

Serum phenylalanine in patients post trauma and with sepsis correlate to neopterin concentrations

M. Ploder¹, G. Neurauder², A. Spittler¹, K. Schroecksnadel², E. Roth¹, and D. Fuchs²

¹ Department of Surgery, Medical University of Vienna, Vienna, Austria

² Biocentre, Division of Biological Chemistry, Innsbruck Medical University, Innsbruck, Austria

Received September 4, 2007

Accepted October 25, 2007

Published online December 28, 2007; © Springer-Verlag 2007

Summary. Increased blood concentrations of phenylalanine in patients with trauma and sepsis are common but unexplained. We examined the potential relationship between serum concentrations of phenylalanine and the immune activation marker neopterin in 84 specimens of 18 patients (14 males and 4 females) post-trauma during 12–14 days of follow up. Compared to healthy controls, average phenylalanine and neopterin concentrations were elevated in patients, and there existed a positive correlation between concentrations of the two analytes ($r_s = 0.375, p < 0.001$). No such association existed between neopterin and tyrosine concentrations ($r_s = -0.018$), but neopterin concentrations correlated to the phenylalanine to tyrosine ratio ($r_s = 0.328, p = 0.001$).

Increased phenylalanine implies insufficient conversion by phenylalanine (4)-hydroxylase (PAH). Oxidative stress due to immune activation and inflammation may destroy cofactor 5,6,7,8-tetrahydrobiopterin and impair PAH activity. This assumption is further supported by the correlation found between higher neopterin concentrations and higher phenylalanine to tyrosine ratio, which estimates efficacy of PAH.

Keywords: Phenylalanine – Neopterin – Immune activation – Oxidative stress – Trauma

Introduction

The aromatic amino acid phenylalanine is essential for humans and is substrate for phenylalanine (4)-hydroxylase (PAH). PAH produces tyrosine, another important amino acid, which is precursor for the biosynthesis of DOPA and catecholamines dopamine, epinephrine, and norepinephrine (Fig. 1) (Shintaku, 2002). Increased blood levels of phenylalanine were reported in patients with HIV infection, with cancer and with trauma, sepsis, and after burns (Watanabe et al., 1984; Roth et al., 1985; Rath et al., 1987; Ollenschläger et al., 1988), in burned patients, the increase of phenylalanine was found to correlate with clinical course and to be predictive for non-survival (Chang et al., 1983; Rath et al., 1987). All the

above-mentioned clinical conditions are known to be linked with inflammation and immune activation and with increased concentrations of immune activation marker neopterin, e.g., increased neopterin concentrations predict poor prognosis in patients with HIV infection, in patients with various types of malignancy and in patients after multiple trauma (Strohmaier et al., 1987; Fahey et al., 1990; Murr et al., 2002; Melichar et al., 2006).

Increased serum phenylalanine concentrations in patients are probably due to a reduced conversion rate of phenylalanine by PAH (Shintaku, 2002). This should be reflected by an increase of the phenylalanine to tyrosine ratio (phe/tyr), which can serve as an estimate of PAH activity (Anderson et al., 1994). Indeed, not only higher phenylalanine but also higher phe/tyr has been described in patients after burns to be associated with reduced survival (Chang et al., 1983; Rath et al., 1987).

In our study, the concentrations of phenylalanine and tyrosine were examined in patients post-trauma and with sepsis. Phe/tyr was calculated, and results were compared to neopterin concentrations.

Materials and methods

Patients

Eighteen patients (14 males, 4 females; aged mean \pm S.D.: 45 ± 19 years, range: 20–77 years, 15 post-trauma and 3 with sepsis; apache score: 18.9 ± 6.75 range: 8–34; ISS: 39 ± 13.1 , range: 18–57) were included in this study (Table 1). Samples were collected during 12–14 days of follow up every third day. In total every patient contributed 3–5 samples to the total number of 84 sera analyzed over the whole period of follow-up. All patients received standard parenteral nutrition (1700kcal/d, 100g amino acids/d).

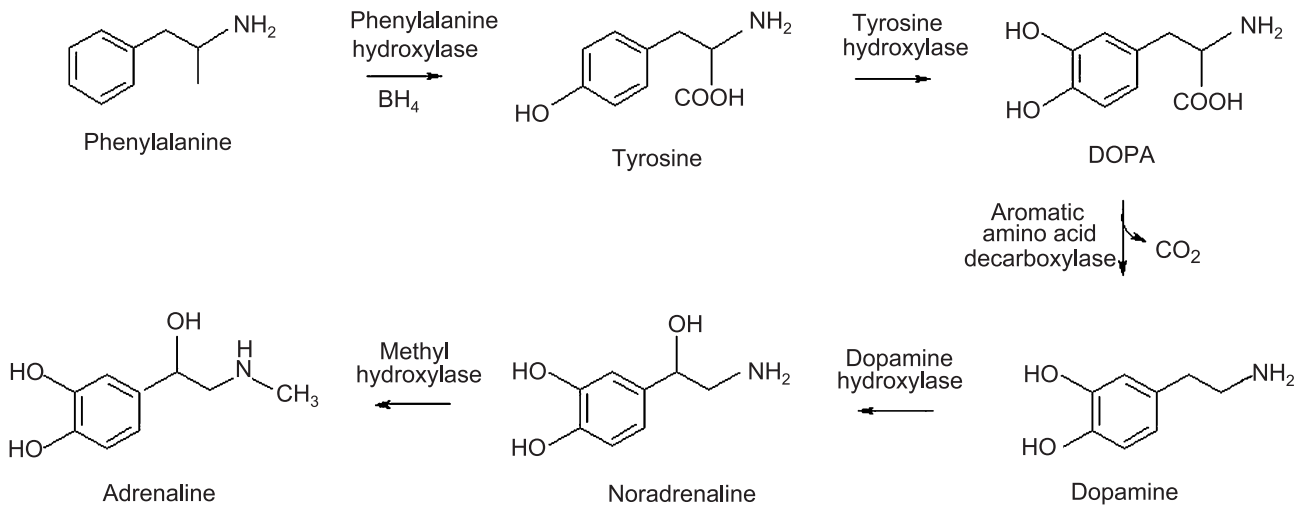


Fig. 1. Biosynthesis of tyrosine from phenylalanine by phenylalanine (5)-hydroxylase requires cofactor 5,6,7,8-tetrahydrobiopterin (BH₄). Product tyrosine is further converted by tyrosine hydroxylase to DOPA, this second enzymatic step again requires cofactor BH₄. Within both enzymatic reactions, BH₄ is oxidized to quinonoid dihydrobiopterin and recycled (= reduced) by dihydropteridine reductase employing NADPH as hydrogen donor. Upon further conversion of DOPA, biogenic amines adrenaline and noradrenaline are produced

Table 1. Demographic data of patients

Number	18
Age (mean ± S.D.)	45 ± 19
Gender (female/male)	4/14 (22.2%/77.8%)
Diagnosis (trauma/sepsis)	15/3 (83.3%/16.7%)
Days at ICU (mean ± S.D.)	26.4 ± 19.6
Inotrop (yes/no)	17/1 (94.4%/5.6%)
Comorbidity (yes/no)	3/15 (83.3%/16.7%)
Apache II score (mean ± S.D.)	18.9 ± 6.75
ISS (mean ± S.D.)	39.0 ± 13.1
Survival (yes/no)	11/7 (61.1%/38.9%)

Assays

Concentrations of both amino acids phenylalanine and tyrosine were determined by HPLC (Ollenschläger et al., 1988). The phenylalanine to tyrosine ratio was calculated. Neopterin was measured by ELISA (BRAHMS, Hennigsdorf/Berlin, Germany) according to the manufacturer's instructions with a detection limit of 2 nM.

The observed values of all study parameters had no influence whatsoever on the course of the therapy. The protocol was approved by the local ethics committee, and written consent was granted by the next of kin.

Statistical evaluation

Because some of the parameters in this study did not show normal distribution, non-parametric Mann-Whitney *U*-test was applied for comparison of grouped data and Spearman's rank correlation coefficients were calculated. *p*-values <0.05 were considered to indicate significant differences.

Results

Absolute concentrations of phenylalanine and tyrosine were increased in the patients after trauma compared to the normal range in healthy controls (Table 2). Neopterin concentrations were increased in patients, and 67/84 (79.8%) measurements revealed concentrations above 8.7 nM, which represents the 95th percentile of healthy controls (Murr et al., 2002). Throughout follow-up, phenylalanine concentrations increased compared to day 1 (*p* < 0.05 from days 4–11; see Fig. 2).

Phenylalanine concentrations correlated with the Apache score at day 1 (*r_s* = 0.494, *p* < 0.05) but not thereafter. Tyrosine concentrations did not correlate with Apache scores throughout the study, whereas the associa-

Table 2. Concentrations of phenylalanine and tyrosine, the phenylalanine to tyrosine ratio (Phe/tyr) and neopterin concentrations (mean ± S.D., and range in brackets) in patients post-trauma (18 patients, 84 measurements) compared to the normal range in healthy controls

	Patients	Healthy controls
Phenylalanine [μM]	78.9 ± 20.5 (39.0–141)*	59.0 ± 15.0 (Roth, 1985)
Tyrosine [μM]	72.7 ± 34.3 (34.0–224)*	51.7 ± 11.3 (Roth, 1985)
Phe/tyr [μmol/mmol]	1.21 ± 0.39 (0.25–2.27)	1.16 ± 0.24 (Roth, 1985)
Neopterin [nM]	30.3 ± 36.4 (4.0–168)*	5.3 ± 2.7 (Murr, 2002)

**p* < 0.01 compared to controls

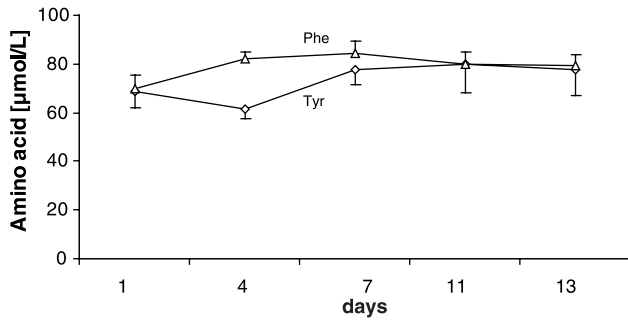


Fig. 2. Concentrations (mean \pm S.E.M.) of serum phenylalanine (Phe, triangles) and tyrosine (Tyr, diamonds) in 18 patients during follow-up for 2 weeks (phenylalanine concentrations on days 4–11 were significantly higher compared to day 1; all $p < 0.05$, changes of tyrosine were not significant)

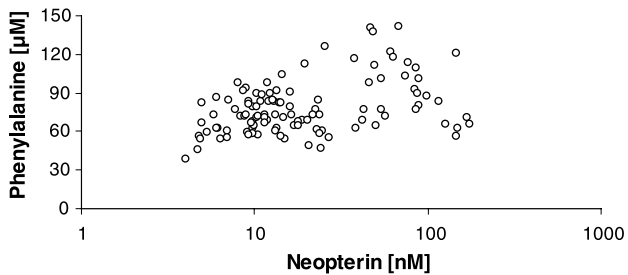


Fig. 3. Correlation between serum neopterin and phenylalanine concentrations ($r_s = 0.374$, $p < 0.001$) in 18 patients during follow-up for 2 weeks (84 specimens; note: graph shows log-scale of neopterin concentrations)

tion between Apache scores and neopterin concentrations became significant after day 4 (day 4: $r_s = 0.378$, $p = 0.068$; day 7: $r_s = 0.594$, $p < 0.01$; day 11: $r_s = 0.541$, $p < 0.05$; day 14: $r_s = 0.527$, $p < 0.05$). In the total set of measurements ($n = 84$), phenylalanine concentrations were higher in non-survivors (87.2 ± 23.1 mmol/L) compared to survivors (73.8 ± 17.0 µmol/L; $U = 2.55$, $p = 0.01$), and significantly higher phenylalanine concentrations were observed in non-survivors compared to survivors on day 1 ($p < 0.05$), later on the differences in individual days did not reach the level of statistical significance. No differences existed for tyrosine concentrations or for phe/tyr.

Significant correlation existed between phenylalanine concentrations and levels of neopterin when all results were considered ($r_s = 0.374$, $p < 0.001$; Fig. 2). Although phenylalanine and tyrosine concentrations were correlated ($r_s = 0.435$, $p < 0.001$), there was no correlation between neopterin and tyrosine ($r_s = -0.018$, n.s.). However, phe/tyr was found to correlate with neopterin concentrations as well ($r_s = 0.328$, $p = 0.001$). When comparing results of measurements on day 1 only, phenylalanine

($r_s = 0.618$) and phe/tyr ($r_s = 0.631$, both $p < 0.01$) correlated with neopterin concentrations.

Discussion

Our study confirms increased serum phenylalanine and neopterin concentrations in patients post trauma or with sepsis. A positive correlation between the serum concentrations of these two parameters was observed. There was no such association of neopterin concentrations with tyrosine levels, but a positive correlation existed between phe/tyr and neopterin concentrations.

Neopterin is produced and released in large amounts from human monocyte-derived macrophages and dendritic cells upon stimulation with Th1-type cytokine interferon- γ (Huber et al., 1984; Wirleitner et al., 2002), and thus, increased neopterin concentrations in patients indicate an activated cellular immune system. Increased neopterin concentrations are well documented during virus infections, in malignant tumor diseases and autoimmune syndromes, as well as after multiple trauma. Thereby, neopterin concentrations follow the extent, the activity and thus, the clinical course of the disease, and higher neopterin levels were found to predict disease progression in patients (Strohmaier et al., 1987; Fahey et al., 1990; Murr et al., 2002; Melichar et al., 2006). The association found between increase of phenylalanine and neopterin concentrations in our study together with the findings of increased phenylalanine levels in diseases (Watanabe et al., 1984; Roth et al., 1985; Rath et al., 1987; Ollenschläger et al., 1988), which are known to be associated with increased neopterin levels, suggests a more general relationship between immune system activation and the accumulation of phenylalanine (Widner et al., 2001). However, further studies are required to demonstrate such a general association.

In macrophages stimulated by interferon- γ , neopterin formation is paralleled by the release of toxic reactive oxygen species (ROS), including hydrogen peroxide (H_2O_2) or hypochlorous acid (HOCl) (Nathan, 1986). Neopterin itself was found to enhance oxidizing capacity of ROS (Hoffmann et al., 2003; Cirillo et al., 2007), and higher neopterin concentrations are thus often associated with, e.g., increased protein oxidation products (Witko-Sarsat et al., 1996) and the depletion of antioxidants and development of oxidative stress (Murr et al., 1999, 2007). The chronic immune system activation appears to be a major cause of the loss of antioxidants and the development of oxidative stress, e.g., subnormal concentrations of antioxidants, like vitamin C and E, have been observed in diseases associated with immune activation and inflam-

mation (Halliwell, 1991; Galli et al., 2005), and it could also lead to an impaired availability of BH₄ in patients post trauma and with sepsis.

Increase of essential amino acid phenylalanine is likely a result from impaired conversion of phenylalanine to tyrosine by PAH (Ponzzone et al., 1993; Anderson et al., 1994; Shintaku, 2002). Phe/tyr allows to estimate the activity of PAH (Anderson et al., 1994; Hoekstra et al., 2002). Accordingly, the correlation between phe/tyr and neopterin concentrations suggests that impaired PAH activity in our patients is related to immune activation. An impaired PAH activity could be due to insufficient supply with cofactor 5,6,7,8-tetrahydrobiopterin (BH₄) which is required as a hydrogen donor within the enzymatic hydroxylation reaction (Fig. 1). BH₄ is very sensitive to oxidative stress, and its reaction with oxygen is irreversible because the side-chain of the pteridine structure is eliminated (Connor et al., 1979). One might speculate that in the situation of oxidative stress, depletion of BH₄ will occur, and in turn, BH₄-dependent enzymes like PAH may suffer from insufficient cofactor supply (Widner et al., 2001). In case of PAH deficiency, concentration of phenylalanine is supposed to increase. Notably, PAH-product tyrosine itself is substrate for a subsequent enzymatic reaction: tyrosine hydroxylase forms DOPA and represents another BH₄-dependent enzyme (Fig. 1). This fact may explain why also tyrosine concentrations were increased in our patients compared to control values. As a sum effect, BH₄ deficiency results in impaired production of DOPA and neurotransmitters like dopamine, epinephrine and norepinephrine (Shintaku, 2002).

Antioxidant vitamins like vitamin C (ascorbic acid), vitamin E (α -tocopherol) and carotenoides are known to stabilize BH₄ (Martinez-Cruz et al., 2002; Kuzkaya et al., 2003; Heller et al., 2004). It would be interesting to know whether supplementation with antioxidant vitamins is able to normalize the elevated phenylalanine concentrations in patients post trauma.

In conclusion, our study demonstrates an association between increased phenylalanine and neopterin concentrations post trauma. Although a correlation found does not necessarily reflect a cause-effect relationship, our data could be explained by a decline of PAH activity in patients as a consequence of oxidative stress.

Acknowledgements

This study was financially supported by the government of the State of the Austrian Tyrol and the „Stiftung Propter Homines, Vaduz – Fürstentum Liechtenstein“. The authors thank Miss Astrid Haara for excellent technical assistance.

References

- Anderson DN, Wilkinson AM, Abou-Saleh MT, Blair JA (1994) Recovery from depression after electroconvulsive therapy is accompanied by evidence of increased tetrahydrobiopterin-dependent hydroxylation. *Acta Psychiatr Scand* 90: 10–13
- Chang XJ, Yang CC, Hsu WS, Xu WZ, Shih TS (1983) Serum and erythrocyte amino acid pattern: studies on major burn cases. *Burns Incl Therm Inj* 9: 240–248
- Cirillo P, Pacileo M, DE Rosa S, Calabrò P, Gargiulo A, Angri V, Granato-Corigliano F, Fiorentino I, Prevete N, DE Palma R, Mauro C, Leonardi A, Chiariello M (2007) Neopterin induces pro-atherothrombotic phenotype in human coronary endothelial cells. *J Thromb Haemost* 5: 248–255
- Connor MJ, Pheasant AE, Blair JA (1979) The identification of p-acetamiobenzoate as a folate degradation product in rat urine. *Biochem J* 178: 795–797
- Fahey JL, Taylor JM, Detels R, Hofmann B, Melmed R, Nishanian P, Giorgi JV (1990) The prognostic value of cellular and serologic markers in infection with human immunodeficiency virus type 1. *N Engl J Med* 322: 166–172
- Galli F, Piroddi M, Annetti C, Aisa C, Floridi E, Floridi A (2005) Oxidative stress and reactive oxygen species. *Contrib Nephrol* 149: 240–260
- Halliwell B (1991) Reactive oxygen species in living systems: source, biochemistry, and role in human disease. *Am J Med* 91: 14S–22S
- Heller R, Hecker M, Stahmann N, Thiele JJ, Werner-Felmayer G, Werner ER (2004) Alpha-tocopherol amplifies phosphorylation of endothelial nitric oxide synthase at serine 1177 and its short-chain derivative trolox stabilizes tetrahydrobiopterin. *Free Radic Biol Med* 37: 620–631
- Hoekstra R, van den Broek WW, Fekkes D, Buijnd JA, Mulder PG, Peppinkhuizen L (2002) Effect of electroconvulsive therapy on biop-terin and large neutral amino acids in severe, medication-resistant depression. *Psychiatry Res* 103: 115–123
- Hoffmann G, Wirleitner B, Fuchs D (2003) Potential role of immune system activation associated production of neopterin derivatives in humans. *Inflammation Res* 52: 313–321
- Huber C, Batchelor JR, Fuchs D, Hausen A, Lang A, Niederwieser D, Reibnegger G, Swetly P, Troppmair J, Wachter H (1984) Immune response-associated production of neopterin. Release from macrophages primarily under control of interferon-gamma. *J Exp Med* 160: 310–316
- Kuzkaya N, Weissmann N, Harrison DG, Dikalov S (2003) Interactions of peroxynitrite, tetrahydrobiopterin, ascorbic acid, and thiols: implications for uncoupling endothelial nitric-oxide synthase. *J Biol Chem* 278: 22546–22554
- Martinez-Cruz F, Pozo D, Osuna C, Espinar A, Marchante C, Guerrero JM (2002) Oxidative stress induced by phenylketonuria in the rat: prevention by melatonin, vitamin E, and vitamin C. *J Neurosci Res* 69: 550–558
- Melichar B, Solichova D, Freedman RS (2006) Neopterin as an indicator of immune activation and prognosis in patients with gynecological malignancies. *Int J Gynecol Cancer* 16: 240–252
- Murr C, Fuiht LC, Widner B, Wirleitner B, Baier-Bitterlich G, Fuchs D (1999) Increased neopterin concentrations in patients with cancer: indicator of oxidative stress? *Anticancer Res* 19: 1721–1728
- Murr C, Widner B, Wirleitner B, Fuchs D (2002) Neopterin as a marker for immune system activation. *Current Drug Metab* 3: 175–187
- Murr C, Talasz H, Artner-Dworzak E, Schroecksnadel K, Fiegl M, Fuchs D, Denz HA (2007) Inverse association between serum selenium concentrations and parameters of immune activation in patients with cardiac disorders. *Clin Chem Lab Med* (in press)
- Nathan CF (1986) Peroxide and pteridine: a hypothesis on the regulation of macrophage antimicrobial activity by interferon gamma. *Interferon* 7: 125–143

- Ollenschläger G, Jansen S, Schindler J, Rasokat H, Schrappe-Bächer M, Roth E (1988) Plasma amino acid pattern of patients with HIV infection. *Clin Chem* 34: 1787–1789
- Ponzone A, Guardamagna O, Spada M, Ponzone R, Sartore M, Kierat L, Heizmann CW, Blau N (1993) Hyperphenylalaninemia and pterin metabolism in serum and erythrocytes. *Clin Chim Acta* 216: 63–71
- Rath T, Roth E, Keidl R, Meissl G (1987) Phenylalanine: total amino acid ratio in 45 burn patients. *Scand J Plast Reconstr Surg Hand Surg* 21: 297–300
- Roth E, Zöch G, Schulz F, Karner J, Muhlbacher F, Hamilton G, Mauritz W, Sporn P, Funovics J (1985) Amino acid concentrations in plasma and skeletal muscle of patients with acute hemorrhagic necrotizing pancreatitis. *Clin Chem* 31: 1305–1309
- Shintaku H (2002) Disorders of tetrahydrobiopterin metabolism and their treatment. *Curr Drug Metab* 3: 123–131
- Strohmaier W, Redl H, Schlag G, Inthorn D (1987) Elevated D-erythro-neopterin levels in intensive care patients with septic complications. *Prog Clin Biol Res* 236B: 59–66
- Watanabe A, Higashi T, Sakata T, Nagashima H (1984) Serum amino acid levels in patients with hepatocellular carcinoma. *Cancer* 54: 1875–1882
- Widner B, Leblhuber F, Sperner-Unterweger B, Fuchs D (2001) Does disturbed homocysteine and folate metabolism in depression result from enhanced oxidative stress? *J Neurol Neurosurg Psychiatry* 70: 419
- Wirleitner B, Reider D, Ebner S, Boeck G, Widner B, Jaeger M, Schennach H, Romani N, Fuchs D (2002) Monocyte-derived dendritic cells release neopterin. *J Leukoc Biol* 72: 1148–1153
- Witko-Sarsat V, Friedlander M, Capeillere-Blandin C, Nguyen-Khoa T, Nguyen AT, Zingraff J, Jungers P, Descamps-Latscha B (1996) Advanced oxidation protein products as a novel marker of oxidative stress in uremia. *Kidney Int* 49: 1304–1313

Authors' address: Dietmar Fuchs, Biocentre, Division of Biological Chemistry, Innsbruck Medical University, Fritz Pregl Strasse 3, Innsbruck, Austria,
Fax: +43 512 9003 73330, E-mail: dietmar.fuchs@i-med.ac.at