**REVIEW ARTICLE** 

# The role of methionine on metabolism, oxidative stress, and diseases

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**Abstract** Methionine is an aliphatic, sulfur-containing, essential amino acid, and a precursor of succinyl-CoA, homocysteine, cysteine, creatine, and carnitine. Recent research has demonstrated that methionine can regulate metabolic processes, the innate immune system, and digestive functioning in mammals. It also intervenes in lipid metabolism, activation of endogenous antioxidant enzymes such as methionine sulfoxide reductase A, and the biosynthesis of glutathione to counteract oxidative stress. In addition, methionine restriction prevents altered methionine/

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transmethylation metabolism, thereby decreasing DNA damage and carcinogenic processes and possibly preventing arterial, neuropsychiatric, and neurodegenerative diseases. This review focuses on the role of methionine in metabolism, oxidative stress, and related diseases.

Keywords Methionine  $\cdot$  Mammalian  $\cdot$  Endogenous antioxidant enzyme  $\cdot$  ROS  $\cdot$  Cancer  $\cdot$  Disease

## Introduction

Mammals rely on nutrients such as amino acids, fatty acids, vitamins, and minerals from ingested food to maintain an adequate nutritional status for the regulation of metabolic, physiological, and neuronal homeostasis, as well as for the prevention of diseases (Trumbo 2008; Stover et al. 2017). Amino acids are natural compounds involved in various important biological processes, such as metabolism, growth, and immunity (He et al. 2011). They are the building blocks of proteins and precursors to functional molecules (Kim et al. 2007; Li et al. 2007; Wu et al. 2007). Some amino acids are conditionally indispensable/essential for certain developmental and physiological situations. Amino acids are crucial for normal physiology and must be supplied in sufficient amounts by the diet (Blachier et al. 2013).

In the field of animal production, it is common for synthetic amino acids to be added to feed to achieve rapid correction of any nutrient deficiencies. These "ideal compounds" also help to reduce the emission of nitrogen into the environment. In addition, some essential amino acids are currently used as nutraceutical supplements to control enteric processes, reduce pathogenic microorganisms and harmful lipids, and improve growth performance (Vieira et al. 2004).



Many factors can influence the concentrations of amino acids required in the diet, such as the chemical composition of the feed, the ambient temperature of the feeding environment, the sex and age of the animal, and the grain size of the feed. All these parameters can alter the consumption of amino acids and their subsequent metabolic processing. Sulfur amino acids are important in mammalian nutrition, because they are limiting nutrients, especially when crystalline amino acids, such as lysine, tryptophan, and threonine, are supplemented (Dauer and Przedborski 2003). Methionine sulfoxide reductase A (MsrA) is a key endogenous antioxidant enzyme that can promote longevity in animals. Many papers have reported that methionine plays a key role in antioxidant processes (Soares et al. 2017).

Oxidative stress is the result of an imbalance of prooxidant and antioxidant homeostasis, causing irreversible damage to macromolecules and cells and resulting in serious damage to the organism as a whole (Gonsette 2008). Mammalian tissue has an innate antioxidant capacity that is made up of non-enzymatic systems and endogenous enzymes, such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx) (Gonsette 2008; Lowe 2014; Del et al. 2015).

Mitochondria are cellular organelles that are implicated in several physiological processes that are essential for cell survival, such as the synthesis of adenosine triphosphate (ATP), the control of intracellular Ca<sup>2+</sup> homeostasis, the regulation of cell cycles, and neurotransmission and synaptic activity in the brain (Serviddio et al. 2011; Kasahara and Scorrano 2014; Romano et al. 2017). The brain is highly susceptible to mitochondrial impairment because of its high energy requirements and vulnerability to oxidative stress (Uttara et al. 2009). Thus, oxidative stress and mitochondrial dysfunction can trigger many neuropsychiatric disorders and neurodegenerative diseases, such as schizophrenia (Ben-Shachar and Laifenfeld 2004) and Alzheimer's disease (Dauer and Przedborski 2003; Clay et al. 2011; Serviddio et al. 2011; Yao and Keshavan 2011; Cassano et al. 2012; Manji et al. 2012; Scola et al. 2013).

Stover et al. (2017) reported that it may be necessary to supplement the diet with essential nutrients (such as amino acids and fatty acids) to stem the development of disease, because the nutritional requirements change according to the health of the individual in question. Brain-specific nutrient deficiencies can initiate disease, its progression, and related comorbidities (Ho et al. 2010; Molero-Luis et al. 2015). The clinical management of brain nutrients can provide meaningful therapeutic benefits (Papakostas et al. 2012).

The WHO has reported that cancer is the second leading cause of death globally, responsible for around 9 million deaths annually. The leading risk factors for cancer are a high body mass index, low fruit and vegetable intake, lack of physical activity, and the consumption of tobacco and alcohol (Torre et al. 2015). Recently, there has been great interest in the use of antioxidants such as vitamins (C and E), polyphenols (flavonoids), minerals (Zn), and amino acids (Met) to decrease oxidative stress and delay the onset of cancer (Nimse and Pal 2015). Because of the regulatory role of methionine in endogenous antioxidant enzymes and other metabolic processes, this amino acid may play a leading role in reducing the prevalence of cancer. The role of methionine restriction in methionine/transmethylation metabolism which directly influences the risk of cancer occurring and could be used as a palliative treatment—is currently being discussed (Xiao et al. 2014; Maddocks et al. 2016). This paper addresses the role of methionine in metabolism, oxidative stress, and diseases.

## Methionine in metabolism

An organism has many different proteins, which are made up of 20 amino acids in different sequences and combinations. Other non-protein amino acids can also be found in living organisms. Of the 20 amino acids that are the basic components of the body's proteins, nine are considered to be essential, as they cannot be synthesized endogenously via metabolic pathways and thus must be provided by dietary sources. Amino acids can be classified as aliphatic, aromatic, or heterocyclic, with aliphatic amino acids being the most common (Blachier et al. 2013).

It is known that beneficial gut microorganisms affect the digestion of proteins and the metabolism of amino acids (Li et al. 2004; Libao-Mercado et al. 2006; Yin and Bie 2010). The efficiency of proteins and amino acids is limited by the catabolism of luminal microbes (Li et al. 2008; Deng et al. 2009; Fang et al. 2010; Yin et al. 2010). Furthermore, during some infections, the intestinal mucosa may need additional energy resources such as amino acids (Blachier et al. 2013). In these instances, branched-chain amino acids (leucine, isoleucine, and valine), histidine, lysine, methionine, phenylalanine, threonine, and tryptophan are used by the intestinal cells (Yin et al. 2004). However, it is not yet fully understood how these amino acids are transported to and processed in the intestinal tract (Chen et al. 2007; Wang et al. 2009).

Methionine, a precursor of succinyl-CoA, homocysteine, cysteine, creatine, and carnitine, is an essential sulfur-containing amino acid. It is necessary for the metabolism of polyamines, creatine, and phosphatidylcholine. Methionine is the precursor for cellular methylation and the synthesis of cysteine, and can thus decrease the dietary cysteine requirement (Finkelstein et al. 1988; Mackay et al. 2012). The cysteine produced can be used in protein translation and the synthesis of the antioxidant glutathione and the osmolyte taurine (Rezzi et al. 2007; Nicholson et al. 2008). Methionine also participates in the recycling of the sulfur that is assimilated in energy-consuming reactions (Pirkov et al. 2008; Albers 2009). A deficiency of this amino acid suppresses epithelial growth in newborn animals by decreasing intestinal activity in the L-methionine cycle (Bauchart-Thevret et al. 2009).

Stoll et al. (1999) reported that the net portal balance of methionine in piglets represents 48% of the intake, which indicates that some of the methionine is used by the intestine. The parenteral methionine requirement is thus approximately 69% of the enteral requirement in newborn piglets (Shoveller et al. 2003). In addition, Shoveller et al. (2003) found that cysteine is effective in conserving methionine: in the presence of excess dietary cysteine, the methionine requirement is around 70% of the enteral requirement. Moreover, Bauchart-Thevret et al. (2009) showed that a deficiency of sulfur-containing amino acids leads to fewer goblet cells and lower glutathione content in the small intestine. Similarly, Riedijk et al. (2007) reported that approximately 20% of dietary methionine is used in the gastrointestinal tract (GIT), which is also a site for homocysteine production and whole-body transmethylation and transsulfuration. However, studies by Blachier et al. (2007) have revealed that less methionine is catabolized in pig enterocytes, but it is substantially catabolized in other cells of the portal-drained viscera and intestinal mucosa.

With regard to other animal species, Tsiagbe et al. (1987) reported that methionine directly influences growth and immunity response in broiler chickens. Moreover, Swain and Johri (2000) found a synergic relationship between methionine and choline in antibody production; dietary methionine levels from 322 to 580 mg/day improved the serum levels of IgG. Likewise, Carew et al. (2003) demonstrated that methionine deficiency decreases the relative weight of the lymphoid organs, which in turn harms growth. However, excess consumption of methionine can also have an adverse effect on growth. In the study above, supplementation with 20 or 40 g/kg excess methionine was found to decrease food intake and reduce weight gain (D'Mello and D'Mello 2003).

#### Methionine in oxidative stress

Reactive oxygen species (ROS) are produced by a variety of both physiological and non-physiological events, including the Fenton reaction, cellular respiration, mitochondrial dysfunctions, pathologies, phagocytes, neutrophils, and stress. However, cellular systems of detoxification easily eliminate low concentrations of ROS and free radicals by activating endogenous antioxidants such as SOD, CAT, GSH peroxidase, and GSH reductase. Nonetheless, excessive production of ROS causes extensive cellular damage and affects DNA and membrane phospholipids, causing cell death, tissue injury, chronic inflammatory responses, and fibrogenesis (Freitas et al. 2016). High concentrations of free radicals and ROS-saturated cellular systems of detoxification can induce cellular damage in two ways: cell senescence or cell death. Cell senescence is characterized by the induction of autophagy and the arrest of the cell cycle, whereas cell death occurs through necrosis or apoptosis. Cell necrosis is induced by a deficiency in ATP content, whereas apoptosis is induced under good energy conditions (Romano et al. 2017).

Methionine plays an essential role in the immune system through its metabolites. In this regard, Blachier et al. (2013) found that this amino acid directly influences the functioning of the immune system because of methionine catabolism leading to an increase in the production of glutathione, taurine, and other metabolites. Methionine is also readily used by the hepatocytes for the direct synthesis of glutathione, which is a low-molecular-weight antioxidant (Blachier et al. 2013). Meanwhile, methionine has been shown to chelate lead and removes it from tissues, which decreases oxidative stress (Patra et al. 2001). It has been also demonstrated that a lower concentration of methionine can prompt transsulfuration. When the intake of methionine is increased, substrate flux through the transmethylation pathway decreases, and flux through the transsulfuration pathway increases (Garg et al. 2011; Hosseini et al. 2012).

Some researchers are currently investigating the impact of methionine restriction on immune system function and oxidative stress in mammals. It has been shown that restricting this amino acid stimulates the production of glutathione and reduces oxidative stress (Hosseini et al. 2012, Liu et al. 2017). Campbell et al. (2016) observed an alteration of the oxidative activity in a branch of the pentose phosphate pathway (PPP) after increasing methionine supplementation. They also found that pre-incubating cells with methionine increased cellular tolerance to the thiol oxidizing agent diamide with relation to oxidative pentose phosphate (Campbell et al. 2016). However, studies by Maddineni et al. (2013) revealed that mice with restricted dietary methionine intake display reduced oxidative stress but no changes in the activity of their antioxidant enzymes. This suggests that further studies are needed to determine the effect of methionine restriction on antioxidant activity (Fig. 1; Table 1).

## Methionine in disease

Many studies have linked oxidative stress with the pathogenesis of several hepatic and renal diseases (Hyelin et al. 2010; Li et al. 2015). Stefanello et al. (2009) showed that chronic exposure to methionine leads to oxidative stress and histological changes in the liver in rat models. The acute administration of methionine and/or methionine sulfoxide (MetO) also significantly changes oxidative stress, as shown by the

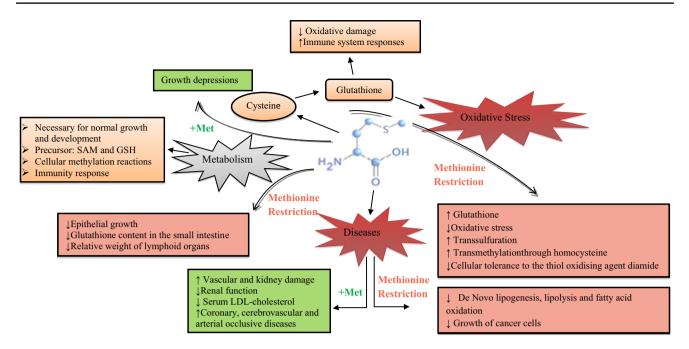


Fig. 1 Main effects of the biological activity of the Met and methionine restriction on metabolism, oxidative stress, and diseases

thiobarbituric acid reactive substances (TBARS), total thiol content, and enzymatic antioxidant defense results (Costa et al. 2013). Meanwhile, chronic dietary methionine may induce vascular (Troen et al. 2003) and kidney damage with tubular hypertrophy (Kumagai et al. 2002). However, the effects of hypermethioninemia on the kidneys are still under investigated; this tissue can regulate plasma concentrations of amino acids, urea, and ammonia. In addition, high serum methionine concentrations have been found in patients with coronary, cerebrovascular, and arterial occlusive diseases. High concentrations of methionine and MetO in the plasma may cause a progressive increase in the rate of glomerular filtration, thus impairing renal function. These results may contribute to understanding the effects of hypermethioninemia on the mechanisms involved in hepatic and renal diseases (Soares et al. 2017).

A direct relationship has been found between oxidative stress and cancer, because oxidative stress is present in various cancer cells. The imbalance of redox pathways could be related to the stimulation of oncogenes. Furthermore, an increased level of 8-OH-G, an indicator of oxidative DNA lesions, induces DNA mutation, a critical element in carcinogenesis and various tumors. Therefore, oxidation is strongly implicated in the etiology of cancer (Jackson and Bartek 2009). According to Valko et al. (2006), DNA damage and genome stability are predominantly associated with the carcinogenic initiation process.

It has been demonstrated that cancer cells are altered by methionine metabolism and transmethylation. When methionine is replaced by homocysteine, the growth of cancer cells is inhibited. Furthermore, methionine dependence may indicate an overall imbalance in transmethylation. Thus, the prevention of altered methionine/transmethylation metabolism or compensation of the altered metabolism may be the main reason for the barrier effect of methionine against cancer (Hoffman 1985; Jackson and Bartek 2009). Recent research has shown that cancer cells have a "methyl-sink", whereby methyl groups are diverted from DNA (Dash et al. 2016). In addition, methionine is seen to provoke alteration and excessive transmethylation in cancer cells. However, Dash et al. (2016) have reported that decreased levels of methionine and its metabolites may decrease cellular function in multiple organs at a systemic level. The effectiveness of methionine restriction for treating cancer is dependent upon many factors, such as age, innate immunity, type and severity of cancer, intestinal health, diet, and nutritional requirements.

The role of methionine in lipid metabolism has also been discussed, mainly as a means of reducing obesity, type 2 diabetes, and insulin resistance. In this sense, methionine restriction can reduce fat accumulation by caloric restriction, which increases de novo lipogenesis, lipolysis, and fatty acid oxidation. However, the physiological mechanisms in the adipose tissue and liver are not well known (Zhou et al. 2016). In addition, the studies of Soares et al. (2017) found that MetO increases the serum triglyceride levels in rats due to increased production of acetyl-CoA by a higher circulation of Met in bloodstream. Likewise, Hidiroglou et al. (2004) demonstrated that hypermethioninemia causes a decrease in serum LDL cholesterol and Stefanello et al. (2007) reported that excess Met reduced serum and brain

Table 1	Biological	function of	f methionine or	ı metabolism,	oxidative stress	, and diseases

Items	Biological function	References
Metabolism	It is necessary for normal growth and development in mammals	Blachier et al. (2013)
	It is precursor of succinyl-CoA, homocysteine, cysteine, creatine, and carnitine. Participates in the biosynthesis of S-adenosyl-methionine (SAM), which is involved in polyamine, creatine, and phosphatidylcholine metabolism	Finkelstein et al. (1988) and Mackay et al. (2012)
	It is precursor for cellular methylation reactions and participates in the recycling of the sulfur. It is also converted to L-methionine sulfoxide (MetO)	Finkelstein et al. (1988), Troen et al. (2003) and Mackay et al. (2012)
	The cysteine produced is used in protein translation, and synthesis of glu- tathione and the osmolyte taurine	Rezzi et al. (2007); Nicholson et al. (2008)
	A deficiency atrophies the small intestine and suppresses epithelial growth in newborn animals, as well as fewer goblet cells and lower glutathione content in the small intestine	Bauchart-Thevret et al. (2009)
	The 20% of the dietary methionine is used in the GIT	Riedijk et al. (2007)
	The parenteral methionine requirement is thus approximately 69% of the enteral requirement in newborn piglets	Shoveller et al. (2003)
	It is less catabolized in pig enterocytes; it is substantially catabolized in other cells of the portal-drained viscera and intestinal mucosa	Blachier et al. (2007)
	Directly influence of Met on growth and immunity response in broiler chickens	Tsiagbe et al. (1987)
	Synergistic effect of Met with choline on the production of antibodies (IgG)	Swain and Johri (2000)
	The deficiency of Met decreases relative weight of lymphoid organs in turn the growth performance	Carew et al. (2003)
	Excess intakes of Met causes growth depressions	D'Mello and D'Mello (2003)
Oxidative stress	It is readily catalyzed by the hepatocytes for the direct synthesis of glutathione, which is a low-molecular-weight antioxidant	Blachier et al. (2013)
	It has been shown to chelate lead and removes it from tissues	Patra et al. (2001)
	An increase in Met intake decreases the substrate flux through the transmethyla- tion pathway	Garg et al. (2011) and Hosseini et al. (2012)
	An increase of Met changes the oxidative activity in the branch of the pentose phosphate pathway and increases cellular tolerance to the thiol oxidizing agent diamide	Campbell et al. (2016)
	Methionine restriction (MR) stimulates the production of glutathione and reduces oxidative stress	Hosseini et al. (2012)
	MR reduces oxidative stress, but no changes in the activity of their antioxidant enzymes	Maddineni et al. (2013)
	MR provokes a transsulfuration which leads to Met catabolism and remethyla- tion, through homocysteine	Hosseini et al. (2012) and Romano et al. (2017)
Diseases	The acute administration of Met and/or MetO modifies oxidative stress param- eters, as shown in TBARS, total thiol content, and enzymatic antioxidant defense results	Costa et al. (2013)
	Chronic exposure to Met induces oxidative stress and promotes histological changes in the liver of young rats	Stefanello et al. (2009)
	Chronic dietary Met may induce vascular and kidney damage with tubular hypertrophy	Kumagai et al. (2002) and Troen et al. (2003)
	High concentrations of Met and MetO in plasma cause a progressive increase in the rate of glomerular filtration, thus impairing renal function	Soares et al. (2017)
	A high serum concentration of Met is associated with patients with coronary, cerebrovascular and arterial occlusive diseases	Soares et al. (2017)
	Cancer cells are altered by Met metabolism and transmethylation, and when is replaced by homocysteine, the growth of cancer cells is inhibited	Hoffman (1985) and Jackson and Bartek (2009)
	Cancer cells have a "methyl-sink", whereby methyl groups are diverted from DNA	Dash et al. (2016)
	Excessively low levels of Met and its metabolic products diminish cellular func- tion in multiple organs at the system level	Dash et al. (2016)
	MR increases de novo lipogenesis, lipolysis, and fatty acid oxidation	Zhou et al. (2016)
	Hypermethioninemia provokes a decreases of serum LDL cholesterol	Hidiroglou et al. (2004)
	Excess Met reduced serum cholesterol and brain in rat	Stefanello et al. (2007) and Soares et al. (2017)

total cholesterol in the rats; thus, more studies are needed to determine the regulatory effects.

This review has discussed the importance of methionine as a sulfur-containing essential amino acid that is indispensable for various physiological and biochemical processes in organisms. In addition, methionine supplementation or restriction can intervene in the natural antioxidant capacity of an organism by leading to the production of endogenous enzymes that reduce oxidative stress and, in turn, DNA damage, cancer, cardiovascular disease, neuropsychiatric disorders, and neurodegenerative diseases.

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#### Compliance with ethical standards

**Conflict of interest** The author declares that there is no potential conflict of interest regarding the publication of this article.

**Research involving human and animal rights** This review article does not involve any human participants and animal work.

### References

- Albers E (2009) Metabolic characteristics and importance of the universal methionine salvage pathway recycling methionine from 5'-methylthioadenosine. IUBMB Life 61(12):1132–1142. doi:10.1002/iub.278
- Bauchart-Thevret C, Stoll B, Chacko S, Burrin DG (2009) Sulfur amino acid deficiency upregulates intestinal methionine cycle activity and suppresses epithelial growth in neonatal pigs. Am J Physiol Endocrinol Metab 296(6):1239–1250. doi:10.1152/ ajpendo.91021.2008
- Ben-Shachar D, Laifenfeld D (2004) Mitochondria, synaptic plasticity, and schizophrenia. Int Rev Neurobiol 59:273–296. doi:10.1016/ S0074-7742(04)59011-6
- Blachier F, Mariotti F, Huneau JF, Tome D (2007) Effects of amino acid-derived luminal metabolites on the colonic epithelium and physiopathological consequences. Amino Acids 33(4):547–562. doi:10.1007/s00726-006-0477-9
- Blachier F, Wu G, Yin Y (2013) Nutritional and physiological functions of amino acids in pigs. Springer, Vienna
- Campbell K, Vowinckel J, Keller MA, Ralser M (2016) Methionine metabolism alters oxidative stress resistance via the pentose phosphate pathway. Antioxid Redox Sign 24(10):543–547. doi:10.1089/ars.2015.6516
- Carew LB, McMurtry JP, Alster FA (2003) Effects of methionine deficiencies on plasma levels of thyroid hormones, insulin-like growth factors-I and -II, liver and body weights, and feed intake in growing chickens. Poult Sci 82(12):1932–1938. doi:10.1093/ ps/82.12.1932
- Cassano T, Serviddio G, Gaetani S, Romano A, Dipasquale P, Cianci S, Bellanti F, Laconca L, Romano AD, Padalino I (2012) Glutamatergic alterations and mitochondrial impairment in a murine

- Chen L, Yin YL, Jobgen WS, Jobgen SC, Knabe DA, Hu WX, Wu G (2007) In vitro oxidation of essential amino acids by jejunal mucosal cells of growing pigs. Livest Sci 109(1–3):19–23. doi:10.1016/j.livsci.2007.01.027
- Clay H, Sillivan S, Konradi C (2011) Mitochondrial dysfunction and pathology in bipolar disorder and schizophrenia. Int J Dev Neurosci 29(3):311–324. doi:10.1016/j.ijdevneu.2010.08.007
- Costa MZ, Silva TMD, Flores NP, Schmitz F, Scherer EBDS, Viau CM, Saffi J, Barschak AG, Wyse ATDS, Spanevello RM (2013) Methionine and methionine sulfoxide alter parameters of oxidative stress in the liver of young rats: in vitro and in vivo studies. Mol Cell Biochem 384(1):21–28. doi:10.1007/s11010-013-1777-5
- D'Mello JPF, D'Mello J (2003) An outline of pathways in amino acid metabolism. In: D'Mello (ed) Amino acids in animal nutrition. CABI Publishing, Wallingford, pp 71–86
- Dash PK, Hergenroeder GW, Jeter CB, Choi HA, Kobori N, Moore AN (2016) Traumatic brain injury alters methionine metabolism: implications for pathophysiology. Front Syst Neurosci 10:36. doi:10.3389/fnsys.2016.00036
- Dauer W, Przedborski S (2003) Parkinson's disease: mechanisms and models. Neuron 39(6):889–909
- Del BC, Martini D, Porrini M, Klimiszacas D, Riso P (2015) Berries and oxidative stress markers: an overview of human intervention studies. Food Funct 6(9):2890–2917. doi:10.1039/c5fo00657k
- Deng D, Yao K, Chu W, Li T, Huang R, Yin Y, Liu Z, Zhang J, Wu G (2009) Impaired translation initiation activation and reduced protein synthesis in weaned piglets fed a low-protein diet. J Nutr Biochem 20(7):544–552. doi:10.1016/j.jnutbio.2008.05.014
- Fang Z, Yao K, Zhang X, Zhao S, Sun Z, Tian G, Yu B, Lin Y, Zhu B, Jia G (2010) Nutrition and health relevant regulation of intestinal sulfur amino acid metabolism. Amino Acids 39(3):633–640. doi:10.1007/s00726-010-0502-x
- Finkelstein JD, Martin JJ, Harris BJ (1988) Methionine metabolism in mammals. The methionine-sparing effect of cystine. J Biol Chem 263(24):11750–11754
- Freitas I, Boncompagni E, Tarantola E, Gruppi C, Bertone V, Ferrigno A, Milanesi G, Vaccarone R, Tira ME, Vairetti M (2016) In situ evaluation of oxidative stress in rat fatty liver induced by a methionine- and choline-deficient diet. Oxid Med Cell Longev 201(2016):9307064. doi:10.1155/2016/9307064
- Garg SK, Yan Z, Vitvitsky V, Banerjee R (2011) Differential dependence on cysteine from transsulfuration versus transport during T cell activation. Antioxid Redox Sign 15(1):39–47. doi:10.1089/ ars.2010.3496
- Gonsette RE (2008) Neurodegeneration in multiple sclerosis: the role of oxidative stress and excitotoxicity. J Neurol Sci 274(1–2):48– 53. doi:10.1016/j.jns.2008.06.029
- He Q, Yin Y, Zhao F, Kong X, Wu G, Ren P (2011) Metabonomics and its role in amino acid nutrition research. Front Biosci Landmark 16(26):2451–2460
- Hidiroglou N, Gilani GS, Long L, Zhao X, Madere R, Cockell K, Belonge B, Ratnayake WMN, Peace R (2004) The influence of dietary vitamin E, fat, and methionine on blood cholesterol profile, homocysteine levels, and oxidizability of low density lipoprotein in the gerbil. J Nutr Biochem 15(12):730–740. doi:10.1016/j. jnutbio.2004.04.009
- Ho A, Michelson D, Aaen G, Ashwal S (2010) Cerebral folate deficiency presenting as adolescent catatonic schizophrenia: a case report. J Child Neurol 25(7):898–900. doi:10.1177/0883073809343475
- Hoffman RM (1985) Altered methionine metabolism and transmethylation in cancer. Anticancer Res 5(1):1–30
- Hosseini SA, Zaghari M, Lotfollahian H, Shivazad M, Moravaj H (2012) Reevaluation of methionine requirement based on

performance and immune responses in broiler breeder hens. J Poult Sci 49(1):26–33. doi:10.2141/jpsa.011021

- Hyelin H, Hyejun S, Feitelson MA, Yu DY (2010) Oxidative stress and antioxidants in hepatic pathogenesis. World J Gastroenterol 16(48):6035–6043
- Jackson SP, Bartek J (2009) The DNA-damage response in human biology and disease. Nature 461(7267):1071–1078. doi:10.1038/ nature08467
- Kasahara A, Scorrano L (2014) Mitochondria: from cell death executioners to regulators of cell differentiation. Trends Cell Biol 24(12):761–770. doi:10.1016/j.tcb.2014.08.005
- Kim SW, Mateo RD, Yin YL, Wu G (2007) Functional amino acids and fatty acids for enhancing production performance of sows and piglets. Asian Austral J Anim 20(2):295–306. doi:10.5713/ ajas.2007.295
- Kumagai H, Katoh S, Hirosawa K, Kimura M, Hishida A, Ikegaya N (2002) Renal tubulointerstitial injury in weanling rats with hyperhomocysteinemia. Kidney Int 62(4):1219–1228. doi:10.1111/j.1523-1755.2002.kid558.x
- Li T, Yin Y, Huang R, Zhong H (2004) Dietary celluloses effect on the flow of amino acid and endogenous amino acid at the terminal ileum of pig. Acta Veterinaria et Zootechnica Sinica 35(4):473–476
- Li P, Yin YL, Li D, Kim SW, Wu G (2007) Amino acids and immune function. Br J Nutr 98(2):237–252. doi:10.1017/ S000711450769936X
- Li TJ, Dai QZ, Yin YL, Zhang J, Huang RL, Ruan Z, Deng Z, Xie M (2008) Dietary starch sources affect net portal appearance of amino acids and glucose in growing pigs. Animal 2(5):723–729. doi:10.1017/S1751731108001614
- Li S, Tan HY, Wang N, Zhang ZJ, Lao L, Wong CW, Feng Y (2015) The role of oxidative stress and antioxidants in liver diseases. Int J Mol Sci 16(11):26087–26124. doi:10.1111/j.1523-1755.2002. kid558.x
- Liu G, Yu L, Fang J, Hu CA, Yin J, Ni H, Ren W, Duraipandiyan V, Chen S, Al-Dhabi NA, Yin Y (2017) Methionine restriction on oxidative stress and immune response in dss-induced colitis mice. Oncotarget 8 (27):44511–44520. doi:10.18632/oncotarget.17812
- Libao-Mercado AJ, Yin Y, Van EJ, de Lange CF (2006) True ileal amino acid digestibility and endogenous ileal amino acid losses in growing pigs fed wheat shorts- or casein-based diets. J Anim Sci 84(6):1351–1361
- Lowe F (2014) Biomarkers of oxidative stress. In: Laher I (ed) Systems biology of free radicals and antioxidants, vol 2, 1st edn. Springer, Berlin, pp 65–87 (chap 3)
- Mackay DS, Brophy JD, Mcbreairty LE, Mcgowan RA, Bertolo RF (2012) Intrauterine growth restriction leads to changes in sulfur amino acid metabolism, but not global DNA methylation. Yucatan miniature piglets. J Nutr Biochem 23(9):1121–1127. doi:10.1016/j.jnutbio.2011.06.005
- Maddineni S, Nichenametla S, Sinha R, Wilson RP, Richie JP (2013) Methionine restriction affects oxidative stress and glutathionerelated redox pathways in the rat. Exp Biol Med 238(4):392–399. doi:10.1177/1535370213477988
- Maddocks ODK, Labuschagne CF, Adams PD, Vousden KH (2016) Serine metabolism supports the methionine cycle and DNA/RNA methylation through de novo ATP synthesis in cancer cells. Mol Cell 61(2):210–221. doi:10.1016/j.molcel.2015.12.014
- Manji H, Kato T, Di PN, Ness S, Beal MF, Krams M, Chen G (2012) Impaired mitochondrial function in psychiatric disorders. Nat Rev Neurosci 13(5):293–307. doi:10.1038/nrn3229
- Molero-Luis M, Serrano M, O'Callaghan MM, Sierra C, Pérez-Dueñas B, García-Cazorla A, Artuch R (2015) Clinical, etiological and therapeutic aspects of cerebral folate deficiency. Expert Rev Neurother 15(7):793–802. doi:10.1586/14737175.2015.1055322

- Nicholson JK, Lindon JC, Holmes E (2008) 'Metabonomics': understanding the metabolic responses of living systems to pathophysiological stimuli via multivariate statistical analysis of biological NMR spectroscopic data. Xenobiotica 29(11):1181–1189. doi:10.1080/004982599238047
- Nimse SB, Pal D (2015) Free radicals, natural antioxidants, and their reaction mechanisms. RSC Adv 5(35):27986–28006. doi:10.1039/ C4RA13315C
- Papakostas GI, Shelton RC, Zajecka JM, Etemad B, Rickels K, Clain A, Baer L, Dalton ED, Sacco GR, Schoenfeld D (2012) L-Methylfolate as adjunctive therapy for SSRI-resistant major depression: results of two randomized, double-blind, parallelsequential trials. Am J Psychiatry 169(12):1267. doi:10.1176/ appi.ajp.2012.11071114
- Patra RC, Swarup D, Dwivedi SK (2001) Antioxidant effects of  $\alpha$  tocopherol, ascorbic acid and L-methionine on lead induced oxidative stress to the liver, kidney and brain in rats. Toxicology 162(2):81–88
- Pirkov I, Norbeck J, Gustafsson L, Albers E (2008) A complete inventory of all enzymes in the eukaryotic methionine salvage pathway. FEBS J 275(16):4111-4120. doi:10.1111/j.1742-4658.2008.06552.x
- Rezzi S, Ramadan Z, Fay LB, Kochhar S (2007) Nutritional metabonomics: applications and perspectives. J Proteome Res 6(2):513–525. doi:10.1021/pr060522z
- Riedijk MA, Stoll B, Chacko S, Schierbeek H, Sunehag AL, van Goundoever JB, Burrin DG (2007) Methionine transmethylation and transsulfuration in the piglet gastrointestinal tract. Proc Natl Acad Sci USA 104(9):3408–3413. doi:10.1073/pnas.0607965104
- Romano A, Serviddio G, Calcagnini S, Villani R, Giudetti AM, Cassano T, Gaetani S (2017) Linking lipid peroxidation and neuropsychiatric disorders: focus on 4-hydroxy-2-nonenal. Free Radic Biol Med 111:281–293. doi:10.1016/j.freeradbiomed.2016.12.046
- Scola G, Kim HK, Young LT, Andreazza AC (2013) A fresh look at complex I in microarray data: clues to understanding disease-specific mitochondrial alterations in bipolar disorder. Biol Psychiatry 73(2):e4–e5. doi:10.1016/j.biopsych.2012.06.028
- Serviddio G, Romano AD, Cassano T, Bellanti F, Altomare E, Vendemiale G (2011) Principles and therapeutic relevance for targeting mitochondria in aging and neurodegenerative diseases. Curr Pharm Des 17(20):2036–2055
- Shoveller AK, Brunton JA, Pencharz PB, Bal RO (2003) The methionine requirement is lower in neonatal piglets fed parenterally than in those fed enterally. J Nutr 133(5):1390–1397
- Soares MS, Oliveira PS, Debom GN, da Silveira Mattos B, Polachini CR, Baldissarelli J, Morsch VM, Schetinger MR, Tavares RG, Stefanello FM, Spanevello RM (2017) Chronic administration of methionine and/or methionine sulfoxide alters oxidative stress parameters and ALA-D activity in liver and kidney of young rats. Amino Acids 49(1):129–138. doi:10.1007/s00726-016-2340-y
- Stefanello FM, Kreutz F, Scherer EBS, Breier AC, Vianna LP, Trindade VMT, Wyse ATS (2007) Reduction of gangliosides, phospholipids and cholesterol content in cerebral cortex of rats caused by chronic hypermethioninemia. Int J Dev Neurosci 25(7):473–477. doi:10.1016/j.ijdevneu.2007.08.004
- Stefanello FM, Matte C, Pederzolli CD, Kolling J, Mescka CP, Lamers ML, de Assis AM, Perry ML, dos Santos MF, Dutra CS, Wyse ATS (2009) Hypermethioninemia provokes oxidative damage and histological changes in liver of rats. Biochimie 91(8):961–968. doi:10.1016/j.biochi.2009.04.018
- Stoll B, Burrin DG, Henry J, Yu H, Jahoor F, Reeds PJ (1999) Substrate oxidation by the portal drained viscera of fed piglets. Am J Physiol 277(1 Pt 1):E168–E175

- Stover PJ, Durga J, Field MS (2017) Folate nutrition and blood– brain barrier dysfunction. Curr Opin Biotechnol 44:146–152. doi:10.1016/j.copbio.2017.01.006
- Swain BK, Johri TS (2000) Effect of supplemental methionine, choline and their combinations on the performance and immune response of broilers. Br Poult Sci 41(1):83–88
- Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A (2015) Global cancer statistics, 2012. Cancer J Clin 65(2):87–108. doi:10.3322/caac.21262
- Troen AM, Lutgens E, Smith DE, Rosenberg IH, Selhub J (2003) The atherogenic effect of excess methionine intake. Proc Natl Acad Sci USA 100(25):15089–15094. doi:10.1073/pnas.2436385100
- Trumbo PR (2008) Challenges with using chronic disease endpoints in setting dietary reference intakes. Nutr Rev 66(8):459–464. doi:10.1111/j.1753-4887.2008.00077.x
- Tsiagbe VK, Cook ME, Harper AE, Sunde ML (1987) Enhanced immune-responses in broiler chicks fed methionine-supplemented diets. Poult Sci 66(7):1147–1154
- Uttara B, Singh AV, Zamboni P, Mahajan RT (2009) Oxidative stress and neurodegenerative diseases: a review of upstream and downstream antioxidant therapeutic options. Curr Neuropharmacol 7(1):65–74. doi:10.2174/157015909787602823
- Valko M, Rhodes CJ, Moncol J, Izakovic M, Mazur M (2006) Free radicals, metals and antioxidants in oxidative stress-induced cancer. Chem Biol Interact 160(1):1–40. doi:10.1016/j.cbi.2005.12.009
- Vieira SL, Lemme A, Goldenberg DB, Brugalli I (2004) Responses of growing broilers to diets with increased sulfur amino acids to lysine ratios at two dietary protein levels. Poult Sci 83(8):1307–1313
- Wang W, Shi C, Zhang J, Gu W, Li T, Gen M, Chu W, Huang R, Liu Y, Hou Y (2009) Molecular cloning, distribution and

ontogenetic expression of the oligopeptide transporter PepT1 mRNA in Tibetan suckling piglets. Amino Acids 37(4):593–601. doi:10.1007/s00726-008-0178-7

- Wu G, Bazer FW, Davis TA, Jaeger LA, Johnson GA, Kim SW, Knabe DA, Meininger CJ, Spencer TE, Yin YL (2007) Important roles for arginine family amino acids in swine nutrition and production. Livest Sci 112(1–2):8–22. doi:10.1016/j.livsci.2007.07.003
- Xiao Q, Freedman ND, Ren J, Hollenbeck AR, Abnet CC, Park Y (2014) Intakes of folate, methionine, vitamin B6, and vitamin B12 with risk of esophageal and gastric cancer in a large cohort study. Br J Cancer 110(5):1328–1333. doi:10.1038/bjc.2014.17
- Yao JK, Keshavan MS (2011) Antioxidants, redox signaling, and pathophysiology in schizophrenia: an integrative view. Antioxid Redox Sign 15(7):2011–2035. doi:10.1089/ars.2010.3603
- Yin YL, Bie T (2010) Manipulation of dietary nitrogen, amino acids and phosphorous to reduce environmental impact of swine production and enhance animal health. J Food Agric Environ 8(3):447–462
- Yin YL, Deng ZY, Huang HL, Zhong HY, Hou ZP, Gong J, Liu Q (2004) Nutritional and health functions of carbohydrate for pigs. J Anim Feed Sci 13(4):523–538. doi:10.22358/jafs/67623/2004
- Yin FG, Zhang ZZ, Ju H, Yin YL (2010) Digestion rate of dietary starch affects systemic circulation of amino acids in weaned pigs. Br J Nutr 103(10):1404. doi:10.1017/S0007114509993321
- Zhou XH, He LQ, Wan D, Yang HS, Yao K, Wu GY, Wu X, Yin YL (2016) Methionine restriction on lipid metabolism and its possible mechanisms. Amino Acids 48(7):1533–1540. doi:10.1007/ s00726-016-2247-7