



Electrochemical sensor using graphene/Fe₃O₄ nanosheets functionalized with garlic extract for the detection of lead ion

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Abstract

Based on the modulated electronic properties of Fe₃O₄-graphene (Fe₃O₄/GN composite) as well as the outstanding complexation between Pb²⁺ and natural substances garlic extract (GE), a novel electrochemical sensor for the determination of Pb²⁺ in wastewater was prepared by immobilization of Fe₃O₄/GN composite integrated with GE onto the surface of glassy carbon electrode (GCE). Fe₃O₄/GN composite was employed as an electrochemical active probe for enhancing electrical response by facilitating charge transfer while GE was used to improve the selectivity and sensitivity of the proposed sensor to Pb²⁺ assay. The electrochemical sensing performance toward Pb²⁺ was appraised by cyclic voltammetry (CV), electrochemical impedance spectroscopy (EIS), and square wave voltammetry (SWV). Under the optimized condition, the sensor exhibited two dynamic linear ranges (LDR) including 0.001 to 0.5 nM and 0.5 to 1000 nM with excellent low detection limit (LOD) of 0.0123 pM ($S/N = 3$) and quantification limit (LOQ) of 0.41 pM ($S/N = 10$). Meanwhile, it displayed remarkable stability, reproducibility (RSD of 3.61%, $n = 3$), and selectivity toward the assay for the 100-fold higher concentration of other heavy metal ions. Furthermore, the novel sensor has been successfully employed to detect Pb²⁺ from real water samples with satisfactory results.

Keywords Garlic extract · Lead(II) · Fe₃O₄/GN · Electrochemical sensor

Introduction

Garlic (*Allium sativum* L.) is a worldwide cultivated Alliaceae species and of economic benefits in food and medicine. In the

previous research, garlic is effective in treating lead intoxication for workers who are exposed to environmental lead. In general, garlic and its derivatives have been gradually recommended as a promising agent for lead treatment because of the negative effect of the chelating agents that can deplete the body of essential metals resulting in secondary injury for health [1]. The positive effects of garlic on human health are ascribed to the presence of bioactive substances such as organosulfur compounds [2–5], flavonoids, and vitamins [6]. The organosulfur compounds mainly include diallyl trisulfide, diallyl tetrasulfide, s-allylcysteine, vinylthiines, allylpropyl disulfide, ajoenes ((E)- and (Z)-4, 5, 9-trithiadodeca-1, 6, 11-triene-9-oxides), and allicin [5, 7–9]. The components are likely to work synergistically to provide the largest range of garlic's health benefits. To the best of our knowledge, Fatima and Ahmad [10] have utilized the antioxidant enzymes of garlic as biomarkers of heavy metals for detecting the heavy metals in wastewater. Cheng et al. [11] and Zhou et al. [12] have fabricated electrochemical sensors for determining Pb²⁺ based on L-cysteine. However, garlic has not been explored in the field of electrochemical sensor to detect the presence of Pb²⁺ in wastewater, which is expected to attract particular attention for the detection of Pb²⁺ by virtue of their simple operation modes, rapid

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response, high sensitivity, and selectivity [13]. Hence, in this study, garlic extract (GE), as a natural product, may provide a sensitive electrochemical interface in the field of electrochemical sensor and pre-concentrate Pb^{2+} to produce stable complexes for the recognition of Pb^{2+} in wastewater.

Nonetheless, the preponderances of GE heavily rely on the supporting materials which are in possession of high specific surface area, superior electronic conductivity, and easy functionalization. Based on these criterions, graphene is an ideal supporting material [14]. Graphene (GN) has a unique nanostructure and interesting properties (such as large surface, excellent conductive property, prominent thermal stability) [15], which results in widespread potential applications, like supercapacitors [16] or serving as absorbent of heavy metal ions [17]. However, it has been widely reported that the graphene-metal oxide nanomaterials have better functionalization and performance in their applications than graphene or metal oxide nanomaterials solely [14, 18], resulting from inheriting the advantages of two component materials [19–21]. Therefore, it is an interesting and appealing method that Fe_3O_4 nanoparticles are grown and anchored on GN sheets as supporting material for the electrode of electrochemical sensor, combining with the fascinating merits of Fe_3O_4 NPs and GN and exhibiting a better electrochemical performance compared with the native phase [21–27]. Suryawanshi et al. [28] have successfully prepared a Fe_3O_4 hierarchically perforated graphene nanosheets composite with enhanced electronic conductivity. Zhou et al. [29] have synthesized a well-organized flexible interleaved GNS/ Fe_3O_4 composite through in situ reduction of iron hydroxide between graphene nanosheets (GNS), behaving superior electrochemical performance. Furthermore, the incorporation of Fe_3O_4 NPs onto graphene sheets tackled the aggregation of Fe_3O_4 nanoparticles and preserved the electrochemically active surface of Fe_3O_4 NPs [30]. Thus, the Fe_3O_4 /GN composite is a remarkable electrode material for electrochemical sensor. Based on this background, in this study, Fe_3O_4 /GN composite as an electrochemical active probe could enhance the electrical response by facilitating charge transfer.

The main purpose of this investigation was to develop a simple, sensitive, selective, and inexpensive electrochemical method for the determination of Pb^{2+} in wastewater. In the current work, a simple and novel electrochemical sensor with high sensitivity and selectivity has been on the spotlight for Pb^{2+} determination in wastewater by using Fe_3O_4 /GN/GE-modified glassy carbon electrode (GCE). The electrode preparation is described in Scheme 1. The GE and Fe_3O_4 /GN composite are chosen as the modifier for GE is an economical nature product and Fe_3O_4 /GN composite is environmentally friendly, respectively. To the best of our knowledge, this is the first work describing and integrating the unique features of both Fe_3O_4 /GN composite and GE through fabricating a

sensor to exploit their synergy for the electrochemical detection of Pb^{2+} . It was found that the proposed sensor is allowed to select optimal conditions for the detection of Pb^{2+} , showing satisfactory linearity, detection limit, sensitivity, selectivity, and high reproducibility and stability. Relative to the reported methods [31–34], the established method has superior electrochemical properties. Furthermore, the experiment results of determination Pb^{2+} in real samples by the developed sensor are also acceptable. Thus, not only does Fe_3O_4 /GN/GE sensor represent a new electrochemical platform for designing environment-friendly sensors but also could meet the needs of practical analysis.

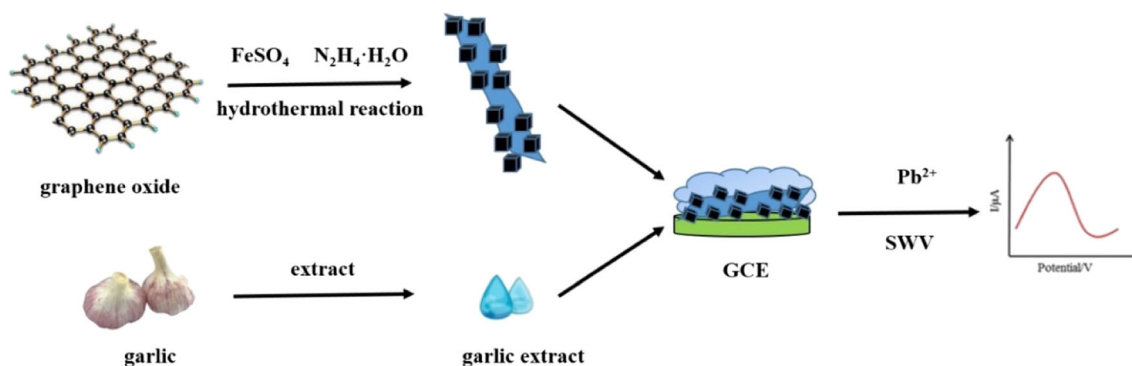
Experiment

Chemicals and apparatus

Aged garlic was purchased from a local market (Chongqing, China). Graphene oxide (GO) was provided by Sigma-Aldrich. All other chemicals, such as $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$, NaOH, and $\text{N}_2\text{H}_4 \cdot \text{H}_2\text{O}$, were of analytical grade purity and purchased from Tianjin Chemical Technology Co., Ltd. (China) without further purification. Unless otherwise stated, the supporting electrolyte is a 0.1 mol/L acetate buffer solution (ABS, pH 5.50). The double-distilled water was used for all solutions.

The morphologies of the as-prepared samples were acquired on a JSM-7500F (Hitachi Co., Ltd., Japan) scanning electron microscope and Tecnai G20 transmission electron microscope (FEI Company, USA). The crystallographic information was recorded by a XRD-6000 classical powder diffractometer with Cu $k\alpha$ radiation ($\lambda = 0.154$ nm). The function groups of the samples were gathered by a Nicolet 550 FTIR Spectrometer (Shimadzu Scientific Instruments) from 400 to 4000 cm^{-1} at room temperature.

GC–MS analysis of the garlic extract (GE) was performed with a 6890N gas chromatograph (Agilent, American) system equipped with a fused silica capillary Agilent Technology HP-5 ms (5% phenyl methyl siloxane) column (30 m \times 0.25 mm i.d., film thickness 0.1 μm) and a 5975C plus mass spectrometer (Agilent, American). For GC–MS detection, 1 μL sample was injected for analysis. In the GC system, injector and detection temperature were set at 280 and 290 $^\circ\text{C}$, respectively. The initial temperature was kept at 40 $^\circ\text{C}$ for 2 min and ramped to 130 $^\circ\text{C}$ with a rate of 8 $^\circ\text{C}/\text{min}$, then increased to 290 $^\circ\text{C}$ (10 min) at a rate of 10 $^\circ\text{C}/\text{min}$. Carrier gas was helium with flow rate of 1 mL/min. A 70-eV EI mode with an ionic source temperature of 230 $^\circ\text{C}$ was used in the MS system. The standard mass spectra of organosulfur compounds were provided by literature data and the software of the GC–MS system



Scheme 1 The preparation process of Fe₃O₄/GN/GE electrochemical detection of Pb²⁺.

(National Institute of Standards and Technology (NIST 05. LIB) libraries date).

A CHI660E electrochemical workstation (ChenHua Instrument Co., Shanghai, China) was used to perform the electrochemical experiments, including cyclic voltammetry (CV) and square wave voltammetry (SWV). Different modified glassy carbon electrodes (Fe₃O₄/GCE, Fe₃O₄/GN/GCE, Fe₃O₄/GN/GE/GCE) or a bare glassy carbon electrode (GCE, 3 mm diameter) were used as the working electrode, with an Ag/AgCl/KCl (3 mol/L KCl saturated with AgCl) and platinum wire serving as the reference and counter electrodes, respectively.

Synthesis of Fe₃O₄ and Fe₃O₄/GN

The Fe₃O₄/GN nanocomposites were synthesized by a solvothermal process with Fe₃O₄ nanoparticles attached to reduced graphene oxide (GN) sheets, which combine the controllable growth of Fe₃O₄ NPs and the reduction of GO in one single step. It was slightly modified from the method as reported by Wang et al. [35]. For preparing Fe₃O₄/GN, 20 mL of FeSO₄·7H₂O solution (a concentration of ~69.5 mg/mL) and 5 mL of NaOH solution (a concentration of ~80 mg/mL) were slowly added into the prepared 30-mL GO water suspension (a concentration of ~2.67 mg/mL) in order with vigorous stirring. The mixture was stirred at room temperature for 1 h. Then, 20 mL N₂H₄·H₂O was injected into the solution space and then the mixture was loaded into a 100-mL Teflon lined stainless steel autoclave for hydrothermal reaction at 180 °C for 8 h. The Fe₃O₄/GN separated by magnetic force from solution was washed with deionized water several times and then dried at 60 °C for 8 h.

For comparison, the same synthetic procedures were carried out in the preparation of Fe₃O₄ NPs but without adding GO, as the reference.

Extraction of GE

The garlic was peeled, cleaned, and dried at room temperature, followed by chopping. A 10-g chopped edible portion was kept at 50 °C for 2 h, then mixed for 1 h

after the addition of 70 mL ethanol solution (anhydrous ethanol/deionized water = 7/1, V/V) at 30 °C for leaching repeatedly. After that, the mixture was filtrated and centrifugated to remove the insoluble substance. Next, the ethanol solution was evaporated from the supernate to obtain the garlic extract (GE). The GE stored at 4 °C until subsequent experiments.

Fabrication of the electrochemical sensor

The GCE ($\Phi = 3$ mm) was polished by 1.0, 0.3, and 0.05 μm alumina slurry, along with successive washing with anhydrous ethanol and deionized water, then dried through nitrogen blowing. One milligram of the synthesized Fe₃O₄/GN was dispersed in 1 mL deionized water with ultra-sonication for 2 h to acquire a homogenous suspension. Then, another 1 h of ultra-sonication was kept after 50 μL of GE mixing until a homogeneously dispersed solution (Fe₃O₄/GN/GE). Afterwards, 5 μL of the resulting homogenous suspension was drop-casted onto the surface of a polished GCE and dried at room temperature. A Fe₃O₄-modified GCE sensor (Fe₃O₄/GCE) and Fe₃O₄/GN-modified GCE sensor (Fe₃O₄/GN/GCE) were also fabricated using similar procedures for comparison studies.

Electrochemical measurements

For CV experiments, 5.0 mmol/L K₃[Fe(CN)₆] with 0.1 mol/L KCl standard solutions were used for evaluating electrochemical sensor performance with a scan rate of 50 mV/s from -0.2 to +0.7 V. Electrochemical impedance spectroscopy (EIS) measurements were performed in 5.0 mmol/L K₃[Fe(CN)₆]/K₄[Fe(CN)₆] solution to determinate the charge transfer resistance of the modified electrodes over a frequency range of 0.1 Hz~100 kHz with amplitude of 10 mV. SWV measurements were taken in 0.1 mol/L ABS containing different concentrations of Pb²⁺ solutions (pH 5.50) in the absence of dissolved oxygen.

Results and discussion

Physicochemical characterization

GC–MS analysis of GE

The components of the garlic extract (GE) were analyzed by GC–MS. Figure 1 is a representative total ion current chromatogram of GE and Table 1 summarizes the identified compounds. Table 1 also provides the identified structure of chemical compounds and GC retention time. The garlic extract included mainly ethyl acetate, 3-methyl-butanal, 1,1-diethoxy-ethan homopolymer, 1,3-dithiane, allyl methyl disulphide, diallyl disulphide, 3-vinyl-1,2-dithiacyclohex-4-ene, 3-vinyl-1,2-dithiacyclohex-5-ene, and diallyl trisulfide, which consisted with the previous researches [4, 5].

SEM and TEM

The morphologies of as-prepared Fe_3O_4 NPs, $\text{Fe}_3\text{O}_4/\text{GN}$ composite, and $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ composite determined by SEM and TEM were shown in Fig. 2. Figure 2a, b is the SEM and TEM images of the Fe_3O_4 NPs, respectively. It can be found in the images that the Fe_3O_4 NPs consisted of cube-shaped structures of NPs with the edge length of 50–150 nm and spherical structures of NPs with the diameter of ~ 50 nm. Figure 2c, d provides the micrographs of $\text{Fe}_3\text{O}_4/\text{GN}$ composite. It is clearly observed from SEM image that Fe_3O_4 NPs with hybrid structure were densely and uniformly anchored over the crumpled surface of GN. Meanwhile, the presence of GN did not alter the mixture cubic and spherical morphology of Fe_3O_4 NPs, but effectively decreased their particles after the composite formation. The TEM image in Fig. 2d further confirmed the present clearer structural information of the $\text{Fe}_3\text{O}_4/\text{GN}$ sample. Valuably, the synergistic effect between the Fe_3O_4 and GN in the formation retains the unique structure and prevents

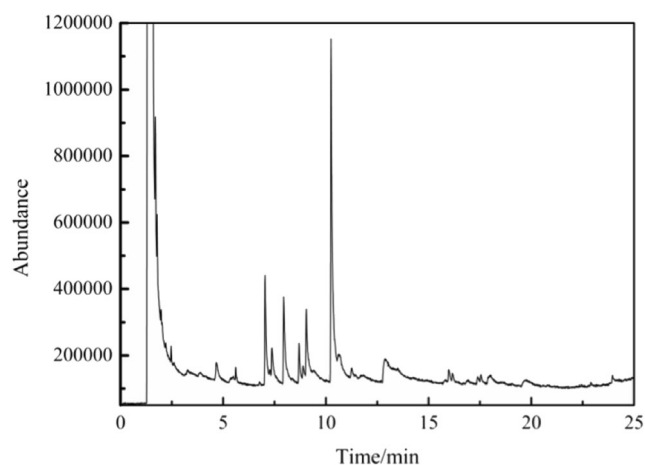


Fig. 1 Total ion current chromatogram of garlic organic components extracted by ethanol

the agglomeration of Fe_3O_4 NPs. In addition, the interaction between Fe_3O_4 NPs and GN makes possible Fe_3O_4 NPs strongly anchored on the surface of GN even after a long-time ultra-sonication and does not need additional molecular linker to bridge the Fe_3O_4 NPs and GN. It is noted that the GN provides large areas. Thereby, we can believe that the $\text{Fe}_3\text{O}_4/\text{GN}$ composite can facilitate the electron transfer and remain the active surface areas to further boost the electrochemical performance of $\text{Fe}_3\text{O}_4/\text{GN}$ electrodes benefiting from the unique structure [36]. The SEM image of $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ composite shown in Fig. 2e presents an obvious change in comparison to $\text{Fe}_3\text{O}_4/\text{GN}$ composite, which has been decorated with a GE layer on the surface of $\text{Fe}_3\text{O}_4/\text{GN}$ composite. All of these results jointly attest the successful preparation of $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ composite, which could be utilized as a remarkable biosensing platform for Pb^{2+} determination.

XRD and FTIR

Figure 3a has studied the crystal phase and structure information of the prepared Fe_3O_4 NPs, $\text{Fe}_3\text{O}_4/\text{GN}$, and $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ nanocomposite by XRD measurement. The diffraction peaks at $2\theta \approx 30.19^\circ$, 35.62° , 43.39° , 57.01° , and 62.79° can be indexed to (2 2 0), (3 1 1), (4 0 0), (5 1 1), and (4 4 0) reflection planes of Fe_3O_4 , which guaranteed the cubic structure of as-synthesized Fe_3O_4 nanostructure (JCPDS No. 01-082-1533) [15, 37]. The structure of Fe_3O_4 in $\text{Fe}_3\text{O}_4/\text{GN}$ and $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ are consistent with the cubic structure of free Fe_3O_4 nanostructures, implying that there is no structural change even after the composite preparation. In addition, there is a new diffraction peak at $2\theta \approx 25.77^\circ$ in $\text{Fe}_3\text{O}_4/\text{GN}$ and $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ composite, which could be credited to the GN, ensuring the composite fabrication of Fe_3O_4 with GN sheets. Moreover, there is no change in the crystalline morphology of $\text{Fe}_3\text{O}_4/\text{GN}$ and $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ composite, enunciating that the GE layer does not change the crystalline morphology of $\text{Fe}_3\text{O}_4/\text{GN}$.

The components responsible for the capping on the prepared $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ nanocomposite are analyzed by FTIR and the FTIR spectrums of Fe_3O_4 NPs, $\text{Fe}_3\text{O}_4/\text{GN}$, and $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ nanocomposite and GE are shown in Fig. 3b. There is an intense peak at $\sim 583 \text{ cm}^{-1}$ in the spectrum of Fe_3O_4 NPs, $\text{Fe}_3\text{O}_4/\text{GN}$, and $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ nanocomposite, as shown in Fig. 3b, corresponding to the Fe–O stretching vibration [38]. From GE, the characteristic adsorption peaks located at 524, 1023, 1121, 1270, 1632, 2937, and 3416 cm^{-1} identify the S–S, C–S, C=S, S–CH₂–, –NH–, –CH₂–, and –CO–NH vibration of GE, respectively, which manifested that the components of GE include organic sulfide, sulfoether compounds, and acid amides. Furthermore, these entire characteristic peaks exit in the FTIR spectrum of $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$, declaring the valid incorporation of GE in the $\text{Fe}_3\text{O}_4/\text{GN}$ nanocomposite.

Table 1 Identity and chemical structures of compounds in garlic extract

Peak No.	Retention time (min)	Compound	Structure
1	1.78	Ethyl Acetate	
2	1.97	3-methyl-Butanal	
3	2.46	1,1-diethoxy-ethan homopolymer	
4	4.67	1,3-Dithiane	
5	4.69	Allyl methyl disulphide	
6	7.03	Diallyl disulphide	
7	8.69	3-vinyl-1,2-dithiacyclohex-4-ene	
8	9.05	3-vinyl-1,2-dithiacyclohex-5-ene	
9	10.32	Diallyl trisulfide	

Electrochemical characterizations

The cyclic voltammeteries (CVs) of different modified electrodes in 5.0 mmol/L $K_3[Fe(CN)_6]$ containing 0.1 mol/L KCl are shown in Fig. 4a, which have studied the change of electrode behavior of each modified electrodes. All of the electrodes exhibited two well-defined redox peaks of $[Fe(CN)_6]^{3-/4-}$. However, the redox peak currents of each electrode were different. The redox peak current of $Fe_3O_4/GN/$

GCE was higher than the redox peak current of GCE, while the redox peak current of $Fe_3O_4/GN/GE/GCE$ was lower than the redox peak current of $Fe_3O_4/GN/GCE$. This phenomenon can be ascribed to the Fe_3O_4/GN enabled fast electron transport through the underlying GN layer to Fe_3O_4 nanoparticles [35, 39] and the GE layer obstructed the electron transport, respectively (EIS see [Supplementary information](#)).

We have studied the electrochemical features and the function for Pb^{2+} analysis of the $Fe_3O_4/GN/GE$ by comparing

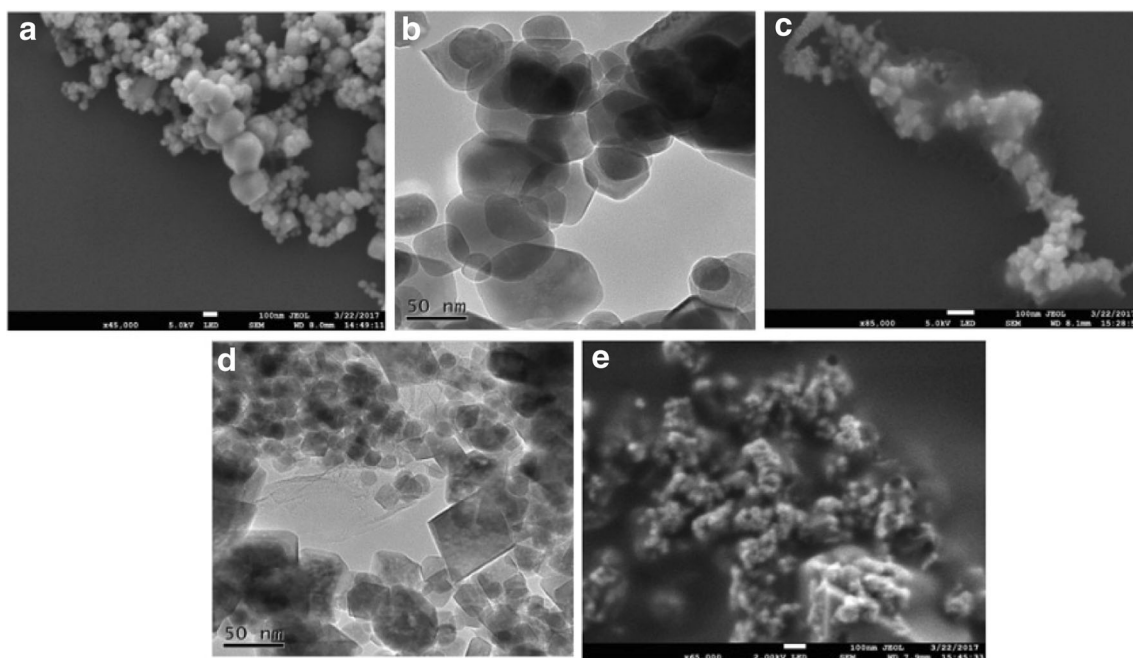


Fig. 2 SEM and TEM images of **a, b** Fe_3O_4 NPs and **c, d** Fe_3O_4/GN composite. SEM images of $Fe_3O_4/GN/GE$ composite

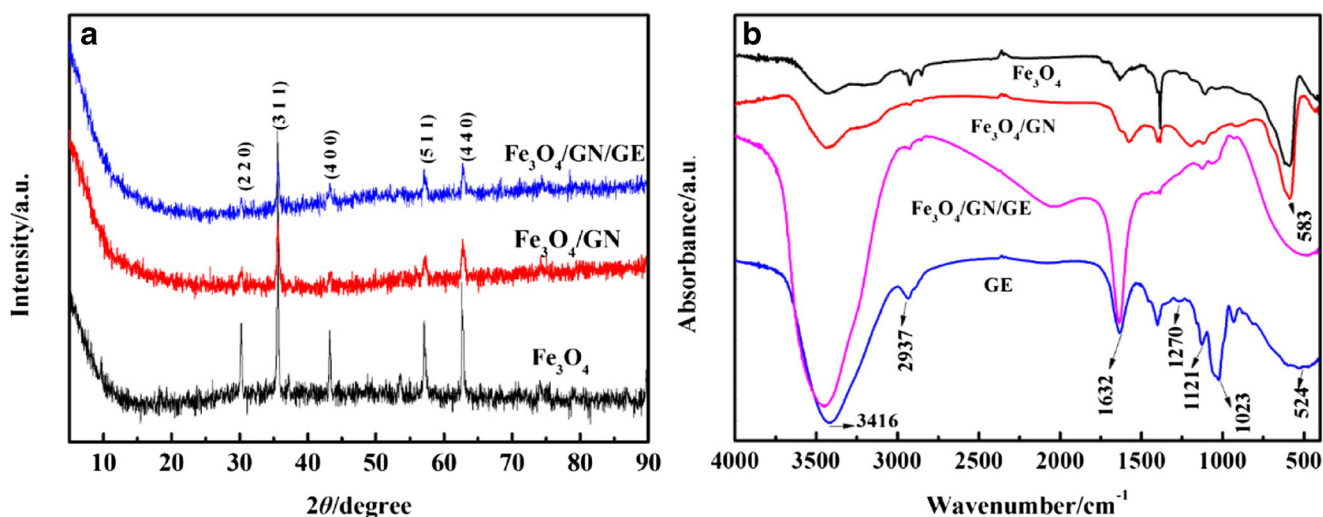


Fig. 3 a XRD patterns of Fe₃O₄ NPs, Fe₃O₄/GN, and Fe₃O₄/GN/GE. b FTIR spectra of Fe₃O₄ NPs, Fe₃O₄/GN, and Fe₃O₄/GN/GE

the electrochemical SWV responses on several modified electrodes in the presence of 1 μ M target Pb²⁺ in pH 5.50 ABS, as seen in Fig. 5. After electrochemical accumulation at -1.0 V for 180 s, a well-defined stripping peak appears at the potential values between -0.55 and -0.60 V with different stripping peak current on all electrodes. There is an inconspicuous stripping peak at -0.57 V on bare GCE, and its stripping peak current for Pb²⁺ is far less than that of other electrodes. The Fe₃O₄ and Fe₃O₄/GN have a larger surface area between the electrode and electrolyte solution. In contrast, the peak current sharply increased upon addition of GE on the Fe₃O₄/GN, which can form strong complex with Pb²⁺ and provide more active sites for Pb²⁺ accumulation. All of those results could certify that the interaction between target Pb²⁺ and Fe₃O₄/GN/GE has changed the electronic signal, thus providing a possibility for the recognition of Pb²⁺.

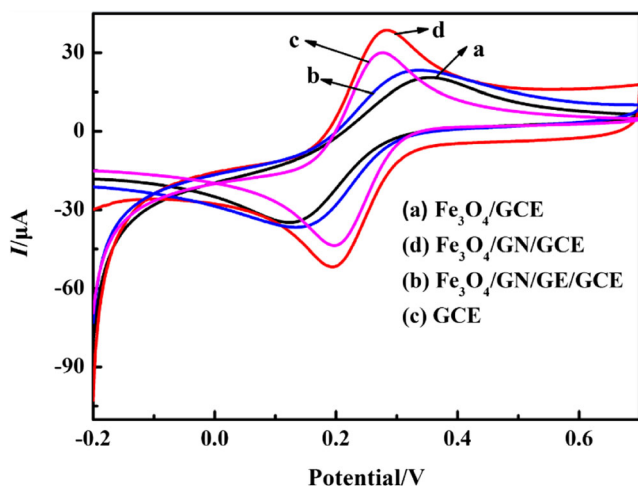


Fig. 4 Cyclic voltammograms for each immobilization step in 5.0 mmol/L K₃[Fe(CN)₆] containing 0.1 mol/L KCl solution

Optimization of assay conditions

Various experimental parameters were optimized for exploiting the maximum efficacy of Fe₃O₄/GN/GE/GCE sensor with care, loading of Fe₃O₄/GN/GE, analyte accumulation time, and pH of buffer solution included. As portrayed in Fig. 6a, the effect of the coated Fe₃O₄/GN/GE composite on electrode was shown. The peak current increased with the volume of the coated Fe₃O₄/GN/GE composite on electrode, and then reached maximum when the volume of Fe₃O₄/GN/GE composite on electrode is 5 μ L. Therefore, in the subsequent measurements, 5 μ L Fe₃O₄/GN/GE composite are coated on electrode. As seen from Fig. 6b, the electrochemical signal of the developed biosensor enhanced rapidly as the analyte accumulation time increased and achieved a plateau at 180 s, which may be owing to the increase of composites film and decrease the electron transfer rate of metal stripping.

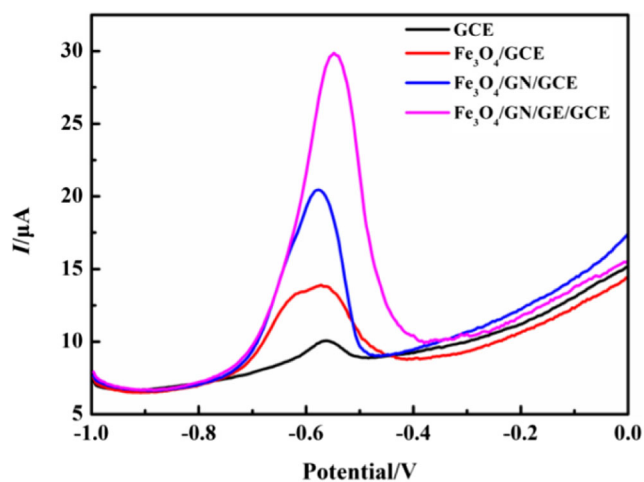


Fig. 5 SWVs recorded at Fe₃O₄/GCE, Fe₃O₄/GN/GCE, and Fe₃O₄/GN/GE/GCE in the presence of 1 μ M Pb²⁺ in pH 5.50 ABS

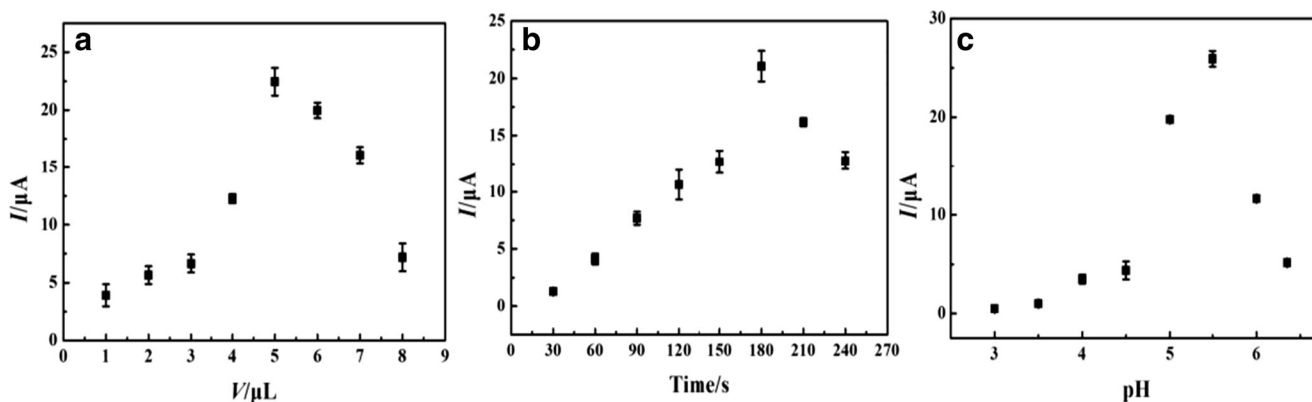


Fig. 6 Effects of **a** volume of coated composites on GCE, **b** accumulation time, and **c** pH value on the stripping peak current. Error bars are the

standard deviation for three consecutive measurements. Error bars are the standard deviation for three consecutive measurements

So, an analyte accumulation time of 180 s was employed as the optimum analyte accumulation time. The influence of the acidity of the electrolyte on the determination of Pb^{2+} in the range of 3.00~6.50 was optimized. As shown in Fig. 6c, it can be seen that the current value increased tardily with the pH of electrolyte in the range from 3.00 to 4.50, while the peak current enhanced rapidly with the pH of electrolyte in the range from 4.50 to 5.50 and obtained the maximum at 5.50. Thus, a pH of 5.50 was chosen as the optimized pH of electrolyte.

Analytical performance

Figure 7 depicts the SWV responses of the $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ biosensor at different concentrations of Pb^{2+} under the optimal conditions. As portrayed in Fig. 7a, the SWV

peaks increased with the increasing target Pb^{2+} concentrations and reached to plateau at concentration of 1000 nM Pb^{2+} . Furthermore, it is linear with the concentrations of Pb^{2+} from 0.001 to 0.5 nM and 0.5 to 1000 nM with correlation coefficients of 0.9999 and 0.9975, respectively (Fig. 7b). The linear regression equations were $y(\mu\text{A}) = 2.7262 + 23.0585x(\text{nM})$ and $y(\mu\text{A}) = 14.2501 + 0.0102x(\text{nM})$. The detection limit (LOD) and quantification limit (LOQ) of the biosensor were determined to be 0.0123 and 0.041 pM based on three times the standard deviation of the blank sample/slope and ten times the standard deviation of the blank sample/slope, respectively. The as-prepared biosensor based on novel $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ showed favorable Pb^{2+} detection performances, compared with that of other Pb^{2+} biosensors in Table 2, highlighting its sensitivity and acceptability further. Our biosensor

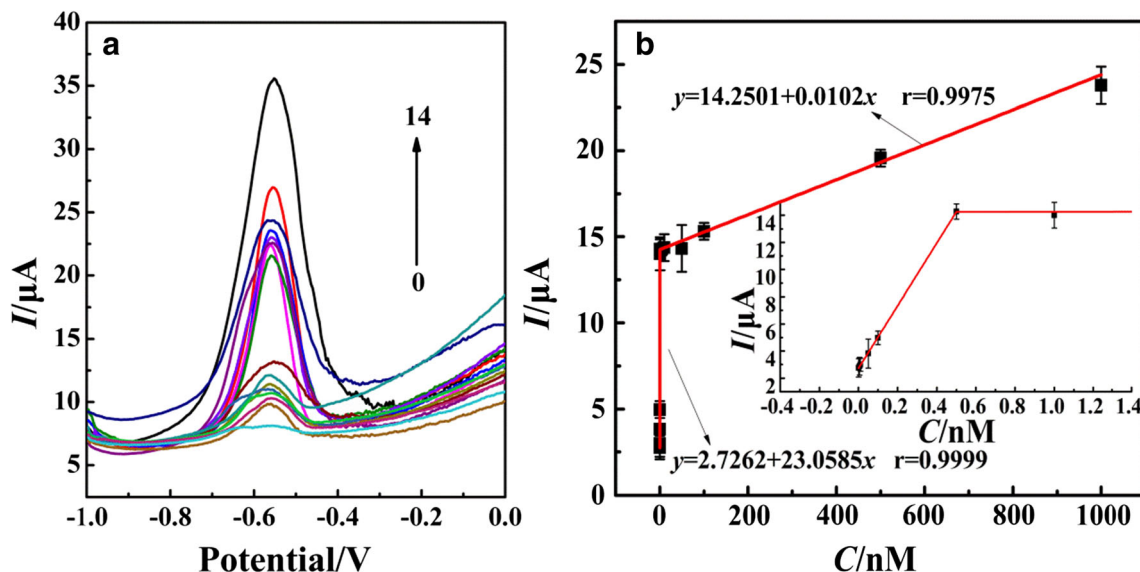


Fig. 7 **a** SWV curves of $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}/\text{GCE}$ electrodes in 0.1 mol/L ABS (pH = 5.50) containing different concentrations of Pb^{2+} solution: 0, 0.001, 0.005, 0.01, 0.05, 0.1, 0.5, 1, 5, 10, 50, 100, 500, and 1000 nM. **b**

Plot of the peak current against the concentration of Pb^{2+} . Error bars are the standard deviation for three consecutive measurements

Table 2 Comparison of analytical properties of different modified electrode toward Pb^{2+}

Lead sensor	Methods	Sensitivity ($\mu\text{A}/\text{nM}$)	LDR (nM)	LOD (pM)	References
NPG/AQDS/GR-5DNAzyme/GCE	DPV	-0.35839	1–120	20	[40]
Fc-ssDNAzyme/Au	DPV	–	0.5–5000	25	[41]
Au-Pd/hemin/Gquadruplex-based DNAzyme	DPV	0.07263	0.001–100	0.34	[42]
OMC-GNPs/DNAzyme	EIS	–	0.5–50,000	200	[43]
Au/FrGO/Au-PWE	DPV	-0.28233	0.005–2000	2	[44]
AuNPs-Apt-CS-modified	DPV	0.1498	0.6–50	312	[45]
5-Br-PADAP/MWCNT/GCEs	DPA	0.031	4.4–553.6	483	[46]
CPEa-CD and CPEb-CD	CV	0.00137 0.00305	10,000–1,000,000	–	[47]
Glassy carbon spheres	DPV	–	1000–10,000	180,000	[48]
PDMS-PA-DB18C6	DPASV	0.0039	96–3381.6	16,908	[49]
PMOs	SWASV	0.036	9.6–480	2415	[50]
$\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}/\text{GCE}$	SWV	23.0585 0.0102	0.001–0.5 0.5–1000	0.0123	This work

NPG nanoporous gold, AQDS disodium-anthraquinone-2, 6-disulfonate, Fc redox-active ferrocene, OMC-GNPs ordered mesoporous carbon-gold nanoparticle, Apt aptamer, CS complementary strand, CD cyclodextrins, PDMS-PA-DB18C6 A siloxane-crown ether polyamide copolymer, PMOs periodic mesoporous organosilica

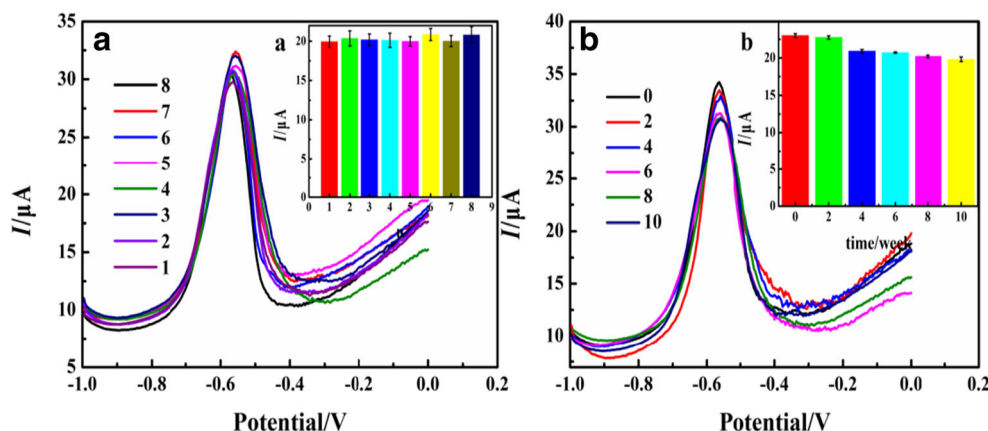
provides an attractive detection limit and a broader working range. Besides, the developed biosensor has advantages in fabrication with a facile hydrothermal technique without any surfactants or templates, economy without the use of DNAzyme and with the utilization of the natural substance GE for Pb^{2+} ion-specific, and environmentally friendly without any toxicant and with the employ of the environmental benignity $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ composite. All of these comparison manifest that the sensing performance of the designed method is superior to the mentioned biosensor for Pb^{2+} determination.

Reproducibility and stability of the proposed sensor

The reproducibility and stability are the extremely important features of the biosensor in its practical applications and

development. In our work, the reproducibility of the as-proposed Pb^{2+} biosensor was evaluated by estimating the variation of the SWV response to 1000 nM Pb^{2+} for eight different $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}$ electrodes. According to the experimental results, as shown in Fig. 8a, the SWV response variation of the assays with the same-batch biosensors is 3.61%, revealing the good reproducibility of this method. Additionally, the stability of the as-prepared biosensor was also explored on a 10-week period. The biosensor was stored in the refrigerator (4 °C) when not in use and measured intermittently (per 7 days). The SWV response remained stable (RSD 6.28%) and preserved 86.14% of the initial response after a storage period of 10 weeks for the recognition of 1000 nM Pb^{2+} , as portrayed in Fig. 8b. All of these results indicated that the reproducibility and stability of the developed biosensor were desirable.

Fig. 8 a SWV curves of eight different $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}/\text{GCE}$ in 0.1 mol/L ABS (pH = 5.50) containing 1000 nM Pb^{2+} . Inset (a) is the reproducibility of the eight different modified electrodes by SWV. b SWV curves of the different preserved time $\text{Fe}_3\text{O}_4/\text{GN}/\text{GE}/\text{GCE}$ in 0.1 mol/L ABS (pH = 5.50) containing 1000 nM Pb^{2+} . Inset (b) is the stability of the proposed sensor. Error bars are the standard deviation for three consecutive measurements



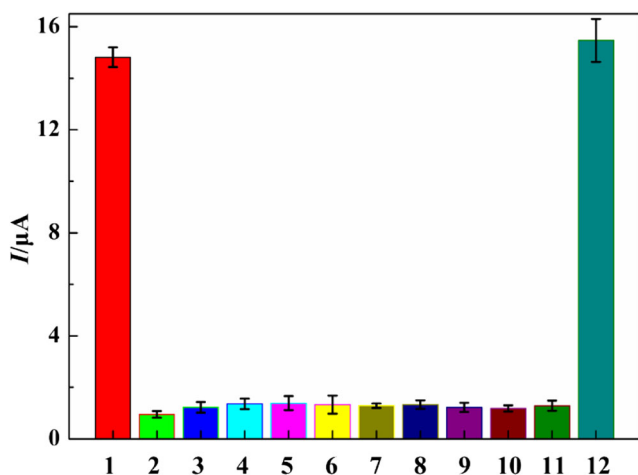


Fig. 9 The selectivity of the proposed sensor. The interfering cations are listed in order (1 to 12): (1) Pb²⁺, (2) blank, (3) Co²⁺, (4) Fe³⁺, (5) Cu²⁺, (6) Mn²⁺, (7) Zn²⁺, (8) Ni²⁺, (9) Ag⁺, (10) Hg²⁺, (11) Cd²⁺, and (12) all the interference mix with 0.5 nM Pb(II)

Specificity of the proposed sensor

The specificity of the designed strategy is supposed to be validated to ensure that the biosensor was acceptable before it was applied to the real sample analysis. We challenged the system against other ions including Co²⁺, Fe³⁺, Cu²⁺, Mn²⁺, Zn²⁺, Ni²⁺, Ag⁺, Hg²⁺, and Cd²⁺ to investigate the specificity of our designed strategy. Figure 9 depicts the histograms of peak current changes for the biosensor after reaction with other interfering metal ions. Obviously, the current response was much higher with the target Pb²⁺ of 0.5 nM than those of other interfering metal ions (10 nM). The results clearly demonstrated that the sensing strategy could monitor Pb²⁺ in the presence of other metal ions selectively.

Application for the analysis of samples

We have determined Pb²⁺ in real samples (tap water, rain water, and river water from Jialing River located in Chongqing, China) by measuring the recovery of Pb²⁺ to evaluate the utility of the fabricated biosensor. To get rid of insoluble substance, all samples were filtered by a filter membrane before detected. We have summarized the recovery values of the real samples, as shown in Table 3. All recovery values range from 91.7 to 99.8% and RSD were 2.33~5.7%, which verify that the developed sensing strategy was potentially applicable for practical detection in real samples.

Conclusions

In this study, we have fabricated a novel cheap and sensitive electrochemical sensor using Fe₃O₄/GN composite and GE as electrode modification materials for Pb²⁺ determination. It is first reported that natural substance garlic extract was utilized as Pb²⁺ specific-receptor and modified with Fe₃O₄-graphene composite for establishing advanced sensing devices. The novel sensor not only showed high sensitivity and selectivity for Pb²⁺ but also exhibited broad linearity, low detection limit, and satisfactory reproducibility and stability. Additionally, the practical application in detecting real water samples shows a satisfactory result. In general, this novel sensor possesses superior electrochemical detection performance and less cost compared with other published electrochemical sensor.

Table 3 Determination of Pb(II) recovery by the suggested sensor in different water samples (nmol/L)

Samples	Pb ²⁺ added	Pb ²⁺ found ^a	Recovery (%)	RSD (%)
Tap water	0	0.0263 (±0.0011)		3.23
	0.1	0.118 (±0.031)	91.7 ± 0.6	5.7
	1000	956 (±7.34)	95.6 ± 0.3	3.04
	0	0.0996(±0.0012)		2.33
Rain water	0.1	0.0197(±0.0036)	97.4 ± 0.5	5.01
	1000	975(±11.97)	97.5 ± 0.49	4.95
	0	0.177(±0.021)		3.15
River water	0.1	0.274(±0.03)	97.1 ± 0.33	3.28
	1000	999(±13.7)	99.8 ± 0.56	5.61
	0	0.253(±0.043)		2.64
Industrial effluents	0.1	0.356(±0.057)	103.9 ± 0.57	3.86
	1000	1003(±12.36)	100.3 ± 0.52	4.92

^a Average of three replicate measurements

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