaf thickness (mm)	0.152(0.005)	0.159(0.003)	0.150(0.007)	1.51	0.471
Leaf curl (mm)	32.8(2.7)	27.6(1.3)	27.6(2.0)	1.62	0.221
Leaf perimeter (cm)	60.3(3.9)	52.7 (1.6)	50.8(1.4)	3.39	0.053
Surface area-to-volume ratio $\rm (cm^2/cm^3)$	134.1(4.5)	127.8(2.6)	138.4 (5.8)	1.07	0.360
Perimeter-to-surface area ratio $\text{(cm/cm}^2\text{)}$	$0.85 (0.03)^{ab}$	$0.95(0.03)^{a}$	$0.75(0.04)^{b}$	10.36	0.006
Specific leaf area $\rm (cm^2/g)$	161.3 $(8.1)^a$	126.4 $(4.1)^{b}$	148.4 $(4.7)^a$	11.32	0.003
Fuelbed depth (cm)	5.3 $(0.6)$	5.1(0.3)	4.8(0.5)	0.30	0.754

<span id="page-5-0"></span>Table 2. Mean (Standard Error) Leaf and Fuelbed Characteristics by Chestnut Type

Comparisons are based on a one-way ANOVA (df = 2, 21) with a Tukey HSD multiple comparison test. Significant differences in characteristics are depicted in bold, with differences among types denoted with superscripted letters.

had a 13.2% shorter maximum leaf length than American chestnut. The backcross hybrid had an intermediate length but was narrower than either American or Chinese chestnut (Figure 2). Perimeter-to-surface area ratio was the highest for the hybrid and was significantly greater than Chinese chestnut, which had the lowest values. Maximum leaf length was the leaf trait most associated with flammability measures and had a positive relationship with flame height ( $r^2 = 0.32$ ,  $p = 0.004$ ),



Figure 2. Representative leaves that highlight measured differences in litter characteristics among American chestnut (left), backcross hybrid (center), and Chinese chestnut (right).

where American chestnut had longer leaves and taller flame heights compared to Chinese chestnut, with the backcross hybrid having intermediate values (Figure [3](#page-6-0)). Fuelbeds of American chestnut were deeper than the hybrid or Chinese chestnut but did not differ among species  $(p = 0.754)$ . Flaming time was negatively associated with leaf curling  $(r^2 = 0.22, p = 0.02)$  and consumption was negatively associated with leaf thickness  $(r^2 = 0.18)$ ,  $p = 0.04$ ) and fuelbed depth ( $r^2 = 0.17$ ,  $p = 0.049$ ), but none of these characteristics differed significantly among chestnut types. Differences in flammability were detected among chestnut types when examined in a multivariate analysis  $(F = 3.9)$ ,  $p = 0.002$ ). Consistent with the univariate analyses, American chestnut had taller flames, shorter flaming times, longer smoldering times, and greater consumption compared to Chinese chestnut and the backcross hybrid (Figure [4\)](#page-7-0). Differences in the multivariate flammability scores among chestnut types corresponded with maximum leaf length  $(r^2 = 0.37, p = 0.007)$ , leaf curl  $(r^2 = 0.29)$ ,  $p = 0.025$ ), and leaf perimeter ( $r^2 = 0.31$ ,  $p = 0.03$ ).

#### **DISCUSSION**

To our knowledge, this is the first study to examine the potential changes in flammability associated with the loss of a foundation species and the implications of resurrecting a genetically similar, disease-resistant genotype. Resurrection of American chestnut through the introduction of a diseaseresistant hybrid would likely serve as an acceptable ecological proxy with respect to litter flammability because the American chestnut and

<span id="page-6-0"></span>

Figure 3. Positive relationship ( $r^2 = 0.32$ ,  $p = 0.004$ ) between maximum leaf length and maximum flame height by chestnut type.

the backcross hybrid burned similarly. Where differences were detected (for example, maximum flame height), these did not seem to be strictly related to differences in leaf traits among the chestnut types. However, consideration of flammability across multiple measures indicated that the backcross hybrid might confer a slight dampening of flammability relative to American chestnut. These findings suggest that selection of disease-resistant genotypes may also consider, among others (for example, Diskin and others [2006](#page-9-0)), traits that can promote litter flammability (for example, maximum leaf length, curl) most similar to the pure American chestnut. The specific impacts to flammability will also depend on the particular families (that is, genotypes), and the level of intercross used to develop the resistant hybrids, which we did not examine, warrants further research. For instance, our results were based on a first intercross of third-generation backcross hybrids  $(BC_3F_2)$  instead of the recommended use of a third generation to confer pathogen resistance (Steiner and others [2017\)](#page-11-0). In addition, the culling of less resistant  $BC_3F_2$  parent plants is still ongoing

and the process of selecting the final resistant genotypes in the resurrection of American chestnut may result in differences in litter flammability, suggesting the need for follow-up research.

Previous research that has examined compositional changes in American chestnut forest types before and after blight-induced mortality has mostly shown substantial increases in mesophytic tree species such as maples (Acer spp.) (for example, Keever [1953](#page-10-0); Agrawal and Stephenson [1995](#page-9-0); Myers and others [2004;](#page-10-0) Hawkins [2006](#page-10-0); Elliott and Swank [2008\)](#page-9-0), although in some locations pyrophytic oak (for example, Quercus alba) or hickory species (for example, Carya glabra) replaced American chestnut (Korstian and Stickel [1927](#page-10-0); McCormick and Platt [1980](#page-10-0)). The degree to which the replacement species would dampen or facilitate regional fire regimes needs more examination, but a recent study by Varner and others [\(2017](#page-11-0)) found that changes in flammability were contingent on the litter characteristics of the replacing species, where replacement by mesophytes had a stronger dampening effect on flammability. Based on our results of substantive litter flammability during

<span id="page-7-0"></span>

Figure 4. Separation of chestnut types along two flammability axes. Each type differed in flammability based on a PERMANOVA test ( $F = 3.9$ ,  $p = 0.002$ ). The first PCA axis (Flam1) explained 49.0% of the variation in flammability and was negatively associated with flaming time (FlmT;  $r = -0.67$ ) and positively associated with consumption (Cnsm;  $r = 0.6$ ). The second PCA axis (Flam2) explained 24.1% of the variation in flammability and was positively associated with smoldering time (SmlT;  $r = 0.94$ ). A third flammability axis (not shown) explained 21.8% and was positively associated with maximum flaming height (FlmH;  $r = 0.7$ ). The ellipses represent 68% of the data for each chestnut type. The maroon join plots refer to relative strength of significant relationships among leaf traits and flammability scores.

laboratory experiments, we propose that the loss of American chestnut significantly contributed to reduced ignition and spread of fires in chestnut forests now dominated by mesophytic species. These impacts would likely be exacerbated when the loss of American chestnut was coupled with decreases in other pyrophytic species (for example, oak and hickory species; Nowacki and Abrams [2008\)](#page-11-0).

Resurrection of American chestnut through the introduction of a disease-resistant backcross hybrid has the potential to influence several ecological processes in these forests, including the potential restoration of their historic fire regimes. The slight dampening in flammability in the backcross hybrid compared to American chestnut may cause reduced ignition and fire spread where introduced, yet other traits may confer persistence. Similar to American chestnut, the hybrid is capable of basal resprouting following fire and other disturbances (Wang and others [2013;](#page-11-0) Belair and others In press). Recent evidence has shown that resprouting of

 $BC_3F_2$  and  $BC_3F_2$  hybrids following dieback of planted seedlings was similar to American chestnut, but less vigorous than Chinese chestnut (Thomas-Van Gundy and others [2017](#page-11-0)) though specific responses to fire have not yet been examined. Persistence of the hybrid will also depend on the rate and amount of bark produced. Some have suggested that American chestnut may have relatively thin bark (for example, Ashe [1912](#page-9-0)); however, these studies are largely based on anecdotal observations, focused on smaller individuals (that is, seedlings and saplings), and occurred during a time period that predated the concept of fire ecology, thus indicating a need to quantify bark thickness within and among chestnut types. American chestnut and the backcross hybrid litter dried at moderate rates and burned with substantial flammability (Table [1](#page-4-0)), relatively consistent with many other pyrophytic species (that is, species with rapidly drying and combustible litter) from fireprone ecosystems. For instance, maximum flame height for American chestnut litter was just below 100 cm, values similar to or exceeding pyrophytic pines and oaks from historically fire-prone ecosystems in the southeastern US. These maximum flame height values were consistent with eastern white oak (Quercus alba; 98.7 cm) and were greater than longleaf pine (Pinus palustris; 92.3 cm) and shortleaf pine (Pinus taeda; 78 cm) (Varner and others [2015a](#page-11-0)). American chestnut litter also had a mean time lag of 6.9 h, indicating moderate drying rates that were slightly higher than other pyrophytic species, such as eastern white oak (4.3 h) and shortleaf pine (3.4 h) (Kreye and others [2013](#page-10-0)).

The importance of fire to American chestnut is not well understood, and conflicting reports are abundant in the literature (for example, McEwan and others [2011;](#page-10-0) Lafon and others [2017\)](#page-10-0). The high flammability of American chestnut litter observed in our study contributes further evidence that American chestnut has a suite of traits to promote persistence in a frequent-fire forest. The species is a prolific resprouter, capable of producing numerous, fast-growing basal sprouts and can rapidly respond to new growing space caused by disturbances (Belair and others [2014\)](#page-9-0). The degree of shade tolerance has differed widely among studies (Niinemets and Valladares [2006;](#page-10-0) Wang and others [2006\)](#page-11-0), but most indicate that American chestnut regeneration is moderately shade tolerant and highly plastic in response to disturbance (Rhoades and others [2009](#page-11-0); Knapp and others [2014](#page-10-0); Belair and others [2018](#page-9-0)). Larger trees that would occupy the overstory of fire-frequent forests may have traits that better allow them to persist under these conditions, such as lower shade tolerance, higher drought tolerance, and greater bark thickness.

Our observations of American chestnut flammability provide additional support that oakchestnut forests historically experienced a frequent-fire regime (Guyette and others [2012](#page-10-0); Stambaugh and others [2015](#page-11-0); Varner and others [2016;](#page-11-0) Lafon and others [2017\)](#page-10-0). A comprehensive review of existing fire scar studies throughout much of the former American chestnut range highlighted a high-frequency fire regime with historic fire return intervals prior to 1900 between 4 and 13 years (Lafon and others [2017\)](#page-10-0). Native American ignitions were likely the primary source for maintaining such a historically frequent-fire regime (Hessl and others [2011;](#page-10-0) Aldrich and others [2014\)](#page-9-0); however, models reconstructing historic fire frequency based on climate–fire relationships independent of contributions from human ignitions also corroborate a frequent-fire regime for oak-chestnut forests (Guyette and others [2012](#page-10-0);

Stambaugh and others [2015](#page-11-0)). The mixture of fireresistant and fire-neutral traits, with fire history evidence aside, suggests the possibility that the fire regime of forests containing American chestnut ranged from moderate to high frequency, depending on the climate, topographic conditions, co-occurring tree species, and other factors for a given site.

Given our findings that American chestnut has characteristics consistent with other pyrophytic species, we propose that the loss of this foundation species may have contributed to the known reductions in fire frequency of these forests following fire exclusion in the early to mid-1900s (Guyette and others [2012](#page-10-0); Lafon and others [2017](#page-10-0)). A positive feedback mechanism termed ''mesophication'' posits that the prolonged exclusion of fire shifted forest structure and composition from lower density, shade-intolerant, pyrophytic species to higher density communities of shade-tolerant, mesophytic (fire-sensitive) species in many eastern US forests (Nowacki and Abrams [2008](#page-11-0)). The increased presence of mesophytes promotes a greater proportion of litter that retains more moisture and has lower flammability (Kreye and others [2013](#page-10-0), [2018\)](#page-10-0), further promoting less flammable conditions and reducing the incidence of fire. In forests containing American chestnut, mesophication could have been facilitated by the loss of American chestnut to blight that reduced the proportional dominance of pyrophytes and their more flammable litter (McEwan and others [2011](#page-10-0)), though more investigation is needed.

Understanding the potential changes in litter flammability due to chestnut blight has important implications for other non-native pathogens introduced to fire-prone forests. Two modern examples of non-native pathogens that have resulted in substantial tree mortality include laurel wilt (Raffaelea lauricola) that affects red bay (Persea burbonia) and other members of the Lauraceae in southeastern US. (Fraedrich and others [2008](#page-10-0)), and sudden oak death (Phytophthora ramorum) that affects tanoak (Notholithocarpus densiflorus) and some true oaks (Quercus spp.) in California and Oregon (Rizzo and Garbelotto [2003](#page-11-0)). Although these species have not yet become functionally extinct, they both, among others, have high potential to spread throughout the species' ranges over the coming decades (Meentemeyer and others [2011;](#page-10-0) Kendra and others [2013\)](#page-10-0). Previous research has focused on the potential impacts of pathogen-induced mortality on changes in fuel characteristics (Forrestel and others [2011](#page-10-0); Kuljian and Varner [2013](#page-10-0)) and the observed synergistic interactions between patho<span id="page-9-0"></span>gen-induced and fire-induced tree mortality (Metz and others [2011,](#page-10-0) [2013](#page-10-0)). Our study provides one means to anticipating potential impacts to changes and the hopeful resurrection of these species through introduction of resistant genotypes.

Resurrection of American chestnut would serve as a resounding ecological success story of a species that has been functionally extinct for nearly a century and potentially could provide many lost or degraded ecosystem services in hardwood forests of eastern North America. American chestnut litter is quite flammable and loss of this species conceivably contributed to inhibition of fire and the greater abundance of mesophytic species in some forests. Reintroduction of a disease-resistant hybrid, with subtly dampened fire behavior compared to American chestnut, may still increase flammability in these forests, but broader ecological implications of the functional differences between American chestnut and hybrid genotypes are not well understood. Insights from this study and others will be needed to adequately assess the broader ecological consequences of resurrecting the once-iconic American chestnut, with the potential to inform the resurrection of other foundation species in frequent-fire ecosystems.

#### ACKNOWLEDGEMENTS

Laboratory burning and moisture trials were assisted by L. Rios, A. Cowen, K. Dedrick, T. Bradley, and J. Avitia. Field collections of samples and leaf scans were conducted by S. Greenler. Discussions with J. Kreye and D. Orwig were helpful in advancing our thoughts on the topic. Two anonymous reviewers also provided helpful suggestions and comments to an earlier draft. Financial support for this research was provided by the USDA National Institute of Food and Agriculture, McIntire-Stennis Cooperative Forestry Research Program at Humboldt State University (Project #CALZ-157) and Purdue University (IND011557MS), the National Fire Plan, and Hardwood Tree Improvement and Regeneration Center.

#### REFERENCES

- Agrawal A, Stephenson SL. 1995. Recent successional changes in a former chestnut-dominated forest in Southwestern Virginia. Castanea:107–113.
- Aldrich SR, Lafon CW, Grissino-Mayer HD, DeWeese GG. 2014. Fire history and its relations with land use and climate over three centuries in the central Appalachian Mountains, USA. Williams J, editor. J Biogeogr 41:2093–104.
- Anagnostakis SL. 1987. Chestnut blight: the classical problem of an introduced pathogen. Mycologia 79:23.
- Ashe WW. 1912. Chestnut in Tennessee. Nashville, Tennessee: Baird-Ward Printing Co.
- Aukema JE, McCullough DG, Von Holle B, Liebhold AM, Britton K, Frankel SJ. 2010. Historical accumulation of nonindigenous forest pests in the continental United States. BioScience 60:886–97.
- Bailey JK, Schweitzer JA, Rehill BJ, Lindroth RL, Martinsen GD, Whitham TG. 2004. Beavers as molecular geneticists: a genetic basis to the foraging of an ecosystem engineer. Ecology 85:603–8.
- Beckage B, Platt WJ, Gross LJ. 2009. Vegetation, fire, and feedbacks: A disturbance-mediated model of savannas. Am Nat 174:805–18.
- Belair ED, Saunders MR, Bailey BG. 2014. Four-year response of underplanted American chestnut (Castanea dentata) and three competitors to midstory removal, root trenching, and weeding treatments in an oak-hickory forest. For Ecol Manag 329:21– 9.
- Belair ED, Saunders MR, Landhäusser SM. 2018. Growth traits of juvenile American chestnut and red oak as adaptations to disturbance. Restor Ecol 26:712–19.
- Brooks ML, D'antonio CM, Richardson DM, Grace JB, Keeley JE, DiTomaso JM, Hobbs RJ, Pellant M, Pyke D. 2004. Effects of invasive alien plants on fire regimes. BioScience 54:677–88.
- Brose PH, Dey DC, Waldrop TA. 2014. The fire—oak literature of eastern North America: synthesis and guidelines. Delaware, OH, USA: USDA Forest Service Northern Research Station [http://www.academia.edu/download/42004891/The\\_fire-oak\\_](http://www.academia.edu/download/42004891/The_fire-oak_literature_of_eastern_North20160203-30232-10ioxmj.pdf) [literature\\_of\\_eastern\\_North20160203-30232-10ioxmj.pdf.](http://www.academia.edu/download/42004891/The_fire-oak_literature_of_eastern_North20160203-30232-10ioxmj.pdf) Last accessed 27/06/2017.
- Brown CE, Mickelbart MV, Jacobs DF. 2014. Leaf physiology and biomass allocation of backcross hybrid American chestnut (Castanea dentata) seedlings in response to light and water availability. Tree Physiol 34:1362–75.
- Clark S, McNab H, Loftis D, Zarnoch S. 2012. American chestnut growth and survival five years after planting in two silvicultural treatments in the southern Appalachians, USA. Forests 3:1017–33.
- Clark SL, Schlarbaum SE, Pinchot CC, Anagnostakis SL, Saunders MR, Thomas-Van Gundy M, Schaberg P, McKenna J, Bard JF, Berrang PC, Casey DM, Casey CE, Crane B, Jackson BD, Kochenderfer JD, Lewis RF, MacFarlane R, Makowski R, Miller MD, Rodrigue JA, Stelick J, Thornton CD, Williamson TS. 2014. Reintroduction of American Chestnut in the National Forest System. J For 112:502–12.
- Clark SL, Schlarbaum SE, Saxton AM, Hebard FV. 2016. Establishment of American chestnuts (Castanea dentata) bred for blight (Cryphonectria parasitica) resistance: influence of breeding and nursery grading. New For 47:243–70.
- Dayton PK. 1972. Toward an understanding of community resilience and the potential effects of enrichments to the benthos at McMurdo Sound, Antarctica. Lawrence, KS: Allen Press.
- Diskin M, Steiner KC, Hebard FV. 2006. Recovery of American chestnut characteristics following hybridization and backcross breeding to restore blight-ravaged Castanea dentata. For Ecol Manag 223:439–47.
- Elliott KJ, Swank WT. 2008. Long-term changes in forest composition and diversity following early logging (1919–1923) and the decline of American chestnut (Castanea dentata). Plant Ecol 197:155–72.
- <span id="page-10-0"></span>Ellison AM, Bank MS, Clinton BD, Colburn EA, Elliott K, Ford CR, Foster DR, Kloeppel BD, Knoepp JD, Lovett GM, Mohan J, Orwig DA, Rodenhouse NL, Sobczak WV, Stinson KA, Stone JK, Swan CM, Thompson J, Von Holle B, Webster JR. 2005. Loss of foundation species: consequences for the structure and dynamics of forested ecosystems. Front Ecol Environ 3:479–86.
- Elton CS. 1958. The Ecology of Invasions by Animals and Plants. Chicago, Illinois, USA: University of Chicago Press.
- Engber EA, Varner JM. 2012. Patterns of flammability of the California oaks: the role of leaf traits. Can J For Res 42:1965– 75.
- Fonda RW. 2001. Burning characteristics of needles from eight pine species. For Sci 47:390–6.
- Forrestel AB, Moritz MA, Stephens SL. 2011. Landscape-scale vegetation change following fire in Point Reyes, California, USA. Fire Ecol 7:114–28.
- Fosberg MA, Lancaster JW, Schroeder MJ. 1970. Fuel moisture response- drying relationships under standard and field conditions. For Sci 16:121–8.
- Foster DR, Clayden S, Orwig DA, Hall B, Barry S. 2002. Oak, chestnut and fire: climatic and cultural controls of long-term forest dynamics in New England, USA. J Biogeogr 29:1359–79.
- Fraedrich SW, Harrington TC, Rabaglia RJ, Ulyshen MD, Mayfield Iii AE, Hanula JL, Eickwort JM, Miller DR. 2008. A fungal symbiont of the redbay ambrosia beetle causes a lethal wilt in redbay and other Lauraceae in the southeastern United States. Plant Dis 92:215–24.
- Guyette RP, Stambaugh MC, Dey DC, Muzika R-M. 2012. Predicting fire frequency with chemistry and climate. Ecosystems 15:322–35.
- Hawkins TS. 2006. A forest transect of Pine Mountain, Kentucky: changes since E. Lucy Braun and chestnut blight. J Ky Acad Sci 67:73–80.
- Hessl AE, Saladyga T, Schuler T, Clark P, Wixom J. 2011. Fire history from three species on a central Appalachian ridgetop. Can J For Res 41:2031–9.
- Jacobs D. 2007. Toward development of silvical strategies for forest restoration of American chestnut (Castanea dentata) using blight-resistant hybrids. Biol Conserv 137:497–506.
- Jacobs DF, Dalgleish HJ, Nelson CD. 2013. A conceptual framework for restoration of threatened plants: the effective model of American chestnut (Castanea dentata) reintroduction. New Phytol 197:378–93.
- Kane JM, Varner JM, Hiers JK. 2008. The burning characteristics of southeastern oaks: Discriminating fire facilitators from fire impeders. For Ecol Manag 256:2039–45.
- Keever C. 1953. Present composition of some stands of the former oak-chestnut forest in the southern Blue Ridge Mountains. Ecology 34:44–54.
- Kendra PE, Montgomery WS, Niogret J, Epsky ND. 2013. An Uncertain Future for American Lauraceae: A Lethal Threat from Redbay Ambrosia Beetle and Laurel Wilt Disease (A Review). Am J Plant Sci 04:727–38.
- Knapp BO, Wang GG, Clark SL, Pile LS, Schlarbaum SE. 2014. Leaf physiology and morphology of Castanea dentata (Marsh.) Borkh., Castanea mollissima Blume, and three backcross breeding generations planted in the southern Appalachians. USA. New For 45:283–93.
- Korstian CF, Stickel PW. 1927. The natural replacement of blight-killed chestnut in the hardwood forests of the northeast. J Agric Res 34:631–48.
- Kreye JK, Varner JM, Hamby GW, Kane JM. 2018. Mesophytic litter dampens flammability in fire-excluded pyrophytic oakhickory woodlands. Ecosphere 9:e02078.
- Kreye JK, Varner JM, Hiers JK, Mola J. 2013. Toward a mechanism for eastern North American forest mesophication: differential litter drying across 17 species. Ecol Appl 23:1976–86.
- Kuljian H, Varner JM. 2013. Foliar consumption across a sudden oak death chronosequence in laboratory fires. Fire Ecol 9:33– 44.
- Lafon CW, Naito AT, Grissino-Mayer HD, Horn SP, Waldrop TA. 2017. Fire history of the Appalachian region: a review and synthesis. Asheville, NC: USDA Forest Service, Southern Research Station [http://trace.tennessee.edu/utk\\_geogpubs/14/.](http://trace.tennessee.edu/utk_geogpubs/14/) Last accessed 31/07/2017.
- Levine JM, D'Antonio CM. 2003. Forecasting biological invasions with increasing international trade. Conserv Biol 17:322–6.
- Liebhold AM, MacDonald WL, Bergdahl D, Mastro VC. 1995. Invasion by exotic forest pests: a threat to forest ecosystems. For Sci Monogr 30:1–49.
- Lovett GM, Canham CD, Arthur MA, Weathers KC, Fitzhugh RD. 2006. Forest ecosystem responses to exotic pests and pathogens in eastern North America. BioScience 56:395–405.
- Mauseth JD. 2000. Theoretical aspects of surface-to-volume ratios and water-storage capacities of succulent shoots. Am J Bot 87:1107–15.
- McCormick JF, Platt RB. 1980. Recovery of an Appalachian forest following the chestnut blight or Catherine Keever-you were right!. Am Midl Nat 104:264.
- McEwan RW, Dyer JM, Pederson N. 2011. Multiple interacting ecosystem drivers: toward an encompassing hypothesis of oak forest dynamics across eastern North America. Ecography 34:244–56.
- Meentemeyer RK, Cunniffe NJ, Cook AR, Filipe JAN, Hunter RD, Rizzo DM, Gilligan CA. 2011. Epidemiological modeling of invasion in heterogeneous landscapes: spread of sudden oak death in California (1990–2030). Ecosphere 2:art17.
- Metz MR, Frangioso KM, Meentemeyer RK, Rizzo DM. 2011. Interacting disturbances: wildfire severity affected by stage of forest disease invasion. Ecol Appl 21:313–20.
- Metz MR, Varner JM, Frangioso KM, Meentemeyer RK, Rizzo DM. 2013. Unexpected redwood mortality from synergies between wildfire and an emerging infectious disease. Ecology 94:2152–9.
- Mola JM, Varner JM, Jules ES, Spector T. 2014. Altered community flammability in Florida's Apalachicola Ravines and implications for the persistence of the endangered conifer Torreya taxifolia. Bond-Lamberty B, editor. PLoS ONE 9:e103933.
- Muggeo VM. 2008. Segmented: an R package to fit regression models with broken-line relationships. R News 8:20–5.
- Murrill WA. 1906. A new chestnut disease. Torreya 6:186–9.
- Myers BR, Walck JL, Blum KE. 2004. Vegetation change in a former chestnut stand on the Cumberland Plateau of Tennessee during an 80-year period (1921–2000). Castanea 69:81–91.
- Nelson RM, Hiers JK. 2008. The influence of fuelbed properties on moisture drying rates and timelags of longleaf pine litter. Can J For Res 38:2394–404.
- Niinemets Ü, Valladares F. 2006. Tolerance to shade, drought, and waterlogging of temperate Northern Hemisphere trees and shrubs. Ecol Monogr 76:521–47.
- <span id="page-11-0"></span>Nowacki GJ, Abrams MD. 2008. The demise of fire and ''mesophication'' of forests in the eastern United States. BioScience 58:123–38.
- Oksanen J, Blanchet G, Friendly M, Kindt R, Legendre P, McGlinn D, Minchin PR, O'Hara RB, Simpson GL, Solymos P, Stevens HH, Szoecs E, Wagner H. 2017. Vegan: Community Ecology Package. R package version 2.4-4. [https://cran.r-proj](https://cran.r-project.org/package=vegan) [ect.org/package=vegan](https://cran.r-project.org/package=vegan).
- Paillet FL. 2002. Chestnut: history and ecology of a transformed species. J Biogeogr 29:1517–30.
- Pinchot C, Clark S, Schlarbaum S, Saxton A, Sung S-J, Hebard F. 2015. Effects of temporal dynamics, nut weight and nut size on growth of American chestnut, Chinese chestnut and backcross generations in a commercial nursery. Forests 6:1537–56.
- R Development Core Team. 2017. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing.
- Rhoades C, Loftis D, Lewis J, Clark S. 2009. The influence of silvicultural treatments and site conditions on American chestnut (Castanea dentata) seedling establishment in eastern Kentucky, USA. For Ecol Manag 258:1211–18.
- Rizzo DM, Garbelotto M. 2003. Sudden oak death: endangering California and Oregon forest ecosystems. Front Ecol Environ 1:197–204.
- Rothermel RC. 1983. How to predict the spread and intensity of forest and range fires. Ogden, UT: USDA Forest Service Intermountain Forest and Range Experiment Station.
- Saterberg T, Sellman S, Ebenman B. 2013. High frequency of functional extinctions in ecological networks. Nature 499:468–70.
- Scarff FR, Westoby M. 2006. Leaf litter flammability in some semi-arid Australian woodlands. Funct Ecol 20:745–52.
- Schweitzer JA, Bailey JK, Rehill BJ, Martinsen GD, Hart SC, Lindroth RL, Keim P, Whitham TG. 2004. Genetically based trait in a dominant tree affects ecosystem processes: Plant genetics impact ecosystems. Ecol Lett 7:127–34.
- Shapiro B. 2017. Pathways to de-extinction: how close can we get to resurrection of an extinct species? Seddon P, editor. Funct Ecol 31:996–1002.
- Sherkow JS, Greely HT. 2013. What if extinction is not forever? Science 340:32–3.
- Sniezko RA. 2006. Resistance breeding against nonnative pathogens in forest trees—current successes in North America. Can J Plant Pathol 28:S270–9.
- Stambaugh MC, Varner JM, Noss RF, Dey DC, Christensen NL, Baldwin RF, Guyette RP, Hanberry BB, Harper CA, Lindblom SG, Waldrop TA. 2015. Clarifying the role of fire in the deciduous forests of eastern North America: reply to Matlack: Fire in Deciduous Forests. Conserv Biol 29:942–6.
- Steiner KC, Carlson JE, editors. 2006. Restoration of American chestnut to forest lands: Proceedings of a conference and workshop. In: Restoration of American chestnut to forest lands: proceedings of a conference and workshop. Washington DC, USA: National Park Service [http://sfr.psu.edu/publi](http://sfr.psu.edu/public/chestnut/information/conference-2004/conference/conference-proceedings) [c/chestnut/information/conference-2004/conference/conferen](http://sfr.psu.edu/public/chestnut/information/conference-2004/conference/conference-proceedings) [ce-proceedings.](http://sfr.psu.edu/public/chestnut/information/conference-2004/conference/conference-proceedings)
- Steiner KC, Westbrook JW, Hebard FV, Georgi LL, Powell WA, Fitzsimmons SF. 2017. Rescue of American chestnut with

extraspecific genes following its destruction by a naturalized pathogen. New For 48:317–36.

- Sturrock RN, Frankel SJ, Brown AV, Hennon PE, Kliejunas JT, Lewis KJ, Worrall JJ, Woods AJ. 2011. Climate change and forest diseases: Climate change and forest diseases. Plant Pathol 60:133–49.
- Thomas-Van Gundy M, Bard J, Kochenderfer J, Berrang P. 2017. Mortality, early growth, and blight occurrence in hybrid, Chinese, and American chestnut seedlings in West Virginia. In: Proceedings 20th Central Hardwood Forest Conference. Vol. GTR NRS-P-167. Columbia, MO, USA: USDA Forest Service, Northern Research Station. pp 222–39.
- Varner JM, Arthur MA, Clark SL, Dey DC, Hart JL, Schweitzer CJ. 2016. Fire in eastern North American oak ecosystems: filling the gaps. Fire Ecol 12:1–5.
- Varner JM, Kane JM, Banwell EM, Kreye JK. 2015a. Flammability of litter from southeastern trees: A preliminary assessment. In: Holley GA, Connor KF, Haywood JD, editors. Proceedings of the 17th Southern Silvicultural Research Conference. Vol. eGTR-SRS-203. Ashville, NC: USDA Forest Service, Southern Research Station. pp 183–7.
- Varner JM, Kane JM, Kreye JK, Engber E. 2015b. The flammability of forest and woodland litter: a synthesis. Curr For Rep 1:91–9.
- Varner JM, Kuljian HG, Kreye JK. 2017. Fires without tanoak: the effects of a non-native disease on future community flammability. Biol Invasions 19:2307–17.
- Veldman JW, Mattingly WB, Brudvig LA. 2013. Understory plant communities and the functional distinction between savanna trees, forest trees, and pines. Ecology 94:424–34.
- Wang GG, Bauerle WL, Mudder BT. 2006. Effects of light acclimation on the photosynthesis, growth, and biomass allocation in American chestnut (Castanea dentata) seedlings. For Ecol Manag 226:173–80.
- Wang GG, Knapp BO, Clark SL, Mudder BT. 2013. The silvics of Castanea dentata (Marsh.) Borkh., American chestnut, Fagaceae (beech family). Asheville, NC, USA: USDA Forest Service Southern Research Station [https://www.researchgate.net/prof](https://www.researchgate.net/profile/Stacy_Clark3/publication/260779839_The_Silvics_of_Castanea_dentata_Marsh_Borkh_American_Chestnut_Fagaceae_Beech_Family/links/54806fa40cf2ccc7f8bd0477.pdf) [ile/Stacy\\_Clark3/publication/260779839\\_The\\_Silvics\\_of\\_Casta](https://www.researchgate.net/profile/Stacy_Clark3/publication/260779839_The_Silvics_of_Castanea_dentata_Marsh_Borkh_American_Chestnut_Fagaceae_Beech_Family/links/54806fa40cf2ccc7f8bd0477.pdf) [nea\\_dentata\\_Marsh\\_Borkh\\_American\\_Chestnut\\_Fagaceae\\_Be](https://www.researchgate.net/profile/Stacy_Clark3/publication/260779839_The_Silvics_of_Castanea_dentata_Marsh_Borkh_American_Chestnut_Fagaceae_Beech_Family/links/54806fa40cf2ccc7f8bd0477.pdf) [ech\\_Family/links/54806fa40cf2ccc7f8bd0477.pdf.](https://www.researchgate.net/profile/Stacy_Clark3/publication/260779839_The_Silvics_of_Castanea_dentata_Marsh_Borkh_American_Chestnut_Fagaceae_Beech_Family/links/54806fa40cf2ccc7f8bd0477.pdf) Last accessed 24/08/2017.
- Weed AS, Ayres MP, Hicke JA. 2013. Consequences of climate change for biotic disturbances in North American forests. Ecol Monogr 83:441–70.
- Whitham TG, Bailey JK, Schweitzer JA, Shuster SM, Bangert RK, LeRoy CJ, Lonsdorf EV, Allan GJ, DiFazio SP, Potts BM, Fischer DG, Gehring CA, Lindroth RL, Marks JC, Hart SC, Wimp GM, Wooley SC. 2006. A framework for community and ecosystem genetics: from genes to ecosystems. Nat Rev Genet 7:510–23.
- Whitham TG, Young WP, Martinsen GD, Gehring CA, Schweitzer JA, Shuster SM, Wimp GM, Fischer DG, Bailey JK, Lindroth RL, Woolbright S, Kuske CR. 2003. Community and ecosystem genetics: a consequence of the extended phenotype. Ecology 84:559–73.
- Youngs RL. 2000. A right smart little jolt: loss of the chestnut and a way of life. J For 98:17–21.