### **CASE REPORT**



# **A long‑term survivor of pediatric midline glioma with** *H3F3A* **K27M and** *BRAF* **V600E double mutations**

YoshikoNakano<sup>1,2</sup>® · Kai Yamasaki<sup>1,2</sup> · Hiroaki Sakamoto<sup>3</sup> · Yasuhiro Matsusaka<sup>3</sup> · Noritsugu Kunihiro<sup>3</sup> · **Hiroko Fukushima4 · Takeshi Inoue4 · Mai Honda‑Kitahara<sup>1</sup> · Junichi Hara2 · Akihiko Yoshida5 · Koichi Ichimura1**

Received: 8 February 2019 / Accepted: 24 June 2019 / Published online: 28 June 2019 © The Japan Society of Brain Tumor Pathology 2019

#### **Abstract**

We report a case of 2-year-old female with lateral ventricular glioma harboring both *H3F3A* K27M and *BRAF* V600E mutations. By the methylation analysis, the tumor was classifed as a difuse midline glioma H3 K27M mutant, WHO grade IV. However, the tumor was pathologically low-grade and likely localized rather than difusely infltrating. Further, the patient has survived more than 8 years after gross total resection of the tumor. Whereas both *H3F3A* K27M and *BRAF* V600E have been reported as poor prognostic markers in pediatric glioma, our case, along with several other reported cases, suggests that the coexistence of these two mutations might not indicate poor prognosis. The case emphasizes the importance of comprehensive assessment based on pathological, genetic and clinical fndings and calls for further investigations of non-difuse glioma with *H3F3A* K27M and glioma with *H3F3A* K27M and *BRAF* V600E.

**Keywords** *BRAF* V600E · *H3F3A* K27M · Pediatric glioma · Double mutations · Prognosis

# **Introduction**

The importance of genetic analysis of pediatric brain tumors has dramatically increased in recent years. First, diagnosis based on the revised 2016 WHO classifcation of tumors of the central nervous system requires the analysis of certain missense mutations, fusion genes, and amplifcations/ deletions of certain chromosome regions [[1](#page-5-0), [2\]](#page-5-1). Second, for

Yoshiko Nakano and Kai Yamasaki contributed equally to this work.

 $\boxtimes$  Yoshiko Nakano yonakano@ncc.go.jp

- <sup>1</sup> Division of Brain Tumor Translational Research, National Cancer Center Research Institute, 5-1-1, Tsukiji, Chuo-ku, Tokyo 104-0045, Japan
- <sup>2</sup> Department of Pediatric Hematology/Oncology, Osaka City General Hospital, Osaka, Japan
- <sup>3</sup> Department of Pediatric Neurosurgery, Osaka City General Hospital, Osaka, Japan
- <sup>4</sup> Department of Pathology, Osaka City General Hospital, Osaka, Japan
- <sup>5</sup> Department of Pathology and Clinical Laboratories, National Cancer Center Hospital, Tokyo, Japan

 $\circled{2}$  Springer

medulloblastomas, a molecular classifcation is used both as a prognostic marker and for stratifcation into risk groups [[3\]](#page-5-2). Third, some genetic abnormalities, such as the *BRAF* V600E mutation, and the *NTRK*, *MET*, and *ALK/ROS1* fusion genes, can be efective therapeutic targets [[4–](#page-5-3)[7\]](#page-5-4). In addition, methylation-based classifcation has become a powerful tool [[8\]](#page-5-5). For example, posterior fossa ependymoma is molecularly classifed into the PFA and PFB subgroups, which have distinct clinical features [[9–](#page-5-6)[11](#page-5-7)]. Strum et al. reported that CNS–PNET could be reclassifed into several novel subgroups [[12\]](#page-5-8). Further, in 2018, the German Cancer Research Center (DKFZ) demonstrated the utility and accuracy of methylation-based classifcation of all brain tumor entities, and this classifcation tool is now available on their website [[13\]](#page-5-9).

Here, we report a case of pediatric midline low-grade glioma (LGG) in the lateral ventricle harboring both the *H3F3A* K27M and *BRAF* V600E mutations. Based on the methylation analysis, the tumor was classifed as a "difuse midline glioma with an H3 K27M mutant." However, the pathological and clinical features were distinct from those of this entity.

<span id="page-1-0"></span>



## **Clinical summary**

A 2-year-old female was referred to our hospital because of an intraventricular mass. The patient was born at 33 weeks gestation and she stayed in the hospital for 1 month because of her low birth weight. At the age of 1 year, she presented with epilepsy, which was controlled with an antiepileptic drug. MRI showed a nodular mass in the left ventricle (Fig. [1](#page-1-0)a–d). The patient underwent gross total resection of the mass because it was slowly growing. Based on the intraoperative fndings, the border between the tumor and the normal thalamic tissues was partially unclear, whereas that between the tumor and ventricular lumen was clear. The local pathological diagnosis based on the 2007 WHO classifcation was oligoastrocytoma. She received no chemotherapy or radiotherapy and has survived without the evidence of disease for more than 8.5 years. After the publication of the revised fourth edition of the WHO classifcation in 2016, a molecular analysis and a review of the central pathology were performed (Fig. [2](#page-1-1), [3](#page-2-0)a–f).



<span id="page-1-1"></span>**Fig. 2** Pyrosequencing results showing the *BRAF* V600E and *H3F3A* K27M mutations

# **Pathological fndings**

The tumor consisted of moderately cellular glial proliferation within a fbrillary background. The lesional cells

were relatively uniform in appearance and were round, oval, and spindle shaped. Pleomorphic, giant, epithelioid, rhabdoid, and piloid cells were lacking. The round cell component was associated with a perinuclear halo and resembled oligodendroglial tumors, whereas the spindle cells resembled the astrocytic lineage (Fig. [3](#page-2-0)a, b). There were a small number of rosette-like glial structures (Fig. [3](#page-2-0)c). Dysmorphic ganglion cells were absent. Mitotic fgures were not detected. Necrosis, microvascular proliferation, Rosenthal fbers, and eosinophilic granular bodies were not observed. Focal microcalcifcation was present. Immunohistochemically, the tumor cells were positive for glial fbrillary acidic protein and S100 protein but were negative for epithelial membrane antigen and synaptophysin. In agreement with the molecular fndings, they were immunopositive for H3 K27M, with a concomitant loss of H3K27me3 expression (Fig. [3d](#page-2-0), e). BRAF V600E immunohistochemistry was positive (Fig. [3f](#page-2-0)), and H3 K27M and BRAF V600E staining appeared to be present in the same cells. The tumor retained ATRX expression, and p53

<span id="page-2-0"></span>**Fig. 3** Histological and immunohistochemical fndings. The tumor consisted of glial proliferation with oligodendroglial (**a**) and astrocytic (**b**) morphology. There were a small number of rosette-like glial structures (**c**). Immunohistochemically, the tumor cells were positive for H3 K27M (**d**), with a concomitant loss of H3K27me3 expression (**e**). BRAF (V600E) immunohistochemistry was also difusely positive (**f**)

staining showed wild-type labeling. The MIB1 labeling index was 6.8%. The tumor mostly lacked transgressing neuroflament–positive fbers, and there was no evidence of difuse infltration.

# **Molecular analysis**

DNA and RNA were extracted from frozen tumor tissue using the DNeasy Blood and Tissue Kit (Qiagen, Tokyo, Japan) and the miRNeasy Mini Kit (Qiagen, Tokyo, Japan), respectively. Hot spot mutations, including *IDH1* R132, *IDH2* R172, *BRAF* T599, *BRAF* V600, *H3F3A* K27, *H3F3A* G34, *TERT p*romoter C250, *TERT* promoter C228, *FGFR1* N546, and *FGFR1* K656, were analyzed by pyrosequencing using the AQ assay with a PyroMark Q96 (Qiagen, Tokyo, Japan). The polymerase chain reaction (PCR) for pyrosequencing and the pyrosequencing assay were performed as previously described [[14\]](#page-5-10). Primer sequences are listed in supplementary Table [1](#page-3-0). The *BRAF* V600E and *H3F3A*



K27M mutations were detected at frequencies of 40% and 46%, respectively (Fig. [2\)](#page-1-1). The hot spot mutation in other analyzed genes was not detected. Multiplex ligationdependent probe amplifcation (MLPA) using the SALSA MLPA probemix P088-C2 (MRC-Holland, Amsterdam, the Netherlands) revealed no *CDKN2A* deletion. The existence of *KIAA1549*-*BRAF* fusion was assessed by reverse transcriptase PCR using previously reported primers and it was not detected [[15\]](#page-5-11).

A methylation classifer assay was then performed. DNA methylation was analyzed using an Infnium HumanMethylation450 BeadChip array (Illumina, San Diego, CA, USA), and the IDAT-fles from the sample were uploaded to the online classifer developed by the DKFZ ([https://www.molec](https://www.molecularneuropathology.org/mnp)

[ularneuropathology.org/mnp](https://www.molecularneuropathology.org/mnp)). The tumor was classifed as a "difuse midline glioma, H3 K27M mutant" with a calibrated score of 0.98.

## **Discussion**

Here, we report a case of midline glioma harboring concurrent *H3F3A* K27M and *BRAF* V600E mutations. The tumor was classifed as a "difuse midline glioma, H3 K27M mutant, WHO grade IV" by methylation analysis. However, in several aspects, the case was not entirely compatible with typical difuse midline gliomas with *H3F3A* K27M mutations. Importantly, the present



<span id="page-3-0"></span>**Table 1** Reported cases of pediatric intracranial tumors harboring the *H3F3A* K27M and *BRAF* V600E double mutations

*AA* anaplastic astrocytoma, *NA* no data available, *LGG* low grade glioma, *NOS* no otherwise specifed, *GTR* gross total resection, *GG* ganglioglioma, *DA* difuse astrocytoma, *PA* pilocytic astrocytoma, *HGG* high grade glioma, *GBM* glioblastoma, *PXA* pleomorphic xanthoastrocytoma, *AGG* anaplastic ganglioglioma

tumor was likely localized rather than difusely infltrating. Radiologically, it seemed well circumscribed; that is most of the tumor surface being surrounded by cerebral fuid. However, the border between tumor and normal thalamic tissue was partly unclear. Histologically, there was a paucity of transgressing neuroflament, suggesting a non-difuse process, although this fnding needs to be interpreted with caution because it may simply refect the intra-ventricular growth. "Diffuse midline glioma with H3 K27M mutant WHO grade IV" was introduced as a new entity of WHO classifcation 2016, based on the knowledge at that time that recurrent mutation at K27 in *H3F3A, HIST1H3B,* and *HIST1H3C* is detected in high-grade glioma (HGG) from the pons, thalamus and spinal cord and that these mutations occur exclusively in difuse midline gliomas, as stated in WHO blue book  $[2, 16]$  $[2, 16]$  $[2, 16]$ . However, some cases of non-difuse glioma with H3F3A K27M were subsequently reported [[17](#page-5-15), [18\]](#page-5-16). Accordingly, Consortium to Inform Molecular and Practical Approaches to CNS Tumor Taxonomy-Not Official WHO (cIMPACT-NOW) up date 2 emphasizes that "difuse," "midline," "glioma" and "H3 K27M-mutant" are requirements for the diagnosis of difuse midline glioma with H3 K27M [[16](#page-5-14)]. Therefore, based on the cIMPACT-NOW-modifed view of the WHO Classifcation, the present case seems incompatible with the "difuse mid-line glioma with H3 K27M" designation. Whereas some of the reported localized gliomas with H3 K27M demonstrated the histology of well-established entities, the present case was difficult to classify histologically. The original diagnosis of oligoastrocytoma is likely invalid due to its mostly localized nature. Although histological fndings, along with the presence of the *BRAF* mutation, may suggest a possibility of pilocytic astrocytoma, characteristic features of that entity were mostly lacking, including piloid or microcystic loose tissues, Rosenthal fbers, eosinophilic granular bodies, and microvascular proliferation. The morphology did not ft well with other *BRAF*-mutant localized gliomas either, such as gangliogioma and pleomorphic xanthoastrocytoma. Taken together, the present case would be best labeled descriptively as low-grade glioma, likely localized, not elsewhere classifed as per the Consortium to Inform Molecular and Practical Approaches to CNS Tumor Taxonomy-Not Official WHO cIMPACT-NOW update 1 [\[16,](#page-5-14) [19](#page-5-17)].

Classifcation based on methylation profles is a highly robust method for facilitating diagnosis and subgrouping brain tumors. However, the interpretation of methylationbased classifcation may sometimes require caution. H3 K27M would cause a considerable change in the tumor's epigenetic landscape, which may mask fne diferences among K27M-harboring tumors, some of which are not compatible with the defnition of difuse midline glioma with H3 K27M mutation, as in our case. Other mutations, such as

*BRAF*, may on the other hand less affect epigenetic landscape, which could make them more difficult to distinguish by methylation profling alone. The principle of defning a tumor entity, whether using should histology, genetics, methylation, or their combination, warrants further discussion.

In addition to the location and histological fndings presented above, the patient's age at diagnosis and her relatively long survival of the present case are also unusual for H3F3A K27M–positive glioma. The *H3F3A* K27M mutation is characteristically detected in pediatric gliomas. The mean age at diagnosis is 6–10 years [[6,](#page-5-18) [20](#page-5-12)[–22\]](#page-5-13). Most *H3F3A* K27M -positive gliomas arise in the pons or thalamus, and only approximately 5% arise in the ventricle  $[6, 20]$  $[6, 20]$  $[6, 20]$  $[6, 20]$ . In most cases, the histological grade is high. The median overall survival of *H3F3A* K27M-positive midline glioma is less than 1 year, which is signifcantly worse than that of *H3F3A* K27 wild-type glioma. Based on an analysis of 77 pediatric patients with difuse midline glioma over the age of 3 years, Karremman et al. reported that H3 K27M status has a negative impact on survival, regardless of tumor location or pathological grade [[20](#page-5-12)]. There is a case report of pediatric LGG in the thalamus harboring H3 K27M transformed to HGG [\[23\]](#page-5-19). Pratt et al. also reported that H3 K27M has a negative impact on survival in circumscribed/non-difuse glioma [[24](#page-5-20)].

In contrast to *H3F3A* K27M, *BRAF* V600E is detected in a variety of tumors arising in patients of all ages, including thyroid and colorectal cancers as well as Langerhans cell histiocytosis [[3,](#page-5-2) [25–](#page-5-21)[31](#page-6-5)]. It is also detected in various glioma subtypes arising in any location, and the frequency is high in some subtypes, such as pleomorphic xanthoastrocytoma. The prognostic implications of *BRAF* V600E are dependent on various factors, including patient age, patient gender, tumor type, and the presence of other concurrent molecular abnormalities [\[3](#page-5-2), [25–](#page-5-21)[32\]](#page-6-6). For pediatric LGG, several reports suggest that *BRAF* V600E is associated with an aggressive clinical course, although this is somewhat controversial [\[3,](#page-5-2) [28,](#page-6-7) [32,](#page-6-6) [33\]](#page-6-8). Lassaletta et al. reported that the progression-free survival of patients with *BRAF* V600E–positive LGG is significantly worse than that of patients with wild-type *BRAF* LGG [\[3](#page-5-2)]. Mistry et al. reported that the frequency of *BRAF* V600E in pediatric LGG that underwent malignant transformation was higher than the frequency in non-transformed LGG (44% vs. 6%) [\[31](#page-6-5)].

Although the coexistence of *H3F3A* K27M and *BRAF* V600E mutations is rare, there have been several reported cases of pediatric glioma harboring these concurrent mutations (Table [1\)](#page-3-0) [\[20,](#page-5-12) [22](#page-5-13), [32](#page-6-6), [34](#page-6-4)[–38\]](#page-6-3). Interestingly, these include both difuse midline glioma and non-difuse glioma. In view of the diagnostic and prognostic information of this alternation, the recently published cIMPACT-NOW update 4 proposed the new classifcation name "Difuse glioma, *BRAF* V600E mutant." The diagnosis of some cases shown in Table [1](#page-3-0) may warrant further discussion [\[39](#page-6-9)]. It is also noteworthy that the

prognosis of these patients was not necessarily dismal but instead appeared to be somewhat better than that of patients with midline glioma harboring only the *H3F3A* K27M mutation, and that no patients died within the frst year after the initial diagnosis. Notably, four patients, including all three patients under the age of 3 years, survived for more than 5 years. However, the number of reported cases with concurrent mutations is small and histological diagnosis, grading and age of diagnosis are variable. The follow-up periods are too short and recent widespread application of BRAF-targeted therapy may change their survival [[40](#page-6-10)]. Therefore, further analysis of more cases is required to clarify these points.

In summary, we report a case of a relatively long-term survivor of pediatric midline glioma harboring concurrent *BRAF* V600E and *H3F3A* K27M. Although the tumor was classifed as "difuse midline glioma with H3K27M mt, WHO grade IV" based on methylation analysis, the pathological and clinical features of this case did not ft those of "difuse midline glioma with H3 K27M mt" as defined in the current WHO classification (2016). Our case points to certain limitation of the current methylation classifer and it calls for further discussion of the integrated diagnosis of brain tumor. Further investigations of non-difuse glioma with *H3F3A* K27M and glioma with both *BRAF* V600E and *H3F3A* K27 M are warranted.

**Acknowledgements** The authors thank the patient and her family for participating in this research. We also thank Y. Matsushita for technical support with the molecular analysis and K. Fukuoka for insightful comments.

#### **Compliance with ethical standards**

**Conflict of interest** The authors declare no conficts of interest associated with this manuscript.

### **References**

- <span id="page-5-0"></span>1. Komori T (2017) Updated 2016 WHO classifcation of tumors of the CNS: turning the corner where molecule meets pathology. Brain Tumor Pathol 34(4):139–140
- <span id="page-5-1"></span>2. Louis DN, Ohgaki H, Wiestler OD, Cavenee WK (2016) WHO classifcation of tumours of the central nervous system, 2nd edn. IARC Press, Lyon
- <span id="page-5-2"></span>3. Ramaswamy V, Taylor MD (2017) Medulloblastoma: from myth to molecular. J Clin Oncol 35(21):2355–2363
- <span id="page-5-3"></span>4. Lassaletta A, Zapotocky M, Mistry M et al (2017) Therapeutic and prognostic implications of BRAF V600E in pediatric low-grade gliomas. J Clin Oncol 35(25):2934–2941
- 5. Yoshihara K, Wang Q, Torres-Garcia W, Zheng S, Vegesna R, Kim H, Verhaak RG (2015) The landscape and therapeutic relevance of cancer-associated transcript fusions. Oncogene 34(37):4845–4854
- <span id="page-5-18"></span>6. Nakano Y, Tomiyama A, Kohno T et al (2018) Identifcation of a novel KLC1-ROS1 fusion in a case of pediatric low-grade localized glioma. Brain Tumor Pathol 36(1):14–19
- <span id="page-5-4"></span>7. Wu G, Diaz AK, Paugh BS et al (2014) The genomic landscape of difuse intrinsic pontine glioma and pediatric non-brainstem high-grade glioma. Nat Genet 46(5):444–450
- <span id="page-5-5"></span>8. Mack SC, Northcott PA (2017) Genomic analysis of childhood brain tumors: methods for genome-wide discovery and precision medicine become mainstream. J Clin Oncol 35(21):2346–2354
- <span id="page-5-6"></span>9. Witt H, Mack SC, Ryzhova M et al (2011) Delineation of two clinically and molecularly distinct subgroups of posterior fossa ependymoma. Cancer Cell 20(2):143–157
- 10. Mack SC, Witt H, Piro RM et al (2014) Epigenomic alterations defne lethal CIMP-positive ependymomas of infancy. Nature 506(7489):445–450
- <span id="page-5-7"></span>11. Fukuoka K, Kanemura Y, Shofuda T et al (2018) Signifcance of molecular classifcation of ependymomas: c11orf95-RELA fusion-negative supratentorial ependymomas are a heterogeneous group of tumors. Acta Neuropathol Commun 6(1):134
- <span id="page-5-8"></span>12. Sturm D, Orr BA, Toprak UH et al (2016) New brain tumor entities emerge from molecular classifcation of CNS-PNETs. Cell 164(5):1060–1072
- <span id="page-5-9"></span>13. Capper D, Jones DTW, Sill M et al (2018) DNA methylationbased classifcation of central nervous system tumours. Nature 555(7697):469–474
- <span id="page-5-10"></span>14. Arita H, Narita Y, Matsushita Y et al (2015) Development of a robust and sensitive pyrosequencing assay for the detection of IDH1/2 mutations in gliomas. Brain Tumor Pathol 32(1):22–30
- <span id="page-5-11"></span>15. Jones DT, Kocialkowski S, Liu L et al (2008) Tandem duplication producing a novel oncogenic BRAF fusion gene defnes the majority of pilocytic astrocytomas. Cancer Res 68(21):8673–8677
- <span id="page-5-14"></span>16. Louis DN, Giannini C, Capper D et al (2018) cIMPACT-NOW update 2: diagnostic clarifcations for difuse midline glioma, H3 K27M-mutant and difuse astrocytoma/anaplastic astrocytoma, IDH-mutant. Acta Neuropathol 135(4):639–642
- <span id="page-5-15"></span>17. Kleinschmidt-DeMasters BK, Mulcahy Levy JM (2018) H3 K27 M-mutant gliomas in adults vs. children share similar histological features and adverse prognosis. Clin Neuropathol 37:53–63
- <span id="page-5-16"></span>18. Morita S, Nitta M, Muragaki Y et al (2018) Brainstem pilocytic astrocytoma with H3 K27M mutation: case report. J Neurosurg 129(3):593–597
- <span id="page-5-17"></span>19. Louis DN, Wesseling P, Paulus W et al (2018) cIMPACT-NOW update 1: not otherwise specifed (NOS) and not elsewhere classifed (NEC). Acta Neuropathol 135(3):481–484
- <span id="page-5-12"></span>20. Karremann M, Gielen GH, Hofmann M et al (2018) Difuse highgrade gliomas with H3 K27M mutations carry a dismal prognosis independent of tumor location. Neuro Oncol 20(1):123–131
- 21. Khuong-Quang DA, Buczkowicz P, Rakopoulos P et al (2012) K27M mutation in histone H3.3 defnes clinically and biologically distinct subgroups of pediatric difuse intrinsic pontine gliomas. Acta Neuropathol 124(3):439–447
- <span id="page-5-13"></span>22. Mackay A, Burford A, Molinari V et al (2018) Molecular, pathological, radiological, and immune profling of non-brainstem pediatric high-grade glioma from the HERBY phase II randomized trial. Cancer Cell 33(5):829–842 **(e825)**
- <span id="page-5-19"></span>23. Ishibashi K, Inoue T, Fukushima H, Watanabe Y et al (2016) Pediatric thalamic glioma with H3F3A K27M mutation, which was detected before and after malignant transformation: a case report. Childs Nerv Syst 32(12):2433–2438
- <span id="page-5-20"></span>24. Pratt D, Natarajan SK, Banda A et al (2018) Circumscribed/nondifuse histology confers a better prognosis in H3K27M-mutant gliomas. Acta Neuropathol 135(2):299–301
- <span id="page-5-21"></span>25. Tabouret E, Bequet C, Denicolai E et al (2015) BRAF mutation and anaplasia may be predictive factors of progression-free survival in adult pleomorphic xanthoastrocytoma. Eur J Surg Oncol 41(12):1685–1690
- 26. Vuong HG, Altibi AMA, Duong UNP et al (2018) BRAF mutation is associated with an improved survival in glioma-a systematic review and meta-analysis. Mol Neurobiol 55(5):3718–3724
- 27. Chen X, Pan C, Zhang P et al (2017) BRAF V600E mutation is a signifcant prognosticator of the tumour regrowth rate in brainstem gangliogliomas. J Clin Neurosci 46:50–57
- <span id="page-6-7"></span>28. Jones DTW, Witt O, Pfster SM (2018) BRAF V600E status alone is not sufficient as a prognostic biomarker in pediatric low-grade glioma. J Clin Oncol 36(1):96
- 29. Kim TH, Park YJ, Lim JA et al (2012) The association of the BRAF (V600E) mutation with prognostic factors and poor clinical outcome in papillary thyroid cancer: a meta-analysis. Cancer 118(7):1764–1773
- 30. Xing M, Liu R, Liu X, Murugan AK, Zhu G, Zeiger MA, Pai S, Bishop J (2014) BRAF V600E and TERT promoter mutations cooperatively identify the most aggressive papillary thyroid cancer with highest recurrence. J Clin Oncol 32(25):2718–2726
- <span id="page-6-5"></span>31. Heritier S, Emile JF, Barkaoui MA et al (2016) BRAF mutation correlates with high-risk Langerhans cell histiocytosis and increased resistance to first-line therapy. J Clin Oncol 34(25):3023–3030
- <span id="page-6-6"></span>32. Mistry M, Zhukova N, Merico D et al (2015) BRAF mutation and CDKN2A deletion defne a clinically distinct subgroup of childhood secondary high-grade glioma. J Clin Oncol 33(9):1015–1022
- <span id="page-6-8"></span>33. Ho CY, Mobley BC, Gordish-Dressman H et al (2015) A clinicopathologic study of diencephalic pediatric low-grade gliomas with BRAF V600 mutation. Acta Neuropathol 130(4):575–585
- <span id="page-6-4"></span>34. Zhang J, Wu G, Miller CP et al (2013) Whole-genome sequencing identifes genetic alterations in pediatric low-grade gliomas. Nat Genet 45(6):602-612
- <span id="page-6-2"></span>35. Pages M, Beccaria K, Boddaert N et al (2018) Co-occurrence of histone H3 K27M and BRAF V600E mutations in paediatric midline grade I ganglioglioma. Brain Pathol 28(1):103–111
- <span id="page-6-0"></span>36. Nguyen AT, Colin C, Nanni-Metellus I et al (2015) Evidence for BRAF V600E and H3F3A K27M double mutations in paediatric glial and glioneuronal tumours. Neuropathol Appl Neurobiol 41(3):403–408
- <span id="page-6-1"></span>37. Solomon DA, Wood MD, Tihan T, Bollen AW, Gupta N, Phillips JJ, Perry A (2016) Difuse midline gliomas with histone H3-K27M mutation: a series of 47 cases assessing the spectrum of morphologic variation and associated genetic alterations. Brain Pathol 26(5):569–580
- <span id="page-6-3"></span>38. Ryall S, Krishnatry R, Arnoldo A et al (2016) Targeted detection of genetic alterations reveal the prognostic impact of H3K27M and MAPK pathway aberrations in paediatric thalamic glioma. Acta Neuropathol Commun 4(1):93
- <span id="page-6-9"></span>39. Ellison DW, Hawkins C, Jones DTW et al (2019) cIMPACT-NOW update 4: difuse gliomas characterized by MYB, MYBL1, or FGFR1 alterations or BRAF(V600E) mutation. Acta Neuropathol 137(4):683–687
- <span id="page-6-10"></span>40. Penman CL, Faulkner C, Lowis SP, Kurian KM (2015) Current understanding of BRAF alterations in diagnosis, prognosis, and therapeutic targeting in pediatric low-grade gliomas. Front Oncol 5:54

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.